

High-density SNP panel provides little evidence for population structure in European sea bass (*Dicentrarchus labrax*) in waters surrounding the UK

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Abstract

The European sea bass (*Dicentrarchus labrax*) is a commercially and recreationally important fish widely, distributed across the Northeast Atlantic Ocean and Mediterranean Sea. Two distinct lineages that represent the Atlantic and Mediterranean regions have been previously identified, with a hybrid zone close to the Almeria–Oran front. The presence of fine-scale population structure within the Northeast Atlantic region is less clear. Here, we investigated population structure in adult samples obtained from the northern part of the Atlantic range surrounding the UK, Ireland, Belgium, Germany, France, the Netherlands, and Norway, along with outgroups from Portugal and the Mediterranean, using a panel of 41 K single nucleotide polymorphism markers. Population structure among Northeast Atlantic Ocean samples was weak in both spawning– $(F_{ST} = 0.00022)$ and feeding– $(F_{ST} = 0.00032)$ season data sets, with small pairwise F_{ST} values between sample pairs. However, average F_{ST} was larger between spawning samples than between feeding samples, with a pattern of isolation-by-distance among the spawning samples, but not the feeding samples, suggesting some biologically meaningful population structure. The largest pairwise F_{ST} values at both International Council for the Exploration of the Sea (ICES) rectangle and division scales involved a sample from the west of Ireland. We found no evidence of a gradient in "Mediterranean" ancestry among samples collected around the UK in our data set or in a reanalysis of a published data set where such a pattern had been previously identified. In summary, there was no evidence that sea bass in different ICES divisions within the Northeast Atlantic Ocean represents genetically separate populations. Further work is required to reconcile evidence from tagging and modelling studies that suggest the potential for demographic independence with the genetic data.

Keywords: fisheries; SNPs; genetics; sea bass; population structure; connectivity

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Introduction

Managing fish stocks in an environmentally, economically, and socially sustainable manner based on best-available scientific evidence is a legal requirement in both the UK (Fisheries Act 2020) and the EU (European Commission 2013). Achieving these targets is challenging for fisheries managers as they must deal with a legacy of overfishing (Thurstan et al. 2010) and actively plan for future environmental and climatic conditions (Cheung et al. 2012). Accurately delineating fisheries stocks, defined here as groups of individuals with similar demographic and genetic characteristics that respond independently to fishing, is essential for conservation and management as failure to do so can impact stock sustainability (Ovenden et al. 2015).

Genetic methods have been widely used as a tool for delineating populations and have now been used in fisheries management for over five decades (Ovenden et al. 2015). Such methods allow the quantification of gene flow, selection, and genetic drift (Vendrami et al. 2017), and provide insight into genetic connectivity among populations. Furthermore, genetic diversity can be managed in its own right as localized adaptative traits are important to the ongoing health of fish stocks (Limborg et al. 2012). More recently, genetic connectivity has been incorporated into the broader framework of Marine Functional Connectivity, which incorporates information on the "spatial exchange of biomass, individuals, genes, and energy via the collective movements of all marine organisms during their lifetimes" (Darnaude et al. 2024). A multitude of genetic techniques exists to survey genetic diversity; however, single nucleotide polymorphisms (SNPs) are now the most commonly used marker for studies of population structure in a wide range of taxa (Helvar et al. 2011). When genotyped in thousands or millions, they provide considerable statistical power and have accelerated the uptake of genetics into marine fisheries management. SNPs have been successfully employed in many species, such as Atlantic cod Gadus morhua (Johansen et al. 2020), Atlantic herring Clupea harengus (Bekkevold et al. 2023), and Atlantic bluefin tuna Thunnus thynnus (Rodriguez-Ezpeleta 2019). However, despite these successes, even relatively low levels of gene flow can dilute signals of population structure. even when utilising thousands of SNP markers (Limborg et al. 2012). To ameliorate this, the use of SNPs under positive selection have been used to resolve fine-scale population structure in species with large effective population sizes, large migration rates, lack of geographic barriers, and pelagic dispersal phases to their lifecycles (Nielsen et al. 2012, Andersson et al. 2024).

European seabass *Dicentrarchus labrax* (Linnaeus, 1758) (Moronidae, Teleostei), hereafter referred to as "sea bass," is a euryhaline, eurythermal, marine fish whose range extends from the Mediterranean Sea to the Northeast Atlantic Ocean (Pickett and Pawson 1994). Sea bass exhibits several characteristics that have implications for population structure, or lack of it. Adults are highly migratory; tagged individuals have migrated distances greater than 1000 km in a 2-month period (Pawson et al. 2007), suggesting the potential for high dispersal and gene flow. Additionally, sea bass reproduce through mass release of pelagic eggs at spawning sites (Mayer et al. 1990), followed by pelagic larval development for 50–70 days (Jennings and Ellis 2015). At present, genetic evidence suggests two distinct genetic lineages: one in the Mediterranean

Sea and another in the Northeast Atlantic Ocean (Tine et al. 2014). In the Northeast Atlantic Ocean, ICES manages sea bass based on four stocks: "West of Scotland and Ireland"; "Atlantic-Iberian"; "Bay of Biscay", and "North Sea, English Channel, Celtic and Irish Sea." The existence and extent of population structure within these ICES management units are less clear, with considerable effort having been put into elucidating stock structure.

In brief, using allozymes, Child (1992) found evidence for two stocks around the UK: one in the Irish Sea and another in the Thames Estuary and English Channel. Subsequent research using microsatellites (Fritsch et al. 2007), or mitochondrial DNA (mtDNA) found little evidence of genetic structuring within the Atlantic Ocean (Lemaire et al. 2005). However, Coscia and Mariani (2011), discovered low, but significant, F_{ST} variance (i.e. genetic differentiation) between Irish Sea and English Channel, as well as Celtic Sea and southern North Sea specimens. Recent research, predominantly using SNPs identified some genetic structure principally driven by Mediterranean introgression into southern (Portuguese) populations (Souche et al. 2015, Ratcliffe et al. 2022). Robinet et al. (2020) identified no strong structure, but detected subtle barriers to dispersal by analysing the introgression of Mediterranean alleles into the Atlantic population(s). Evidence from genetics, tagging, and larval/juvenile pelagic drift studies revealed strong connectivity between the northern (ICES divisions 4.b and c, 7.a, d-h) and southern (ICES divisions 8.a and b) Atlantic sea bass stock units (Robinet et al. 2020, de Pontual et al. 2023, ICES 2023). Potential boundaries were identified within the northern sea bass stock, particularly between the North Sea and the Celtic/Irish Sea suggesting a single metapopulation structure representing a group of spatially separated populations interacting at some level through migration, reproduction, and/or feeding (Robinet et al. 2020, de Pontual et al. 2023, ICES 2023).

In this study, we investigated the fine-scale structure of sea bass in the northern Atlantic Ocean stocks—principally the "North Sea, English Channel, Celtic and Irish Sea" and "West of Scotland and Ireland" stocks using an SNP genotyping array (Allal et al. 2020). We utilized samples collected from around the UK, Ireland, Belgium, Germany, France, the Netherlands, and Norway, alongside reference samples from Portugal and the Mediterranean Sea to document spatial patterns of Mediterranean ancestry.

Methods

Sample collection and SNP genotyping

Scales or tissue samples from 1152 sea bass were sourced from around the UK, Ireland, Belgium, France, Germany, the Netherlands, Norway, Portugal, and the Mediterranean Sea from scale archives, market sampling, and bespoke capture, during both the spawning and the feeding seasons (Fig. 1a and b; Supplementary Table S1). All samples were adults with the exception of the Portuguese samples, which were juveniles. Scales were dried and stored at room temperature before cutting into approximate 2×2 mm pieces. Dissection equipment was sterilized between samples. One to eight dissected fragments (depending on the size of the original scales) were used for SNP genotyping. DNA extraction and SNP genotyping using the Axiom Dlab chip, a 57 K SNP array developed for sea

(a) Spawning samples and southern outgroups (inset)



(b) Feeding samples

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Figure 1. (a and b) Sampling locations for spawning and feeding seasons labelled with ICES rectangle in yellow circles. ICES divisions are also shown. Southern samples are shown as inset in spawning sample map. See Supplementary Table S1a and b for samples sizes and other information for each site. Figure 1c and d. Pairwise F_{ST} among ICES divisions for spawning and feeding samples. Cells are coloured by F_{ST} value (included as text) and the statistical significance after Bonferroni correction is indicated (*P < 0.05; **P < 0.01; ***P < 0.001).

bass aquaculture studies (Allal et al. 2020) were performed at the GENTYANE INRAE Clermont platform.

SNP calling and filtering

Axiom Analysis Suite (5.1.1, Affymetrix 2011) was used to call genotypes across four 384 well genotyping arrays using "Best Practice Genotyping Analysis Workflow." Array output files with Dish QC values below 0.82, QC call rates below 0.97, and a plate-wide average call rate below 98.5% were excluded from downstream analysis. Further quality filtering was undertaken in PLINK (1.9; Purcell et al. 2007) where SNPs with minor allele frequencies (MAFs) < 0.05 and Hardy Weinberg Expectation (HWE) were significant at $< 1 \times 10^{-10}$ were pruned from the dataset. Pruning SNPs based on linkage disequilibrium was not performed as LD was accounted for during the SNP chip design. Individuals missing >5% of SNPs were removed, along with one of any pair of individuals with relatedness > 0.35.

Data set partitioning

To investigate population structure in Northeast Atlantic Ocean, individuals were divided into two data sets that reflected putative spawning and feeding seasons based on the sampling month (Data sets 1 and 2: Northeast Atlantic Ocean spawning and feeding samples). Individuals caught between February and May were assigned as "spawning," individuals caught between June and November were assigned as "feeding." Individuals caught in December and January were assigned as "spawning" in ICES divisions 7.d and 7.e and as "feeding" in the North Sea (ICES divisions 7.c and north) (Britton et al. 2023). The Mediterranean Sea and Portuguese samples were included as outgroup samples to detect signals of Mediterranean ancestry for both data sets.

Genetic diversity

Average $H_{\rm E}$, $H_{\rm O}$, and $F_{\rm IS}$ (expected and observed heterozygosity and the deficit of heterozygotes in a sample relative to those expected under HWE) were estimated for each ICES rectangle across all SNPs along with % polymorphic loci using the gl.report.heterozygosity of *dartR* (Gruber et al. 2018, Supplementary Table S1).

Outlier identification

A search for SNPs potentially (a) affected by selection in samples surrounding the UK and (b) loci putatively affected by selection between the Northeast Atlantic Ocean and the Mediterranean Sea was conducted using three different methods: Bayescan version 2.0 (Foll and Gaggioti 2008), Out-FLANK (Whitlock and Lotterhos 2015) and pcadapt (Luu et al. 2017). Bayescan parameters were: n = 5000, thinning = 10, pb = 20, pilot = 5000, burn = 50000, and pr-odds = 100. A *q*-value threshold of 0.05 was used as a threshold for out-lier detection. OutFLANK was run using the default parameters (*LeftTrimFraction* = 0.05, *RightTrimFraction* = 0.05, *Hmin* = 0.1, *qthreshold* = 0.05) using the gl.outflank wrapper script from *dartR* (Gruber et al. 2018). Pcaadapt was run using default parameters and a k = 2 and only for the (b) data set that included the Mediterranean and Portugal N sample.

Population structure

Population structure was investigated using several complementary methods. Global FST and associated standard error was estimated using Weir and Cockerham's (1984) theta implemented in the globalFST function of FinePop2 (Kitada et al. 2021) for the spawning and feeding datasets (excluding the Portuguese and Mediterranean Sea samples) at the scale of ICES rectangle and ICES divisions (https://www.ices.dk/ data/maps/pages/default.aspx). Only sites with 10 or greater individuals were included in the analyses. Pairwise Weir and Cockerham's F_{ST} was calculated using the gl.fst.pop function in *dartR* (Gruber et al. 2018) for feeding and spawning samples independently and at two spatial scales: (a) among ICES rectangles and (b) among ICES divisions and also for feeding and spawning samples combined. P values were calculated using 1000 bootstraps for the for the independent spawning and feeding data sets and 10 000 permutations for the larger combined matrix. Sequential Bonferroni correction was applied to P values with the R function p.adjust in stats (R core team 2023) for the ICES area data sets and using Benjamini and Hochberg (1995) correction for the combined

feeding/spawning matrix due to the large number of pairwise comparisons. Differences in the relative magnitude of pairwise theta between spawning and feeding data sets at the ICES rectangle scale were visualized using network graphs (forceplots) implemented in the *qgraph* R package after conversion to similarity matrices by dividing each value in both feeding and spawning Fst matrices by the maximum F_{ST} value across both data sets in order to standardize edge thickness across the two plots, followed by subtracting each standardized F_{ST} from 1 (Epskamp et al. 2012). A permutation test was performed on the magnitude of pairwise theta between the feeding and spawning data ICES rectangle sets using the *lmp* function of *lmperm* R package (Wheeler and Torchiano 2025).

Mantel tests were performed using the *ade4* package in R (Dray and Dufour 2007) with p-values calculated using 1000 permutations on pairwise F_{ST} and pairwise sea distances (km) estimated using the Contour pairwise marine distances script (Assis et al. 2013) on feeding and spawning data sets independently. Portuguese and Mediterranean Sea samples were excluded from the analyses to ensure the results were not driven by these geographically and genetically outlying sites.

Population specific F_{ST} s, which use both differences in allele frequencies and heterozygosity to determine differentiation from a putative ancestral population (Kitada et al. 2021) were calculated using the pop_specificFST function of *FinePop2* package in R (Kitada et al. 2021). This data set included all samples (feeding and spawning) and included the Mediterranean Sea and Northern and Southern Portuguese samples. Principal components analysis was performed using the glPCA function of *adegenet* in R (Jombart 2008) on the spawning and feeding data sets independently, both with, and without, the Mediterranean Sea and Portuguese samples.

Clustering analysis was performed using sparse nonnegative matrix factorization (sNMF) in LEA (3.8.0; Frichot and François 2015). Ten repetitions were run for each k. Cross entropy was used to select the best-supported number of clusters (k = 2-10). sNMF results use the best-fitting repetition for the best-supported number of clusters. Outputs were summarized and plotted using the R package pong (Behr et al. 2016). In addition, the relationship between geographic distance from the Mediterranean Sea and the % of the genome with Mediterranean ancestry was investigated by running *LEA* with K = 2 (Atlantic vs Mediterranean ancestry) for both the feeding and spawning data sets and taking the mean "Mediterranean" ancestry component for individuals in each ICES rectangle, which were plotted against sea distance from the Northern Portugal sample. The lm function in base R was used to fit linear models.

Results

Over one thousand (n = 1152) sea bass samples were genotyped for 57 K SNPs and ~51 K SNPs passed initial Affymetrix quality controls. Subsequent filtering removed ~10 K SNPs, with 3547 removed due to strong deviations from HWE, and 6965 due to MAF. After removing samples with < 10 individuals, 901 individual sea bass (representing 38 geographic localities) and 41 022 SNPs were included in downstream analysis. Observed and expected heterozygosities were similar across all Atlantic Ocean samples with H_E and $H_O \sim 0.35$ for all samples (Supplementary Tables S2a and S2b).



Pairwise F_{ST} spawning and feeding samples from around UK

Figure 2. Pairwise Weir and Cockerham's F_{ST} (theta) for feeding (fe) and spawning (sp) sea bass samples labelled by the ICES rectangle from which they were collected (see Fig 1 a, b). All samples comprise at least 10 individuals). Cells are coloured by F_{ST} value and the statistical significance after Benjamini and Hochberg (1995) correction is indicated by ***= P < 0.001, **P = 0.01, *P = 0.05. https://www.ices.dk/data/maps/Pages/ICES-statistical-rectangles.aspx.

Outlier identification

Bayescan identified no SNPs and OutFLANK identified one SNP locus potentially influenced by selection in the Atlantic + Northern Portugal dataset. Pcadapt was not run in the Atlantic only data set due to lack of structure in PCA plots. Bayescan identified 20 loci putatively under positive selection in the Atlantic Ocean vs Mediterranean Sea data set (Supplementary Fig. S1a), OutFLANK identified 1553 loci (Supplementary Fig. S1b) and pcadapt identified 1242 loci. Overall, there were 2037 loci across the three methods with 738 in common between outflank and pcaadapt and 12 loci in common across the three methods.

Population structure

Global F_{ST} among the Northeast Atlantic Ocean samples was small, with global $F_{ST} = 0.000316$ (standard

error (*se*) = 0.000054) among the UK Atlantic Ocean spawning samples when partitioned at ICES rectangle level and $F_{ST} = 0.0002235$ (*se* = 0.0000414) when individuals were combined into ICES divisions. Global $F_{ST} = 0.00022$ (*se* = 0.000032) among the Atlantic feeding samples when partitioned into ICES rectangles and $F_{ST} = 0.0001$ (*se* = 0.000026) when individuals were combined into ICES divisions. When including the Mediterranean Sea and Portuguese samples, global, F_{ST} was larger, with $F_{ST} = 0.0062$ (*se* = 0.0000796) for the spawning sample data set.

Within the Atlantic samples surrounding the UK, pairwise F_{ST} was small among ICES rectangles, ranging between 0 and 0.0019 for spawning samples and between 0 and 0.0016 for feeding samples. *P* values for permutation tests revealed some pairwise F_{ST} values that were significantly different from zero after Benjamini and Hochberg (1995) correction (Fig. 2) with one feeding sample showing significant differences in almost



Figure 3. Forceplots of feeding and spawning pairwise similarity $(1-F_{ST})$. The line colour saturation and the width of the edges show the absolute weight and scale relative to the strongest weight in the plot. Nodes represent ICES rectangles and are coloured by ICES division.

all pairwise comparisons (32F1; North Sea, ICES division 4.c). Differentiation between ICES divisions was also very low with pairwise theta ranging from 0 to 0.0019 for spawning samples, with Northern Portugal significantly different from all other ICES divisions (Fig. 1c). The Irish Sea (ICES division 7.a) and West Coast of Ireland (ICES division 7.b) showed the greatest differentiation ($F_{ST} = 0.0019$). Pairwise F_{ST} between feeding samples was smaller, ranging from 0 to 0.0005 with a some statistically significant comparisons despite the small F_{ST} (Fig 1d). The West Coast of Ireland (ICES division 7.b) had somewhat larger (~double) pairwise F_{ST} values than other comparisons.

Pairwise F_{ST} were significantly larger between ICES rectangle spawning than between feeding samples (Permutation test, P = 0.015; Supplementary Fig. S2). Differences between pairwise F_{ST} of the spawning and feeding assemblages is illustrated using network forceplots (Fig. 3) where similarity (1- F_{ST}) is plotted. Thicker bars between nodes (ICES rectangles) indicate greater genetic similarity.

Mantel tests revealed a significant relationship between log geographic sea distance and genetic distance (F_{ST}) within the Northeast Atlantic Ocean spawning date set (P = 0.031) but not in the feeding data set (P > 0.05) when excluding the Portuguese and Mediterranean Sea samples (Supplementary Figs S2a and b). See Supplementary Table S1a, b for samples sizes and other information for each site and season.

Population (sample) specific F_{ST}

The Mediterranean Sea sample had the lowest sample specific F_{ST} . Feeding and spawning samples had similar sample specific F_{ST} , with one feeding site (32F1, North Sea, ICES division 4.c) and three spawning sites (26E6, English Channel, ICES division 7.e; 34E0, West Coast of Ireland, ICES division 7.b; and 30E4; Bristol Channel, ICES division 7.f) showing higher values (Fig. 4).

Principal components 1 and 2 accounted for 3.42% and 0.49% of the total variation in the data set, respectively in-

cluding the Mediterranean Sea and Portuguese samples and the PCA identified clear differentiation between the Northeast Atlantic Ocean samples, the Portuguese samples, and the Mediterranean Sea (Fig. 5a). The two Portuguese samples form a cluster with North and South Portugal samples differentiated on PC2 (Fig. 5a) with two individuals from the Southern Portuguese site intermediate with the Mediterranean Sea cluster. The Northeast Atlantic Ocean samples form a single cluster with clear differentiation from the Mediterranean Sea and Portuguese samples. Principal component analysis on solely the Northeast Atlantic Ocean samples did not reveal any site-specific clusters and there was considerable overlap between individuals across both the ICES division (Fig. 5b) and rectangle scales for both feeding and spawning data sets. The first two PCs each explained 0.43% of the total variation.

Model-based clustering

Model-based clustering identified k = 2 as the largest loglikelihood, with the Mediterranean Sea sample with strong Mediterranean ancestry and the Atlantic samples with lower Mediterranean ancestry (Fig. 6). Larger Ks did not reveal additional population structure within the Northeast Atlantic Ocean (Supplementary Fig. S4). The Southern Portuguese individuals showed an average Mediterranean component of 29%, but with a wide range of Mediterranean ancestry (15%– 90%). This suggested the presence of several Mediterranean migrants or admixed migrants in the sample and some individuals that had much lower Mediterranean admixture components typical of Atlantic sites. The Northern Portuguese sample showed an average of 7.8% Mediterranean ancestry, with the samples surrounding the UK showing an average of 1.2%– 2.7% Mediterranean ancestry (Fig. 6).

Mediterranean ancestry and geographic distance

There was no evidence of a relationship between the admixture proportion and distance from the Mediterranean Sea within the samples surrounding the UK and Ireland or any ev-



Figure 4. Population (sample) specific *F*_{ST} values for feeding and spawning sea bass samples by ICES rectangles, where the initial two-digit number indicates degrees latitude. See Supplementary Table S1a. b for samples sizes and other information for each site and season.

idence of genetic breaks associated with ICES division when using solely the samples surrounding the UK and Ireland (Fig. 7). We also investigated the relationship between geographic distance and Mediterranean ancestry using the loci identified by Bayescan, OutFLANK, and pcadapt as putatively affected by selection. Similar patterns were identified as using the whole data set, but confidence intervals were larger (Fig. 8). No genetic breaks among ICES divisions in the English Channel were identified.

Discussion

This study utilized dense sampling of European sea bass from across the Northeast Atlantic Ocean, in waters surrounding the UK, and more than 41 000 SNP markers suggests extremely low levels of population structure. Global and pairwise levels of F_{ST} are smaller than those previously identified in other commercially important marine fishes at similar geographic scales (see below). Measures of population differentiation, as measured by F_{ST} , were small across almost all sample pairs with statistically significant differences identified mainly in comparisons involving a few specific sites. The West of Ireland (ICES division 7.b) spawning sample was the most differentiated of the samples with a higher pairwise F_{ST} value and was statistically significantly differentiated from the Irish Sea (ICES division 7.a) with lower pairwise F_{ST} values compared to other divisions. While pairwise F_{ST} s were small, spawning samples had significantly larger pairwise FST values than feeding samples, suggesting some biologically meaningful structure within the data set. There was evidence of isolation-bydistance among spawning samples surrounding the UK and Ireland, but not within the feeding data set. Principal com-



Figure 5. Principal component analysis of spawning sea bass samples by ICES division, including (a) and excluding (b) samples from the Mediterranean Sea and Portugal.



Figure 6. Output of model-based clustering for spawning (a) and feeding (b) sea bass samples arranged by ICES rectangle [for K = 2 (greatest log likelihood clustering)]. Portuguese and MED outgroups are included in both plots.

ponent analysis and model-based clustering failed to identify any evidence of population structure within the Northeast Atlantic Ocean samples. We identified an expected steep decline in Mediterranean ancestry between the Mediterranean Sea and the Northeast Atlantic Ocean, but there was no evidence of a decrease in Mediterranean ancestry in relation to geographic distance within the Northeast Atlantic Ocean samples (when excluding Portugal).

Population structure

Overall, we identified low levels of population structure among sea bass in the Northeast Atlantic Ocean. Global F_{ST} was small in both spawning and feeding data sets. Global

 F_{ST} ranged between 0.0001 and 0.000316 depending on the scale (ICES rectangles vs divisions), and between spawning and feeding collection periods. This is comparable to the $F_{ST} = 0.0002$ estimated in the SNP-based sea bass study of Robinet et al. (2020), which included a wider sampling area, but fewer SNPs (n = 1012). However, it is smaller than the value of $F_{ST} = 0.009$ found by Coscia and Mariani (2011) using microsatellite markers. F_{ST} values in marine fish are typically considered to be small, with Ward et al. (1994) identifying an average F_{ST} of 0.062 across 57 marine species. This value was estimated using mainly allozyme based studies, but more recent analyses using SNP markers have found comparable values. For example, Wright et al. (2021) found a global



Figure 7. Proportion of Mediterranean ancestry in spawning (a) in relation to geographic sea distance from the Northern Portuguese sample. Error bars are 2x standard error. Populations are coloured by ICES division. Regression line is shown in black (y = 0.02 + 0 * distance + e). Adjusted R^2 value = -0.0484, P = 0.5622.

 F_{ST} of 0.045 in Atlantic cod (*Gadus morhua*) sampled from around the Scottish coastline, which is almost 2 orders of magnitude greater than found here in sea bass suggesting much lower levels of genetic structure in sea bass than typically found in cod.

Pairwise F_{ST} values were also small at both the ICES rectangle and ICES division scales. However statistically significant pairwise comparisons were identified within the data set at both ICES scales. The west of Ireland sample had the highest pairwise F_{ST} value of all comparisons which could indicate this area is more genetically isolated than other areas in the study, but as the result is based on a single sample, caution is needed in any inferences drawn. A single ICES rectangle in the North Sea off Clacton (32F1) during the feeding period was an outlier in both the population specific analyses and had 34 out of 34 pairwise comparisons, which were statistically significant (after Benjamini and Hochberg correction). The rectangle did not have a small sample size nor geographic isolation that could explain the higher levels of structure or higher population specific F_{ST} in this sample. However, the sampled scales were archived in 2007 and the age may have impacted the genotyping. Other outlying samples are the spawning samples from the west coast of Ireland (34E0) which are in a different ICES stock to the other samples in the data set ("West of Scotland and Ireland"). This sample has 18 of 34 statistically significant pairwise comparisons. This sample is a geographical outlier and the sample size is the smallest that we included in the analysis (n = 10). Other parameters (H_E , H_O , and F_{IS}) do not appear unusual, but it is also an outlier on the population specific F_{ST} plot (Fig. 4). Spawning samples 26E6 and 30E4 (English Channel, ICES divisions 7e and 7d) were outliers on the population specific F_{ST} analysis and similar to the outlying feeding sample had high $F_{\rm IS}$ indicating more heterozygotes than expected within the sample.

The low levels of population structure in sea bass identified here are predicted by particle tracking models, which follow the modelled trajectories of planktonic eggs and larvae and individual tagging experiments. Particle tracking models suggest that there is considerable connectivity between putative sea bass spawning areas around the UK (Beraud et al. 2018, Graham et al. 2023) with particles originating in the Western English Channel moving further east and north into the North Sea as well as into the Bristol channel. Interestingly, particles originating in the Bay of Biscay were also shown to settle in the western English Channel. Tagging studies have also demonstrated large-scale movements of adult sea bass in UK waters, showing some individuals moving between the Celtic Sea/Irish Sea and the North Sea (Wright et al. 2024) and some individuals moving between the Bay of Biscay and the Western English Channel (de Pontual et al. 2023).

The significantly higher level of population structure found in spawning samples compared to feeding samples suggests that there is increased population integrity during the spawning season (January–May) than in the feeding season, which is expected when investigating marine fish stocks based on genetic parameters (Carvalho and Hauser 1994). Sea bass are known to form spawning aggregations between February and June in waters surrounding the UK (Pickett and Pawson 1994, Pawson et al. 2007) and migrants can exhibit strong fidelity to summer feeding and winter spawning areas (de Pontual et al. 2023).

Outlier loci can be useful in identifying fine scale population structure in marine species where effective population sizes are large, reducing the rate at which drift can modify allele frequencies (Nielsen et al. 2012). Three methods were used in this study to identify outlying loci, which may have been influenced (directly or indirectly through linkage) by some type of selection. We identified a single locus that was likely affected by selection within the UK Atlantic samples using OutFLANK and none with Bayescan. However, including the Mediterranean Sea sample resulted in larger numbers of putatively selected loci:-20 markers with Bayescan and 1530 with OutFLANK and 1242 with pcadapt, reflecting the effect of numerous genetic barriers to gene flow that exist between Atlantic Ocean and Mediterranean sea bass lineages (Tine et al. 2014). We used SNPs under divergent selection between lineages (i.e. those with reduced introgression rates) to investigate within-Atlantic population structure by investigating the relationship between Mediterranean ancestry and geographic distance from the Mediterranean Sea. Using this strategy, Robinet et al. (2020) identified a break in Mediterranean ancestry between the eastern and western parts of the English Channel, suggesting a barrier to effective dispersal in this area. Robinet et al. (2020) also found a slight increase in Mediterranean ancestry northward to the English Channel, which was attributed to "allele surfing" of rare Mediterranean alleles in the northern part of the species range due to northern expansion, or more efficient selection against weakly deleterious Mediterranean alleles in the larger populations of the Bay of Biscay than in populations around the UK. We did not identify such a break in the English Channel in this study using either the full data set or solely outlier loci (Figs 7 and 8), or a trend of increasing MED ancestry as distance from the Mediterranean Sea increased among the northeastern Atlantic samples. To further investigate this discrepancy, we accessed genotype data and ancestry output files from the Robinet et al.'s (2020) paper (https://zenodo.org/records/ 3989825). We identified a number of cases where unexpectedly high-individual MED ancestry was associated with high



Figure 8. Proportion of Mediterranean ancestry in spawning samples in relation to geographic sea distance from the Portugal North sample using outlier loci. Error bars are 2x standard error. Populations are coloured by ICES division. (a) Bayescan outliers (20 loci), (b) OutFLANK outliers (1553 loci), (c) pcadapt outliers (1242 loci), and (d) all outliers (2037 loci).

levels of individual missing data. Thus, we refiltered the Robinet et al. (2020) data to exclude individuals with >5% missing data (Supplementary Fig. S5a and S5b) and replotted the Robinet et al. (2020) MED ancestry figure (Robinet et al. Fig. 4a and c), which no longer showed an increase in MED ancestry in the northeast Atlantic along with smaller error bars across the whole data set as a result of outlying individuals being removed (Supplementary Fig. S6a and S6b). The finding of increasing Mediterranean ancestry in the samples around the UK, appears to have been driven by a single sample from the North East UK (NEUK), which had a small sample size (n = 8) and large F_{IS} . Two individuals had very high levels of missing

data and large MED ancestry (Supplementary Fig. S5a). Thus, a combination of small sample size and missing data in some individuals appears to have led to an artefactual increase in MED ancestry in the northeastern Atlantic from the English Channel northwards in the Robinet et al. (2020) paper. After filtering the Robinet et al.'s (2020) data, the results for the northeastern Atlantic are similar to those found in our study.

Management implications

This study identified weak structure within the Northeast Atlantic Ocean sea bass stock, but identified some structure between the West of Ireland and Irish Sea/English Channel, indicating the potential for differentiation between the "West Coast of Ireland and Scotland" and "North Sea, English Channel, Celtic and Irish Sea" stocks. Some previous studies have indicated population structuring across similar geographic areas, (e.g. Fritsch et al. 2007), but the results were not consistent (e.g. Child 1992, Coscia and Mariani 2011). Large scale studies have not found distinct population structure within the northern stock (e.g. Souche et al. 2015, Robinet et al. 2020) making the identification of stocks based on genetic structure difficult (ICES 2023). However, high levels of fidelity to feeding and spawning areas as adults (de Pontual et al. 2023) would suggest that smaller management units are needed than are currently in place to effectively manage the stock and ensure sustainability. As a result, it is important to consider connectivity between areas in terms of pelagic drift of eggs and larvae, and adult movements to define subpopulations that could be included within a single metapopulation management unit that could be used to drive future ICES assessments (ICES 2023).

Bringing together all existing data on genetics, pelagic drift, and migration suggest that "Northern" and "Biscay" sea bass are not separate populations but exist as a metapopulation with potentially distinct subpopulation components with connectivity between current ICES advice units (ICES 2023). However, there was insufficient evidence to identify the subpopulations or zones of mixing as the evidence was contradictory (ICES 2023). In addition, it is likely that connectivity exists between all four of the existing stock units (i.e. Northern, Biscay, Atlantic-Iberian, and West of Scotland and Ireland), so further genomics, tagging, biophysical modelling, and otolith microchemistry studies, are needed to understand exactly how sea bass within the Northeast Atlantic Ocean are connected, the subpopulations that should be considered, in order to define the relevant management units. Utilizing additional analytical methods may also be informative, including novel population-specific genotype by phenotype associations to identify introgressed individuals and migrants (Leitwein et al. 2024), species essential habitat modelling to identify suitable spawning habitat and environmental barriers (Dambrine et al. 2021), and dynamic energy budget modelling to predict size and season specific distributions based on physiological parameters (Teal et al. 2012).

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Author contributions

M.I.T., P.D.L., I.C., D.M., and K.H. conceived the study, developed the approach, completed the analysis, and drafted the manuscript. M.B., T.C.C., H.A.F., K.G., F.G., T.H., P.K.K., R.K., T.L., H.L., F.M., I.M., P.P., J.C.O.P., M.P.B., M.R., W.R., V.S., A.S., M.L.T., C.W., P.V., S.V., and M.S.W. provided samples for analysis. All authors gave critical comments on each draft of the manuscript, as well as guidance on interpretation of the results.

Supplementary material

Supplementary material is available at the *ICESJMS Journal* online version of the manuscript.

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Data availability

Data files underpinning the analyses contained in this study have been archived on Zenodo (doi: 10.5281/zen-odo.15025594). R Scripts for analyses and plots are available from Github (https://github.com/martiniantaylor/ICES_sea_b ass).

References

- Affymetrix. Axiom Analysis Suite. 2011 Available from: https://www.thermofisher.com/uk/en/home/life-science/microa rray-analysis/microarray-analysis-instruments-software-services/ microarray-analysis-software/axiom-analysis-suite.html [Accessed January 2024].
- Allal F, Griot R, Phocas F et al. Axiom DlabCHIP, a 57 K SNPs genotyping array for European sea bass (*Dicentrarchus labrax*): array design and genetic map. SEANOE 2020. https://doi.org/10.17882/75680
- Andersson L, Bekkevold D, Berg F et al. How fish population genomics can promote sustainable fisheries: a road map. Annu Rev Anim Biosci 2024;12:1–20. https://doi.org/10.1146/annurev-animal-021 122-102933
- Assis J, Castilho Coelho N, Alberto F et al. High and distinct range-edge genetic diversity despite local bottlenecks. PLoS One 2013;8:e68646. https://doi.org/10.1371/journal.pone.0068646
- Behr AA, Liu KZ, Liu-Fang G et al. pong: fast analysis and visualization of latent clusters in population genetic data. *Bioinformatics* 2016;32:2817–23. https://doi.org/10.1093/bioinformatics/btw327
- Bekkevold D, Berg F, Polte P et al. Mixed-stock analysis of Atlantic herring (*Clupea harengus*): a tool for identifying management units and complex migration dynamics. *ICES J Mar Sci* 2023;80:173–84. https://doi.org/10.1093/icesjms/fsac223
- Benjamini Y, Hochberg Y. Controlling the False Discovery Rate: A Practical and Powerful Approach to Multiple Testing. J R Stat Soc Ser B Stat Method, 1995;57:289–300. https://doi.org/10.1111/j.2517-6 161.1995.tb02031.x
- Beraud C, Van Der Molen J, Armstrong M et al. The influence of oceanographic conditions and larval behaviour on settlement success—the European sea bass Dicentrarchus labrax (L.). ICES J Mar Sci 2018;75:455–70. https://doi.org/10.1093/icesjms/fsx195
- Britton JR, Harrison A, Andreou D et al. Review and Synthesis of Current Evidence on the Biology, Ecology and Fisheries for Sea Bass and Assessment of Evidence Gaps. BU Global Environmental Solutions (BUG) Report (BUG2931a) to Cefas; 2023, 95.

- Carvalho GR, Hauser L. Molecular genetics and the stock concept in fisheries. *Rev Fish Biol Fish* 1994;4:326–50. https://doi.org/10.100 7/BF00042908
- Cheung WWL, Pinnegar J, Merino G et al. Review of climate change impacts on marine fisheries in the UK and Ireland. Aquat Conserv Mar Freshw Ecosyst 2012;22:368–88. https://doi.org/10.1002/aqc. 2248
- Child AR. Biochemical polymorphism in bass, *Dicentrarchus labrax*, in the waters around the British Isles. J Mar Biol Assoc UK 1992;72:357-64. https://doi.org/10.1017/S0025315400037759
- Coscia I, Mariani S. Phylogeography and population structure of European sea bass in the north-east Atlantic. *Biol J Linn Soc* 2011;104:364–77. https://doi.org/10.1111/j.1095-8312.2011.017 12.x
- Dambrine C, Woillez M, Huret M et al. Characterising Essential Fish Habitat using spatio-temporal analysis of fishery data: a case study of the European seabass spawning areas. Fish Oceanogr 2021;30:413–28. https://doi.org/10.1111/fog.12527
- Darnaude AM, Tanner SE, Hunter E et al. Advancing research in marine functional connectivity for improved policy and management. Mar Ecol Prog Ser 2024;731:1–8. https://doi.org/10.3354/meps14550
- de Pontual H, Heerah K, Goossens J et al. Seasonal migration, site fidelity, and population structure of European seabass (*Dicentrarchus labrax*). ICES J Mar Sci 2023;80:1606–18. https://doi.org/10.1093/ icesjms/fsad087
- Dray S, Dufour A. The ade4 package: implementing the duality diagram for ecologists. J Stat Softw 2007;22:1–20. https://doi.org/10.18637 /jss.v022.i04
- Epskamp S, Cramer AO, Waldorp LJ et al. qgraph: network visualizations of relationships in psychometric data. J Stat Softw 2012;48:1– 18. https://doi.org/10.18637/jss.v048.i04
- European Commission. Regulation (EU) No 1380/2013 of the European Parliament and of the Council of 11 December 2013 on the Common Fisheries Policy, amending Council Regulations (EC) No 1954/2003 and (EC) No 1224/2009 and repealing Council Regulations (EC) No 2371/2002. https://eur-lex.europa.eu/eli/reg/2013/1 380/oj/eng Off J Eur Union 2013;L354.
- Fisheries Act. 2020. Retrieved June 13, 2022, fromhttps://www.legislat ion.gov.uk/ukpga/2020/22/contents/enacted
- Foll M, Gaggiotti OE. A genome scan method to identify selected loci appropriate for both dominant and codominant markers: a Bayesian perspective. *Genetics* 2008;180:977–93. https://doi.org/10.1534/genetics.108.092221
- Frichot E, François O. LEA: an R package for landscape and ecological association studies. *Methods Ecol Evol* 2015;6:925–9. https://doi.or g/10.1111/2041-210X.12382
- Fritsch M, Morizur Y, Lambert E et al. Assessment of sea bass (Dicentrarchus labrax L.) stock delimitation in the Bay of Biscay and the English Channel based on mark-recapture and genetic data. Fish Res 2007;83:123–32. https://doi.org/10.1016/j.fishres.2006.09 .002
- Graham JA, Watson JW, García García LM *et al.* Pelagic connectivity of European sea bass between spawning and nursery grounds. *Front Mar Sci* 2023;9:1–18. https://doi.org/10.3389/fmars.2022.1 046585
- Gruber B, Unmack PJ, Berry OF *et al.* DARTR: an R package to facilitate analysis of SNP data generated from reduced representation genome sequencing. *Mol Ecol Resour* 2018;18:691–9. https://doi.org/10.1 111/1755-0998.12745
- Helyar SJ, Hemmer-Hansen J, Bekkevold D et al. Application of SNPs for population genetics of nonmodel organisms: new opportunities and challenges. Mol Ecol Resour 2011;11:123–36. https://doi.org/ 10.1111/j.1755-0998.2010.02943.x
- ICES. Benchmark on selected sea bass stocks-stock ID workshop (WKBSEABASS-ID). ICES Sci Rep 2023;5:52. https://doi.org/10.1 7895/ices.pub.22794737
- Jennings S, Ellis JR. Bass (Moronidae). In: HJL Heessen, N Daan, JR Ellis (eds). Fish Atlas of the Celtic Sea, North Sea, and Baltic Sea.

Wageningen Academic Publishers/KNNV Publishing, Wageningen, 2015, 324-7.

- Johansen T, Besnier F, Quintela M et al. Genomic analysis reveals neutral and adaptive patterns that challenge the current management regime for East Atlantic cod Gadus morhua L. Evol Appl 2020;13:2673–88. https://doi.org/10.1111/eva.13070
- Jombart T. Adegenet: an R package for the multivariate analysis of genetic markers. *Bioinformatics* 2008;24:1403–5. https://doi.org/10.1 093/bioinformatics/btn129
- Kitada S, Nakamichi R, Kishino H. Understanding population structure in an evolutionary context: population-specific FST and pairwise FST. G3 2021;11:jkab316. https://doi.org/10.1093/g3journal/ jkab316
- Leitwein M, Durif G, Delpuech E *et al.* The fate of a polygenic phenotype within the genomic landscapes of introgression in the European seabass hybrid zone. *Mol Biol Evol* 2024;41:msae194. https: //doi.org/10.1093/molbev/msae194
- Lemaire C, Versini JJ, Bonhomme F. Maintenance of genetic differentiation across a transition zone in the sea: discordance between nuclear and cytoplasmic markers. J Evol Biol 2005;18:70–80. https: //doi.org/10.1111/j.1420-9101.2004.00828.x
- Limborg MT, Helyar SJ, De Bruyn M *et al.* Environmental selection on transcriptome-derived SNPs in a high gene flow marine fish, the Atlantic herring (*Clupea harengus*). Mol Ecol 2012;21:3686–703. https://doi.org/10.1111/j.1365-294X.2012.05639.x
- Luu K, Bazin E, Blum MGB. pcadapt: an R package to perform genome scans for selection based on principal component analysis. *Mol Ecol Resour* 2017;17:67–77. https://doi.org/10.1111/1755-0 998.12592
- Mayer I, Shackley E, Witthames PR. Aspects of the reproductive biology of the bass, *Dicentrarchus labrax* L. II. Fecundity and pattern of oocyte development. J Fish Biol 1990;36:141–8. https://doi.org/10.1111/j.1095-8649.1990.tb05590.x
- Nielsen E, Cariani A, Aoidh E et al. Gene-associated markers provide tools for tackling illegal fishing and false eco-certification. Nat Commun 2012;3:851. https://doi.org/10.1038/ncomms1845
- Ovenden JR, Berry O, Welch DJ *et al.* Ocean's eleven: a critical evaluation of the role of population, evolutionary and molecular genetics in the management of wild fisheries. *Fish* 2015;16:125–59. https://doi.org/10.1111/faf.12052
- Pawson MG, Pickett GD, Leballeur J et al. Migrations, fishery interactions, and management units of sea bass (*Dicentrarchus labrax*) in Northwest Europe. *ICES J Mar Sci* 2007;64:332–45. https://doi.or g/10.1093/ICESjms/fsl035
- Pickett GD, Pawson MG. Sea bass: biology, exploitation and conservation. Am Fish Soc Symp 1994;124:643. https://doi.org/10.1577/15 48-8659-124.4.643
- Purcell S, Neale B, Todd-Brown K et al. PLINK: a tool set for wholegenome association and population-based linkage analyses. Am J Hum Genet 2007;81:559–75. https://doi.org/10.1086/519795
- R Core Team. R: A Language and Environment for Statistical Computing. Vienna, Austria: R Foundation for Statistical Computing, 2023. https://www.R-project.org/
- Ratcliffe FC, Garcia de Leaniz C, Consuegra S. MHC class I-α population differentiation in a commercial fish, the European sea bass (*Dicentrarchus labrax*). Anim Genet 2022;53:340–51. https://doi.org/10.1111/age.13184
- Robinet T, Roussel V, Cheze K et al. Spatial gradients of introgressed ancestry reveal cryptic connectivity patterns in a high gene flow marine fish. Mol Ecol 2020;29:3857–71. https://doi.org/10.1111/mec. 15611
- Rodríguez-Ezpeleta N, Díaz-Arce N, Walter JF et al. Improved Atlantic bluefin tuna management through genetic assignment of natal origin. Front Ecol Environ 2019;17:439–44. https://doi.org/10.1002/ fee.2090
- Souche EL, Hellemans B, Babbucci M et al. Range-wide population structure of European sea bass Dicentrarchus labrax. Biol J Linn Soc 2015;116:86–105. https://doi.org/10.1111/bij.12572

- Teal LR, van Hal R, van Kooten T *et al.* Bio-energetics underpins the spatial distribution of North Sea plaice (*Pleuronectes platessa* L.) and sole (*Solea solea* L.) to climate change. *Glob Change Biol* 2012;18:3291–305. https://doi.org/10.1111/j.1365-2486.2012.027 95.x
- Thurstan RH, Brockington S, Roberts CM. The effects of 118 years of industrial fishing on UK bottom trawl fisheries. *Nat Commun* 2010;1:15. https://doi.org/10.1038/ncomms1013
- Tine M, Kuhl H, Gagnaire PA *et al*. European sea bass genome and its variation provide insights into adaptation to euryhalinity and speciation. *Nat Commun* 2014;5:5770. https://doi.org/10.1038/ncomms 6770
- Vendrami DLJ, Telesca L, Weigand H et al. RAD sequencing resolves fine-scale population structure in a benthic invertebrate: implications for understanding phenotypic plasticity. *Roy Soc Open Sci* 2017;4:160548. https://doi.org/10.1098/rsos.160 548
- Ward RD, Woodwark M, Skibinski DOF. A comparison of genetic diversity levels in marine, freshwater, and anadromous fishes. J Fish

Biol 1994;44:213–232. https://doi.org/10.1111/j.1095-8649.1994 .tb01200.x

- Weir BS, Cockerham CC. Estimating F-statistics for the analysis of population structure. *Evolution* 1984;38:1358–70. https://doi.org/10.2 307/2408641
- Wheeler RE, Torchiano M. *lmPerm: Permutation Tests for Linear Models. R package version 2.1.5*, . 2025 Available from: https://github.com/mtorchiano/lmperm [Accessed: March 2025]
- Whitlock MC, Lotterhos KE. Reliable detection of loci responsible for local adaptation: inference of a null model through trimming the distribution of FST. Am Nat 2015;186:S24–36. https://doi.org/10.1 086/682949
- Wright PJ, Doyle A, Taggart J *et al*. Linking scales of life-history variation with population structure in Atlantic cod. *Front Mar Sci* 2021;18. https://doi.org/10.3389/fmars.2021.630515
- Wright S, Griffiths CA, Bendall V et al. Seasonal migrations of the European bass (*Dicentrarchus labrax* L.) in UK and surrounding waters. *Mov Ecol* 2024;12:45. https://doi.org/10.1186/s40462-024-00482
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