

1 **Foodborne bacterial pathogens: genome-based approaches for enduring and** 2 **emerging threats in a complex and changing world**

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13 14 15 **Abstract**

16 Foodborne illnesses pose a substantial health and economic burden, presenting challenges
17 in prevention due to the diverse microbial hazards that can enter and spread within food
18 systems. Various factors, including natural, political and commercial drivers, influence food
19 production and distribution. The risks of foodborne illness will continue to evolve in step
20 with these drivers and changes to food systems. For example, climate impacts on water
21 availability during agriculture, changes in food sustainability targets, and evolving customer
22 preferences can all have an impact on the ecology of foodborne pathogens and the agrifood
23 niches that can carry microorganisms. Whole genome and metagenome sequencing,
24 combined with microbial surveillance schemes and insights from the food system, can
25 provide authorities and businesses with transformative information to address risks and
26 implement new food safety interventions across the food chain. In this Review, we describe
27 how genome-based approaches have advanced our understanding of the evolution and
28 spread of enduring bacterial foodborne hazards, as well as their role in identifying emerging
29 foodborne hazards. Furthermore, foodborne hazards exist in complex microbial
30 communities across the entire food chain, and consideration of these co-existing organisms
31 is essential to understanding the entire ecology supporting pathogen persistence and
32 transmission in an evolving food system.

33 34 35 36 **Introduction**

37 The world is changing at an unprecedented pace, with the current decade being defined by
38 our realisations of the near- and long-term impacts of a changing climate, set amidst the
39 societal and economic influences of the COVID-19 pandemic. The impacts to our food may
40 be just as widespread, with an increasingly globalised food supply reaching 9 billion
41 consumers that ultimately relies on the success of localised production activities.

42 Food systems comprise widespread but connected activities to produce, distribute, store,
43 and sell foods, with food businesses and regulators seeking economic outcomes through

44 strong enterprise, health outcomes through safe and nutritious diets, and environmental
45 outcomes through improved land use^{1,2}. These food system activities and outcomes are
46 impacted by similarly diverse drivers — for example, the availability of natural resources,
47 changes in consumer preference for different food types, and the flux of supply chains and
48 food pricing³. Bacteria are present throughout these systems.

49 There is a connection between how bacteria transmit through our natural world and the
50 complex agrifood structures used to produce foods. In this Review, we describe how an
51 understanding of microbial ecology can be used to inform and prevent food safety risks.
52 Foodborne pathogens such as *Salmonella enterica* and *Campylobacter* spp. are hazards
53 synonymous with food safety, with food safety itself being an integral part of food security,
54 because ‘if it’s not safe, it’s not food’⁴. As such, the Food and Agriculture Organization (FAO)
55 notes that food safety activities result in a multitude of benefits. These benefits extend
56 beyond merely preventing illness, but also encompass the reduction of food waste and the
57 adaptation of systems to support resilient and healthy ways of producing foods⁵. Within the
58 UK, both food safety and food security feature as key strategies to promote resilience to the
59 impacts of climate change⁶.

60 Considering the connection between ecological and socioeconomic drivers within food
61 systems, we take the view that there are multiple consequences for food safety that may
62 stem from new patterns of microbial transmission within agrifood settings (Figure 1). For
63 example, climate change can alter the prevalence, abundance and distribution of bacterial
64 species in aquatic and marine systems, and may lead to widespread impacts to human,
65 plant and animal health^{7,8}. The detection of foodborne pathogens has significantly evolved;
66 the development and increasing affordability of whole genome and metagenome
67 sequencing now make it feasible to apply these techniques within programmes conducting
68 microbial surveillance, outbreak detection, and regulatory testing, allowing the observation
69 of new patterns. Genomics and metagenomics are tools that have been identified to cut
70 across different sectors — including in health, agriculture and the environment, in both high
71 and low resource settings — offering enhanced insights into how to manage foodborne
72 diseases⁹.

73 In this Review, we put these contemporary tools into context in relation to both enduring
74 and emerging foodborne bacterial hazards. We base our definition of emerging hazards on
75 those “that have newly appeared in a population or have existed but are rapidly increasing
76 in incidence or geographic range”¹⁰; some foodborne or potential foodborne pathogens
77 fitting this definition include Shiga toxin-producing *Escherichia coli* O104:H4 and *Salmonella*
78 *enterica* subsp. *enterica* serovar Typhimurium sequence type 313 (ST313), which emerged
79 due to changes in food production practices and amplification through the food chain, as
80 well as pathogen evolution¹¹⁻¹⁴. We also extend this definition to include hazards which
81 likely have long existed but have only recently been identified due to advances in
82 sequencing and other technologies, examples being Candidatus *Campylobacter infans*¹⁵
83 (associated with breastfeeding) and *Arcobacter* spp.¹⁶ (Figure 2). Enduring hazards are
84 those that have persisted for decades and, from a scientific standpoint, have often been
85 observed and classified at taxonomic levels such as genus and species, as well as subspecies
86 classifications such as serotype and sequence type. However, the level of observable
87 resolution within subspecies classifications is being re-written by whole genome sequencing

88 (WGS), and as will be explored in this Review, as are the definitions of enduring hazards that
89 may now be viewed as emerging hazards based upon the revelation of specific genetic traits
90 that explain the expansion of subspecies in agrifood niches. Although this definition is
91 dichotomised, the true nature of foodborne hazards is a continuum, and when viewed
92 under the lens of genomics, what is classified as emerging can shift to enduring, or *vice*
93 *versa*. The US Centers for Disease Control and Prevention uses WGS to define bacterial
94 strains causing disease as ‘Reoccurring’ (strains that repeatedly but periodically cause acute
95 outbreaks), ‘Emerging’ (previously novel or rare strains that are increasingly causing illness)
96 or ‘Persisting’ (strains that consistently cause disease over time)¹⁷. In this Review, we will
97 cover how advances in genomics and metagenomics have revolutionised how we identify
98 bacterial foodborne pathogens, track their evolution and spread, and how our
99 understanding of what hazards are present and how to control them can change.

100 **Bacterial foodborne human pathogens**

101 Foodborne illness remains a significant threat to human health and wealth despite advances
102 in awareness, food hygiene and sanitation. In 2010, the World Health Organization (WHO)
103 estimated that almost 1 in 10 people suffered foodborne illness, with 420,000 dying as a
104 result¹⁸. This was not evenly distributed across geographical regions or demographics, with
105 Asia, Africa (Figure 3a) and children bearing the heaviest burden. The WHO estimate
106 includes cases caused by 31 foodborne hazards, of which 12 were bacterial. However,
107 bacterial agents have an enormous impact, causing five, six and seven of the top 10
108 foodborne illnesses, foodborne deaths, and foodborne disability-adjusted life years,
109 respectively¹⁹; therefore, this Review focuses on foodborne bacterial hazards.

110

111 **Enduring hazards**

112 There is good consensus between food and health agencies on the bacterial genera and
113 species considered as key foodborne hazards, with the importance of many being
114 recognised for decades (Figure 3b). Bacterial foodborne pathogens generally cause disease
115 in one of two ways once ingested. First, through colonisation and subsequent penetration
116 through the gastrointestinal mucus, causing damage to the epithelial cells and subsequent
117 diarrhoea. Examples of key pathogens causing disease in this manner are *Salmonella*
118 *enterica*, *Campylobacter jejuni*, and *Listeria monocytogenes*, and in some cases, disease
119 progresses to invasive illness or systemic syndromes (for example, bloodstream infections
120 and sepsis). Second, foodborne pathogens can cause disease through the production of
121 toxins, as seen in cases such as *Clostridium perfringens*, Shiga toxin-producing *Escherichia*
122 *coli* O157, *Bacillus cereus*, and *Clostridium botulinum*. The biology, epidemiology and
123 pathogenesis of these and other foodborne bacterial hazards have been reviewed
124 extensively elsewhere^{20,21}. With the increasingly large numbers of whole genome sequences
125 of bacterial pathogens being produced in research and coordinated surveillance
126 programmes, a more nuanced picture is emerging on the population structures and
127 underlying genetic determinants associated with their transmission in their associated
128 niches.

129

130 **Emerging hazards**

131

132 Although bacterial foodborne pathogens like *Salmonella enterica*. and *C. jejuni* have been
133 recognised for decades, the threat they pose to human health is compounded by the
134 complex systemic risks associated with the production and global distribution of current and
135 future foods, influenced by continually changing natural and political environments (Figure
136 1). These factors not only mean that foodborne hazards may evolve and change, but they
137 also provide opportunities for the emergence and spread of new or previously unrecognised
138 hazards that cause disease.

139

140 *Whole genome sequencing — not all foodborne pathogens are created equal*

141 Historically, surveillance systems for foodborne pathogens relied on techniques such as
142 serotyping or pulsed-field gel electrophoresis, which enabled differentiation of pathogens
143 beyond genus and species levels and facilitated the linking of isolates to common sources in
144 cases where these subspecies characterisations correlated with epidemiological findings^{22,23}.
145 These techniques often lack high discriminatory power; the advent of higher resolution WGS
146 data has enhanced our ability to define epidemiologically-related clusters of isolates while
147 also understanding the specific mechanisms and diverse ability of individual bacterial strains
148 to cause disease within species or genus²⁴. Bacterial evolution, such as the acquisition of
149 mutations or new genetic elements, can result in increased pathogenicity, virulence, or host
150 range, and subsequent emergence of a new pathogen. For example, successful waves of
151 different subtypes of *S. Typhimurium*, which were originally distinguished by phenotypic
152 methods including serotyping and phage typing including definitive type (DT) 204c and
153 DT104²⁵, have dominated the cases of human disease caused by this organism. The use of
154 WGS has provided insight into the genetic mechanisms facilitating the success of the various
155 subtypes, including the current pandemic subtype monophasic *S. Typhimurium* ST34. In this
156 case, the acquisition of antimicrobial resistance (AMR) genes, a novel prophage and a
157 genomic island encoding metal tolerance genes likely facilitated the transition of this
158 organism from an emerging hazard, with pigs being the most likely reservoir, to an enduring
159 hazard. Monophasic *S. Typhimurium* ST34 is now one of the most common *Salmonella*
160 subtypes globally, found across multiple host species and countries²⁶. Similarly, in the
161 Brazilian poultry industry where antimicrobial selection pressure is high, two distinct
162 serovars of *Salmonella* (*Salmonella enterica* subsp. *enterica* serovar Minnesota and
163 *Salmonella enterica* subsp. *enterica* serovar Heidelberg) became dominant through
164 acquisition of AMR plasmids carrying genes conferring resistance to β -lactams, tetracyclines
165 and sulphonamides²⁷. Common genomic features including loss-of-function mutations and
166 gain of pathogenicity islands have also been identified in *Salmonella* serovars adapted to
167 specific hosts^{12,28}, which offers potential for predicting how future pathogens may emerge.

168

169 Other examples are acquisition of a gene encoding Shiga toxin by an enteroaggregative *E.*
170 *coli* strain, leading to the largest outbreak of Shiga toxin-producing *E. coli* ever recorded¹¹,
171 and *L. monocytogenes*, a highly heterogeneous organism for which the use of WGS has

172 identified different evolutionary lineages. These evolutionary lineages have been divided
173 into clonal complexes which are associated with different sources and levels of virulence²⁹.
174 The generation of increasingly larger numbers of genome sequences allows the application
175 of genomic epidemiology concepts of source attribution and outbreak response²⁴, topics
176 that will be explored later in this Review.
177

178 Threats to our food system and human health from bacterial pathogens arise at various
179 points across the food chain, from post-primary production to the point of consumption.
180 These threats often manifest in conditions that are very different from the original
181 reservoirs, such as animal gastrointestinal tracts, farm soils or crops³⁰. Therefore, key for a
182 successful foodborne pathogen is the ability to survive these diverse conditions, and the use
183 of 'omics technologies is enabling our understanding of the underlying genetic mechanisms.
184 For example, *Cronobacter sakazakii* is a neonatal bacterial pathogen known to persist in
185 low-moisture matrices, such as powdered infant formula. RNA sequencing identified the
186 expression of many genes that were significantly up- or down-regulated as a response to
187 survive desiccation³¹. Phenotypic plasticity has also been observed in major foodborne
188 pathogens, such as *C. jejuni*, where fluoroquinolone resistance has been associated with
189 enhanced survival in the chicken host even in the absence of antimicrobial pressure³². Other
190 mechanisms to support survival through the food chain (reviewed elsewhere³³) can function
191 at the individual bacterium level, such as sporulation³⁴ and metabolic or transcriptomic
192 changes³⁵, or at the community or population level, such as single- or multi-species biofilm
193 formation^{36,37}.

194

195 *Contemporary drivers shaping the microbial landscape*

196

197 The emergence of foodborne bacterial hazards is driven by many factors, which not only
198 include bacterial evolution itself, but also ecological and socioeconomic drivers such as
199 consumer preference, economy and trade, climate change and technology innovation
200 (Figure 1). Market demands shaped by factors such as shifts in consumer preference is
201 changing the way food is produced in many high-income countries (HICs), leading to new
202 routes for foodborne hazards to enter the food chain. For example, the global demand for
203 sustainable protein and policies to reduce greenhouse gas emissions have led to the
204 development of cell-based foods that derive animal agricultural products such as meat
205 directly from cell cultures; along the production stages, multiple entry points for pathogens
206 have been identified through elicitation of expert opinion^{38,39}, such as contamination of raw
207 materials and added reagents with bacteria and other microorganisms that can contaminate
208 cells. However, there remains a lack of empirical data. Although little research has been
209 conducted yet on the potential foodborne hazards associated with these new foods, a
210 recent study demonstrated that growth of *Listeria* spp. and *Salmonella enterica* was greater
211 in plant-based milks than in cow milk⁴⁰. A contrasting trend towards increasing consumption
212 of animal source foods is expected in low- and middle-income countries (LMICs) with
213 economic development, which may result in greater numbers of foodborne illness if
214 investment in food safety management does not keep pace^{41,42}. The increasingly globalised

215 nature of food chains can spread and amplify foodborne hazards, which is explored in more
216 detail later in this Review.

217

218 Climate change has more extensive impacts than merely the impetus for the development
219 of new food types, affecting the dispersal and persistence of bacterial foodborne
220 hazards^{43,44}. In 2022, it was estimated that over half of human pathogenic diseases are
221 exacerbated by climate change, with *Vibrio* spp. as the bacterial foodborne pathogen with
222 the strongest evidence of aggravation by climate hazards⁴⁵. Projections estimate that
223 warming coastal waters will increase the numbers and distribution of vibriosis cases, not
224 only through the extended geographical range of the bacteria but also through increased
225 recreational use of coastal waters⁴⁶⁻⁴⁸.

226

227 Technology innovation in the generation of genome-based data has greatly contributed to
228 our recognition of emerging foodborne hazards. While the increasing cost effectiveness of
229 large scale WGS has revolutionised our ability to understand the biology and characteristics
230 of enduring bacterial pathogens, the majority of foodborne illness is of unknown cause. In
231 the UK, foodborne illness is responsible for an estimated 2.4 million cases, costing in excess
232 of £9bn annually; crucially, £6bn of this cannot be attributed to specific agents and so
233 interventions aimed at reduction are limited by knowledge⁴⁹. The development and use of
234 untargeted, culture-free approaches has been used to investigate both known, and hitherto
235 unknown, bacterial causes of foodborne illness. Whole metagenome sequencing of faecal
236 samples led to the identification of a new candidate species of *Campylobacter*, *Candidatus*
237 *Campylobacter infans*, which has been found in breastfed infants in sub-Saharan Africa and
238 South Asia¹⁵. The use of culture-free methods has also highlighted the widespread presence
239 of *Arcobacter* spp. including *Arcobacter cryaerophilus* and *Arcobacter butzleri* in
240 wastewater⁵⁰, suggesting previously underappreciated reservoirs of these emerging
241 pathogens (Figure 2). Culture-independent diagnostic tests (CIDTs) provide an agnostic and
242 more rapid approach to identifying potential causative agents of foodborne illness, and
243 facilitate the identification of bacteria which cannot be cultured⁵¹. However, barriers to its
244 widespread implementation in food safety applications still exist. The proportion of the food
245 metagenome comprised of pathogens is very low⁵², and so may not be detected, depending
246 on the amount of DNA from food itself within samples and depth of sequencing. The lack of
247 an isolate presents a barrier to the implementation of genome-based evaluations of
248 outbreaks and transmission at multiple scales^{53,54}, explored in the next section. Further
249 challenges arise when attempting to classify bacteria present beyond the species level, and
250 to identify the presence of multiple lineages of the same bacterium within a sample;
251 metagenome-assembled genomes (MAGs) can facilitate such identification, particularly with
252 long-read metagenome sequencing, but do not completely solve this problem. As such,
253 'strain-resolved metagenomics' is an active area of research^{55,56}.

254

255 In terms of food products, all foods will carry bacteria unless specifically sterilised; another
256 advantage of culture-free approaches is that an understanding of the total microbiota — the
257 entire microbial content present — can be obtained. These bacteria may be pathogens, may
258 cause food spoilage or support the persistence of pathogens, or may act as reservoirs of

259 AMR genes; previous work has shown the concentration of AMR genes on different retail
260 foods to be between 10^4 – 10^{10} AMR genes per gram of food⁵². The majority of our
261 knowledge of foodborne pathogens is based on investigation of pure cultures of single
262 strains within the laboratory, but in the food chain pathogens exist within complex microbial
263 communities. These interactions are often critical to the success of foodborne pathogens;
264 for example, previous studies have shown how *C. jejuni* benefits from passive protection of
265 existing *Pseudomonas aeruginosa* biofilms on abiotic surfaces³⁶ and that multi-species
266 biofilms impart biocide tolerance to *L. monocytogenes*⁵⁷.

267

268 The novel insights into bacterial foodborne hazards through technological innovation also
269 extends to the analysis of the vast amount of data generated by genome-based approaches.
270 Leveraging the recent advances in artificial intelligence (AI), big data and machine learning,
271 novel genetic factors associated with successful foodborne pathogens have been identified.
272 Such approaches have been used to identify specific subsets of cattle-associated Shiga-toxin
273 producing *E. coli* O157 strains that are more likely to cause human disease, even within
274 previously defined pathogenic lineages⁵⁸, and to identify new genes potentially associated
275 with *E. coli* pathogenicity^{59,60}. Other applications of machine learning for food safety include
276 the analysis of public search records as a proxy for on-site inspection of restaurants⁶¹.

277

278

279 **Transmission pathways and propagation of pathogens along the food chain**

280

281 The prevention and control of both enduring and emerging foodborne bacterial hazards
282 requires an understanding of the factors influencing the propagation of pathogens along the
283 food chain. This extends across multiple scales, both spatial and temporal, encompassing
284 the transmission of pathogens through increasingly interconnected global supply chains, the
285 movement of people and animals across international borders, and localised outbreaks or
286 sporadic cases arising from contaminated food products. Many foodborne pathogens, both
287 enduring and emerging, are amplified in animal reservoirs (maintenance hosts) before
288 contaminating the food supply chain. Hence, the need to adopt a 'One Health' approach³⁰ in
289 the surveillance of bacterial foodborne hazards that considers sampling across time and
290 space in different environments (Figure 4) to identify robustly the pathways that move
291 pathogens from animal reservoirs to the point of ingestion, infection and disease in human
292 hosts (Figure 5).

293

294 Genome-based approaches provide greater resolution for the investigation of transmission
295 at multiple scales, from global to local, and through complex food supply chains and
296 networks. Examples of the application of these approaches are described below. These
297 include determining historical and contemporary patterns of transmission on a global scale,
298 outbreak investigation at local and international scales, and source attribution and source
299 tracking along the food chain.

300

301 *Global-scale transmission*

302 Increased global trade in food products is a major driver of improved food security, notably
303 for countries with rapid population growth and natural resource constraints. However,
304 historical trade in livestock has spread foodborne pathogens between continents and trade
305 increases the potential for contaminated food to be moved further and more rapidly
306 between countries. Genome-based tools provide evidence of the role of historical trade in
307 live cattle in the international transmission of Shiga toxin-producing *E. coli*⁶², and insight into
308 the role of transatlantic livestock trade followed by the growth of cattle farming and
309 industrialisation of food production, in the spread of *L. monocytogenes*⁶³. Recent advances
310 in machine learning, combined with genome sequencing, have enabled rapid global source
311 tracking of *Salmonella enterica*, identifying the geographical origin of cases arising from
312 contaminated food products in the global supply chain⁶⁴.

313 *Outbreak investigation*

314 Cases of foodborne illness are often categorised as either outbreak-associated or sporadic,
315 where the latter are not considered to be part of a temporal and/or spatial cluster of cases
316 associated with a common source. By aiding hypothesis-generation and serving as high-
317 quality evidence in public health investigations, high resolution genome-based methods
318 have helped to reduce the size and duration of outbreaks in the United States compared to
319 lower resolution molecular tools⁶⁵ (Figure 6). This is achieved by comparison of isolates from
320 clinical cases with isolates from food or food production environments; isolates from food
321 sources that cluster together with human isolates, as a result of differing by a small number
322 of single nucleotide polymorphisms (SNPs) or alleles, are more likely to have a recent
323 common ancestor and be implicated as the source of the outbreak (for example, the large
324 scale outbreak of listeriosis in South Africa⁶⁶). Genome sequencing of presumptive sporadic
325 cases has also identified previously undetected clusters of cases that are more likely to
326 share a common source, providing evidence of covert outbreaks^{67,68}.

327
328 Determining membership of a cluster of cases with a putative common origin can be
329 difficult. A study⁶⁹ indicates that three metrics can provide evidence for a common origin of
330 sequenced isolates, thereby indicating their association with of an outbreak and links to
331 food sources. These metrics are the number of SNPs, the location of isolates relative to
332 others in the tree topology (for instance, monophyly provides greater support than
333 paraphyly and polyphyly) and bootstrap support for the branch containing the isolates in
334 question. The number of SNPs or shared alleles that are consistent with a recent common
335 ancestor depends on the organism and period of sampling. A study²⁹ used the distribution
336 of core genome multilocus sequence typing (cgMLST) allele differences between isolates
337 from known outbreak sets and all cases of listeriosis caused by *L. monocytogenes* to propose
338 a cutoff of <7 allele differences for membership of an outbreak, whereas a recent modelling
339 study identified a wider range of epidemiologically relevant cutoffs not only within *L.*
340 *monocytogenes* but also a range of SNP or cgMLST allele cutoffs within other common
341 bacterial foodborne hazards⁷⁰.

342
343 Technology for high throughput sequencing has advanced rapidly, but spatial resolution and
344 speed is important to pinpoint the origin of an outbreak to a specific locale⁷¹. This

345 underlines the importance of surveillance, facilitated by global collaboration and data
346 sharing among researchers, public health agencies and industry, including shared data
347 repositories of genome sequences and accompanying metadata⁷². Examples include:
348 National Center for Biotechnology Information⁷³; GenomeTrakr Network⁷⁴; Global Microbial
349 Identifier⁷⁵; and PubMLST⁷⁶.

350

351 Rapid sharing of genomic and associated data helps to resolve local and international
352 outbreaks of enduring pathogens, and aid the characterisation, detection and control of
353 emerging pathogens¹¹ (for example, aggregate datasets from multiple programmes,
354 informing interventions such as novel vaccine designs⁷⁷).

355

356 The ability to apply the techniques described above depends on the availability of large
357 repositories of bacterial whole genome sequences with high quality associated
358 epidemiological metadata, such as date of isolation, location, and host species. Considerable
359 global efforts by publicly-funded and commercial laboratories and public health and food
360 safety authorities have helped to develop the strategic and operational requirements to
361 support widespread implementation and routine use of genomics as a tool for food safety
362 purposes, including sharing of metadata and the standardisation of terminology,
363 bioinformatic analytics, databases and data access (reviewed in refs.^{24,51,78,79}). The
364 decreasing cost of sequencing has allowed the implementation of WGS-based surveillance
365 of bacterial foodborne hazards in many high resource settings^{23,80,81}. However, given the
366 global nature of travel, food supply chains and bacterial foodborne hazards, the lack of
367 substantial similar data from low resource settings is a barrier to applying these methods in
368 such settings or even understanding the major food safety concerns there^{82,83}. Furthermore,
369 the identification of emerging hazards will be even more difficult if they arise in a region
370 with no or limited (sequencing) surveillance capacity. The main barrier to implementation of
371 WGS for food safety in low resource settings identified in a recent Review was lack of
372 governmental dedication of resources for WGS implementation⁸³. Additional issues
373 identified were a lack of bioinformatics expertise, computing resources and insufficient
374 resources and infrastructures^{82,83}, although fundamentally, countries without an established
375 surveillance system are unlikely to see the benefit of adding WGS capacity⁸³.

376

377 The development of more accessible tools for analysing and interpreting genomic data has
378 improved the accessibility, visualisation and presentation of public and private genome
379 sequencing and related metadata. Examples of such platforms include interactive web-
380 based tools such as Nextstrain⁸⁴, Microreact⁸⁵ and Pathogenwatch⁸⁶. These are dynamic and
381 scalable, and provide a variety of phylogenetic, genomic and epidemiological tools. The
382 developers of Nextstrain and Microreact provide open-source software for end-users to
383 design bespoke applications. As more genomics data with associated metadata and
384 improved bioinformatics and statistical tools become available, the utility of these
385 applications for decision makers will increase. This was evident during the COVID-19
386 pandemic, where pathogen-specific genomic tools were rapidly developed, providing easy
387 to interpret, interactive websites that were designed to aid public health decision making⁸⁷,
388 and which have also been used for foodborne bacterial pathogens⁸⁸⁻⁹⁰.

389

390 *Source attribution and source tracking: determining reservoirs, pathways and risk factors for*
391 *foodborne illness*

392 The availability of higher resolution genomic data has resulted in development of new
393 model-based approaches for unravelling the complex chain of events that lead to foodborne
394 illness, from animal reservoirs to the host factors that lead to ingestion, infection and
395 disease (Figure 5). Determining the 'source' of contamination of food with foodborne
396 pathogens helps the implementation of appropriate responses to reduce the burden of
397 foodborne illness. This includes understanding the relative importance of different animal
398 reservoirs and infection pathways, identifying the source of contamination in food
399 processing and determining host-associated risk factors.

400

401 Reservoirs and pathways. Determining the relative contribution of different animal
402 reservoirs to the overall burden of disease caused by foodborne pathogens helps the
403 implementation of effective control measures and can lead to measurable improvements in
404 food safety and public health⁹¹. However, there are often multiple animal reservoirs and
405 transmission pathways involved in cases of illness associated with enduring pathogens such
406 as *C. jejuni* and *Salmonella enterica*. Animal reservoirs include wildlife⁹² as well as food-
407 producing animals, and faeces from the same reservoir can contaminate multiple pathways,
408 for example leading to both food and waterborne infections (Figure 5). Many techniques
409 have been devised for determining the relative contribution of different animal reservoirs
410 and pathways, including expert opinion, exposure modelling and, more recently, molecular
411 and genomic methods^{93,94}.

412

413 Advances in modelling have enabled DNA sequence data from isolates from putative
414 sources to be combined with sequence data from clinical cases to determine the relative
415 contribution of, for example, chickens, cattle and sheep, to the burden of
416 campylobacteriosis and salmonellosis⁹⁵⁻⁹⁷. Earlier applications of these models used
417 multilocus sequence typing, typically involving loci in seven genes that were amplified and
418 sequenced using Sanger sequencing⁹⁸. Reservoir attribution models have recently been
419 extended to include covariates, highlighting the differences in attribution in urban and rural
420 areas, as well as the changes occurring before and after interventions in the food chain⁹⁹.

421 The advent of WGS has resulted in the availability of considerably more, higher resolution
422 genomic data for source attribution modelling, including cgMLST and k-mer analyses. The
423 application of machine learning tools for reservoir attribution using WGS data has the
424 potential to provide more accurate and precise estimates of source attribution^{100,101}. Other
425 approaches include the use of network analysis¹⁰² and genome-wide identification of host
426 associated SNPs¹⁰³. Non-culture based metagenomic approaches have also been used for
427 reservoir attribution, with analysis conducted using machine learning tools¹⁰⁴. However,
428 many technical challenges remain in utilizing WGS data for source attribution modelling¹⁰⁵.

429

430 Processing plant tracking and food source attribution. The application of WGS was described
431 for identifying the ingredient that was the source of *L. monocytogenes* in ice cream and for

432 determining whether or not a strain of *Salmonella* was 'resident' in a processing plant⁶⁹.
433 Machine learning and *Listeria* spp. genomic data have been used to identify food sources¹⁰⁶
434 and to distinguish environmental (for example, soil) from food factory isolates¹⁰⁷. The latter
435 provides insight into barriers that would limit but not prevent transmission between these
436 two niches.

437 Recent food industry initiatives have explored the use of metagenomic approaches for
438 monitoring hygiene and source tracking in food chains, processing plants and food
439 production facilities. The approach aims to characterise the 'normal' microbiome of food
440 ingredients and final products to provide a framework for identifying anomalies that may be
441 associated with contamination and potential food safety risks¹⁰⁸. Further applications of
442 metagenomics in food safety, including food and environmental sampling, have been
443 reviewed in ref.⁵¹. The Review also describes a number of risks and benefits associated with
444 the application of metagenomics for food safety, and the barriers to application, such as
445 standardisation of methods, cost, and lack of representative datasets and bioinformatics
446 expertise.

447 Host-associated risk factors. Reservoir attribution models have been combined with a
448 traditional case-control study design to identify risk factors for human cases arising from
449 different pathways. These studies provide both reservoir attribution and risk factor
450 analyses, and include epidemiological studies of campylobacteriosis using 7-gene MLST¹⁰⁹
451 and genome sequencing¹¹⁰

452 **Food safety interventions informed by genomics**

453

454 Genomics is now widely used by many government agencies to gather the detailed evidence
455 needed to support surveillance activities and take informed action in the multiple
456 transmission pathways affecting food safety. Likewise, food businesses are now widely
457 considering the application of ever-maturing genomics technologies to improve their
458 individualised circumstances and processes to produce food^{24,111,112}. Both government and
459 food business have the same motivation — the safety of foods — with overlapping
460 perspectives on how to reduce the risk of exposure to consumers. Government agencies
461 such as those responsible for public health and food safety commonly take the view of the
462 general population, whereas food businesses such as those producing or serving finished
463 foods take the view of their customers. Both views are connected because the opportunity
464 to understand and act on risks in localised production settings informs, and in turn, is
465 informed by, global food systems and the wider population of food consumers (Figure 4).

466 The consideration or adoption of new technological advances is a core component to the
467 food safety culture within food businesses, as they seek to identify, understand, and then
468 control foodborne risks¹¹³. Moreover, in many countries it is a regulatory requirement that
469 food businesses establish a food safety management plan founded on the principles of
470 Hazard Analysis and Critical Control Point (HACCP) that include the establishment of
471 monitoring procedures and a process for corrective action. HACCP sampling plans generally
472 involve laboratory testing for microorganisms such as *Salmonella* spp., *L. monocytogenes*,
473 *Campylobacter* spp. and Shiga toxin-producing *E. coli*¹¹⁴ where detection of these pathogens
474 can necessitate further investigation and action on root causes¹¹⁵. Therefore, within a
475 producer's own food safety culture and best practices, the HACCP principles and resulting
476 management plan are the most rigorous measures that food producers use to ensure food
477 safety risks can be identified and prevented, representing localised change. Given that
478 genomics is now becoming feasible to deploy in these settings, similar to their use in public
479 health programmes where the depth of knowledge surpasses that of current routine
480 microbiology surveillance testing, it is likely that any new genomic testing system will likely
481 be applied within the context of HACCP.

482

483 Food businesses that consider incorporating genomics into their testing programmes
484 demonstrate an innovative management approach, which reflects a forward-thinking vision
485 with a willingness to think differently about the nature of microbial risks and how they can
486 be controlled with new behaviours and actions. To develop a genomics capacity that is
487 suited for application in food businesses — such that the methodology can sample across
488 diverse food and environmental matrices¹¹⁶ while being customisable to the individualised
489 circumstances on how a business produces food — it will be important that there is mutual
490 engagement between food businesses and institutes with expertise in genomics to develop
491 most use-case scenarios. As a future perspective and informed by stakeholder engagements
492 with the food industry (partly through the UK Food Safety Research Network), we propose
493 illustrative examples of questions that food businesses may now ask of their processes and
494 the possible actions they may then take (Box 1). The depth of knowledge gained from

495 genomics offers those responsible for the supply into food chains improved compliance,
496 consumer assurance, and market and brand protection, through the gained confidence in
497 the safety of products going to market^{52,114,115}.
498

499 **Conclusions and perspectives**

500 Bacterial foodborne pathogens remain a significant threat to human health and economies
501 across the world. The enhanced resolution information provided by WGS and metagenomics
502 has advanced our understanding of these hazards, and demonstrated that enduring and
503 emerging foodborne hazards represent a continuum, with bacterial hazards shifting along
504 this continuum. Furthermore, the complex biological, socioeconomic and ecological drivers
505 shaping the evolution and transmission of bacterial pathogens, and the wider microbial
506 communities in which they exist, emphasise the need for surveillance systems that are
507 sensitive enough to identify both enduring and emerging hazards. Key to these surveillance
508 systems is not only sensitivity, but also timeliness, so that pre-emptive action to prevent
509 foodborne illness can be taken rather than using the information to respond to incidents
510 once they have occurred. Currently, such surveillance systems exist predominantly in high
511 resource settings; despite the decreasing cost of technologies, there is still an inconsistency
512 across the international landscape and a disparity in resource availability between high and
513 low resource settings. Although there have been very few economic evaluations of the
514 application of WGS for foodborne pathogen surveillance¹¹⁷, the net benefit attributed to the
515 introduction of WGS in some high resource settings is estimated to be substantial^{65,118}. In a
516 food supply and trading ecosystem that is now global, these technologies also need to be
517 enabled in low resource settings to obtain a true understanding of the nature and risks of
518 foodborne hazards globally. This requires collaboration between high and low resource
519 settings, underpinned by data sharing and common standards for the generation and
520 interpretation of genome-based approaches to food safety, involving researchers, public
521 health agencies and industry across the various One Health compartments.
522

523 As routine microbial testing supports the assessment of hazards within food systems and
524 serves as a signal for further action when hazards are detected, there is a need to
525 understand the more complicated scale of information produced by genome-based testing
526 and then to translate those findings into interventions. For example, identification of
527 specific lineages or genes associated with pathogen success or persistence through the use
528 of WGS and other 'omics technologies can suggest targets to design future intervention
529 strategies. However, the use of metagenomics may identify the presence of pathogens but
530 it cannot determine whether these organisms are viable or if their identification is an
531 artefact⁵². This may have ethical, commercial and legal implications which need to be
532 resolved. Monitoring microbial communities along the food chain may serve as an early
533 warning system for food safety issues or may help identify candidates for biocontrol agents,
534 but requires a step change in thinking about food safety: "...a modified approach to hazard
535 analysis and risk assessment for food safety may be required; for instance, shifting from risk
536 assessments of individual pathogens to assessments of the entire microbial community and
537 food chain", as stated in ref.⁵¹. To exploit the full value of genomic and metagenomic data, it
538 is important to remember that these data should not be considered in isolation. Instead,
539 they need to be integrated with epidemiological and other information across all spatial and
540 temporal scales, from local (for example, movement of people and equipment within a food

541 processing facility) to global (for example, long-term climate change patterns or
542 international trade practices), in order to effect change.

543

544 The true burden of foodborne illness is difficult to estimate due to lack of surveillance, as
545 well as underreporting and challenges in definitively identifying causes of foodborne
546 illness¹¹⁹. Food safety is a key component of food security, and in a complex and changing
547 world shaped by dynamic ecological and socioeconomic drivers, improved implementation
548 of genome-based technologies offers an opportunity to strengthen the prevention,
549 detection and response to foodborne bacterial hazards throughout the entire food chain.

550

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552 **References**

- 553 1 Zurek, M. *et al.* Assessing sustainable food and nutrition security of the EU food system-an
554 integrated approach. *Sustainability* **10**, doi:10.3390/su10114271 (2018).
- 555 2 Parsons, K., Hawkes, C. & Wells, R. Brief 2. Understanding the food system: Why it matters
556 for food policy. (Centre for Food Policy, London, 2019).
- 557 3 Hasnain, S., Ingram, J. & Zurek, M. Mapping the UK Food System - a report for the UKRI
558 Transforming UK Food Systems Programme. (Environmental Change Institute, University of
559 Oxford, Oxford, 2020).
- 560 4 FAO. *If it isn't safe, it isn't food*, <<https://www.fao.org/fao-stories/article/en/c/1179647/>>
561 (2019).
- 562 5 FAO. FAO strategic priorities for food safety within the FAO strategic framework 2022-2031.
563 (Food and Agricultural Organization of the United Nations, Rome, 2023).
- 564 6 Committee, C. C. Progress in adapting to climate change; 2023 Report to Parliament. (2023).
- 565 7 EFSA, Maggiore, A., Afonso, A., Barrucci, F. & De Sanctis, G. Climate change as a driver of
566 emerging risks for food and feed safety, plant, animal health and nutritional quality.
567 (European Food Safety Authority, 2020).
- 568 8 Morgado, M. E. *et al.* Climate change, extreme events, and increased risk of salmonellosis:
569 foodborne diseases active surveillance network (FoodNet), 2004-2014. *Environ Health* **20**,
570 105, doi:10.1186/s12940-021-00787-y (2021).
- 571 9 FAO. Final meeting report: Technical meeting on the impact of whole genome sequencing
572 (WGS) on food safety management: within a One Health approach. (Food and Agriculture
573 Organization of the United Nations, 2016).
- 574 10 Morse, S. S. Factors in the emergence of infectious diseases. *Emerg Infect Dis* **1**, 7-15,
575 doi:10.3201/eid0101.950102 (1995).
- 576 11 Grad, Y. H. *et al.* Genomic epidemiology of the *Escherichia coli* O104:H4 outbreaks in Europe,
577 2011. *Proc Natl Acad Sci U S A*, doi:10.1073/pnas.1121491109 (2012).
- 578 12 Kingsley, R. A. *et al.* Epidemic multiple drug resistant *Salmonella* Typhimurium causing
579 invasive disease in sub-Saharan Africa have a distinct genotype. *Genome Res* **19**, 2279-2287,
580 doi:10.1101/gr.091017.109 (2009).
- 581 13 Seribelli, A. A. *et al.* Phylogenetic analysis revealed that *Salmonella* Typhimurium ST313
582 isolated from humans and food in Brazil presented a high genomic similarity. *Braz J*
583 *Microbiol* **51**, 53-64, doi:10.1007/s42770-019-00155-6 (2020).
- 584 14 Almeida, F. *et al.* Multilocus sequence typing of *Salmonella* Typhimurium reveals the
585 presence of the highly invasive ST313 in Brazil. *Infect Genet Evol* **51**, 41-44,
586 doi:10.1016/j.meegid.2017.03.009 (2017).
- 587 15 Bian, X. *et al.* *Campylobacter* abundance in breastfed infants and identification of a new
588 species in the Global Enterics Multicenter Study. *mSphere* **5**, doi:10.1128/mSphere.00735-19
589 (2020).

590 16 Ramees, T. P. *et al.* *Arcobacter*: an emerging food-borne zoonotic pathogen, its public health
591 concerns and advances in diagnosis and control - a comprehensive review. *Vet Q* **37**, 136-
592 161, doi:10.1080/01652176.2017.1323355 (2017).

593 17 CDC. *Reoccurring, emerging, and persisting enteric bacterial strains*,
594 <<https://www.cdc.gov/ncezid/dfwed/outbreak-response/rep-strains.html>> (2023).

595 18 WHO. WHO estimates of the global burden of foodborne diseases: foodborne disease
596 burden epidemiology reference group 2007-2015., (World Health Organization, Switzerland,
597 2015).

598 19 Devleeschauwer, B., Haagsma, J. A., Mangen, M.-J. J., Lake, R. J. & Havelaar, A. H. in *Food*
599 *Safety Economics* (ed T. Roberts) (Springer, 2018).

600 20 Abebe, E., Gugsu, G. & Ahmed, M. Review on major food-borne zoonotic bacterial
601 pathogens. *J Trop Med* **2020**, 4674235, doi:10.1155/2020/4674235 (2020).

602 21 Bintsis, T. Foodborne pathogens. *AIMS Microbiol* **3**, 529-563,
603 doi:10.3934/microbiol.2017.3.529 (2017).

604 22 Foley, S. L., Lynne, A. M. & Nayak, R. Molecular typing methodologies for microbial source
605 tracking and epidemiological investigations of Gram-negative bacterial foodborne
606 pathogens. *Infect Genet Evol* **9**, 430-440, doi:10.1016/j.meegid.2009.03.004 (2009).

607 23 Deng, X., den Bakker, H. C. & Hendriksen, R. S. Genomic epidemiology: Whole-genome-
608 sequencing-powered surveillance and outbreak investigation of foodborne bacterial
609 pathogens. *Annu Rev Food Sci Technol* **7**, 353-374, doi:10.1146/annurev-food-041715-
610 033259 (2016).

611 24 Taboada, E. N., Graham, M. R., Carrico, J. A. & Van Domselaar, G. Food safety in the age of
612 next generation sequencing, bioinformatics, and open data access. *Front Microbiol* **8**, 909,
613 doi:10.3389/fmicb.2017.00909 (2017).

614 25 Threlfall, E. J. Epidemic *Salmonella* Typhimurium DT104 - a truly international multiresistant
615 clone. *J Antimicrob Chemother* **46**, 7-10 (2000).

616 26 Petrovska, L. *et al.* Microevolution of monophasic *Salmonella* Typhimurium during epidemic,
617 United Kingdom, 2005-2010. *Emerg Infect Dis* **22**, 617-624, doi:10.3201/eid2204.150531
618 (2016).

619 27 Alikhan, N. F. *et al.* Dynamics of *Salmonella enterica* and antimicrobial resistance in the
620 Brazilian poultry industry and global impacts on public health. *PLoS Genet* **18**, e1010174,
621 doi:10.1371/journal.pgen.1010174 (2022).

622 28 Langridge, G. C. *et al.* Patterns of genome evolution that have accompanied host adaptation
623 in *Salmonella*. *Proc Natl Acad Sci U S A* **112**, 863-868, doi:10.1073/pnas.1416707112 (2015).

624 29 Moura, A. *et al.* Whole genome-based population biology and epidemiological surveillance
625 of *Listeria monocytogenes*. *Nat Microbiol* **2**, 16185, doi:10.1038/nmicrobiol.2016.185 (2016).

626 30 Ma, L.-C., Zhao, H.-Q., Wu, L. B., Cheng, Z.-L. & Liu, C. Impact of the microbiome on human,
627 animal and environmental health from a One Health perspective. *Science in One Health* **2**,
628 100037 (2023).

629 31 Srikumar, S. *et al.* RNA sequencing-based transcriptional overview of xerotolerance in
630 *Cronobacter sakazakii* SP291. *Appl Environ Microbiol* **85**, doi:10.1128/AEM.01993-18 (2019).

631 32 Luo, N. *et al.* Enhanced in vivo fitness of fluoroquinolone-resistant *Campylobacter jejuni* in
632 the absence of antibiotic selection pressure. *Proc Natl Acad Sci U S A* **102**, 541-546,
633 doi:10.1073/pnas.0408966102 (2005).

634 33 Marmion, M., Macori, G., Ferone, M., Whyte, P. & Scannell, A. G. M. Survive and thrive:
635 Control mechanisms that facilitate bacterial adaptation to survive manufacturing-related
636 stress. *Int J Food Microbiol* **368**, 109612, doi:10.1016/j.ijfoodmicro.2022.109612 (2022).

637 34 Gauvry, E. *et al.* Knowledge of the physiology of spore-forming bacteria can explain the
638 origin of spores in the food environment. *Res Microbiol* **168**, 369-378,
639 doi:10.1016/j.resmic.2016.10.006 (2017).

640 35 NicAogain, K. & O'Byrne, C. P. The role of stress and stress adaptations in determining the
641 fate of the bacterial pathogen *Listeria monocytogenes* in the food chain. *Front Microbiol* **7**,
642 1865, doi:10.3389/fmicb.2016.01865 (2016).

643 36 Teh, A. H. T., Lee, S. M. & Dykes, G. A. Association of some *Campylobacter jejuni* with
644 *Pseudomonas aeruginosa* biofilms increases attachment under conditions mimicking those
645 in the environment. *PLoS One* **14**, e0215275, doi:10.1371/journal.pone.0215275 (2019).

646 37 Whelan, M. V. X. *et al.* Acquisition of fluoroquinolone resistance leads to increased biofilm
647 formation and pathogenicity in *Campylobacter jejuni*. *Sci Rep* **9**, 18216, doi:10.1038/s41598-
648 019-54620-1 (2019).

649 38 Ong, K. J. *et al.* Food safety considerations and research priorities for the cultured meat and
650 seafood industry. *Compr Rev Food Sci Food Saf* **20**, 5421-5448, doi:10.1111/1541-
651 4337.12853 (2021).

652 39 FAO & WHO. Food safety aspects of cell-based food. (Food and Agriculture Organization of
653 the United Nations; World Health Organization, Rome, Italy, 2023).

654 40 Bartula, K., Begley, M., Latour, N. & Callanan, M. Growth of food-borne pathogens *Listeria*
655 and *Salmonella* and spore-forming *Paenibacillus* and *Bacillus* in commercial plant-based milk
656 alternatives. *Food Microbiol* **109**, 104143, doi:10.1016/j.fm.2022.104143 (2023).

657 41 Li, M. *et al.* Global disease burden of pathogens in animal source foods, 2010. *PLoS One* **14**,
658 e0216545, doi:10.1371/journal.pone.0216545 (2019).

659 42 Jaffee, S., Henson, S., Unnevehr, L., Grace, D. & Cassou, E. The safe food imperative:
660 Accelerating progress in low- and middle-income countries. (World Bank, Washington, D.C.,
661 2019).

662 43 Hellberg, R. S. & Chu, E. Effects of climate change on the persistence and dispersal of
663 foodborne bacterial pathogens in the outdoor environment: A review. *Crit Rev Microbiol* **42**,
664 548-572, doi:10.3109/1040841X.2014.972335 (2016).

665 44 Jones, B. A. *et al.* Zoonosis emergence linked to agricultural intensification and
666 environmental change. *Proc Natl Acad Sci U S A*, doi:10.1073/pnas.1208059110 (2013).

667 45 Mora, C. *et al.* Over half of known human pathogenic diseases can be aggravated by climate
668 change. *Nat Clim Chang* **12**, 869-875, doi:10.1038/s41558-022-01426-1 (2022).

669 46 Archer, E. J. *et al.* Climate warming and increasing *Vibrio vulnificus* infections in North
670 America. *Sci Rep* **13**, 3893, doi:10.1038/s41598-023-28247-2 (2023).

671 47 Vezzulli, L. *et al.* Climate influence on *Vibrio* and associated human diseases during the past
672 half-century in the coastal North Atlantic. *Proc Natl Acad Sci U S A* **113**, E5062-5071,
673 doi:10.1073/pnas.1609157113 (2016).

674 48 Baker-Austin, C. *et al.* Emerging *Vibrio* risk at high latitudes in response to ocean warming.
675 *Nat Clim Chang* **3**, 73-77 (2013).

676 49 FSA. The burden of foodborne disease in the UK 2018. (Food Standards Agency, 2020).

677 50 Kristensen, J. M., Nierychlo, M., Albertsen, M. & Nielsen, P. H. Bacteria from the genus
678 *Arcobacter* are abundant in effluent from wastewater treatment plants. *Appl Environ*
679 *Microbiol* **86**, doi:10.1128/AEM.03044-19 (2020).

680 51 Billington, C., Kingsbury, J. M. & Rivas, L. Metagenomics approaches for improving food
681 safety: A review. *J Food Prot* **85**, 448-464, doi:10.4315/JFP-21-301 (2022).

682 52 Bloomfield, S. J. *et al.* Determination and quantification of microbial communities and
683 antimicrobial resistance on food through host DNA-depleted metagenomics. *Food Microbiol*
684 **110**, 104162, doi:10.1016/j.fm.2022.104162 (2023).

685 53 Carleton, H. A. *et al.* Metagenomic approaches for public health surveillance of foodborne
686 infections: Opportunities and challenges. *Foodborne Pathog Dis* **16**, 474-479,
687 doi:10.1089/fpd.2019.2636 (2019).

688 54 Ray, L. C. *et al.* Changing diagnostic testing practices for foodborne pathogens, Foodborne
689 Diseases Active Surveillance Network, 2012-2019. *Open Forum Infect Dis* **9**, ofac344,
690 doi:10.1093/ofid/ofac344 (2022).

691 55 Anyansi, C., Straub, T. J., Manson, A. L., Earl, A. M. & Abeel, T. Computational methods for
692 strain-level microbial detection in colony and metagenome sequencing data. *Front Microbiol*
693 **11**, 1925, doi:10.3389/fmicb.2020.01925 (2020).

694 56 Kang, X., Luo, X. & Schonhuth, A. StrainXpress: strain aware metagenome assembly from
695 short reads. *Nucleic Acids Res* **50**, e101, doi:10.1093/nar/gkac543 (2022).

696 57 Rolon, M. L., Voloshchuk, O., Bartlett, K. V., LaBorde, L. F. & Kovac, J. Multi-species biofilms
697 of environmental microbiota isolated from fruit packing facilities promoted tolerance of
698 *Listeria monocytogenes* to benzalkonium chloride. *Biofilm* **7**, 100177,
699 doi:10.1016/j.bioflm.2024.100177 (2024).

700 58 Lupolova, N., Dallman, T. J., Matthews, L., Bono, J. L. & Gally, D. L. Support vector machine
701 applied to predict the zoonotic potential of *E. coli* O157 cattle isolates. *Proc Natl Acad Sci U S*
702 *A* **113**, 11312-11317, doi:10.1073/pnas.1606567113 (2016).

703 59 Im, H., Hwang, S. H., Kim, B. S. & Choi, S. H. Pathogenic potential assessment of the Shiga
704 toxin-producing *Escherichia coli* by a source attribution-considered machine learning model.
705 *Proc Natl Acad Sci U S A* **118**, doi:10.1073/pnas.2018877118 (2021).

706 60 Burgaya, J. *et al.* The bacterial genetic determinants of *Escherichia coli* capacity to cause
707 bloodstream infections in humans. *PLoS Genet* **19**, e1010842,
708 doi:10.1371/journal.pgen.1010842 (2023).

709 61 Sadilek, A. *et al.* Machine-learned epidemiology: real-time detection of foodborne illness at
710 scale. *NPJ Digit Med* **1**, 36, doi:10.1038/s41746-018-0045-1 (2018).

711 62 Franz, E. *et al.* Phylogeographic analysis reveals multiple international transmission events
712 have driven the global emergence of *Escherichia coli* O157:H7. *Clin Infect Dis* **69**, 428-437,
713 doi:10.1093/cid/ciy919 (2019).

714 63 Moura, A. *et al.* Emergence and global spread of *Listeria monocytogenes* main clinical clonal
715 complex. *Sci Adv* **7**, eabj9805, doi:10.1126/sciadv.abj9805 (2021).

716 64 Bayliss, S. C. *et al.* Rapid geographical source attribution of *Salmonella enterica* serovar
717 Enteritidis genomes using hierarchical machine learning. *Elife* **12**, doi:10.7554/eLife.84167
718 (2023).

719 65 Brown, B., Allard, M., Bazaco, M. C., Blankenship, J. & Minor, T. An economic evaluation of
720 the Whole Genome Sequencing source tracking program in the U.S. *PLoS One* **16**, e0258262,
721 doi:10.1371/journal.pone.0258262 (2021).

722 66 Thomas, J. *et al.* Outbreak of listeriosis in South Africa associated with processed meat. *N*
723 *Engl J Med* **382**, 632-643, doi:10.1056/NEJMoa1907462 (2020).

724 67 Kovanen, S. M. *et al.* Multilocus sequence typing (MLST) and whole-genome MLST of
725 *Campylobacter jejuni* isolates from human infections in three districts during a seasonal peak
726 in Finland. *J Clin Microbiol* **52**, 4147-4154, doi:10.1128/JCM.01959-14 (2014).

727 68 Yang, C. *et al.* Outbreak dynamics of foodborne pathogen *Vibrio parahaemolyticus* over a
728 seventeen year period implies hidden reservoirs. *Nat Microbiol* **7**, 1221-1229,
729 doi:10.1038/s41564-022-01182-0 (2022).

730 69 Pightling, A. W. *et al.* Interpreting whole-genome sequence analyses of foodborne bacteria
731 for regulatory applications and outbreak investigations. *Front Microbiol* **9**, 1482,
732 doi:10.3389/fmicb.2018.01482 (2018).

733 70 Duval, A., Opatowski, L. & Brisse, S. Defining genomic epidemiology thresholds for common-
734 source bacterial outbreaks: a modelling study. *Lancet Microbe* **4**, e349-e357,
735 doi:10.1016/S2666-5247(22)00380-9 (2023).

736 71 Hoffmann, M. *et al.* Tracing origins of the *Salmonella* Bareilly strain causing a food-borne
737 outbreak in the United States. *J Infect Dis* **213**, 502-508, doi:10.1093/infdis/jiv297 (2016).

738 72 Dooley, D. M. *et al.* FoodOn: a harmonized food ontology to increase global food
739 traceability, quality control and data integration. *NPJ Sci Food* **2**, 23, doi:10.1038/s41538-
740 018-0032-6 (2018).

741 73 NCBI. *National Center for Biotechnology Information datasets*,
742 <<https://www.ncbi.nlm.nih.gov/datasets/>> (2024).

743 74 Timme, R. E. *et al.* GenomeTrakr proficiency testing for foodborne pathogen surveillance: an
744 exercise from 2015. *Microb Genom* **4**, doi:10.1099/mgen.0.000185 (2018).

745 75 Moran-Gilad, J. *et al.* Proficiency testing for bacterial whole genome sequencing: an end-
746 user survey of current capabilities, requirements and priorities. *BMC Infect Dis* **15**, 174,
747 doi:10.1186/s12879-015-0902-3 (2015).

748 76 Jolley, K. A., Bray, J. E. & Maiden, M. C. J. Open-access bacterial population genomics:
749 BIGSdb software, the PubMLST.org website and their applications. *Wellcome Open Res* **3**,
750 124, doi:10.12688/wellcomeopenres.14826.1 (2018).

751 77 Baker, K. S. *et al.* Genomics for public health and international surveillance of antimicrobial
752 resistance. *Lancet Microbe* **4**, e1047-e1055, doi:10.1016/S2666-5247(23)00283-5 (2023).

753 78 Kovac, J., den Bakker, H., Carroll, L. M. & Wiedmann, M. Precision food safety: A systems
754 approach to food safety facilitated by genomics tools. *Trac-Trends in Analytical Chemistry*
755 **96**, 52-61, doi:10.1016/j.trac.2017.06.001 (2017).

756 79 WHO. Whole genome sequencing as a tool to strengthen foodborne disease surveillance and
757 response: module 1: introductory module. (World Health Organization, 2023).

758 80 Jackson, B. R. *et al.* Implementation of nationwide real-time whole-genome sequencing to
759 enhance listeriosis outbreak detection and investigation. *Clin Infect Dis* **63**, 380-386,
760 doi:10.1093/cid/ciw242 (2016).

761 81 Dallman, T. J. *et al.* Whole-genome sequencing for national surveillance of Shiga toxin-
762 producing *Escherichia coli* O157. *Clin Infect Dis* **61**, 305-312, doi:10.1093/cid/civ318 (2015).

763 82 Grace, D. Food safety in low and middle income countries. *Int J Environ Res Public Health* **12**,
764 10490-10507, doi:10.3390/ijerph120910490 (2015).

765 83 Apruzzese, I. *et al.* Investing in food safety for developing countries: Opportunities and
766 challenges in applying whole-genome sequencing for food safety management. *Foodborne*
767 *Pathog Dis* **16**, 463-473, doi:10.1089/fpd.2018.2599 (2019).

768 84 Hadfield, J. *et al.* Nextstrain: real-time tracking of pathogen evolution. *Bioinformatics* **34**,
769 4121-4123, doi:10.1093/bioinformatics/bty407 (2018).

770 85 Argimon, S. *et al.* Microreact: visualizing and sharing data for genomic epidemiology and
771 phylogeography. *Microb Genom* **2**, e000093, doi:10.1099/mgen.0.000093 (2016).

772 86 Argimon, S. *et al.* A global resource for genomic predictions of antimicrobial resistance and
773 surveillance of *Salmonella* Typhi at pathogenwatch. *Nat Commun* **12**, 2879,
774 doi:10.1038/s41467-021-23091-2 (2021).

775 87 Gangavarapu, K. *et al.* Outbreak.info genomic reports: scalable and dynamic surveillance of
776 SARS-CoV-2 variants and mutations. *Nat Methods* **20**, 512-522, doi:10.1038/s41592-023-
777 01769-3 (2023).

778 88 Reuter, S. *et al.* Directional gene flow and ecological separation in *Yersinia enterocolitica*.
779 *Microb Genom* **1**, e000030, doi:10.1099/mgen.0.000030 (2015).

780 89 Rodrigues, J. A. *et al.* Pangenomic analyses of antibiotic-resistant *Campylobacter jejuni*
781 reveal unique lineage distributions and epidemiological associations. *Microb Genom* **9**,
782 doi:10.1099/mgen.0.001073 (2023).

783 90 Neves, A. *et al.* The Swiss Pathogen Surveillance Platform - towards a nation-wide One
784 Health data exchange platform for bacterial, viral and fungal genomics and associated
785 metadata. *Microb Genom* **9**, doi:10.1099/mgen.0.001001 (2023).

786 91 Sears, A. *et al.* Marked campylobacteriosis decline after interventions aimed at poultry, New
787 Zealand. *Emerg Infect Dis* **17**, 1007-1015, doi:10.3201/eid1706.101272 (2011).

788 92 Gardner, T. J. *et al.* Outbreak of campylobacteriosis associated with consumption of raw
789 peas. *Clin Infect Dis* **53**, 26-32, doi:10.1093/cid/cir249 (2011).

790 93 Cody, A. J., Maiden, M. C., Strachan, N. J. & McCarthy, N. D. A systematic review of source
791 attribution of human campylobacteriosis using multilocus sequence typing. *Euro Surveill* **24**,
792 doi:10.2807/1560-7917.ES.2019.24.43.1800696 (2019).

793 94 Pires, S. M., Vieira, A. R., Hald, T. & Cole, D. Source attribution of human salmonellosis: An
794 overview of methods and estimates. *Foodborne Pathog Dis*, doi:10.1089/fpd.2014.1744
795 (2014).

796 95 Mullner, P. *et al.* Assigning the source of human campylobacteriosis in New Zealand: a
797 comparative genetic and epidemiological approach. *Infect Genet Evol* **9**, 1311-1319,
798 doi:10.1016/j.meegid.2009.09.003 (2009).

799 96 Sheppard, S. K. *et al.* *Campylobacter* genotyping to determine the source of human
800 infection. *Clin Infect Dis* **48**, 1072-1078, doi:10.1086/597402 (2009).

801 97 Mughini-Gras, L. *et al.* Tracing the sources of human salmonellosis: a multi-model
802 comparison of phenotyping and genotyping methods. *Infect Genet Evol* **28**, 251-260,
803 doi:10.1016/j.meegid.2014.10.003 (2014).

804 98 Wilson, D. J. *et al.* Tracing the source of campylobacteriosis. *PLoS Genet* **4**, e1000203,
805 doi:10.1371/journal.pgen.1000203 (2008).

806 99 Liao, S. J., Marshall, J., Hazelton, M. L. & French, N. P. Extending statistical models for source
807 attribution of zoonotic diseases: a study of campylobacteriosis. *J R Soc Interface* **16**,
808 20180534, doi:10.1098/rsif.2018.0534 (2019).

809 100 Arning, N., Sheppard, S. K., Bayliss, S., Clifton, D. A. & Wilson, D. J. Machine learning to
810 predict the source of campylobacteriosis using whole genome data. *PLoS Genet* **17**,
811 e1009436, doi:10.1371/journal.pgen.1009436 (2021).

812 101 Munck, N., Njage, P. M. K., Leekitcharoenphon, P., Litrup, E. & Hald, T. Application of whole-
813 genome sequences and machine learning in source attribution of *Salmonella* Typhimurium.
814 *Risk Anal* **40**, 1693-1705, doi:10.1111/risa.13510 (2020).

815 102 Wainaina, L. *et al.* Source attribution of human campylobacteriosis using whole-genome
816 sequencing data and network analysis. *Pathogens* **11**, doi:10.3390/pathogens11060645
817 (2022).

818 103 Jehanne, Q. *et al.* Genome-wide identification of host-segregating single-nucleotide
819 polymorphisms for source attribution of clinical *Campylobacter coli* isolates. *Appl Environ*
820 *Microbiol* **86**, doi:10.1128/AEM.01787-20 (2020).

821 104 Duarte, A. S. R. *et al.* Metagenomics-based approach to source-attribution of antimicrobial
822 resistance determinants - Identification of reservoir resistome signatures. *Front Microbiol*
823 **11**, 601407, doi:10.3389/fmicb.2020.601407 (2020).

824 105 Pasquali, F., Remondini, D., Snary, E. L., Hald, T. & Guillier, L. Editorial: Integrating whole
825 genome sequencing into source attribution and risk assessment of foodborne bacterial
826 pathogens. *Front Microbiol* **12**, 795098, doi:10.3389/fmicb.2021.795098 (2021).

827 106 Tanui, C. K., Benefo, E. O., Karanth, S. & Pradhan, A. K. A machine learning model for food
828 source attribution of *Listeria monocytogenes*. *Pathogens* **11**,
829 doi:10.3390/pathogens11060691 (2022).

830 107 Liao, J. *et al.* Comparative genomics unveils extensive genomic variation between
831 populations of *Listeria* species in natural and food-associated environments. *ISME Commun*
832 **3**, 85, doi:10.1038/s43705-023-00293-x (2023).

833 108 Beck, K. L. *et al.* Monitoring the microbiome for food safety and quality using deep shotgun
834 sequencing. *NPJ Sci Food* **5**, 3, doi:10.1038/s41538-020-00083-y (2021).

835 109 Mughini-Gras, L. *et al.* Risk factors for human salmonellosis originating from pigs, cattle,
836 broiler chickens and egg laying hens: a combined case-control and source attribution
837 analysis. *PLoS One* **9**, e87933, doi:10.1371/journal.pone.0087933 (2014).

838 110 Lake, R. J. *et al.* Source attributed case-control study of campylobacteriosis in New Zealand.
839 *Int J Infect Dis* **103**, 268-277, doi:10.1016/j.ijid.2020.11.167 (2021).

840 111 Amini, S. in *FoodNavigator Europe* (William Reed, 2018).

841 112 Gerner-Smidt, P. in *Food Safety Magazine* (2021).
842 113 GFSI. A culture of food safety: A position paper from the Global Food Safety Initiative (GFSI).
843 (2018).
844 114 Espinoza, M. S. A., Flink, C., Boisen, N., Scheutz, F. & Käsbohrer, A. Microbiological sampling
845 and analyses in the food business operators' HACCP-based self-control programmes.
846 *Frontiers in Food Science and Technology* **3**, doi:10.3389/frfst.2023.1110359 (2023).
847 115 CFA. Principles of an environmental monitoring program for the management of *Listeria*
848 *monocytogenes*. (2023).
849 116 Jagadeesan, B. *et al.* The use of next generation sequencing for improving food safety:
850 Translation into practice. *Food Microbiol* **79**, 96-115, doi:10.1016/j.fm.2018.11.005 (2019).
851 117 Tran, M. *et al.* Economic evaluations of whole-genome sequencing for pathogen
852 identification in public health surveillance and health-care-associated infections: a
853 systematic review. *Lancet Microbe* **4**, e953-e962, doi:10.1016/S2666-5247(23)00180-5
854 (2023).
855 118 Jain, S., Mukhopadhyay, K. & Thomassin, P. J. An economic analysis of *Salmonella* detection
856 in fresh produce, poultry, and eggs using whole genome sequencing technology in Canada.
857 *Food Research International* **116**, 802-809, doi:10.1016/j.foodres.2018.09.014 (2019).
858 119 WHO. *Food safety*, <<https://www.who.int/news-room/fact-sheets/detail/food-safety>>
859 (2022).
860 120 Ogden, N. H., AbdelMalik, P. & Pulliam, J. Emerging infectious diseases: prediction and
861 detection. *Can Commun Dis Rep* **43**, 206-211, doi:10.14745/ccdr.v43i10a03 (2017).
862 121 Plowright, R. K. *et al.* Pathways to zoonotic spillover. *Nat Rev Microbiol* **15**, 502-510,
863 doi:10.1038/nrmicro.2017.45 (2017).
864 122 Falloon, P. *et al.* What do changing weather and climate shocks and stresses mean for the
865 UK food system? *Environ Res Lett* **17**, 051001, doi:10.1088/1748-9326/ac68f9 (2022).
866 123 Elliott, C. in *New Food* (Russell Publishing Ltd, Brasted, Kent, UK, 2022).
867 124 McLauchlin, J. *et al.* An outbreak of human listeriosis associated with frozen sweet corn
868 consumption: Investigations in the UK. *Int J Food Microbiol* **338**, 108994,
869 doi:10.1016/j.ijfoodmicro.2020.108994 (2021).
870 125 Kindle, P., Nuesch-Inderbinen, M., Cernela, N. & Stephan, R. Detection, isolation, and
871 characterization of Shiga toxin-producing *Escherichia coli* in flour. *J Food Prot* **82**, 164-167,
872 doi:10.4315/0362-028X.JFP-18-256 (2019).
873 126 Bloomfield, S. J. *et al.* Genomic analysis of *Salmonella enterica* serovar Typhimurium DT160
874 associated with a 14-year outbreak, New Zealand, 1998-2012. *Emerg Infect Dis* **23**, 906-913,
875 doi:10.3201/eid2306.161934 (2017).
876 127 Pijnacker, R. *et al.* An international outbreak of *Salmonella enterica* serotype Enteritidis
877 linked to eggs from Poland: a microbiological and epidemiological study. *Lancet Infect Dis* **19**,
878 778-786, doi:10.1016/S1473-3099(19)30047-7 (2019).
879 128 CDC. *A-Z index for foodborne illness*,
880 <<https://www.cdc.gov/foodsafety/diseases/index.html>> (2021).
881 129 EFSA & ECDC. The European Union One Health 2021 Zoonoses Report. *EFSA J* **20**, e07666,
882 doi:10.2903/j.efsa.2022.7666 (2022).
883 130 Wagenaar, J. A., French, N. P. & Havelaar, A. H. Preventing *Campylobacter* at the source:
884 why is it so difficult? *Clin Infect Dis* **57**, 1600-1606, doi:10.1093/cid/cit555 (2013).
885 131 CDC. *Detecting outbreaks with whole genome sequencing*, <[https://www.cdc.gov/amd/how-](https://www.cdc.gov/amd/how-it-works/detecting-outbreaks-wgs.html)
886 [it-works/detecting-outbreaks-wgs.html](https://www.cdc.gov/amd/how-it-works/detecting-outbreaks-wgs.html)> (2019).

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891 **Acknowledgements**

892 A.E.M. and M.W.G. are supported by the Biotechnology and Biological Sciences Research
893 Council (BBSRC) Institute Strategic Programme Microbes and Food Safety BB/X011011/1
894 and its constituent project BBS/E/F/000PR13634 (Theme 1, Microbial threats from foods in
895 established and evolving food systems). This work was also supported in part by BBSRC
896 grants BB/V018221/1 and BB/X002985/1 (UK Food Safety Research Network). The funders
897 had no role in study design, data collection and analysis, decision to publish or preparation
898 of the manuscript.

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900 **Author contributions**

901 The authors contributed equally to all aspects of the article.

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903 **Competing interests**

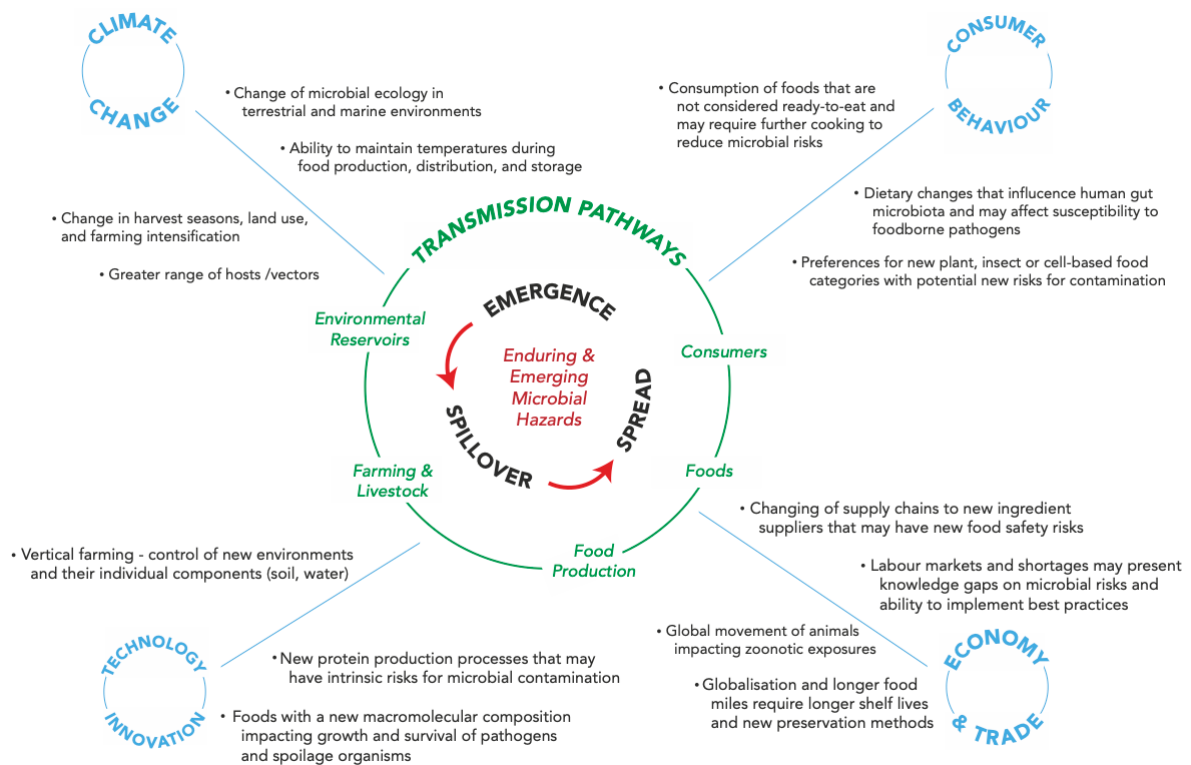
904 International Patent Application No. PCT/GB2023/050906 entitled “Determination and
905 quantification of the microbial communities and antimicrobial resistance genes on food” in
906 the name of Quadram Institute Bioscience has been filed (priority date 05/04/2022) and is
907 currently in the international phase; A.E.M. is an inventor. This relates to the aspect of the
908 Review where it is mentioned that the potential detection of pathogens through
909 metagenomics can be affected by the amount of contaminating host DNA and sequencing
910 depth. N.F. is a member of the International Commission on Microbiological Specifications
911 for Foods (ICMSF) and an Emeritus Director of the New Zealand Food Safety Science and
912 Research Centre. Both organisations receive support from the food industry and
913 government agencies. Both roles are unpaid and advisory, and related to food safety
914 research, and neither organisation influenced the content contributed by N.F. to the
915 Review. The other authors declare no competing interests.

916

917 **Peer review information**

918 *Nature Reviews Microbiology* thanks Steven Djordjevic, who co-reviewed with Veronica
919 Jarocki; Jasna Kovac; and the other, anonymous, reviewer(s) for their contribution to the
920 peer

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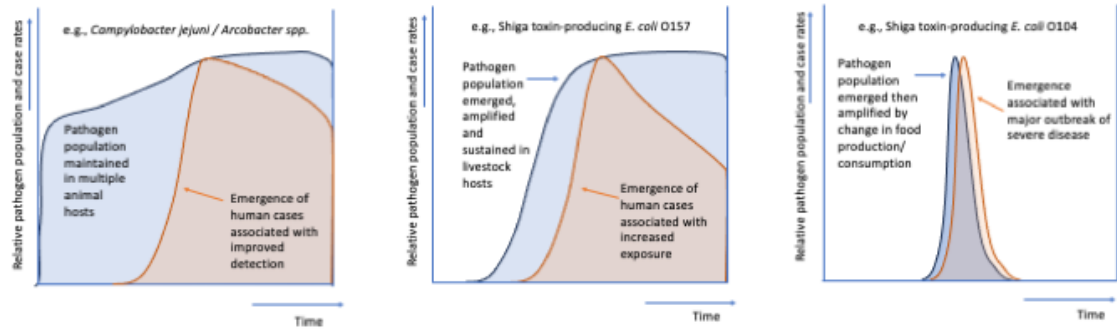
924 **Figure 1. Ecological and socioeconomic drivers that may shape the microbial landscape of food and**
 925 **food production systems.**

926 The risk of human exposure to foodborne pathogens and spoilage microorganisms can be amplified
 927 due to varied drivers that act across environmental, agricultural, agrifood and consumer niches.
 928 These drivers impact and facilitate the emergence and transmission of newly evolved and existing
 929 microorganisms within these niches. Drivers include changes to global and local ecologies,
 930 innovation in food production technologies, pressures imposed by evolving supply chains and
 931 agrifood economies, and the impacts of dietary and consumer choices on food availability,
 932 preparation and consumption. Terminology and drivers adapted from refs.¹²⁰⁻¹²³ and examples are
 933 cited in the main Review; additionally, examples of potential food risks arising from consumption of
 934 foods not considered ready-to-eat include frozen vegetables (with a risk of *Listeria*
 935 *monocytogenes*)¹²⁴ and flour (with a risk of Shiga-toxin producing *Escherichia coli*)¹²⁵.

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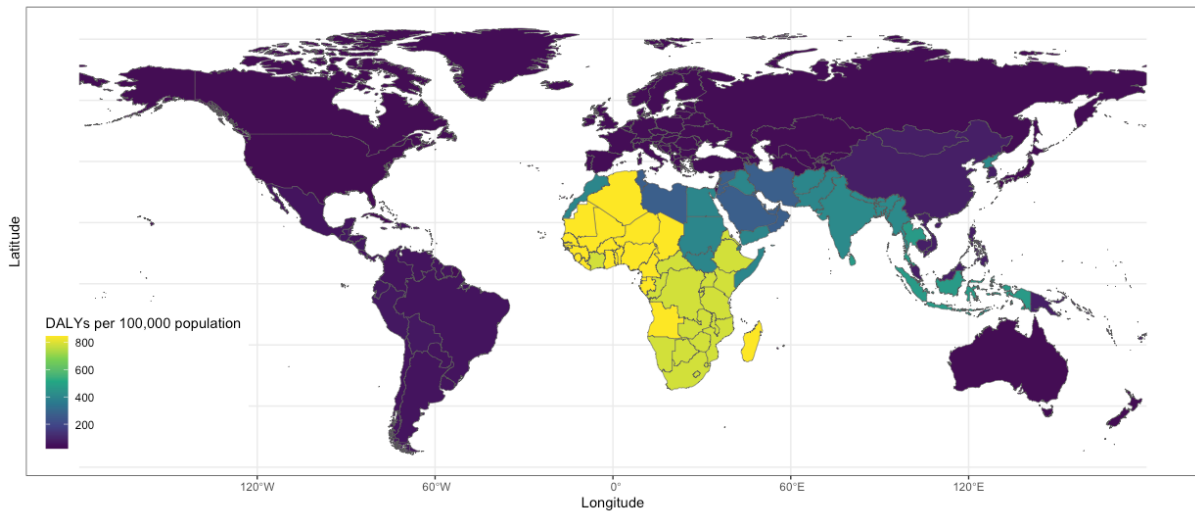


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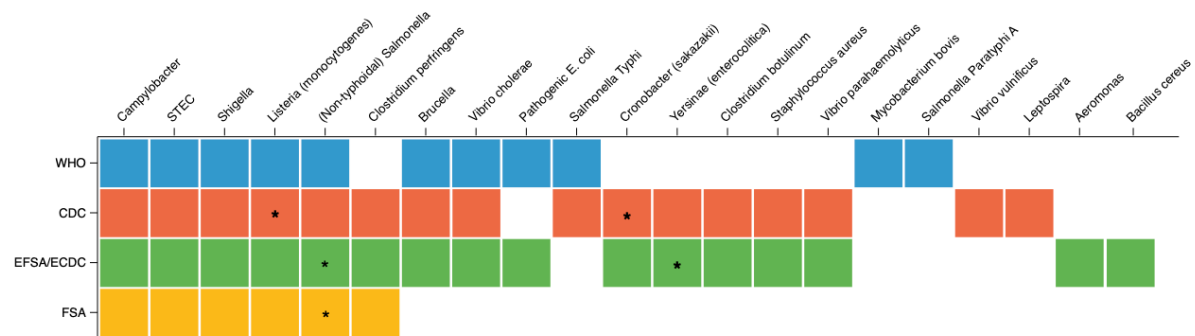
Figure 2. Relationship between pathogen populations and the dynamics of foodborne illness.

Schematics illustrating putative relationships between pathogen populations (blue) and the dynamics of foodborne illness (red). Emergence may be associated with increased exposure and/or improved detection. Examples of pathogens are indicated in each panel. a) The pathogen population emerged historically and had been amplified as a result of host-species crossover and changes in host dynamics (for example, host population increases and changes in farming practices). The pathogen caused historical human cases of foodborne illness, but these were undetected until improvements in diagnostics led to the apparent emergence. Improved detection resulted in reduced transmission to humans, and potentially reduced transmission in animals. b) Pathogen evolution and changes in host dynamics (for example, movement of farm animals associated with global trade and changes in production practices) resulted in increased exposure to the pathogen via the food chain, which may have coincided with improved detection. Subsequent interventions have reduced human exposure and lowered amplification in animals. c) A pathogen evolved to acquire increased virulence and was amplified following contamination of a food production system. Identification of the source results in interventions that lead to a rapid decline in human cases and the pathogen population. Relative pathogen populations can be inferred from effective population sizes estimated using ancestral state reconstruction methods^{126 62 127}. Foodborne illness dynamics can be derived from reported human cases⁶².

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980 b)

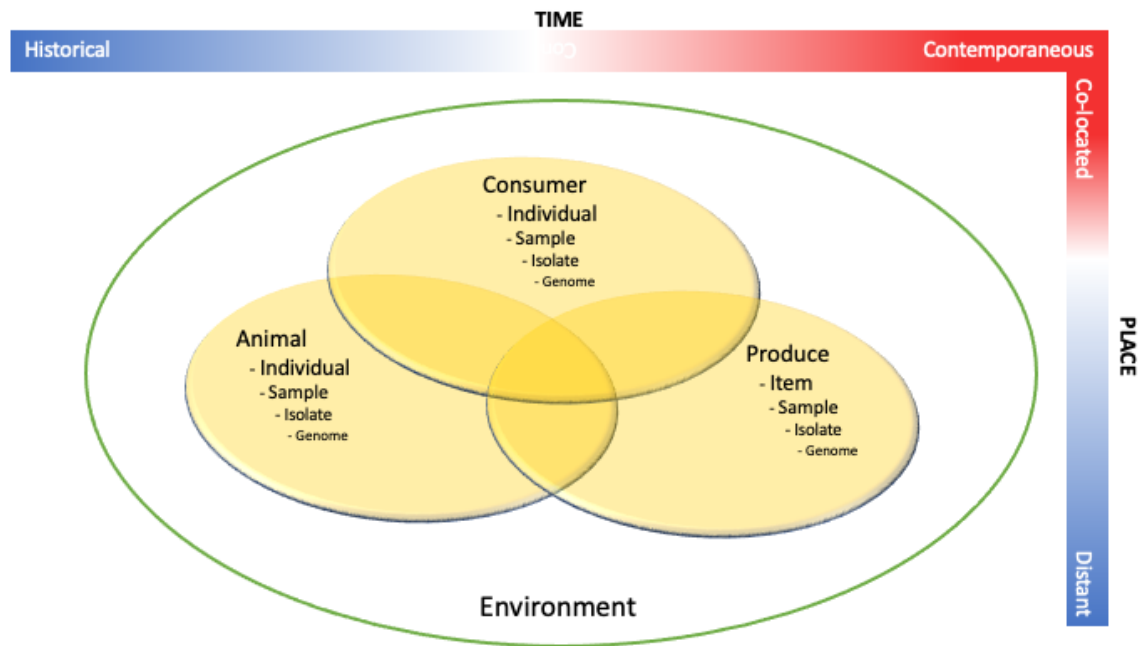


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982 **Figure 3. International impacts and prioritisation of bacterial foodborne illness.**

983 a) World map showing median rates of foodborne Disability Adjusted Life Years (DALYs) per 100,000
 984 population, caused by bacterial pathogens considered by the World Health Organization Foodborne
 985 Disease Burden Epidemiology Reference Group: *Campylobacter* spp., enteropathogenic *Escherichia*
 986 *coli*, enterotoxigenic *E. coli*, Shiga toxin-producing *E. coli* (STEC), non-typhoidal *Salmonella enterica*,
 987 *Shigella* spp., *Vibrio cholerae*, *Brucella* spp., *Listeria monocytogenes*, *Mycobacterium bovis*,
 988 *Salmonella* Paratyphi A, *Salmonella* Typhi. Data represent the estimated total DALYs from these
 989 pathogens in 2010; data obtained from Table 8 in ref.¹⁸. b) Enduring bacterial foodborne hazards
 990 that are widely recognised by public health and food agencies, including the World Health
 991 Organization (WHO)¹⁸, UK Food Standards Agency (FSA)⁴⁹, US Centers for Disease Control (CDC)¹²⁸
 992 and the European Food Safety Authority (EFSA) and European Centre for Disease Prevention and
 993 Control (ECDC)¹²⁹. For the CDC and WHO lists of bacterial foodborne hazards, non-Shiga toxin-
 994 producing *Escherichia coli* (non-STEC) listed were re-categorised as ‘Pathogenic *E. coli*’ to match the
 995 nomenclature of the European Food Safety Authority (EFSA) and the European Centre for Disease
 996 Prevention and Control (ECDC) list. *, indicates when the agency recognises the microorganism to
 997 the genus level.

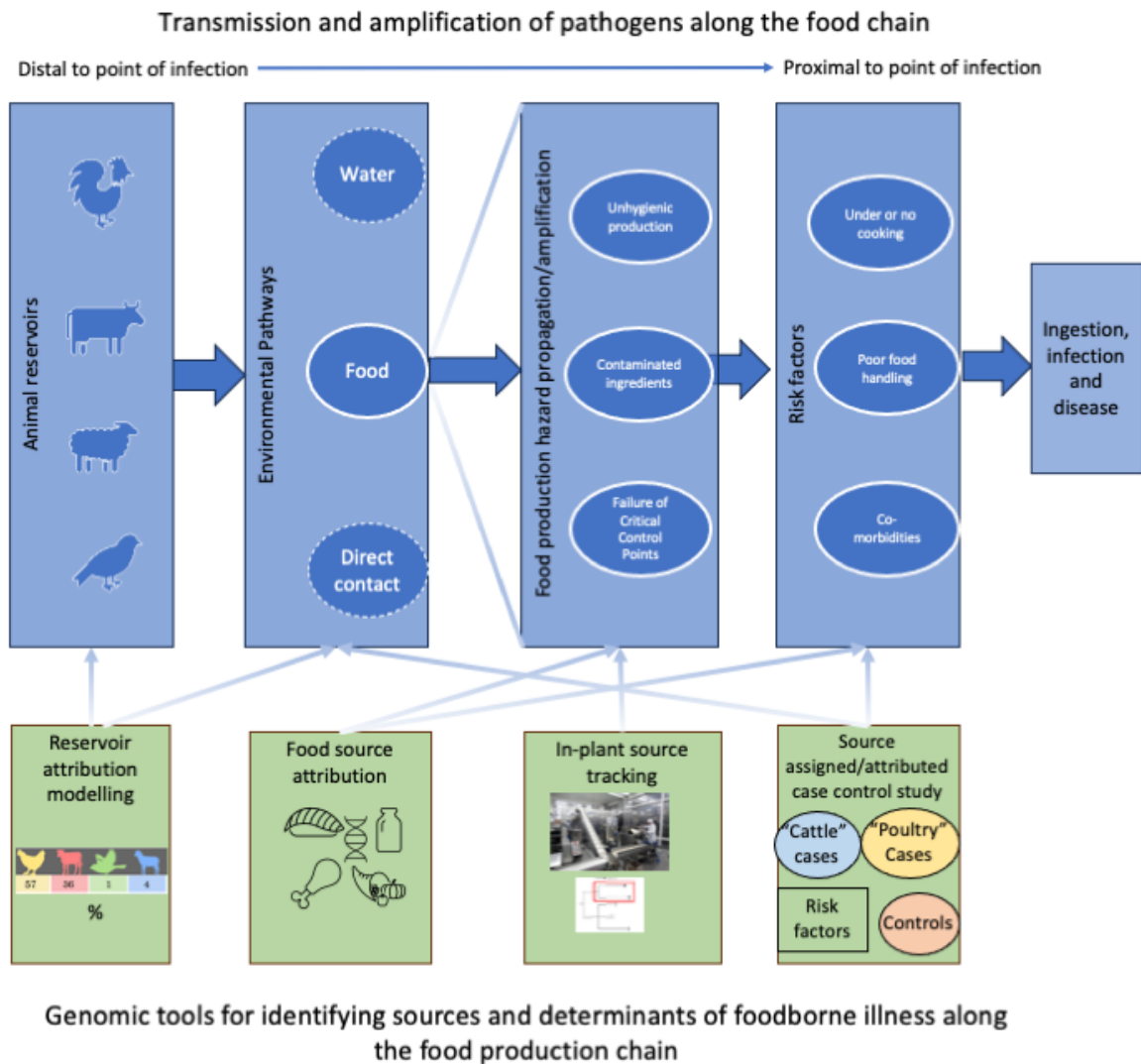
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Figure 4. One Health approach to ecological frameworks and epidemiology in food safety

A One Health approach to microbiological safety of food requires an ecological framework that embraces the interactions of consumers, animals and produce in the environment in which they interact. The resolution of sampling frame and sample size calculations — the number of samples taken at population level, the number of samples taken per subject, the number of isolates per sample phenotyped and the number of genomes sequenced — are necessary considerations for surveillance design, outbreak investigation and quantification of risk. Notwithstanding the advanced technologies now available, association and causality must still be considered within classical epidemiological parameters of spatial relatedness and temporal directionality; contemporaneous, and co-located data remain the cornerstones of causal inference and plausible source attribution.

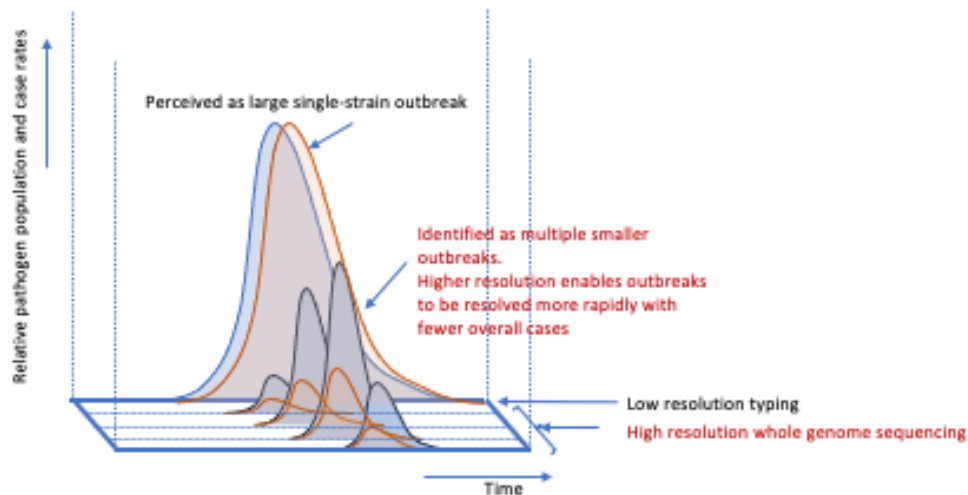


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Figure 5. Tracing bacterial hazards in the food chain.

1014 Many bacterial hazards that enter the food chain are amplified and maintained in animal reservoirs,
 1015 including domestic farmed animals and wildlife. Illustrated are the critical steps along the food chain
 1016 that propagate foodborne pathogens from the animal reservoir to the point of human ingestion,
 1017 infection and disease. For many enduring pathogens, such as *Campylobacter jejuni* and *Salmonella*
 1018 *enterica*, there may be multiple animal reservoirs that can contaminate food and other transmission
 1019 pathways (for example, water and direct faecal contact as indicated in boxes with dashed lines).
 1020 Molecular tools, including whole genome sequencing and metagenomics, combined with
 1021 evolutionary models, are used to determine critical events along the food chain (blue boxes). This
 1022 includes reservoir attribution models, food processing facility source tracking and source attributed
 1023 case-control studies (red boxes). Although these methods are focused on understanding the
 1024 epidemiology and control of enduring pathogens, often defined at the species-level, genomics tools
 1025 are highly valuable for the investigation of outbreaks of emerging pathogens^{11,66}. If the emerging
 1026 pathogen persists and becomes a sustained, multijurisdictional epidemic or pandemic, large-scale
 1027 sequencing will facilitate more comprehensive food-chain and epidemiological investigations.
 1028 Derived from concepts explored in ref.¹³⁰.

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1032 **Figure 6. The impact of high-resolution whole genome sequencing on the detection and resolution**
1033 **of foodborne outbreaks.**

1034 Higher resolution WGS allows more outbreaks to be identified and resolved more rapidly, with fewer
1035 overall cases. Pathogen population sizes are shown in blue, and the number of cases of foodborne
1036 illness are in red. With low-resolution typing, a single large outbreak is detected (back panel), which
1037 may correspond to a seasonal increase in cases or a prolonged, multi-year outbreak. This type of
1038 low-resolution typing tends to group epidemiologically-unlinked cases together, resulting in an
1039 inaccurate understanding of the underlying epidemiology, and less effective interventions. By
1040 contrast, high resolution genome sequence-based typing enables better assignment of cases to
1041 discrete smaller outbreaks with a common epidemiological origin (front four panels). The improved
1042 assignment enables outbreaks to be resolved more rapidly, leading to fewer cases associated with
1043 each outbreak, and a reduction in the overall pathogen population. This example is motivated by the
1044 US Centers for Disease Control description of the analysis of outbreaks using Advanced Molecular
1045 Detection, comparing low resolution methods with WGS¹³¹.

1046
1047 **Box 1**

1048 **Questions that genomics now allows food businesses to ask:**

- 1049 • Is there a predominant pathogen or strain in a facility? Is this strain spilling over to
1050 their food products?
- 1051 • Are there other microorganisms supporting the pathogens? Do these likewise find
1052 their way onto their foods?
- 1053 • How are the pathogens surviving disinfection measures? Are the 'supporting'
1054 microorganisms likewise being selected for with these measures?
- 1055 • Can the possible spread of microorganisms be tracked within individual facilities (and
1056 potentially their foods)?
- 1057 • Can strains of pathogens be identified that might be of a higher risk than other
1058 strains (for example, genetic markers to inform risk, recognising that not all strains of
1059 a particular pathogen are equally hazardous)?
- 1060 • Can the baseline microbial composition of a food process, setting, or ingredient be
1061 determined using metagenomics?
- 1062 • Can the impact of interventions imposed in food businesses be monitored by
1063 determining if there is a change in the distribution or composition of microbiota?⁷⁹

1064

1065 **Actions that can now be considered with the depth of knowledge learned from genomics:**

- 1066 • Modification of chemical disinfection plans based on the observation of susceptible
1067 or tolerant microorganisms
- 1068 • Reconsideration of hygiene best practices related to how equipment and ingredients
1069 are handled, placed, and/or move throughout a facility based on evidence of
1070 microorganisms residing or spreading in a facility (that is, facility-specific pathways)
- 1071 • Redesign or use of innovative materials in food non-contact and contact surfaces
1072 that may mitigate the persistence or transmission of key microorganisms in these at-
1073 risk pathways
- 1074 • Consideration of novel environmental or food biocontrol strategies (for example, the
1075 application of bacteriophages, or the introduction of other microorganisms to
1076 change the microbial community structure).
- 1077 • Implementation of non-chemical interventions at identified critical control points
1078 (for example, ultraviolet light or sonication)
- 1079 • Use of metagenomic information as the guide point to recover key taxa from related
1080 samples for further characterisation and study
- 1081 • Informed and proportional action to urgent levels of risk (for example, observation
1082 of strain with characteristics of high concern)

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