



Multigenerational downregulation of insulin/IGF-1 signaling in adulthood improves lineage survival, reproduction, and fitness in *Caenorhabditis elegans* supporting the developmental theory of ageing

Elizabeth M. L. Duxbury,^{1,2}  Hanne Carlsson,¹  Kris Sales,¹  Zahida Sultanova,¹  Simone Immler,¹ 
 Tracey Chapman,¹  and Alexei A. Maklakov¹ 

¹*School of Biological Sciences, University of East Anglia, Norwich NR4 7TJ, United Kingdom*

²*E-mail: E.Duxbury@uea.ac.uk*

Received March 21, 2022

Accepted September 8, 2022

Adulthood-only downregulation of insulin/IGF-1 signaling (IIS), an evolutionarily conserved pathway regulating resource allocation between somatic maintenance and reproduction, increases life span without fecundity cost in the nematode, *Caenorhabditis elegans*. However, long-term multigenerational effects of reduced IIS remain unexplored and are proposed to carry costs for offspring quality. To test this hypothesis, we ran a mutation accumulation (MA) experiment and downregulated IIS in half of the 400 MA lines by silencing *daf-2* gene expression using RNA interference (RNAi) across 40 generations. Contrary to the prediction, adulthood-only *daf-2* RNAi reduced extinction of MA lines both under UV-induced and spontaneous MA. Fitness of the surviving UV-induced MA lines was higher under *daf-2* RNAi. Reduced IIS increased intergenerational F1 offspring fitness under UV stress but had no quantifiable transgenerational effects. Functional *hrde-1* was required for the benefits of multigenerational *daf-2* RNAi. Overall, we found net benefit to fitness from multigenerational reduction of IIS and the benefits became more apparent under stress. Because reduced *daf-2* expression during development carries fitness costs, we suggest that our findings are best explained by the developmental theory of ageing, which maintains that the decline in the force of selection with age results in poorly regulated gene expression in adulthood.

KEY WORDS: Ageing, germline, life-history evolution, mutation accumulation, senescence.

Ageing, the physiological deterioration of an organism leading to increased probability of death and decreased reproduction with advancing adult age, is taxonomically ubiquitous but remains incompletely understood (Flatt and Partridge 2018; Maklakov and Chapman 2019; Regan et al. 2019; Gaillard and Lemaître 2020). Although there is broad agreement that ageing evolves because natural selection gradients on traits decline after reproductive maturity (Medawar 1952; Williams 1957; Hamilton 1966; Caswell and Shyu 2017; Day and Abrams 2020), the proximate

causes of late-life deterioration are less clear (Gems and Partridge 2013; Flatt and Partridge 2018; Maklakov and Chapman 2019; Flatt 2020). The “disposable soma” theory of ageing suggests that ageing evolves due to competitive resource allocation between the germline and the soma (Kirkwood 1977; Kirkwood and Holliday 1979; Kirkwood 1987). It follows that increased investment in somatic maintenance leading to longer life span trades off with traits associated with reproduction, such as the number or size of progeny (Stearns 1992; Roff 2002). Despite considerable

support for trade-offs between somatic maintenance and reproduction, growing empirical work questions the universality of such resource allocation trade-offs in the evolution of ageing (Edward and Chapman 2012; Maklakov and Immler 2016; Maklakov and Chapman 2019; Flatt 2020). Several studies have shown that experimentally increased life span, often via the downregulation of genes in nutrient-sensing signaling pathways in adulthood, is not detrimental to reproduction (Dillin et al. 2002; Flatt 2011; Maklakov and Immler 2016; Lind et al. 2019; Maklakov and Chapman 2019). These results are in line with the hypothesis that a gradual decline in selection gradients with advancing age after reproductive maturity results in physiological processes that are optimized for successful early-life reproduction, but start causing harm in later life, contributing to somatic deterioration (de Magalhães and Church 2005; de Magalhães 2012; Blagosklonny 2013; Ezcurra et al. 2018; Wang et al. 2018; Maklakov and Chapman 2019; Sala et al. 2020; Carlsson et al. 2021).

However, it has been proposed that reduced offspring quality, for example, via reduced germline maintenance, can be a hidden cost of life span extension via reduced nutrient-sensing signaling (Maklakov and Immler 2016). Germline maintenance, the repair and surveillance of genomic and proteomic integrity in germline stem cells and gametes, is energetically expensive (Maklakov and Immler 2016; Berger et al. 2017) and germline signaling plays a key role in resource allocation to somatic maintenance (Kenyon 2010; Labbadia and Morimoto 2015; Maklakov and Immler 2016; Berger et al. 2017). Germline ablation results in increased somatic maintenance and life span in *Drosophila melanogaster* fruitflies (Flatt et al. 2008) and *Caenorhabditis elegans* nematodes (Hsin and Kenyon 1999; Arantes-Oliveira et al. 2002; Antebi 2013; Ermolaeva et al. 2013; Shemesh et al. 2013), although life span extension in *C. elegans* requires an intact somatic gonad (Hsin and Kenyon 1999). It is important to note that the requirement of the intact somatic gonad for life span extension in germline-less worms does not negate the possibility of a resource allocation trade-off as sometimes implied, but only shows that germline signaling is required to mediate the effect (Arantes-Oliveira et al. 2002; Flatt et al. 2008; Maklakov and Immler 2016). Similarly, recent work in *Danio rerio* zebrafish suggests that germline ablation increases somatic maintenance under stress (Chen et al. 2020). Furthermore, nutritional stress in *C. elegans* results in germline reduction and increased life span (Thondamal et al. 2014). When soma-to-germline communication is disrupted, the number of germ cells is unaffected and life span extension is abolished (Thondamal et al. 2014). Taken together, these results suggest that germline maintenance is costly and can trade-off with life-history traits such as life span and offspring number (Maklakov and Immler 2016; Berger et al. 2017). The corollary is that increased investment in somatic maintenance and longevity can trade-off

with offspring quality by increasing germline mutation load in offspring.

We tested this hypothesis by focusing on life span extension via reduced insulin/IGF-1 signaling (IIS) in *C. elegans*. IIS is an evolutionarily conserved pathway that regulates the physiological response of organisms to their environment (Regan et al. 2019). IIS mechanistically links nutrient intake with development, growth, reproduction, and life span across diverse taxa (Fontana et al. 2010; Kenyon 2010; Regan et al. 2019). Reduced IIS, via genetic and environmental interventions, consistently extends life span (Kenyon 2010). Mutations that reduce the function of *daf-2* gene in *C. elegans*, the homolog of human insulin/IGF-1 receptor, increase life span and reduce fitness (Chen et al. 2007). However, previous work showed that reducing IIS in adulthood via *daf-2* RNA interference (RNAi) knockdown in a single generation increases life span without a cost to offspring number (Dillin et al. 2002), quality (Lind et al. 2019), or fitness in variable environments (Carlsson et al. 2021). It is possible, however, that multigenerational *daf-2* knockdown will expose a hidden cost to offspring number and/or offspring quality via accumulation of germline mutations. Here, we tested this prediction via adulthood-only *daf-2* RNAi knockdown for 40 consecutive generations in 400 spontaneous and UV-induced mutation accumulation (MA) lines. We found that multigenerational adulthood-only *daf-2* knockdown reduced lineage extinction under both spontaneous and UV-induced MA and increased fitness of remaining lineages under UV-induced MA. Reduced IIS protected MA lines from infertility, egg hatching failure, and developmental abnormalities and increased offspring fitness under UV irradiation, but we found no evidence of transgenerational effects of adulthood-only *daf-2* RNAi.

Materials and Methods

Supplementary methods details are included in the Supporting Information.

NEMATODE STOCKS AND CULTURE

The nematode (roundworm) *C. elegans* N2 wild-type (Bristol) and heritable RNAi deficiency 1 (*hrde-1*) mutant strains were defrosted from stocks acquired from Caenorhabditis Genetics Center (University of Minnesota, USA, funded by NIH Office of Research Infrastructure Programs, P40 OD010440) and from the lab of Prof. Eric Miska (Gurdon Institute, University of Cambridge, UK), respectively, and stored at -80°C until use.

Defrosted *C. elegans* strains were reared through two generations prior to setup, on NGM agar supplemented with the fungicide nystatin and antibiotics streptomycin and ampicillin, to prevent infection (each at 100 $\mu\text{g}/\text{mL}$, as standard recipes, e.g.,

Lionaki and Tavernarakis 2013) and seeded with the antibiotic-resistant *Escherichia coli* bacterial strain OP50-1 (pUC4K, from J. Ewbank at the Centre d'Immunologie de Marseille-Luminy, France). We bleached eggs from the grandparents of experimental individuals, to standardize parental age and remove any infection or temperature effects from defrosting, prior to experiments.

REDUCING IIS VIA *daf-2* RNAi FEEDING IN ADULTHOOD

To downregulate adulthood expression of the insulin-like signaling receptor homolog, *daf-2*, we fed late-L4 larvae with *E. coli* bacteria expressing *daf-2* double-stranded RNA, which decreases mRNA levels of the complementary transcribed *daf-2* systemically (Fire et al. 1998; Timmons et al. 2001). The *daf-2* gene is upstream of and inhibits the action of master regulator *daf-16/FOXO*. RNase-III deficient, IPTG-inducible HT115 *E. coli* bacteria with an empty plasmid vector (L4440) was used as the control (as Timmons et al. 2001; Dillin et al. 2002; Lind et al. 2019). RNAi clones were acquired from the Vidal feeding library (Source BioScience, created by M. Vidal lab, Harvard Medical School, USA) and all clones were verified via sequencing prior to delivery.

ULTRAVIOLET WAVELENGTH C IRRADIATION

To induce mutations and stress, we UV-irradiated Day 2 adults, with a calibrated ultraviolet-C (UV-C) radiation dose of 46 J/m² (wavelength 254nm), via 20 s exposure to the UV-C radiation emitted from the lamp of a Thermo Scientific Heraguard ECO Safety Cabinet (calibration details in Supplementary Methods). This dose is in the range of previous UV-C irradiation doses for *C. elegans* adults or eggs (Meyer et al. 2007; Stergiou et al. 2007; Hyun et al. 2008; Boyd et al. 2010). Our pilot work showed that this dose reduced the fecundity of Day 2 adults laying at 20°C by 61% compared with unexposed sham controls ($n = 30$ worms/treatment, mean Day 2 offspring count = 35 for UV, 90 for non-UV; full data not shown). Nonirradiated control worms received a sham-irradiation, by being positioned in identical orientation under the UV-C lamp for 20 s, while it was switched off.

MUTATION ACCUMULATION LINES

To determine the effects of reduced adulthood IIS, via *daf-2* RNAi, on spontaneous and UV-induced MA, we established 800 MA lines, across two genetic backgrounds. The eight experimental treatments were the full-factorial combinations of genotype (N2 or *hrde-1* mutant), RNAi treatment (empty vector or *daf-2* RNAi), and irradiation (UV or sham), with 100 MA lines per treatment. We ran each of the eight treatments in parallel, divided into two time-staggered independent blocks of 50 lines per treatment, for logistical reasons and to capture any between-

block variation. We tracked MA lines until the majority of UV-irradiated lines were extinct—40 generations for N2 and 25 generations for *hrde-1* mutants. Every MA generation, *daf-2* RNAi or empty vector was delivered from the start of adulthood onward, and all lines developed on empty vector. More details are provided in the Supporting Information.

Each MA line was propagated as a single individual hermaphrodite per generation, to create successive genetic bottlenecks, allowing de novo deleterious mutations to accumulate in the relative absence of selection (Keightley and Bataillon 2000; Vassilieva et al. 2000; Ajie et al. 2005), if they had no effect on developmental time or viability, and in the absence of mating. MA is a common approach used in several species, to study the evolutionary genetics of de novo mutation rates (Halligan and Keightley 2009). In wild-type *C. elegans*, the spontaneous mutation rate in MA lines is positively correlated with the rate of fitness decay (Baer et al. 2005).

VALIDATION OF *daf-2* GENE EXPRESSION UNDER RNAi

We quantified, via quantitative reverse transcriptase polymerase chain reaction (qRT-PCR), the extent of downregulation of *daf-2* expression in Day 2 adults from the MA lines at generation 20, following adulthood-only *daf-2* RNAi knockdown, relative to generation 20 empty vector control MA lines. A sample from all eight MA treatments above that had been frozen at generation 20 were used (mean $n = 6$ MA lines per treatment, with mean $n = 3$ individual worms per MA line; Table S1). Previous work has shown that adulthood *daf-2* RNAi reduces *daf-2* expression in Day 2 N2 adults by 50%, relative to controls (Carlsson et al. 2021), equivalent to the first generation of MA. The aim was to test the degree of *daf-2* knockdown after 20 generations of *daf-2* RNAi and MA in both N2 and *hrde-1* mutant backgrounds. Worms for the gene expression assay were maintained under standard conditions (as the main experiment) and were reared for two generations on empty vector prior to the assay, to remove freezing effects and age-synchronize. RNAi was applied from late-L4 stage. Further method details are included in the Supporting Information.

EFFECT OF 20 GENERATIONS OF SPONTANEOUS AND UV-INDUCED MA ON FITNESS AND LIFE HISTORY OF SURVIVING MA LINES

To test for life-history differences between the N2 wild-type MA lines after 20 generations of MA and of adulthood *daf-2* RNAi, we assayed age-specific reproduction, egg size, male production, and adult heat shock resistance ($n = 100$ grand-offspring from each of the four N2 MA treatments). Further details are provided in the Supporting Information.

Individual age-synchronized, late-L4 hermaphrodites were set up on separate empty vector plates, labeled with a unique identifier, to blind experimenters to treatment identity, and thus avoid bias. The two experimenters conducted the life-history assay in two simultaneous time blocks and each block had an identical and equal representation of individual worms from all four treatments ($n = 50$ per treatment per block).

For all experiments, we assayed age-specific offspring production (fecundity) over the first 6 days of adult life (the reproductive window for *C. elegans* hermaphrodites), by transferring to fresh plates every 24 h, to acquire daily reproduction counts, following the standard temporal resolution for *C. elegans* studies (Dillin et al. 2002; Lind et al. 2019; Scharf et al. 2021). Plates were scored for viable adult offspring 2.5 days later. Individual fitness (λ) was calculated—a measure analogous to the intrinsic rate of population growth, which is weighted by early reproduction and fast development (Stearns 1992; Brommer et al. 2002; Lind et al. 2016; Lind et al. 2019). Total lifetime reproduction (lifetime reproductive success) was calculated for each individual as the sum of age-specific offspring counts across the first 6 days of adulthood.

To determine if reduced IIS influenced parental resource allocation into eggs under MA, we measured egg area (in mm^2), with 75 samples per treatment. Eggs measured were laid by the individuals in the generation 20 age-specific reproduction assay at peak reproduction (Day 2). Egg size is commonly used as a proxy for parental investment (Hibshman et al. 2016; Lind et al. 2019). We photographed one egg laid per individual, within 2 h of lay, under $12\times$ magnification light microscopy (Leica M165C with Lumenera Infinity 2–7C digital microscope camera).

TRANSGENERATIONAL EFFECTS OF *daf-2* RNAi ON OFFSPRING FITNESS IN N2 AND *hrde-1* MUTANT BACKGROUNDS

To determine whether *daf-2* RNAi was transferred transgenerationally via the germline, we assayed, in a separate experiment, whether offspring fitness benefits persisted across three generations of offspring (F1, F2, and F3) from Day 2 parents treated with *daf-2* RNAi or empty vector control throughout adulthood, in N2 wild-type and *hrde-1* mutant backgrounds. Offspring of each generation were maintained on empty vector ($n = 30$ individuals per treatment and per genotype) and assayed for daily reproduction.

EFFECTS OF REDUCED IIS ON PARENTAL FITNESS AND TOTAL REPRODUCTION UNDER INTERMITTENT FASTING

To determine whether there was a reproductive cost of adulthood-only *daf-2* RNAi under resource limitation, we exposed N2 wild-

type adults to either an intermittent fasting (IF) regime consisting of 9 h of starvation on Days 1, 3, 5, and 7 of adulthood, or ad libitum (AL) feeding throughout life, and assayed daily reproduction. Full details are in the Supporting Information.

STATISTICAL ANALYSES

All analyses were conducted in R version 4.0.0 (R Core Team 2020). Fitness and total lifetime reproduction data were plotted using the R package “dabestr” (data analysis using bootstrap-coupled estimation [Ho et al. 2019]), for visualization.

Parental survival in the intergenerational (first) experiment was analyzed using a Cox proportional hazards mixed-effects model (Therneau 2019) (“coxme”), fitting RNAi, UV, and their interaction as fixed effects and plate as a random effect. Separate analyses were conducted with matricides either classed as deaths or as censors. The age at death response variable contained a coding variable to distinguish deaths from censored individuals (due to accidental losses). Extinction (survival trajectories) under MA was analyzed using Cox proportional hazards regression analysis (“coxph” function in “survival” package). The generation number at which each MA line went extinct was the response variable (instead of age at death) and this contained a coding variable to distinguish extinctions from censors, as above. To test the effect of experimental block or genotype (either N2 wildtype or *hrde-1* mutant) on extinction and on the interaction between RNAi and UV effects under MA, maximal models were fitted with either a three-way RNAi \times UV \times block interaction or a RNAi \times UV \times genotype interaction, and stepwise model simplification was conducted (Bolker et al. 2009). Block was included as a fixed effect to test for repeatability between the two blocks. For all survival analyses, the z -scores and P -values were reported, with the z -score defined as the log of the hazard ratio (risk of death, $\exp(\text{coef})$) divided by the standard error of the log hazard ratio.

Age-specific reproduction was analyzed using generalized linear mixed-effects models to account for temporal pseudoreplication of repeated measures on the same individuals across lifetime, with the template model builder package (“glmmTMB” in R [Brooks et al. 2017; Brooks et al. 2019]). Models fitted with Poisson, Zero-Inflated Poisson, Generalized Poisson, and Zero-Inflated Generalized Poisson error structure were compared using AIC values of model fit (“AICtab” function in “bbmle” package), following Brooks et al. (2019), to account for under- or overdispersion and for zero-inflation, when it was found to occur in simulated residuals generated with the “DHARMA” package (Hartig 2020). Age and a quadratic form of age (age^2) were fitted as fixed effects in both the conditional and zero-inflation model formula, and significance assessed in each case. The age^2 term controlled for the curved (nonlinear) trajectory of reproduction across age (Bates et al. 2015). UV irradiation, RNAi treatment, and their

three-way interaction with age or age² were fitted as fixed effects, and experimental block, observer, and a unique plate identifier as random effects, the latter to control for repeated measures of fecundity from the same individuals across lifetime. Genotype was substituted with UV as a fixed effect to assess the transgenerational effects of *daf-2* RNAi on N2 versus *hrde-1* mutants. Models that did not converge were not included in model comparison, and the converging model with best AIC fit was presented (as Brooks et al. 2019).

Fitness (lambda) was calculated using the “lambda” function in the “popbio” package. More details are provided in the Supporting Information. Fitness, total reproduction, and egg size data were analyzed using a generalized linear model (GLM) with Gaussian error structure (“lm” function in “stats” package). Locomotion, a measure of stress resistance to heat shock, was coded as a binary response variable (normal or abnormal movement) and analyzed using a GLM with binomial error structure.

Results and Discussion

PARENTAL *daf-2* RNAi IN ADULTHOOD EXTENDS LIFE SPAN AND IMPROVES OFFSPRING FITNESS UNDER UV-INDUCED STRESS

Adulthood-only *daf-2* RNAi in N2 wildtype *C. elegans* nematodes significantly extended parental life span relative to empty vector controls under benign conditions (no irradiation) and under ultraviolet-C (“UV”) irradiation-induced stress (Cox proportional hazards mixed-effects model, *coxme*, with matricides censored, RNAi: $z = 10.530$, $P < 0.001$; UV: $z = 0.070$, $P = 0.940$; RNAi \times UV: $z = 1.500$, $P = 0.130$; matricides classed as dead, RNAi: $z = 10.400$, $P < 0.001$; UV: $z = 0.250$, $P = 0.800$; RNAi \times UV: $z = 1.310$, $P = 0.190$; Fig. 1a). This UV-induced stress encompassed both the mutagen properties of UV radiation and the stress response induced.

There was no cost of *daf-2* RNAi in adulthood to parental reproduction, neither under benign conditions nor when parents were UV irradiated (Fig. S1). Reproduction was estimated as age-specific offspring production (fecundity, Generalized Poisson, RNAi \times UV \times Age: $z = -2.546$, $P = 0.0109$), as individual fitness (lambda, generalized linear model, GLM, RNAi: $t = -0.065$, $df = 1$, $P = 0.948$; UV: $t = -3.328$, $df = 1$, $P = 0.0104$; RNAi \times UV: $t = 0.159$, $df = 1$, $P = 0.874$), and as total lifetime reproduction (GLM, RNAi: $t = -1.887$, $df = 1$, $P = 0.0606$; UV: $t = -3.665$, $df = 1$, $P < 0.001$; RNAi \times UV: $t = 0.066$, $df = 1$, $P = 0.948$). UV irradiation did reduce parental fitness and total lifetime reproduction.

However, *daf-2* RNAi significantly increased offspring fitness both under benign conditions and when parents were UV irradiated (lambda, GLM: RNAi: $t = -2.305$, $df = 1$, $P = 0.0222$; UV: $t = -1.625$, $df = 1$, $P = 0.1059$; RNAi \times UV: $t = -0.772$,

$df = 1$, $P = 0.441$; Fig. 1b). This absence of fitness costs for the offspring of UV-irradiated parents, despite the costs of irradiation for parental reproduction, suggests that irradiated parents may have invested in germline repair mechanisms, which improved the fitness of their offspring (Smelick and Ahmed 2005; Maklakov and Immler 2016; Chen et al. 2020).

The impact of parental *daf-2* RNAi and UV treatment on offspring age-specific reproduction varied with offspring age as did the number of offspring with zero fecundity (zero-inflated generalized Poisson [ZIGP], RNAi \times age: $z = 4.864$, $P < 0.001$; UV \times age: $z = 3.678$, $P < 0.001$; RNAi \times UV: $z = -0.175$, $P = 0.861$; ZI varied with age: $z = -0.227$, $P = 0.00656$ and age²: $z = 3.301$, $P < 0.001$; Fig. S2). There was no effect of parental treatment (neither *daf-2* RNAi, UV irradiation, nor their interaction) on offspring total lifetime reproduction (GLM, RNAi: $t = -1.141$, $df = 1$, $P = 0.255$; UV: $t = -1.626$, $df = 1$, $P = 0.106$; RNAi \times UV: $t = -0.053$, $df = 1$, $P = 0.958$; Fig. S2).

Our findings show that under adulthood-only *daf-2* RNAi, parents increase investment into somatic maintenance resulting in increased life span with no survival or reproductive cost to themselves or their offspring under benign conditions validating previous work in *C. elegans* (Dillin et al. 2002; Lind et al. 2019). Importantly, we reveal that the absence of a longevity-fecundity trade-off in parents persisted under stressful conditions, when organisms have to invest into repairing UV-induced damage. Previous work in *daf-2* mutants has also shown them to be more resistant to UV irradiation (Hyun et al. 2008).

Furthermore, we show that *daf-2* knockdown in adult parents primarily influenced the timing of reproduction in their offspring. Specifically, it caused a shift to increased early-life reproduction, improving offspring individual fitness, rather than an increase in total reproduction.

Timing of reproduction in both laboratory and natural *C. elegans* populations is important as *C. elegans* reproduces very fast and so the strength of natural selection (selection gradient) declines very rapidly with advancing age (Chen et al. 2006; Frezal and Felix 2015). Under standard lab conditions, self-fertilizing *C. elegans* hermaphrodites complete most of their reproduction within the first 3 days of adulthood and in nature, *C. elegans* exhibit boom-and-bust population dynamics involving periods of very rapid population growth, driven by an ephemeral food supply (Frezal and Felix 2015; Schulenburg and Felix 2017). As individual fitness is heavily weighted by early reproduction and fast development (Supporting Information), even relatively small increases in Day 1 and Day 2 reproduction (Fig. S2) can improve individual fitness, despite having no effect on total lifetime reproduction, if later reproduction is correspondingly reduced.

Although an earlier study found increased total reproduction in the first generation of offspring from *C. elegans*

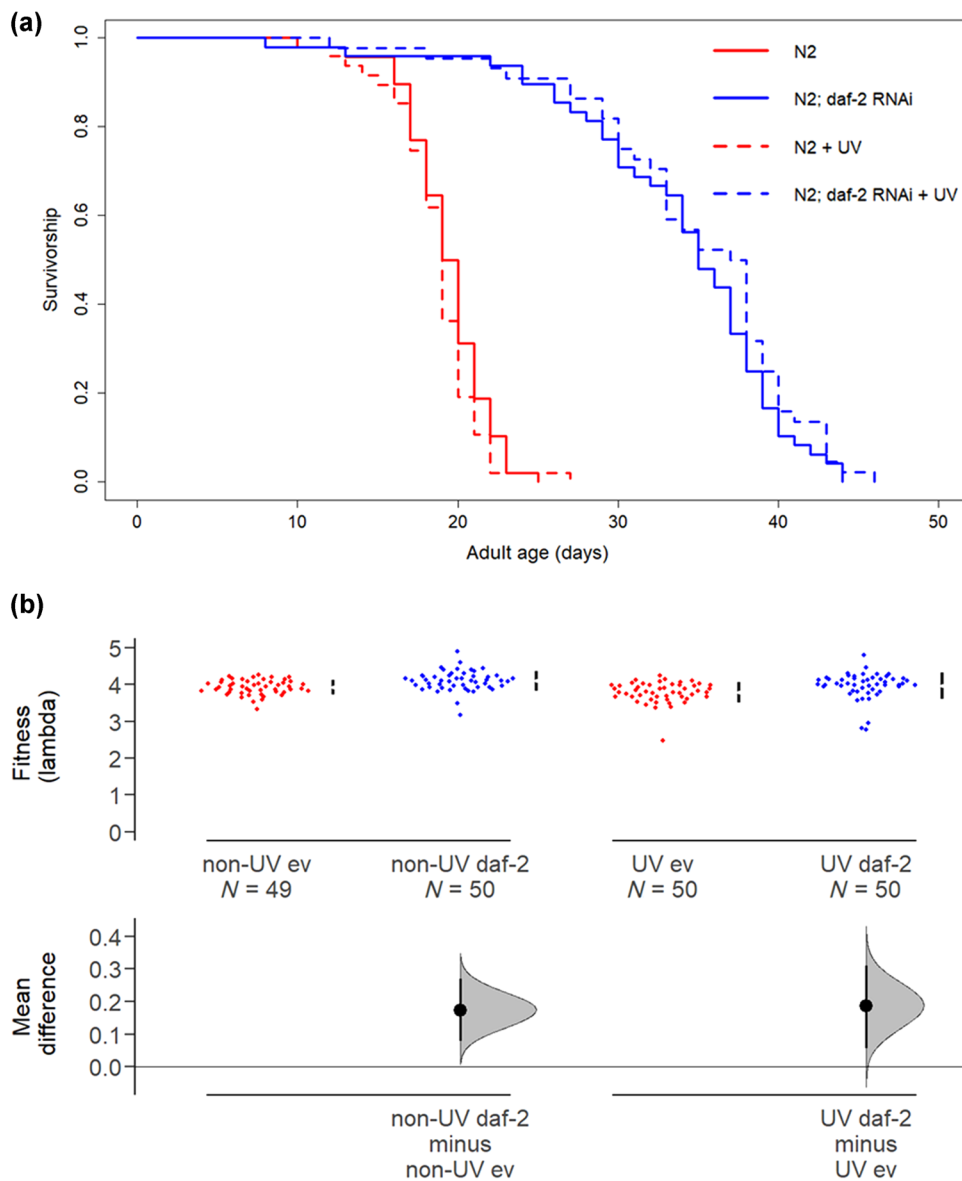


Figure 1. Adulthood *daf-2* RNAi in UV-irradiated and nonirradiated parents increases parental life span and offspring fitness. (a) Parental survival, $n = 50$ per treatment. (b) Offspring fitness. UV irradiation status and adulthood RNAi treatment (either *daf-2* RNAi, “*daf-2*,” or empty vector control, “ev”) of parents are indicated. Offspring were maintained on empty vector and were not irradiated. Mean (effect size) and 95% confidence intervals shown were derived using nonparametric bootstrap resampling in “dabestr” R package (Ho et al. 2019).

parents treated with *daf-2* RNAi, across N2 wild-type and two other genetic backgrounds, this was also accompanied by increased offspring fitness (Lind et al. 2019), in agreement with our results here. Under UV-induced mutagenesis, the costs of germline maintenance would be expected to become more apparent (Sniegowski et al. 2000; Berger et al. 2017; Chen et al. 2020). However, we show that adulthood-only *daf-2* RNAi improves fitness of F1 offspring even when parents are under stress.

PARENTAL *daf-2* RNAi IN ADULTHOOD DOES NOT REDUCE FITNESS OR TOTAL REPRODUCTION UNDER IF

Under resource limitation via IF, there was also no cost of adulthood-only *daf-2* RNAi for individual fitness (lambda, generalized linear mixed-effects model [GLMER], RNAi: $t = -0.964$, $P = 0.337$; Fig. 2a). Despite a small cost of adulthood *daf-2* RNAi for fitness when fed AL in this assay (GLMER, RNAi: $t = 2.375$, $P = 0.019$; RNAi \times diet: $t = -2.067$, $P = 0.040$;

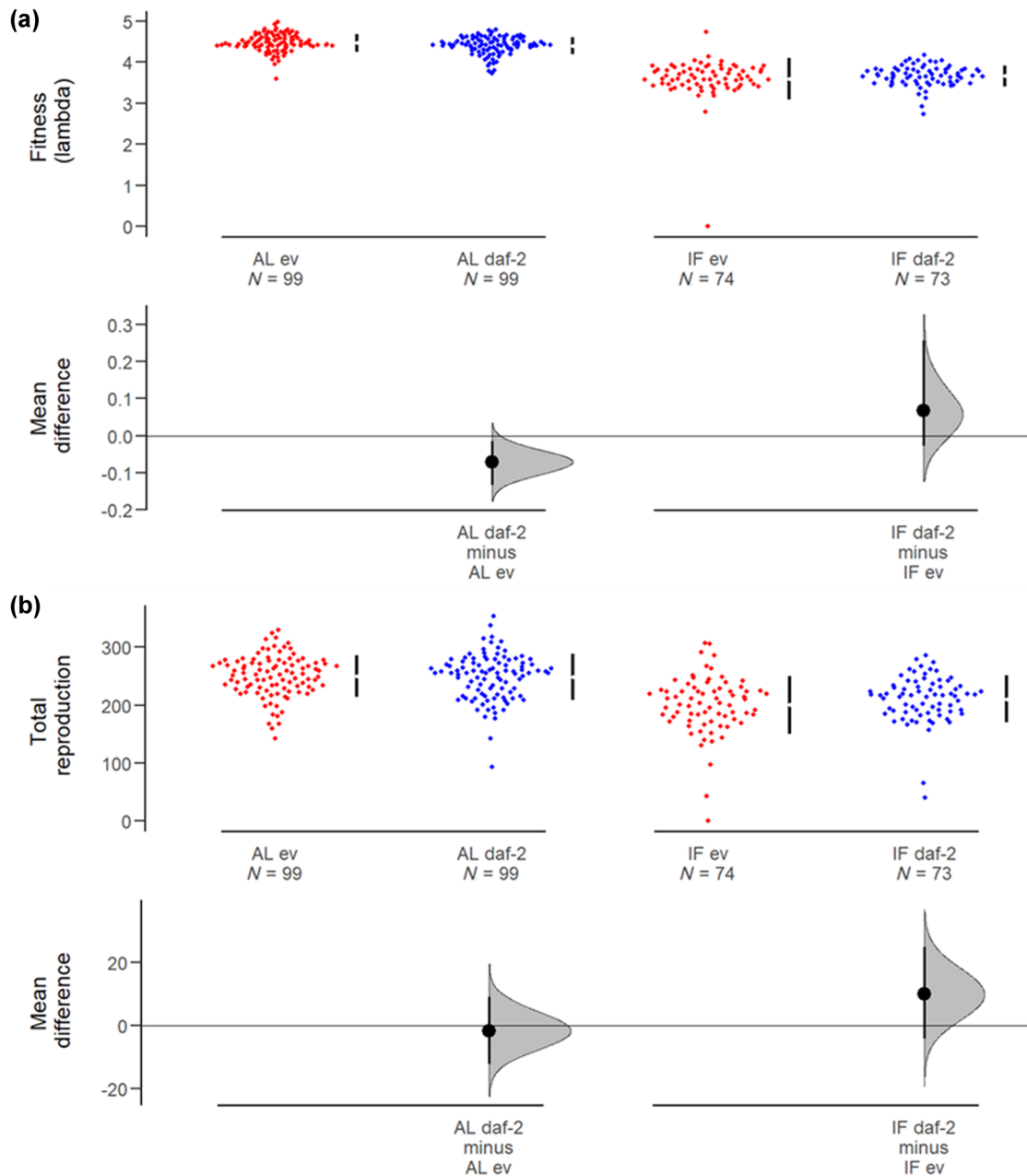


Figure 2. The effect of adulthood *daf-2* RNAi on parental reproduction under intermittent fasting and ad libitum feeding. (a) Fitness (λ). (b) Total lifetime reproduction. Individuals were either intermittently fasted (IF) for 9 h on Days 1, 3, 5, and 7 of adulthood or fed ad libitum (AL) throughout life. Adulthood RNAi treatment (either *daf-2* RNAi, “*daf-2*,” or empty vector control, “*ev*”) of parents is indicated. Adulthood *daf-2* RNAi had no effect on fitness or total reproduction under IF. Mean effect size and 95% confidence intervals are shown.

diet: $t = -15.595$, $P < 0.001$; Fig. 2a), there was no cost of *daf-2* RNAi for total lifetime reproduction under either IF or AL (GLMER, RNAi: $t = 0.290$, $P = 0.772$; diet: $t = -5.979$, $P < 0.001$; RNAi \times diet: $t = -1.387$, $P = 0.16646$; Fig. 2b).

We may have expected a reproductive cost of adulthood *daf-2* RNAi to become apparent under nutritional or UV irradiation

stress, but instead we show no cost to fitness or total lifetime reproduction of adulthood *daf-2* RNAi under IF (Fig. 2) or UV irradiation (Fig. S1). This is in agreement with previous work that showed *daf-2* RNAi in adult N2 *C. elegans* did not reduce lambda fitness under a variable environment of light and temperature stress (Carlsson et al. 2021).

MULTIGENERATIONAL *daf-2* RNAi IN ADULTHOOD PROTECTS AGAINST EXTINCTION

We found that *daf-2* RNAi reduced extinction under both UV-induced and spontaneous MA across 40 generations (Cox proportional hazards regression analysis, *coxph*, RNAi: $z = 2.159$, $P = 0.031$; UV: $z = 11.469$, $P < 0.001$; RNAi \times UV: $z = -0.537$, $P = 0.591$; Fig. 3a). Extinction results did not differ significantly between two independent experimental blocks (*coxph*, $z = -0.942$, $P = 0.346$; Fig. S3). The major causes of MA line extinction (Fig. 3b) were infertility (failure to lay eggs) and sterility (the production of eggs that did not hatch), indicative of underlying germline damage. Infertility and sterility were likely associated with observed reproductive abnormalities such as a deformed vulva, abnormal external growth close to the vulva (possible tumor), and cavities in the reproductive tract in place of embryos or oocytes (Fig. 3c). It is known that vulva-less *C. elegans* mutants are unable to lay eggs and can die from internal hatching (Ferguson and Horvitz 1985; Sternberg and Horvitz 1989).

It is possible that UV irradiation may also have caused non-genetic damage (to proteins or other macromolecules) that was not inherited but may also have contributed to sterility or infertility, and that *daf-2* RNAi additionally may have protected against. Furthermore, *daf-2* RNAi may have provided enhanced stress resistance to protect against stress responses induced by UV irradiation. Despite this, previous work in *C. elegans* has shown that UV irradiation most commonly induces base substitutions, which are subject to the nucleotide excision repair pathway (Meier et al. 2014).

The reduction in extinction observed in our MA lines with *daf-2* RNAi shows the benefits of increased investment into germline maintenance that became more pronounced across multiple generations of UV-induced and spontaneous MA.

MULTIGENERATIONAL *daf-2* RNAi INCREASED FITNESS OF SURVIVING MA LINES

We next tested for the effects of *daf-2* RNAi on life-history traits of surviving MA lines. We were interested in how IIS influences potentially detrimental effects of MA that are not sufficiently severe to cause line extinction. We assayed age-specific reproduction, egg size, male production, and adult heat shock resistance in grand-offspring from the spontaneous and UV-induced MA lines on *daf-2* RNAi versus control treatments, at generation 20 of MA, following two generations of rearing under common garden conditions (no irradiation, on empty vector control) to attenuate direct effects of irradiation and RNAi. Generation 20 was a point at which there was a pronounced benefit of *daf-2* RNAi for protection against extinction in the UV-induced MA lines, but no clear difference in extinction trajectories for the spontaneous (nonirradiated) MA lines (Fig. 3a).

Adulthood-only *daf-2* RNAi across 20 generations of MA significantly increased individual fitness in the surviving irradiated MA lines, but there was no effect of *daf-2* RNAi on the fitness of nonirradiated lines (GLM, irradiated, RNAi: $t = -2.804$, $P = 0.006$; nonirradiated, RNAi: $t = 0.647$, $P = 0.518$; all data, RNAi \times UV: $t = -2.886$, $P = 0.004$; Fig. 4a), in agreement with the results on line extinction at generation 20 (Fig. 3a).

The increased fitness of irradiated *daf-2* RNAi MA lines was driven by their improved early-life fecundity (Day 1 and Day 2 offspring production), relative to irradiated controls (ZIGP, RNAi \times age: $z = -1.981$, $P = 0.0475$; RNAi \times age²: $z = 3.334$, $P < 0.001$; Fig. 4b), an effect absent in the nonirradiated lines (ZIGP, non-UV, RNAi: $z = 0.05$, $P = 0.963$; all data: RNAi \times UV \times age: $z = -2.50$, $P = 0.0125$; RNAi \times UV \times age²: $z = 3.58$, $P < 0.001$; ZI intercept: $z = -18.27$, $P < 0.001$). There was no effect on total reproduction (GLM, UV: $t = -0.811$, $P = 0.419$; non-UV: $t = 0.875$, $P = 0.383$; UV \times RNAi: $t = -1.138$, $P = 0.256$; Fig. S4).

We suggest that the fitness benefits of *daf-2* RNAi for irradiated MA lines were most likely due to genetic differences between the treatments, as the benefits persisted after two generations of common garden rearing and therefore were neither due to the direct exposure of offspring to RNAi nor acute effects of UV (neither as adults nor as eggs). This conclusion is reinforced by the finding that parental *daf-2* RNAi effects on offspring fitness do not persist beyond F1 (see below, Fig. 6). Variation in fitness was considerably greater across irradiated than across nonirradiated MA lines (Fig. 4a), suggesting that irradiation-induced de novo mutations generated greater genetic and phenotypic variation compared to spontaneous MA. The spontaneous mutation rate in N2 wild-type *C. elegans* is estimated as one de novo mutation per individual per generation, under standard conditions (Denver et al. 2012; Meier et al. 2014). The close association between extinction trajectories during the first 20 generations of MA (Fig. 3a) and the fitness of the surviving MA lines (Fig. 4a) suggests that a threshold of accumulating deleterious mutations needs to be crossed before the effect size is sufficiently severe to result in extinction.

The most parsimonious explanation for the reduced extinction and increased fitness of *daf-2* RNAi-irradiated lines would therefore be due to genetic differences in the rate of MA. To directly distinguish between genetic and nongenetic effects associated with MA line extinction, future work could use whole genome sequencing to quantify mutation rates between the MA treatments.

To determine if increased offspring fitness in the surviving *daf-2* RNAi-treated MA lines was associated with greater parental resource allocation into their eggs, we measured egg size, as a proxy for parental investment ($n = 75$ eggs measured, one per individual taken from the Generation 20 fitness assay).

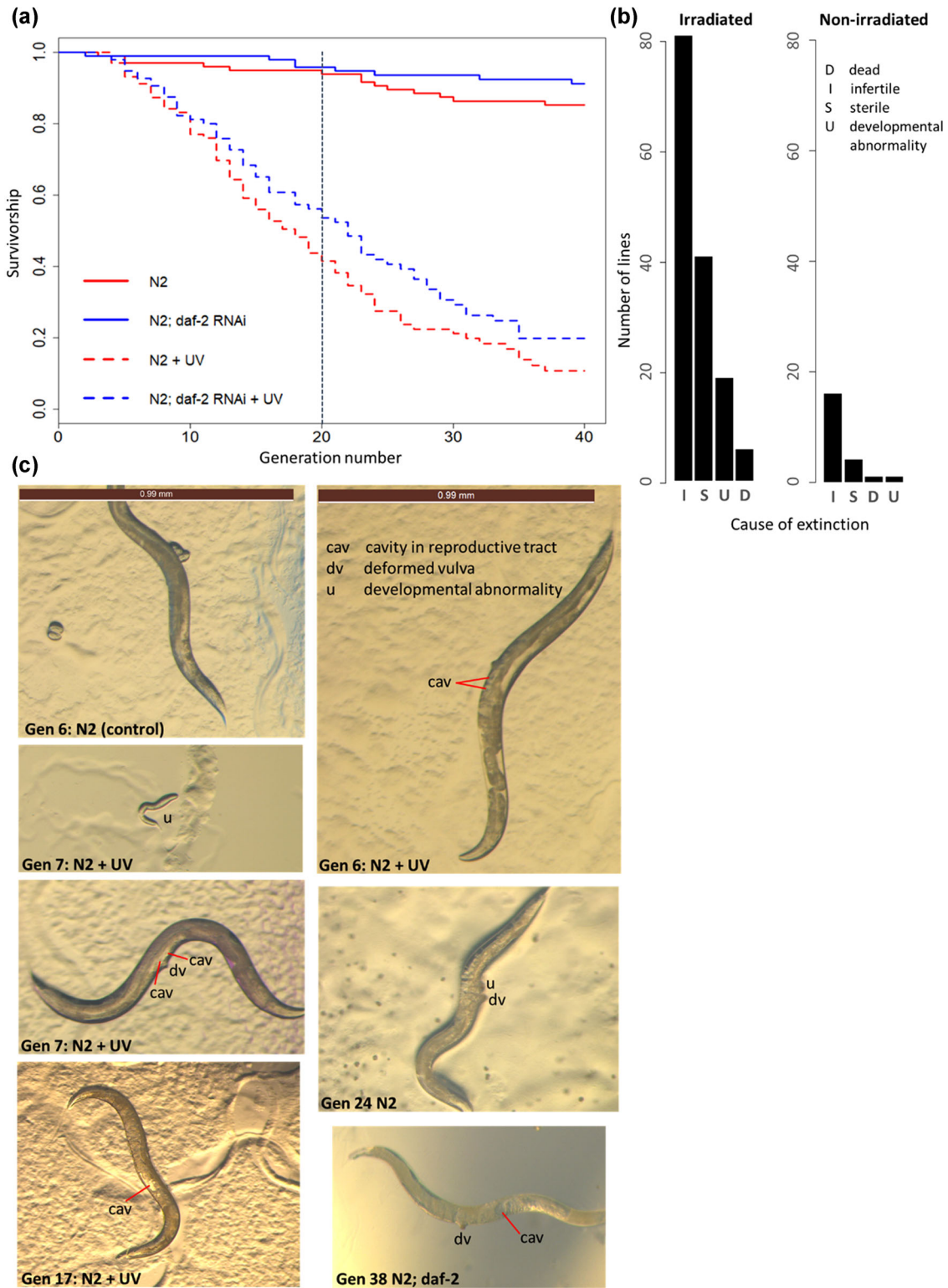


Figure 3. Reduced adulthood IIS, via *daf-2* RNAi, protects against N2 wild-type extinction under mutation accumulation. (a) Transgenerational survival in N2 under spontaneous and UV-induced mutation accumulation. Vertical dotted line at generation 20 indicates timing of life-history assay. Sample size of 100 lines per RNAi strain by irradiation treatment combination. (b) Causes of extinction of N2 wild-type MA lines indicate germline damage. (c) Representative images of germline damage. Brown scale bar of 0.99 mm for all images.

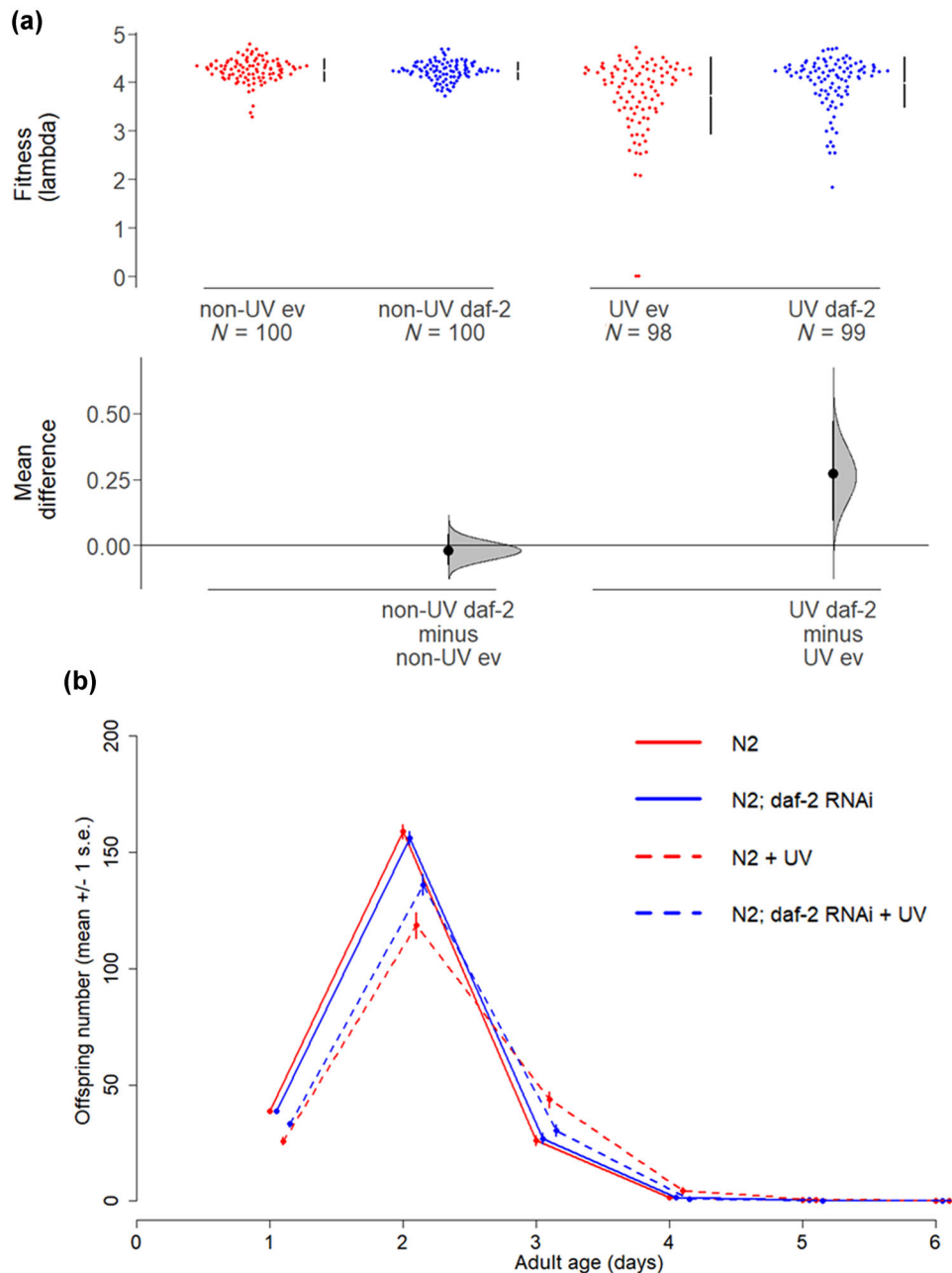


Figure 4. The effect of *daf-2* RNAi on fitness and age-specific reproduction after 20 generations of MA. (a) Individual fitness (lambda) of the grand-offspring of N2 wild-type MA lines at generation 20. Fitness was assayed in a standard common garden environment, on the empty vector control and no irradiation; following two generations of rearing under standard conditions, from MA generation 20. Mean and 95% confidence intervals are shown. (b) Age-specific reproduction in the grand-offspring of N2 MA lines at generation 20.

Previous work has shown that reduced IIS, either via dietary restriction or via *daf-2* RNAi, increases mean egg size (Hibshman et al. 2016; Lind et al. 2019). Egg size has been correlated with offspring fitness (Lind et al. 2019) and larval body length (Ivimey-Cook et al. 2021) in *C. elegans*.

We found that grandparental *daf-2* RNAi resulted in grand-offspring (F2) that laid smaller eggs if their grand-parents from

the MA lines had been irradiated, but there was no significant effect on the size of eggs laid by F2 offspring descended from nonirradiated grandparents treated with *daf-2* RNAi following 20 generations of MA (GLM, UV lines, RNAi: $t = 2.362$, $df = 1$, $P = 0.0195$; non-UV lines, RNAi: $t = -1.246$, $df = 1$, $P = 0.215$; all data, RNAi \times UV: $t = 2.642$, $df = 1$, $P = 0.00869$; RNAi \times UV \times Block: $t = -0.774$, $df = 1$, $P = 0.440$; Fig. 5).

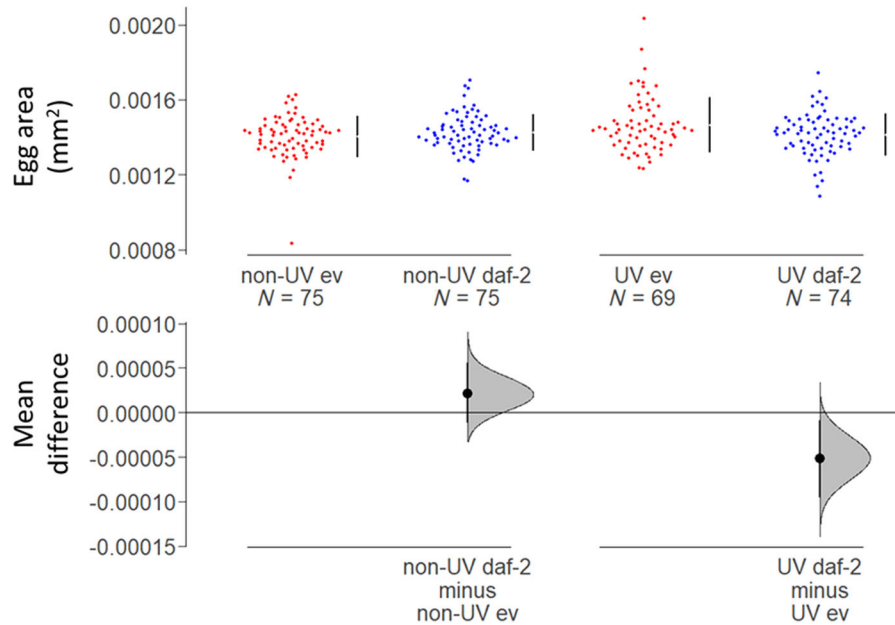


Figure 5. The effect of *daf-2* RNAi on egg size after 20 generations of spontaneous or UV-induced MA. Egg size (area, mm²) was reduced in the grand-offspring of UV-induced MA lines on *daf-2* RNAi (“*daf-2*”) relative to those from UV-irradiated MA lines on empty vector control (“*ev*”). However, this difference in egg size was absent from the spontaneous (“non-UV”) MA lines, following 20 generations of MA. Grand-offspring were all nonirradiated and kept on *ev*. Mean and 95% confidence intervals are shown.

This is contrary to the increase in F1 egg size under reduced parental IIS found in previous work under benign conditions (Hibshman et al. 2016; Lind et al. 2019).

Our results indicate that even though F2 offspring from irradiated *daf-2* RNAi MA lines laid smaller eggs, this did not seem to impact negatively on the quality (fitness) of offspring they produced. It is possible that a smaller egg size could be a phenotypically plastic response to UV radiation, perhaps to improve stress resistance to irradiation. Alternatively, it could be the result of a trade-off between improved investment into germline genetic quality and egg size. At present, it is unclear why grandparental *daf-2* RNAi resulted in smaller eggs in F2.

Exposure to certain environmental stresses (e.g., high temperature [Nigon and Dougherty 1949], starvation [Morran et al. 2009], increased mutation rate [Cutter 2005]) increases male production and outcrossing in *C. elegans*. However, we found no increase in the proportion of males produced by the grand-offspring in irradiated MA lines. Only one male developed from the 293 eggs assayed.

Reduced IIS increases the activity of heat shock factor 1 that mediates life span extension (Hsu et al. 2003; Seo et al. 2013). Heat shock responses are conserved across diverse taxa and positively associated with life span (as reviewed by Muñoz 2003; Åkerfelt et al. 2010). To test the stress resistance of postreproductive (Day 7) adults derived from the Generation 20 MA lines, we assayed survival and locomotion under acute heat shock for

1 h and 45 min at 37°C (following Zevian and Yanowitz 2014). We found no effect of MA line treatment (neither UV irradiation nor *daf-2* RNAi) on the survival of untreated Day 7 F2 offspring following heat shock. Only 4% of the 371 individuals were dead by 24 h after heat shock. This high post-heat-shock survival of Day 7 adults is in line with previous work, which found survival after heat shock to more than double with age, from the first to the fourth day of adulthood in *C. elegans* (Zevian and Yanowitz 2014). However, F2 offspring from nonirradiated *daf-2* RNAi MA lines recovered normal locomotion faster following heat shock than F2 from nonirradiated MA control lines, but this benefit was not seen in descendants from irradiated MA lines (Binomial GLM, 3 h post-heat shock, RNAi: $z = -2.274$, $df = 1$, $P = 0.0229$; UV: $z = -2.078$, $df = 1$, $P = 0.0377$; RNAi \times UV: $z = 2.304$, $df = 1$, $P = 0.0212$; 24 h post-heat shock, UV: $z = -0.206$, $df = 1$, $P = 0.837$; RNAi: $z = -0.045$, $df = 1$, $P = 0.964$; RNAi \times UV: $z = 1.347$, $df = 1$, $P = 0.178$; Fig. S5).

PARENTAL *daf-2* RNAi DOES NOT AFFECT OFFSPRING FITNESS TRANSGENERATIONALLY

Transgenerational epigenetic inheritance of RNAi can occur in *C. elegans* via transmission of small interfering RNAs (Fire et al. 1998; Posner et al. 2019). This transgenerational inheritance of RNAi can last for several generations (Rechavi et al. 2014; Ni et al. 2016; Posner et al. 2019) and requires the germline

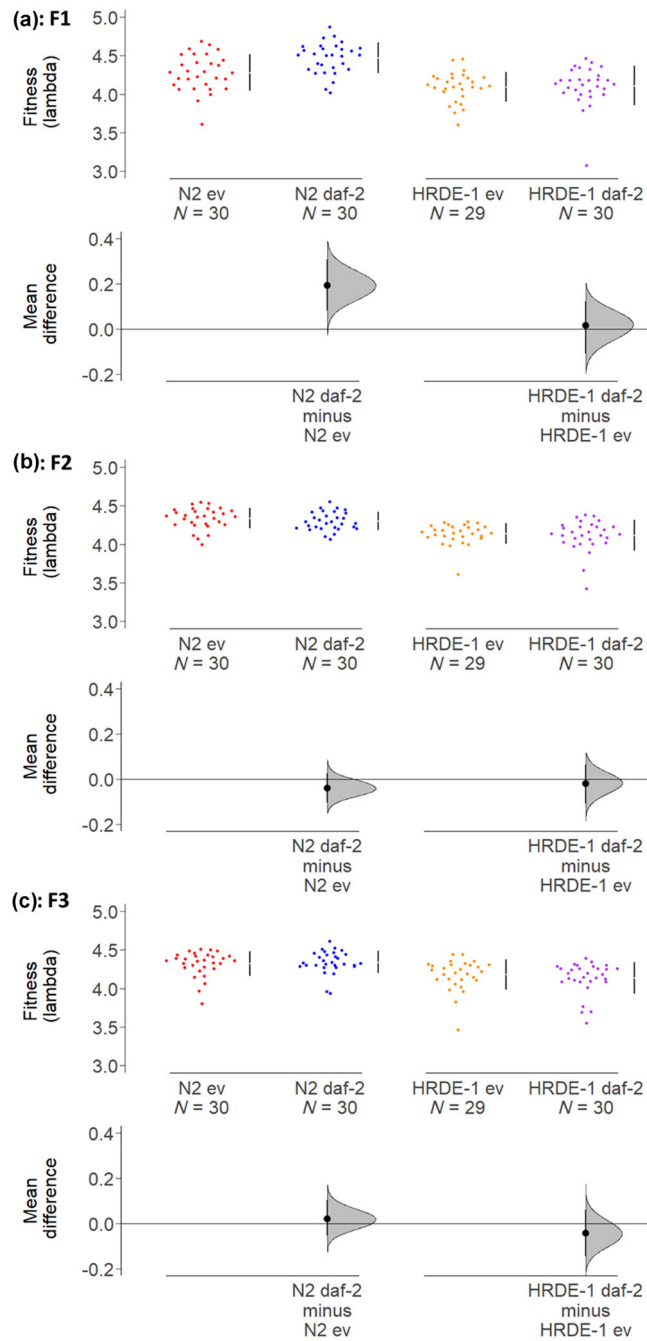


Figure 6. The effects of *daf-2* RNAi on offspring fitness do not persist beyond the first offspring generation. Fitness of the (a) first (F1), (b) second (F2), and (c) third (F3) generations of offspring from parents treated with *daf-2* RNAi (“*daf-2*”) or an empty vector control (“*ev*”) as indicated, in N2 wild-type and RNAi inheritance-deficient *hrde-1* mutant backgrounds. All offspring generations were untreated (kept on *ev*).

argonaut protein, HRDE-1, that is absent in *hrde-1* (heritable RNAi defective-1) mutants (Buckley et al. 2012). RNAi is still functional in somatic cells, as the *hrde-1* mutation only affects the inheritance of RNAi via the germline. To determine whether there was direct transgenerational transfer of *daf-2* RNAi via

the germline and thus for how many generations the effects of parental *daf-2* RNAi could persist, we assayed the fitness effects on three generations of untreated offspring (F1, F2, and F3) from *daf-2* RNAi-treated versus control parents. Two genetic backgrounds were used: *C. elegans* N2 wild-types, and *hrde-1* mutants that did not transfer RNAi transgenerationally.

Effects of parental *daf-2* RNAi on the fitness of untreated descendants were absent after the first generation of offspring in the N2 wild-types and absent across all generations of offspring in the *hrde-1* mutants (GLM, F1, RNAi \times genotype: $t = -2.160$, $df = 1$, $P = 0.0329$; F2, RNAi \times genotype: $t = 0.372$, $df = 1$, $P = 0.711$; RNAi: $t = 1.090$, $df = 1$, $P = 0.278$; genotype: $t = 7.044$, $df = 1$, $P < 0.001$; F3, RNAi \times genotype: $t = -0.973$, $df = 1$, $P = 0.332$; RNAi: $t = 0.359$, $df = 1$, $P = 0.721$; genotype: $t = 5.302$, $df = 1$, $P < 0.001$; all data: RNAi \times generation: $F = 4.450$, $df = 2$, $P = 0.0124$; Fig. 6). Furthermore, there was no effect of parental *daf-2* RNAi on the age-specific reproduction of offspring beyond F1, for either N2 or *hrde-1* backgrounds (Fig. S6; Table S2). There was also no parental *daf-2* RNAi effect on total reproduction for F2 and F3 offspring generations (Fig. S7; Table S2).

The absence of fitness benefits in the second and third generations of offspring from *daf-2* RNAi parents implies that *daf-2* RNAi is not transgenerationally inherited beyond the first generation of offspring in the N2 background. This strongly suggests that the life-history differences between the *daf-2* RNAi and control irradiated MA lines at generation 20 in the common garden experiment were due to genetic differences and not the direct effects of inherited RNAi. Interestingly, the heritable RNAi deficiency 1 (*hrde-1*) gene was required for the fitness benefits of parental *daf-2* RNAi in F1 offspring, as these offspring fitness benefits were absent in the *hrde-1* mutant background. The *hrde-1* mutants had overall lower fitness and total reproduction than N2 wild type.

***hrde-1* IS REQUIRED FOR *daf-2* RNAi TO CONFER GERMLINE PROTECTION UNDER MA**

hrde-1 encodes a germline argonaut protein that plays a key role in nuclear RNAi, RNAi inheritance, and promoting germline immortality (Spracklin et al. 2017). To determine if functional *hrde-1* was necessary for the protective effects of *daf-2* RNAi under spontaneous and UV-induced MA, we ran 400 MA lines in parallel to the N2 MA experiment, using the *C. elegans* *hrde-1* mutant background and reduced IIS, via *daf-2* RNAi, in half of the MA lines. The heritable RNAi deficiency (*hrde-1* mutant) resulted in the rapid extinction of irradiated MA lines and the loss of the protective effects of *daf-2* RNAi under UV-induced MA, across 25 generations (RNAi: $z = -4.016$, $P < 0.001$; UV: $z = 12.370$, $P < 0.001$; RNAi \times UV: $z = -1.758$, $P = 0.079$; Fig. 7). In fact, irradiated *daf-2* RNAi lines went extinct faster than controls, in

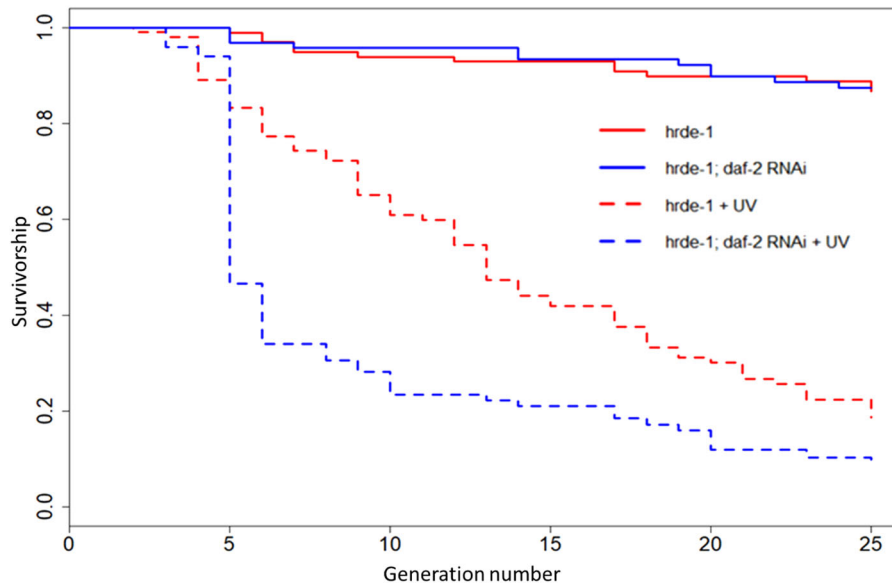


Figure 7. The effect of *daf-2* RNAi on *hrde-1* mutant multigenerational survival under mutation accumulation. Sample size of 100 MA lines for each RNAi by UV irradiation treatment combination.

the *hrde-1* mutant background. This represented a significant difference between *hrde-1* mutant and N2 wild-type backgrounds in the effect of *daf-2* RNAi (although not UV) on MA line extinction (RNAi \times genotype: $z = 4.480$, $P < 0.001$; genotype: $z = -8.083$, $P < 0.001$; RNAi: $z = -4.351$, $P < 0.001$; UV \times genotype: $z = -0.487$, $P = 0.626$; UV: $z = 17.148$, $P < 0.001$).

The major cause of extinction in irradiated *hrde-1* mutant MA lines was developmental arrest, followed by infertility (Fig. S8). This strongly suggests that functional RNAi, mediated by the HRDE-1 germline argonaut protein, is required for *daf-2* RNAi protection against extinction, and for normal development and reproduction under UV-induced MA.

We show the important role of *hrde-1* in the *daf-2* RNAi-mediated protection of irradiated offspring under mutation. Our results suggest that the interaction between UV-induced germline damage and a deficiency in transgenerational inheritance of RNAi reverses the protective benefits of *daf-2* RNAi. This supports previous work suggesting that *hrde-1* mutants suffer from progressive sterility under high temperatures, implying their increased sensitivity to environmental stresses driven in part by defects in gametogenesis (Buckley et al. 2012).

MULTIGENERATIONAL *daf-2* RNAI IN ADULTHOOD DOWNREGULATES *daf-2* EXPRESSION IN MA LINES AT GENERATION 20

We confirmed, using qRT-PCR analysis, that feeding nematodes bacteria that expresses double-stranded RNA for *daf-2* significantly downregulated *daf-2* expression in the MA lines at gen-

eration 20 (Table S3). The extent of *daf-2* downregulation at generation 20 varied between the MA treatments (Table S3; Fig. S9). For N2 wild types, *daf-2* expression was downregulated by 23% in UV-induced MA lines and by 12% in spontaneous MA lines (based on mean gene expression fold change, $2^{-\Delta\Delta CT}$). For *hrde-1* mutants under spontaneous MA, *daf-2* expression was more subtly downregulated (by 6% fold change; Table S3; Fig. S9). We were unable to analyze data from UV-induced MA *hrde-1* mutant lines as they did not recover well from freezing (only $n = 1$ MA line for the *daf-2* RNAi treatment; Fig. S9).

The degree of downregulation of *daf-2* expression between MA treatments interestingly matched the different extent of life-history divergence between *daf-2* RNAi and empty vector treatments for the respective MA treatments at generation 20, in both N2 and *hrde-1* mutant extinction trajectories (Figs. 3a, 7) and in N2 fitness (Fig. 4a). The relatively modest degree of *daf-2* downregulation in the spontaneous MA lines for N2, and particularly for *hrde-1* mutants, in comparison to the 50% knockdown of *daf-2* expression from adulthood *daf-2* RNAi found in previous work on Day 2 N2 adults (Carlsson et al. 2021), could be due to an interaction between RNAi knockdown and genetic background, or a possible “acclimatization” to the multigenerational application of *daf-2* RNAi in adulthood, but the exact reasons would require further investigation.

Technical replicates were highly repeatable for all samples (coefficient of variation, CV $< 2.4\%$ and CV $< 1.5\%$ for 98% of samples). Biological replicates showed more variation in relative gene expression within each MA line (mean CV = 7%; maximum

CV = 8%), but within the expected range for *C. elegans* based on previous qRT-PCR analyses for different genes (Chauve et al. 2020; Lind et al. 2021).

Conclusion

We found that reduced insulin signaling in adulthood, via *daf-2* RNAi, protects against extinction under both UV-induced and spontaneous MA in *C. elegans*. Furthermore, the fitness of the surviving UV-irradiated MA lines was higher under *daf-2* RNAi. Most extinctions occurred because of infertility, egg hatching failure, and developmental abnormalities suggesting that germline MA directly contributed to the observed differences between the RNAi treatments. Future assessment of mutation rates in the MA lines could be useful to distinguish between genetic and nongenetic effects on extinction. Germline protection under *daf-2* RNAi requires nuclear argonaut *hrde-1* because fitness of *hrde-1*; *daf-2* RNAi worms was reduced both in one-generation and in multigeneration experiments. This is in line with previous work suggesting that *hrde-1* is required for trans-generational inheritance and germline immortality (Buckley et al. 2012; Spracklin et al. 2017).

We set out to test whether adulthood-only *daf-2* RNAi, known to extend life span without an obvious cost to parental fecundity (Dillin et al. 2002; Lind et al. 2019), trades off with detrimental fitness effects when applied across multiple generations. However, we found that multigenerational downregulation of *daf-2* via adulthood-only RNAi has positive effects on the germline and protects MA lines from extinction. This was particularly so when germline mutation rate was increased by low-level UV radiation. Our results therefore suggest that although wild-type levels of *daf-2* expression promote development, growth, and early-life reproduction, they are costly for late-life fitness and this cost accumulates across generations leading to lineage extinction.

Antagonistic pleiotropy theory of ageing (AP) maintains that genes that increase fitness in early life at the expense of fitness late in life can be overall beneficial for fitness and go to fixation (Williams 1957). In line with AP, downregulation of *daf-2* expression during development reduces fitness (Dillin et al. 2002; Chen et al. 2007), but what is the physiological basis of the putative trade-off? Our findings that adulthood-only downregulation of *daf-2* expression protects both the germline and the soma under benign conditions and under UV-induced stress argue against the idea that resource allocation underlies this trade-off. However, our results are in line with the hypothesis that selection optimizes physiological processes for development and early-life reproduction but fails to optimize the regulation of gene expression associated with these same physiological processes later in life (de Magalhaes and Church 2005; de Magalhaes 2012;

Blagosklonny 2013; Gems and Partridge 2013; Maklakov and Chapman 2019). This is likely because selection on insulin signaling in adulthood is too weak in *C. elegans* (Chen et al. 2006). Future studies should focus on investigating the contribution of weakening selection on gene expression with age to the evolution of ageing across taxa, on testing the effects of downregulated IIS on survival and fitness under wide range of ecologically relevant stresses, and on developing a comprehensive theoretical framework that links genetic and physiological theories of ageing.

AUTHOR CONTRIBUTIONS

ED, AAM, and HC conceived and designed the study, with TC and SI providing additional advice. ED, HC, KS, and ZS performed the experiments and collected the data, which ED and KS analyzed. ED and AAM wrote the manuscript, with comments from all authors.

ACKNOWLEDGMENTS

The authors thank M. Lind and E. Ivimey-Cook for statistical discussions, and T. van Baalen, N. Edden, and A. Dehem for experimental help. This study was funded by an ERC Consolidator Grant (GermlineAgeing-Soma/724909) to AAM.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

DATA ARCHIVING

The data are available at <https://doi.org/10.6084/m9.figshare.12783650.v1>. The codes for analyses are available at <https://gist.github.com/ElizabethMLDuxbury/1e339bd9ac871461eeb2050b805d19a8>.

LITERATURE CITED

- Ajje, B.C., Estes, S., Lynch, M. & Phillips, P.C.. 2005. Behavioral degradation under mutation accumulation in *Caenorhabditis elegans*. *Genetics*, 170, 655–660.
- Åkerfelt, M., Morimoto, R.I. & Sistonen, L.. 2010. Heat shock factors: integrators of cell stress, development and lifespan. *Nat. Rev. Mol. Cell Biol.*, 11, 545–555.
- Antebi, A. 2013. Regulation of longevity by the reproductive system. *Exp. Gerontol.*, 48, 596–602.
- Arantes-Oliveira, N., Apfield, J., Dillin, A. & Kenyon, C.. 2002. Regulation of life-span by germ-line stem cells in *Caenorhabditis elegans*. *Science*, 295, 502–505.
- Baer, C.F., Shaw, F., Steding, C., Baumgartner, M., Hawkins, A., Houppert, A., Mason, N., Reed, M., Simonelic, K., Woodard, W., et al. 2005. Comparative evolutionary genetics of spontaneous mutations affecting fitness in rhabditid nematodes. *Proc. Natl. Acad. Sci. USA*, 102, 5785–5790.
- Bates, D., Machler, M., Bolker, B.M. & Walker, S.C.. 2015. Fitting linear mixed-effects models using lme4. *J. Stat. Softw.*, 67, 1–48.
- Berger, D., Stangberg, J., Grieshop, K., Martinossi-Allibert, I. & Arnavqvist, G.. 2017. Temperature effects on life-history trade-offs, germline maintenance and mutation rate under simulated climate warming. *Proc. R. Soc. B Biol. Sci.*, 284, 20171721.
- Blagosklonny, M.V. 2013. Ageing is not programmed. *Cell Cycle*, 12, 3736–3742.

- Bolker, B.M., Brooks, M.E., Clark, C.J., Geange, S.W., Poulsen, J.R., Stevens, M.H.H. & White, J.-S.S.. 2009. Generalized linear mixed models: a practical guide for ecology and evolution. *Trends Ecol. Evol.*, 24, 127–135.
- Boyd, W.A., Crocker, T.L., Rodriguez, A.M., Leung, M.C.K., Wade Lehmann, D., Freedman, J.H., Van Houten, B. & Meyer, J.N.. 2010. Nucleotide excision repair genes are expressed at low levels and are not detectably inducible in *Caenorhabditis elegans* somatic tissues, but their function is required for normal adult life after UVC exposure. *Mutat. Res.*, 683, 57–67.
- Brommer, J.E., Merila, J. & Kokko, H.. 2002. Reproductive timing and individual fitness. *Ecol. Lett.*, 5, 802–810.
- Brooks, M.E., Kristensen, K., Darrigo, M.R., Rubim, P., Uriarte, M., Bruna, E. & Bolker, B.M.. 2019. Statistical modelling of patterns in annual reproductive rates. *Ecology*, 100, e02706.
- Brooks, M.E., Kristensen, K., Van Benthem, K.J., Magnusson, A., Berg, C., Nielsen, A., Skaug, H., Machler, M. & Bolker, B.M.. 2017. glmmTMB balances speed and flexibility among packages for zero-inflated generalized linear mixed modeling. *R J.*, 9, 378–400.
- Buckley, B.A., Burkhart, K.B., Gu, S.G., Spracklin, G., Kershner, A., Fritz, H., Kimble, J., Fire, A. & Kennedy, S.. 2012. A nuclear Argonaute promotes multigenerational epigenetic inheritance and germline immortality. *Nature*, 489, 447–451.
- Carlsson, H., Ivimey-Cook, E., Duxbury, E.M.L., Edden, N., Sales, K. & Maklakov, A.A.. 2021. Ageing as “early-life inertia”: disentangling life-history trade-offs along a lifetime of an individual. *Evol. Lett.*, 5, 551–564.
- Caswell, H. & Shyu, E.. 2017. Senescence, selection gradients and mortality. Pp. 56–82 in Shefferson, R.P., Jones, O.R., and Salguero-Gomez, R., eds. *The evolution of senescence in the tree of life*. Cambridge Univ. Press, Cambridge, U.K.
- Chauve, L. et al. 2020. High-throughput quantitative RT-PCR in single and bulk *C. elegans* samples using nanofluidic technology. *J. Vis. Exp.*, 159, e61132.
- Chen, H.-Y., Jolly, C., Bublys, K., Marcu, D. & Immler, S.. 2020. Trade-off between somatic and germline repair in a vertebrate supports the expensive germ line hypothesis. *Proc. Natl. Acad. Sci. USA*, 117, 8973–8979.
- Chen, J., Lewis, E.E., Carey, J.R., Caswell, H. & Caswell-Chen, E.P.. 2006. The ecology and biodemography of *Caenorhabditis elegans*. *Exp. Gerontol.*, 41, 1059–1065
- Chen, J., Lewis, E.E., Carey, J.R., Caswell, H. & Caswell-Chen, E.P.. 2006. The ecology and biodemography of *Caenorhabditis elegans*. *Exp. Gerontol.*, 41, 1059–1065.
- Chen, J., Senturk, D., Wang, J.-L., Muller, H.-G., Carey, J.R., Caswell, H. & Caswell-Chen, E.P.. 2007. A demographic analysis of the fitness cost of extended longevity in *Caenorhabditis elegans*. *J. Gerontol. A Biol. Sci. Med. Sci.*, 62, 126–135.
- Cutter, A. 2005. Mutation and the experimental evolution of outcrossing in *Caenorhabditis elegans*. *J. Evol. Biol.*, 18, 27–34.
- Day, T. & Abrams, P.. 2020. Density dependence, senescence, and Williams’ hypothesis. *Trends Ecol. Evol.*, 35, P300-302.
- de Magalhaes, J.P. 2012. Programmatic features of aging originating in development: aging mechanisms beyond molecular damage? *FASEB J.*, 26, 4821–4826.
- de Magalhaes, J.P. & Church, G.M.. 2005. Genomes optimize reproduction: aging as a consequence of the developmental program. *Physiology*, 20, 252–259.
- Denver, D.R., Wilhelm, L.J., Howe, D.K., Gafner, K., Dolan, P.C. & Baer, C.F.. 2012. Variation in base-substitution mutation in experimental and natural lineages of *Caenorhabditis nematodes*. *Genome Biol. Evol.*, 4, 513–522.
- Dillin, A., Crawford, D.K. & Kenyon, C.. 2002. Timing requirement for insulin/IGF-1 signalling in *C. elegans*. *Science*, 298, 830–834.
- Edward, D.A. & Chapman, T.. 2012. Mechanisms underlying reproductive trade-offs: costs of reproduction. Pp. 137–152 in Flatt, T. and Heyland, A., eds. *Mechanisms of life history evolution: the genetics and physiology of life history traits and trade-offs*. Oxford Univ. Press, Oxford, U.K.
- Ermolaeva, M.A., Segref, A., Dakhovnik, A., Ou, H.-L., Schneider, J.I., Uterm-Hlen, O., Hoppe, T. & Schumacher, B.R.. 2013. DNA damage in germ cells induces immune response triggering systemic stress resistance. *Nature*, 501, 416–420.
- Ezcurra, M., Benedetto, A., Sornda, T., Gilliat, A.F., Au, C., Zhang, Q., van Schelt, S., Petrache, A.L., Wang, H., de la Guardia, Y., et al. 2018. *C. elegans* eats its own intestine to make yolk leading to multiple senescent pathologies. *Curr. Biol.*, 28, 2544–2556.
- Ferguson, E.L. & Horvitz, H.R.. 1985. Identification and characterization of 22 genes that affect the vulval cell lineages of the nematode *Caenorhabditis elegans*. *Genetics*, 110, 17–72.
- Fire, A., Xu, S., Montgomery, M.K., Kostas, S.A., Driver, S.E. & Mello, C.C.. 1998. Potent and specific genetic interference by double-stranded RNA in *Caenorhabditis elegans*. *Nature*, 391, 806–811.
- Flatt, T. 2011. Survival costs of reproduction in *Drosophila*. *Exp. Gerontol.*, 46, 369–375.
- Flatt, T. 2020. Life-history evolution and the genetics of fitness components in *Drosophila melanogaster*. *Genetics*, 214, 3–48.
- Flatt, T., Min, K.-J., D’alterio, C., Villa-Cuesta, E., Cumbers, J., Lehmann, R., Jones, D.n.e. & Tatar, M.. 2008. *Drosophila* germ-line modulation of insulin signalling and lifespan. *Proc. Natl. Acad. Sci. USA*, 105, 6368–6373.
- Flatt, T. & Partridge, L.. 2018. Horizons in the evolution of aging. *BMC Biol.*, 16, 93.
- Fontana, L., Partridge, L. & Longo, V.. 2010. Extending healthy life span - from yeast to humans. *Science*, 328, 321–326.
- Frezal, L. & Felix, M.-A.. 2015. The natural history of model organisms: *C. elegans* outside the Petri dish. *eLife*, 4, e05849.
- Gaillard, J.-M. & Lemaître, J.-F.. 2020. An integrative view of senescence in nature. *Funct. Ecol.*, 34, 4–16.
- Gems, D. & Partridge, L.. 2013. Genetics of longevity in model organisms: debates and paradigm shifts. *Ann. Rev. Physiol.*, 75, 621–644.
- Halligan, D.L. & Keightley, P.D.. 2009. Spontaneous mutation accumulation studies in evolutionary genetics. *Annu. Rev. Ecol. Evol. Syst.*, 40, 151–172.
- Hamilton, W.D. 1966. The moulding of senescence by natural selection. *J. Theor. Biol.*, 12, 12–45.
- Hartig, F. 2020. DHARMA: residual diagnostics for hierarchical (multi-level/mixed) regression models. R package version 0.2.0. <https://cran.r-project.org/web/packages/DHARMA/vignettes/DHARMA.html>.
- Hibshman, J.D., Hung, A. & Baugh, L.R.. 2016. Maternal diet and insulin like signaling control intergenerational plasticity of progeny size and starvation resistance. *PLoS Genet.*, 14, e1007639.
- Ho, J., Tumkaya, T., Aryal, S., Choi, H. & Claridge-Chang, A.. 2019. Moving beyond p values: everyday data analysis with estimation plots. *Nat. Methods*, 16, 565–566.
- Hsin, H. & Kenyon, C.. 1999. Signals from the reproductive system regulate the lifespan of *C. elegans*. *Nature*, 399, 362–366.
- Hsu, A.L., Murphy, C.T. & Kenyon, C.. 2003. Regulation of aging and age-related disease by DAF-16 and heat-shock factor. *Science*, 300, 1142–1145.

- Hyun, M., Lee, J., Lee, K., May, A., Bohr, V.A. & Ahn, B. 2008. Longevity and resistance to stress correlate with DNA repair capacity in *Caenorhabditis elegans*. *Nucleic Acids Res.*, 36, 1380–1389.
- Ivimey-Cook, E.R., Sales, K., Carlsson, H., Immler, S., Chapman, T. & Maklakov, A.A. 2021. Transgenerational fitness effects of lifespan extension by dietary restriction in *Caenorhabditis elegans*. *Proc. R. Soc. B Biol. Sci.*, 288, 20210701.
- Keightley, P.D. & Bataillon, T.M. 2000. Multigeneration maximum-likelihood analysis applied to mutation-accumulation experiments in *Caenorhabditis elegans*. *Genetics*, 154, 1193–1201.
- Kenyon, C.J. 2010. The genetics of ageing. *Nature*, 464, 504–512.
- Kirkwood, T.B.L. 1977. Evolution of ageing. *Nature*, 270, 301–304.
- Kirkwood, T.B.L. 1987. Immortality of the germ-line versus disposability of the soma. Pp. 209–218 in Woodhead, A.D. and Thompson, K.H., eds. *Evolution of longevity in animals: a comparative approach*. Plenum Press, New York.
- Kirkwood, T.B.L. & Holliday, R. 1979. The evolution of ageing and longevity. *Proc. R. Soc. B Biol. Sci.*, 205, 531–546.
- Labbadia, J. & Morimoto, R.I. 2015. Repression of the heat shock response is a programmed event at the onset of reproduction. *Mol. Cell*, 59, 639–650.
- Lind, M.I., Carlsson, H., Duxbury, E.M.L., Ivimey-Cook, E. & Maklakov, A.A. 2021. Cost-free lifespan extension via optimization of gene expression in adulthood aligns with the developmental theory of ageing. *Proc. R. Soc. B Biol. Sci.*, 288, 20201728.
- Lind, M.I., Zwoinska, M.K., Meurling, S., Carlsson, H. & Maklakov, A.A. 2016. Sex-specific trade-offs with growth and fitness following lifespan extension by rapamycin in an outcrossing nematode, *Caenorhabditis remanei*. *J. Gerontol. Ser. A Biol. Sci. Med. Sci.*, 71, 882–890.
- Lind, M.I.L., Ravindran, S., Sekajova, Z., Carlsson, H., Hinas, A. & Maklakov, A.A. 2019. Experimentally reduced insulin/IGF-1 signalling in adulthood extends lifespan of parents and improves Darwinian fitness of their offspring. *Evol. Lett.*, 4, 737–744.
- Lionaki, E. & Tavernarakis, N. 2013. High-throughput and longitudinal analysis of aging and senescent decline in *Caenorhabditis elegans*. Pp. 485–500 in Galluzzi, L., Vitale, I., Kepp, O., and Kroemer, G., eds. *Cell senescence*. Humana Press, Hatfield, U.K.
- Maklakov, A.A. & Chapman, T. 2019. Evolution of ageing as a tangle of trade-offs: energy versus function. *Proc. R. Soc. B Biol. Sci.*, 286, 20191604.
- Maklakov, A.A. & Immler, S. 2016. The expensive germline and the evolution of ageing. *Curr. Biol.*, 26, R577–R586.
- Medawar, P.B. 1952. *An unsolved problem of biology*. H. K. Lewis, Lond.
- Meier, B., Cooke, S.L., Weiss, J., Bailly, A.P., Alexandrov, L.B., Marshall, J., Raine, K., Maddison, M., Anderson, E., Stratton, M.R., et al. 2014. *C. elegans* whole-genome sequencing reveals mutational signatures related to carcinogens and DNA repair deficiency. *Genome Res.*, 24, 1624–1636.
- Meyer, J.N., Boyd, W.A., Azzam, G.A., Haugen, A.C., Freedman, J.H. & Van Houten, B. 2007. Decline of nucleotide excision repair capacity in aging *Caenorhabditis elegans*. *Genome Biol.*, 8, R70.
- Morran, L.T., Cappy, B.J., Anderson, J.L. & Phillips, P.C. 2009. Sexual partners for the stressed: facultative outcrossing in the self-fertilizing nematode *Caenorhabditis elegans*. *Evolution*, 63, 1473–1482.
- Muñoz, M.J. 2003. Longevity and heat stress regulation in *Caenorhabditis elegans*. *Mech. Ageing Dev.*, 124, 43–48.
- Ni, J.Z., Kalinava, N., Chen, E., Huang, A., Trinh, T. & Gu, S.G. 2016. A transgenerational role of the germline nuclear RNAi pathway in repressing heat stress-induced transcriptional activation in *C. elegans*. *Epigenetics Chromatin*, 9, 3.
- Nigon, V. & Dougherty, E.C. 1949. Reproductive patterns and attempts at reciprocal crossing of *Rhabditis elegans* Maupas, 1900, and *Rhabditis briggsae* Dougherty and Nigon, 1949 (Nematoda: Rhabditidae). *J. Exp. Zool.*, 112, 485–503.
- Posner, R., Toker, I.A., Antonova, O., Star, E., Anava, S., Azmon, E., Hendricks, M., Bracha, S., Gingold, H. & Rechavi, O. 2019. Neuronal small RNAs control behaviour transgenerationally. *Cell*, 177, 1814–1826.
- R Core Team. 2020. R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna.
- Rechavi, O., Houriz-Zeevi, L., Anava, S., Goh, W.S.S., Kerk, S.Y., Hannon, G.J. & Hobert, O. 2014. Starvation-induced transgenerational inheritance of small RNAs in *C. elegans*. *Cell*, 158, 277–287.
- Regan, J.C., Froy, H., Walling, C.A., Moatt, J.P. & Nussey, D.H. 2019. Dietary restriction and insulin-like signalling pathways as adaptive plasticity: a synthesis and re-evaluation. *Funct. Ecol.*, 34, 107–128.
- Roff, D. 2002. *Life history evolution*. Sinauer Associates, Sunderland, MA.
- Sala, A.J., Bott, L.C., Brielmann, R.M. & Morimoto, R.I. 2020. Embryo integrity regulates maternal proteostasis and stress resilience. *Genes Dev.*, 34, 1–10.
- Scharf, A., Pohl, F., Egan, B.M., Kocsisova, Z. & Kornfield, K. 2021. Reproductive aging in *Caenorhabditis elegans*: from molecules to ecology. *Front. Cell Dev. Biol.*, 9, 718522.
- Schulenburg, H. & Felix, M.-A. 2017. The natural biotic environment of *Caenorhabditis elegans*. *Genetics*, 206, 55–86.
- Seo, K., Choi, E., Lee, D., Jeong, D.-E., Jang, S.K. & Lee, S.-J. 2013. Heat shock factor 1 mediates the longevity conferred by inhibition of TOR and insulin/IGF-1 signaling pathways in *C. elegans*. *Aging Cell*, 12, 1073–1081.
- Shemesh, N., Shai, N. & Ben-Zvi, A. 2013. Germline stem cell arrest inhibits the collapse of somatic proteostasis early in *Caenorhabditis elegans* adulthood. *Aging Cell*, 12, 814–822.
- Smelick, C. & Ahmed, S. 2005. Achieving immortality in the *C. elegans* germline. *Ageing Res. Rev.*, 4, 67–82.
- Sniegowski, P.D., Gerrish, P.J., Johnson, T. & Shaver, A. 2000. The evolution of mutation rates: separating causes from consequences. *BioEssays*, 22, 1057–1066.
- Spracklin, G., Fields, B., Wan, G., Becker, D., Wallig, A., Shukla, A. & Kennedy, S. 2017. The RNAi inheritance machinery of *Caenorhabditis elegans*. *Genetics*, 206, 1403–1416.
- Stearns, S.C. 1992. *The evolution of life histories*. Oxford Univ. Press, Oxford, U.K.
- Stergiou, L., Doukoumetzidis, K., Sandoel, A. & Hengartner, M.O. 2007. The nucleotide excision repair pathway is required for UV-C-induced apoptosis in *Caenorhabditis elegans*. *Cell Death Differ.*, 14, 1129–1138.
- Sternberg, P.W. & Horvitz, H.R. 1989. The combined action of two intercellular signalling pathways specifies three cell fates during vulval induction in *C. elegans*. *Cell*, 58, 679–693.
- Therneau, T.M. 2019. coxme: mixed effects cox models. R package version 2.2-14.
- Thondamal, M., Witting, M., Schmitt-Kopplin, P. & Aguilani, H. 2014. Steroid hormone signalling links reproduction to lifespan in dietary-restricted *Caenorhabditis elegans*. *Nat. Commun.*, 5, 4879.
- Timmons, L., Court, D.L. & Fire, A. 2001. Ingestion of bacterially expressed dsRNAs can produce specific and potent genetic interference in *Caenorhabditis elegans*. *Gene*, 263, 103–112.
- Vassilieva, L.L., Hook, A.M. & Lynch, M. 2000. The fitness effects of spontaneous mutations in *Caenorhabditis elegans*. *Evolution*, 54, 1234–1246.

Wang, H., Zhao, Y., Ezcurra, M., Benedetto, A., Gilliat, A.F., Hellberg, J., Ren, Z., Galimov, E.R., Athigapanich, T., Girstmair, J., et al. 2018. A parthenogenetic quasi-program causes teratoma-like tumors during aging in wild-type *C. elegans*. *NPJ Aging Mech. Dis.*, 4, 6.

Williams, G.C. 1957. Pleiotropy, natural selection, and the evolution of senescence. *Evolution*, 11, 398–411.

Zevian, S.C. & Yanowitz, J.L.. 2014. Methodological considerations for heat shock of the nematode *Caenorhabditis elegans*. *Methods*, 68, 450–457.

Associate Editor: Prof. Damian K Dowling
Handling Editor: Dr. Andrew G McAdam

Supporting Information

Additional supporting information may be found online in the Supporting Information section at the end of the article.

Supplementary Material