

# Excited-State Barrier Controls $E \rightarrow Z$ Photoisomerization in $p$ -Hydroxycinnamate Biochromophores

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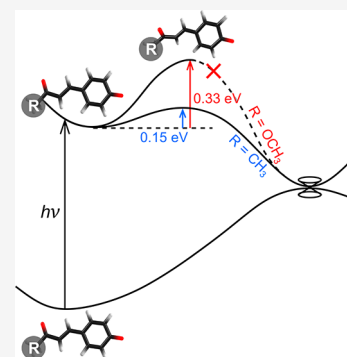


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**ABSTRACT:** Molecules based on the deprotonated  $p$ -hydroxycinnamate moiety are widespread in nature, including serving as UV filters in the leaves of plants and as the biochromophore in photoactive yellow protein. The photophysical behavior of these chromophores is centered around a rapid  $E \rightarrow Z$  photoisomerization by passage through a conical intersection seam. Here, we use photoisomerization and photodissociation action spectroscopies with deprotonated 4-hydroxybenzal acetone ( $pCK^-$ ) to characterize a wavelength-dependent bifurcation between electron autodetachment (spontaneous ejection of an electron from the  $S_1(\pi\pi^*)$  state because it is situated in the detachment continuum) and  $E \rightarrow Z$  photoisomerization. While autodetachment occurs across the entire  $S_1(\pi\pi^*)$  band (370–480 nm),  $E \rightarrow Z$  photoisomerization occurs only over a blue portion of the band (370–430 nm). No  $E \rightarrow Z$  photoisomerization is observed when the ketone functional group in  $pCK^-$  is replaced with an ester or carboxylic acid. The wavelength-dependent bifurcation is consistent with potential energy surface calculations showing that a barrier separates the Franck–Condon region from the  $E \rightarrow Z$  isomerizing conical intersection. The barrier height, which is substantially higher in the gas phase than in solution, depends on the functional group and governs whether  $E \rightarrow Z$  photoisomerization occurs more rapidly than autodetachment.



Molecules possessing the  $p$ -hydroxycinnamate moiety are widespread in nature.<sup>1</sup> Examples include sinapoyl malate, caffeic acid, and ferulic acid in both the free form and covalently bound to cell walls and lignin structures, which are present in the leaves, stems, and seeds of plants where they function as UV-B filters.<sup>2</sup> The UV-B filtering mechanism is, in part, thought to rely on a rapid internal conversion to the ground state, accompanied by  $E \rightarrow Z$  photoisomerization.<sup>3</sup> The efficacy of this nonradiative decay has prompted the skin-care industry to develop commercial sunscreens containing cinnamate-based molecules.<sup>4,5</sup> In another biological context, deprotonated  $p$ -hydroxycinnamates are invoked as models for the chromophore in photoactive yellow protein (PYP), which is a small blue-light sensing protein found in the *Halorhodospira halophila* bacterium.<sup>6–8</sup> In the PYP photocycle, absorption of blue light by a thioester-based hydroxycinnamate chromophore leads to an  $E \rightarrow Z$  photoisomerization of the chromophore, which in turn leads to a change in protein conformation and eventually a negative phototaxis response of the bacterium.<sup>9–12</sup>

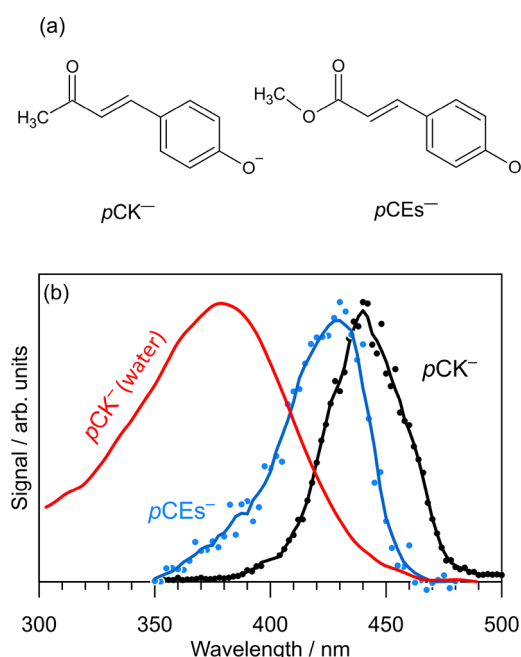
A desire to understand the photophysics of  $p$ -hydroxycinnamates and to develop synthetic derivatives that might be incorporated into optogenetic applications<sup>13–15</sup> or skin-care products<sup>16</sup> has prompted numerous investigations on the inherent photophysics in this class of molecules. Although the excited-state dynamics in  $p$ -hydroxycinnamates has been studied extensively in solution over the past two decades (see, for example, refs 17–22 and references therein), the extent to which solvation perturbs the intrinsic excited-state

dynamics remains unclear. Theoretical investigations have suggested that solvation of anionic  $p$ -hydroxycinnamates significantly perturbs the  $S_1(\pi\pi^*)$  potential energy surfaces and conical intersection seams,<sup>23–29</sup> although experimental strategies capable of directly observing photoisomerization in the gas phase are now starting to emerge.<sup>30–32</sup>

Previous experiments on hydroxycinnamate anions, focusing on the inherent dynamics, utilized techniques including time-resolved photoelectron spectroscopy,<sup>33–35</sup> frequency-resolved photoelectron spectroscopy to fingerprint internal conversion dynamics,<sup>36</sup> and photoisomerization action (PISA) spectroscopy to select precursor deprotonated or geometric isomers and to probe photoisomerization or phototautomerization.<sup>32,37</sup> However, these studies were unable to provide any evidence for  $E \rightarrow Z$  photoisomerization across a series of around 20 hydroxycinnamate anions. The present study provides clear evidence for an  $E \rightarrow Z$  photoisomerization response in deprotonated 4-hydroxybenzal acetone ( $pCK^-$ , Figure 1a), a molecule which has been invoked as a proxy for the PYP chromophore.<sup>18,24,26,33,35,38</sup> Importantly, the  $E \rightarrow Z$  photoisomerization response occurs only following excitation of the

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**Figure 1.** (a)  $p$ -Hydroxycinnamate anions considered in this study. (b) Action spectra for  $pCK^-$  (photodissociation, black) and  $pCEs^-$  (photodetachment, blue, from ref 32) as proxies for the  $S_1 \leftarrow S_0$  absorption bands. The absorption spectrum of  $pCK^-$  in water (at  $T = 300$  K) is shown in red. Solid lines are moving averages over three data points for the gas phase and seven data points for the condensed phase data.

higher photon energy region of the  $S_1 \leftarrow S_0$  absorption band, consistent with earlier molecular dynamics simulations hypothesizing that photoisomerization by passage through a conical intersection is a barrier-controlled process.<sup>39–41</sup> Our study provides experimental confirmation that functional group substitution on the hydroxycinnamate tail critically affects the excited-state barrier height and thus photoisomerization efficacy.<sup>24,26,42</sup> Comparison of time-resolved data for  $pCK^-$  in solution with gas phase data implies that the potential energy surface barrier to isomerization is stabilized in solution.

A photodissociation action spectrum (black), which serves as a proxy for the visible absorption spectrum of  $pCK^-$ , is shown in Figure 1b. The spectrum was recorded by monitoring absorption-induced fragmentation of the anion under ultra-high-vacuum conditions (see the Supporting Information for experimental details). The spectrum spans the 390–480 nm range with maximum response at 440 nm. The spectrum is red-shifted by  $\approx 10$  nm compared with the photodetachment action spectrum for the methyl ester ( $pCEs^-$ ) from ref 32 (see also ref 43), corresponding to absorption-induced electron ejection. The origin of the red-shift for  $pCK^-$  is presumably due to differences in inductive electron donation. The absorption spectrum of  $pCK^-$  in water is blue-shifted by  $\approx 80$  nm relative to the photodissociation spectrum.

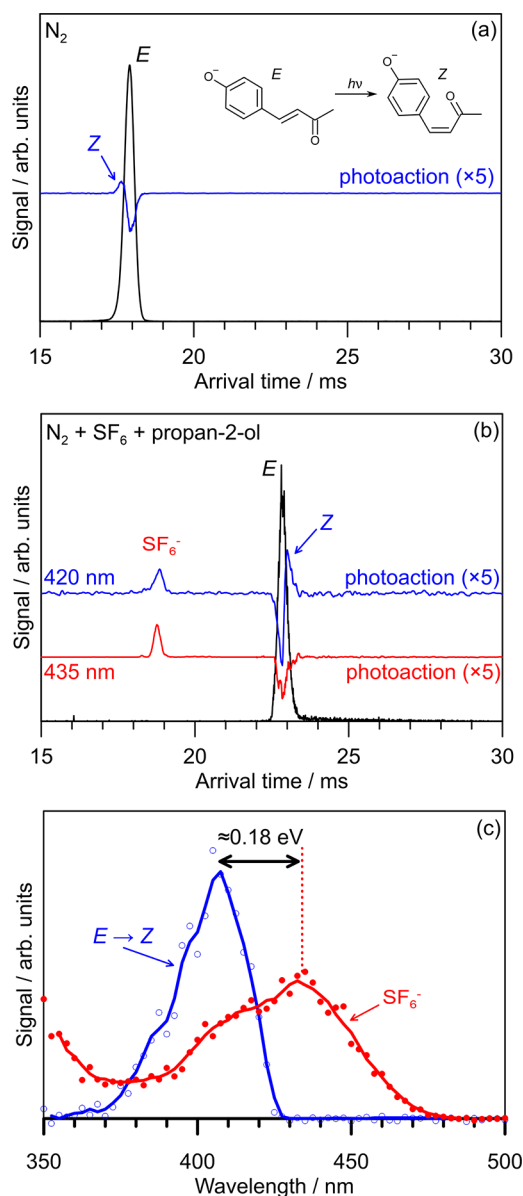
Photoisomerization of isolated  $pCK^-$  was investigated using the emerging technique of PISA spectroscopy. A detailed description and illustration of the PISA spectroscopy technique are available in refs 30 and 44. Briefly, PISA spectroscopy allows for isomer-selected irradiation experiments, isomer-specific product detection, and quantification of photo-detached electrons for anions using an electron scavenger ( $SF_6$ ).<sup>32,44</sup> In an experiment, charged isomers that are drifting under the influence of an electric field through a buffer gas

(e.g.,  $N_2$  or  $CO_2$ ) are separated according to their drift speeds, which depend on their collision cross sections. The target isomer is selected in a primary drift stage and then exposed to wavelength tunable light, with separation of photoisomers or photofragments in a second drift stage. By monitoring the yield of photoisomers and  $SF_6^-$  as a function of wavelength, photoisomerization and photodetachment action spectra are recorded. Complete experimental details are given in the Supporting Information.

Electrospray ionization of  $pCK^-$  in any of the buffer gases considered in this study produced a single arrival time distribution (ATD) peak, consistent with a single isomer (black traces in Figure 2a,b) assigned to the  $E$  configuration. In pure  $N_2$  buffer gas, the photoaction ATD in Figure 2a, corresponding to the difference between “light on” and “light off” ATDs, shows generation of a photoisomer at a slightly shorter arrival time, consistent with the  $Z$  isomer, since cross-section modeling predicts that the  $Z$  isomer has a smaller collision cross section in pure  $N_2$  (Table 1). Similar results were obtained in pure  $CO_2$  buffer gas (see the Supporting Information). The photoaction ATD in  $N_2$  buffer gas doped with  $\approx 1\%$   $SF_6$  and  $\approx 1\%$  propan-2-ol shows generation of a photoisomer at longer arrival time (assigned to  $Z$ ) and electron detachment as detected through  $SF_6^-$  formation when using 420 nm light. Further explanation on isomer-specific interactions with propan-2-ol leading to the increased collision cross section for the  $Z$  isomer compared with the  $E$  isomer is given in the Supporting Information. Because the photo-depletion signal (i.e., bleach of the  $E$  isomer) in Figure 2b is balanced by the sum of photoisomerization and electron detachment signals, the experiment captures all prompt photoaction. This correspondence is true across all wavelengths considered in this study. Notably, there is no photodissociation in this experiment because collisional energy quenching (tens to hundreds of nanoseconds) occurs more rapidly than recovery of the ground electronic state followed by statistical dissociation (microseconds).<sup>32,45</sup>

Photodetachment (red) and photoisomerization (blue) action spectra for  $pCK^-$  are shown in Figure 2c. While electron detachment is observed across the 370–480 nm range with maximum response at  $\approx 435$  nm,  $E \rightarrow Z$  isomerization was observed only over the 360–430 nm range with maximum response at  $\approx 405$  nm. These spectra span the same wavelength range as the photodissociation spectrum (proxy for the absorption spectrum) in Figure 1b, although differing in shape, which is indicative of competitive photochemical pathways. It is worth noting that electron detachment may occur following absorption of a single photon because the onset of the action spectra is situated above the adiabatic detachment energy (Table 1) when allowing for the internal energy associated with temperature of the ions at  $T = 300$  K (0.3 eV). This situation is also true for  $pCEs^-$ .<sup>32,34,36,37</sup>

In PISA spectroscopy, an isomerization signal can result from two mechanisms: (i) a rapid excited-state process associated with passage through a conical intersection and (ii) statistical isomerization on the ground electronic state before collisions in the drift region thermalize the activated ions. In an earlier study considering  $pCEs^-$ ,<sup>32</sup> we used master equation simulations combining RRKM isomerization rates with Langevin collisional energy quenching to explore the possibility of process ii, where it was concluded to be unlikely because of the electronic energy difference between the two isomers, although some  $Z \rightarrow E$  thermal reversion may occur



**Figure 2.** Action spectroscopy of  $pCK^-$ : (a) light-off (black) and photoaction (blue) ATD at 420 nm in pure  $N_2$  buffer gas; (b) light-off (black) and photoaction (blue, 420 nm and red, 435 nm) ATD in  $N_2$  buffer gas seeded with  $\approx 1\%$  propan-2-ol and  $\approx 1\%$   $SF_6$ ; (c) electron photodetachment (red) and  $E \rightarrow Z$  photoisomerization (blue) action spectra. The photoaction spectra show the changes between light-on and light-off ATDs, reflecting any photoinduced processes. The photoisomerization quantum yield is estimated at a 1–2% at 400 nm. See the Supporting Information for  $CO_2$  buffer gas data. The excited-state barrier to isomerization is estimated at  $\approx 0.18$  eV from the difference in spectral maxima in (c); use of thresholds is not reliable because of hot bands and the direct photodetachment contribution to electron detachment because the  $S_1$  state is situated in the detachment threshold.

before collisions stabilize the isomers. To investigate thermal reversion (process ii) for  $pCK^-$ , which may skew the appearance of the photoisomerization action spectra, we considered an experimental approach in which the ion mobility experiments were repeated in  $CO_2$  buffer gas (see the Supporting Information). The rationale is that the vibrational energy quenching collision cross section for  $CO_2$  is an order of magnitude larger than that for  $N_2$ ,<sup>49</sup> providing more rapid

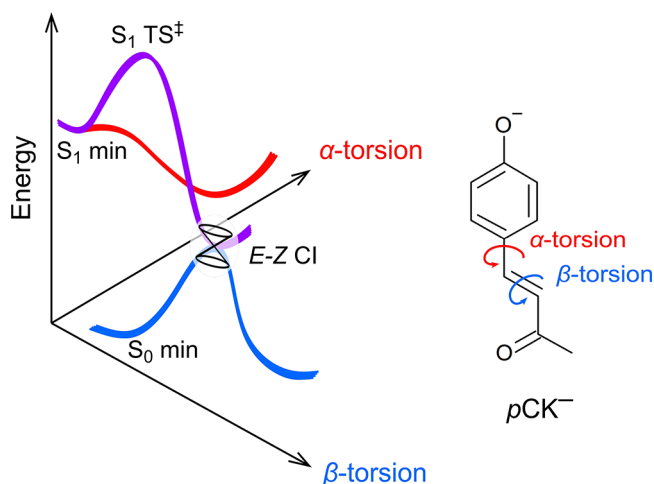
**Table 1.** Calculated Properties for the  $E$  and  $Z$  Isomers of  $pCK^-$

species	$\Delta E^a$	ADE <sup>a</sup>	VDE <sup>a</sup>	$\Omega_c$
( $E$ )- $pCK^-$	0	2.83	2.90	137
( $Z$ )- $pCK^-$	27	2.80	2.86	136
TS <sup>b</sup>	125			
expt		$2.8 \pm 0.1^c$	$3.0 \pm 0.1^c$	$134 \pm 5^d$

<sup>a</sup> $\Delta E$  in units of  $kJ\ mol^{-1}$ ; ADE (adiabatic detachment energy) and VDE (vertical detachment energy) in units of eV. <sup>b</sup>TS is the isomerization transition state on the ground electronic state. All energies at the DLPNO-CCSD(T)/aug-cc-pVTZ level of theory using ORCA 5.0.3.<sup>46</sup>  $\Omega_c$  in units of  $\text{\AA}^2$ , calculated using MOBCAL.<sup>47,48</sup> <sup>c</sup>Reference 35. <sup>d</sup>( $E$ )- $pCK^-$ .

thermalization and suppression of ground-state statistical processes. Because the action spectra in  $CO_2$  buffer gas closely resemble those shown in Figure 2c, it is unlikely that  $Z \rightarrow E$  thermal reversion processes have a significant bearing on the action spectra.

The occurrence of  $E \rightarrow Z$  photoisomerization for  $pCK^-$  in the gas phase contrasts with  $pCEs^-$  and derivatives such as the phenoxide deprotonomer of  $p$ -coumaric acid and ring-substituted derivatives caffeic, ferulic, and sinapinic acid (and methyl esters of each), for which no  $E \rightarrow Z$  isomerization was observed.<sup>32,37</sup> Earlier molecular dynamics simulations and related studies have suggested that there is a barrier on the  $S_1$  state potential energy surface for double-bond rotation ( $\beta$ -torsion coordinate in Figure 3a).<sup>39–41</sup> A recent study of  $pCK^-$  calculated a barrier



**Figure 3.** Schematic illustration of potential energy surfaces for the  $E$  isomer of  $pCK^-$  showing the  $\alpha$  and  $\beta$  coordinates and identifying  $S_0$  and  $S_1$  minimum-energy geometries, the  $\beta$ -coordinate transition state ( $S_1$  TS<sup>‡</sup>), and the  $E$ - $Z$  minimum-energy conical intersection (CI). Calculated energies for these critical points are given in Table 2. The  $\alpha$  coordinate has been considered in ref 35.

of  $\approx 0.25$  eV along the  $\beta$ -torsion coordinate,<sup>35</sup> although this value is based on linear interpolation of internal coordinates between the Franck–Condon geometry and the double-bond twisted minimum-energy structure. This value is lower than the earlier calculated barrier of  $\approx 0.4$  eV, relative to the Franck–Condon geometry, found for  $pCEs^-$  using the DLPNO-STEOM-CCSD/aug-cc-pVDZ method.<sup>34</sup> To enable a robust comparison between  $pCK^-$  and  $pCEs^-$ , we optimized the  $\beta$ -torsion critical points along the  $S_1$  potential energy surfaces (Figure 3) using a CASSCF(10,9) wave function followed by

XMCQDPT2 energy calculations (Table 2) using the Firefly 8.2.0 software package.<sup>50</sup> These calculations gave a barrier of

**Table 2.** Calculated Potential Energy Surface Critical Points in eV at the XMCQDPT2(10,9)/aug-cc-PVDZ Level of Theory for  $pCK^-$  and  $pCEs^-$

	$pCK^-$	$pCEs^-$
VEE <sup>a</sup>	2.87	2.96
$S_1$ min	2.69	2.85
$S_1$ TS <sup>b</sup>	0.15	0.33
E–Z CI	2.62	2.82

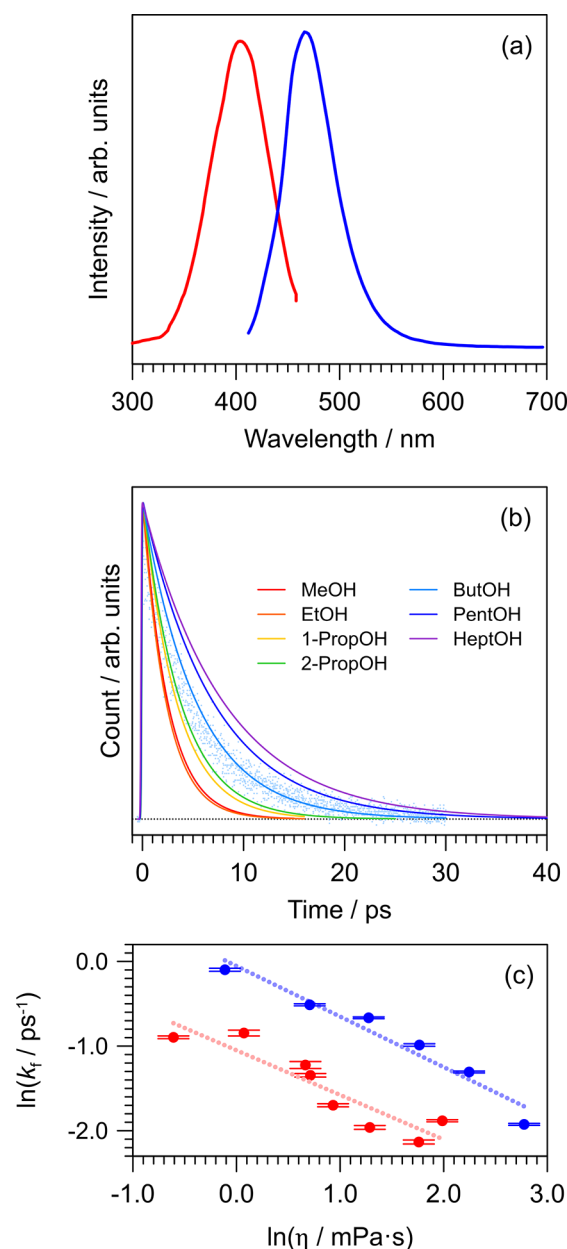
<sup>a</sup>VEE = vertical excitation energy. <sup>b</sup>Relative to  $S_1$  min. Values relative to the Franck–Condon geometry are given in the text.

0.15 eV for  $pCK^-$  and 0.33 eV for  $pCEs^-$ , with torsion of the  $\beta$  coordinate of  $53.5^\circ$  ( $pCK^-$ ,  $-238\text{ cm}^{-1}$ ) and  $49.1^\circ$  ( $pCEs^-$ ,  $-325\text{ cm}^{-1}$ ) at the transition state. Significantly, the computed barrier for  $pCK^-$  is in good agreement with experiment ( $\approx 0.18\text{ eV}$ ) and is substantially lower than that for  $pCEs^-$ .

Having established the intrinsic  $E \rightarrow Z$  photoisomerization response for  $pCK^-$  at  $T = 300\text{ K}$ , we next considered excited-state lifetimes. Following rapid geometric relaxation from the Franck–Condon geometry ( $\tau_1 < 1\text{ ps}$ ), gas phase lifetimes for  $pCK^-$  have been determined to be  $\tau_2 = 52\text{ ps}$  when pumped at  $400\text{ nm}$ <sup>33</sup> and  $\approx 120\text{ ps}$  when pumped at  $444\text{ nm}$ ,<sup>35</sup> with the latter corresponding to exciting near the absorption band maximum. While the  $400\text{ nm}$  study observed ground-state recovery (and assumed isomerization), the  $444\text{ nm}$  study observed only autodetachment (spontaneous ejection of an electron from the  $S_1$  state because it is situated in the detachment continuum) and predicted  $\alpha$ -torsion dynamics based on photoelectron angular distributions. The present action spectra shown in Figure 2c confirm that both time-resolved studies reached correct conclusions: excitation at  $400\text{ nm}$  leads to isomerization while excitation at  $444\text{ nm}$  does not. These dynamics contrast with  $pCEs^-$ , which, when pumped near the maximum in its absorption band ( $438\text{ nm}$ ), decays exclusively thorough autodetachment with a lifetime of  $45 \pm 4\text{ ps}$ .<sup>34</sup> Similar autodetachment processes have been fingerprinted over the entire absorption band for  $pCEs^-$  with no evidence for internal conversion and thus the possibility of  $Z$  isomer formation.<sup>36</sup> The longer excited-state lifetime for  $pCK^-$  compared with  $pCEs^-$  when pumped near the maximum in the photodissociation action spectrum is presumably because, while the two anions have similar electron detachment thresholds, the absorption profile for  $pCK^-$  is red-shifted by  $\approx 0.1\text{ eV}$  (Figure 1), decreasing the propensity for electron autodetachment following rapid nuclear relaxation away from the Franck–Condon geometry.

To facilitate a comparison of gas phase excited-state lifetimes with those in solution, we performed time-resolved fluorescence upconversion ( $\approx 50\text{ fs}$  time resolution) on  $pCK^-$  dissolved in a series of polar solvents at  $T = 300\text{ K}$ .<sup>51,52</sup> Fluorescence upconversion is a time-resolved spectroscopic technique in which the fluorescence emission from a sample is frequency mixed with a probe laser pulse ( $800\text{ nm}$ ), producing an “upconverted” signal. By changing the delay between femtosecond pump and probe pulses, and monitoring the upconverted signal, fluorescence lifetimes are measured. The present upconversion measurements refine an earlier solvent polarity and viscosity upconversion study on  $pCK^-$ <sup>18</sup> and were performed because the earlier study (i) was limited by  $\approx 500\text{ fs}$

time resolution, (ii) used an excitation wavelength ( $340\text{ nm}$ ), which is far from the absorption maximum and likely accesses a  $n\pi^*$  state that gains substantial intensity through Herzberg–Teller coupling,<sup>36</sup> and (iii) assumed static samples that likely gave rise to photostationary states. Steady-state fluorescence excitation and emission spectra for  $pCK^-$  in water are shown in Figure 4a, revealing a large Stokes shift of  $5794 \pm 20\text{ cm}^{-1}$  ( $4143 \pm 20\text{ cm}^{-1}$  in ethanol; see further data in the Supporting Information); the large shift is in part attributed to hydrogen-bond interactions between the phenoxide group and solvent



**Figure 4.** Fluorescence spectroscopy of  $pCK^-$  in solution at  $T = 300\text{ K}$ : (a) excitation (red, monitoring at  $480\text{ nm}$ ) and emission (blue, exciting at  $400\text{ nm}$ ) fluorescence spectra in water; (b) time-resolved fluorescence upconversion decay curves and model fits in a series of alcohols (fitted values are given in the Supporting Information). Experimental data points are shown for ButOH; (c) viscosity ( $\eta$ ) effect of  $k_f \approx \frac{1}{\tau_2}$  in a series of alcohols (red) and water–ethylene glycol mixtures (blue).



molecules weakening upon excitation.<sup>17,19,41</sup> The Stokes shift at  $T = 77$  K is significantly lower at  $2243 \pm 20$  cm<sup>-1</sup> (ethanol), consistent with inhibition of nuclear and/or solvent relaxation to reach the lowest energy fluorescing geometry.

Fluorescence upconversion data following excitation of  $pCK^-$  with 400 nm light, which were measured in a flow cell at  $T = 300$  K, are shown in Figure 4b; results for water–ethylene-glycol mixtures are given in the Supporting Information. The decay curves were fit with a two-component exponential decay model with lifetime  $\tau_1$  dominated by rapid solvent rearrangement (limited by the cross correlation), and  $\tau_2$  is linked to the excited-state lifetime and associated solvent motion, i.e., convoluted with the  $\approx 880$  fs longest time scale dynamics for water rearrangement.<sup>53</sup> Fitted lifetimes in selected solvents are given in Table 3 (see all data in the

**Table 3. Fitted Excited-State Lifetimes ( $\tau_2$  in ps) for  $pCK^-$  in Water and Alcohol Solvents at  $T = 300$  K**

species	$\tau_2$	$\pm$
water	1.17	0.01
MeOH	2.45	0.02
EtOH	2.33	0.04
1-PropOH	3.40	0.07
2-PropOH	3.84	0.04
ButOH	5.47	0.05
PentOH	7.11	0.08
HeptOH	8.44	0.10
OctOH	6.57	0.04

Supporting Information). It is worth noting that the  $\approx 1$  ps lifetime of  $pCK^-$  in water is comparable with the time scale for  $E \rightarrow Z$  photoisomerization of the chromophore in PYP.<sup>11,54</sup> It is striking that the excited-state lifetimes for  $pCK^-$  in solution are 1 or 2 orders of magnitude shorter than in the gas phase, suggesting a considerable reduction of the isomerization barrier in solution or access of an alternative relaxation pathway. This situation contrasts with anionic retinoids in the gas phase that undergo barrier-controlled stereospecific  $E \rightarrow Z$  photoisomerization and have considerably shorter lifetimes than in solution.<sup>55</sup> Studies of derivative hydroxycinnamate chromophores in solution have shown that the excited-state lifetime is sensitive to the identity of the functional group on the carbonyl tail,<sup>19</sup> with the ketone group for  $pCK^-$  giving rise to the shortest lifetimes, although an overall picture is complicated because of solvent polarity effects, differences in charge-transfer character, and hydrogen bonding.<sup>20</sup>

The influence of viscosity on the excited-state lifetime of  $pCK^-$  is shown in Figure 4c revealing a strong effect, consistent with an isomerization-type reaction. See the Supporting Information for the solvent polarity effect. Following ref 18, excited-state lifetimes as a function of viscosity were fit with the phenomenological power law  $k_f \approx \frac{1}{\tau_2} = C\eta^{-\alpha}$ ,<sup>56</sup> where  $k_f$  is assumed as the photoisomerization rate and  $C$  is proportional to the Arrhenius term  $e^{-E_a/kT}$  and is linked to polarity dependence (stabilization) of the transition state. The parameter  $\alpha$  is a measure of the viscosity effect for isomerization, which approaches unity in highly viscous solvents.<sup>57</sup> Fitted values of  $\alpha$  are 0.53 and 0.59 for the alcohols and water–ethylene glycol mixtures, respectively, with the latter being slightly larger than that reported in ref 18. Assuming an Arrhenius relation at  $T = 300$  K and a pre-

exponential factor of  $\approx 10^{13}$  s<sup>-1</sup>, the excited-state barrier height in water is  $\approx 0.07$  eV, which is around half of the gas phase value. This estimate is consistent with potential energy surface calculations on the dianion of  $p$ -coumaric acid with microhydration, showing a reduction of the barrier from 0.70 eV (gas phase) to 0.09 eV. Values of  $\alpha$  have been determined at 0.64 for  $pCEs^-$ <sup>18</sup> and 0.75 for the thioester anion,<sup>17</sup> consistent with  $pCK^-$  having the lowest isomerization barrier in solution. We conclude that solvation significantly stabilizes the barrier to isomerization.

In summary, this study has demonstrated that  $E \rightarrow Z$  photoisomerization of a  $p$ -hydroxycinnamate anion may occur in the gas phase, although a barrier to double-bond torsion on the  $S_1(\pi\pi^*)$  potential energy surface is a key factor in defining if photoisomerization is competitive with electron autodetachment. Substitution on the carbonyl group tunes the barrier height separating the Franck–Condon geometry and the  $E \rightarrow Z$  isomerizing conical intersection seam. Solvation of the chromophore significantly stabilizes the excited-state barrier, leading to rapid nonradiative relaxation. The present experimental strategy is applicable to other charged systems that may photoisomerize and possess excited-state barriers, and is particularly applicable to systems for which there are wavelength-dependent dynamics leading to multiple isomeric products.<sup>55</sup> Future work on deprotonated hydroxycinnamate anions will seek to photogenerate, isolate, and apply frequency and time-resolved action spectroscopy techniques to  $Z$  isomers, as well as other unstable or intermediate isomers such as *keto–enol* tautomers,<sup>32</sup> to map out excited-state potential energy surfaces.

## ■ ASSOCIATED CONTENT

### Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.jpcllett.2c02613>.

Experimental methods; theoretical methods; CASSCF orbitals; vertical excitation energies; action spectroscopy in CO<sub>2</sub> buffer gas;  $pCK^-$ -propan-2-ol complexes; solution spectroscopy; critical point geometries (PDF)

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## Notes

The authors declare no competing financial interest.

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## REFERENCES

- (1) Pei, K.; Ou, J.; Huang, J.; Ou, S. *p*-Coumaric Acid and its Conjugates: Dietary Sources, Pharmacokinetic Properties and Biological Activities. *J. Sci. Food Agric.* **2016**, *96*, 2952–2962.
- (2) Baker, L. A.; Horbury, M. D.; Greenough, S. E.; Allais, F.; Walsh, P. S.; Habershon, S.; Stavros, V. G. Ultrafast Photoprotecting Sunscreens in Natural Plants. *J. Phys. Chem. Lett.* **2016**, *7*, 56–61.
- (3) Baker, L. A.; Marchetti, B.; Karsili, T. N. V.; Stavros, V. G.; Ashfold, M. N. R. Photoprotection: Extending Lessons Learned From Studying Natural Sunscreens to the Design of Artificial Sunscreen Constituents. *Chem. Soc. Rev.* **2017**, *46*, 3770–3791.
- (4) Holt, E. L.; Stavros, V. G. Applications of Ultrafast Spectroscopy to Sunscreen Development, From First Principles to Complex Mixtures. *Int. Rev. Phys. Chem.* **2019**, *38*, 243–285.
- (5) Kinoshita, S.; Harabuchi, Y.; Inokuchi, Y.; Maeda, S.; Ehara, M.; Yamazaki, K.; Ebata, T. Substitution Effect on the Nonradiative Decay and *trans* → *cis* Photoisomerization Route: A Guideline to Develop Efficient Cinnamate-Based Sunscreens. *Phys. Chem. Chem. Phys.* **2021**, *23*, 834–845.
- (6) Meyer, T. Isolation and Characterization of Soluble Cytochromes, Ferredoxins and Other Chromophoric Proteins From the Halophilic Phototrophic Bacterium *Ectothiorhodospira halophila*. *Biochim. Biophys. Acta - Bioenergetics* **1985**, *806*, 175–183.
- (7) Meyer, T. E.; Yakali, E.; Cusanovich, M. A.; Tollin, G. Properties of a Water-Soluble, Yellow Protein Isolated From a Halophilic Phototrophic Bacterium that has Photochemical Activity Analogous to Sensory Rhodopsin. *Biochem* **1987**, *26*, 418–423.
- (8) Sprenger, W. W.; Hoff, W. D.; Armitage, J. P.; Hellingwerf, K. J. The Eubacterium *Ectothiorhodospira halophila* is Negatively Photo-tactic, With a Wavelength Dependence that Fits the Absorption Spectrum of the Photoactive Yellow Protein. *J. Bacteriol.* **1993**, *175*, 3096–3104.
- (9) van Wilderen, L. J. G. W.; van der Horst, M. A.; van Stokkum, I. H. M.; Hellingwerf, K. J.; van Grondelle, R.; Groot, M. L. Ultrafast Infrared Spectroscopy Reveals a Key Step For Successful Entry Into the Photocycle For Photoactive Yellow Protein. *Proc. Nat. Acad. Sci.* **2006**, *103*, 15050–15055.
- (10) Tenboer, J.; Basu, S.; Zatsopin, N.; Pande, K.; Milathianaki, D.; Frank, M.; Hunter, M.; Boutet, S.; Williams, G. J.; Koglin, J. E.; et al. Time-Resolved Serial Crystallography Captures High-Resolution Intermediates of Photoactive Yellow Protein. *Science* **2014**, *346*, 1242–1246.
- (11) Pande, K.; Hutchison, C. D. M.; Groenhof, G.; Aquila, A.; Robinson, J. S.; Tenboer, J.; Basu, S.; Boutet, S.; DePonte, D. P.; Liang, M.; et al. Femtosecond Structural Dynamics Drives the *trans/cis* Isomerization in Photoactive Yellow Protein. *Science* **2016**, *352*, 725–729.
- (12) Kuramochi, H.; Takeuchi, S.; Yonezawa, K.; Kamikubo, H.; Kataoka, M.; Tahara, T. Probing the Early Stages of Photoreception in Photoactive Yellow Protein With Ultrafast Time-Domain Raman Spectroscopy. *Nat. Chem.* **2017**, *9*, 660–666.
- (13) Losi, A.; Gardner, K. H.; Möglich, A. Blue-Light Receptors for Optogenetics. *Chem. Rev.* **2018**, *118*, 10659–10709.
- (14) Jang, J.; Woolley, G. A. Directed Evolution Approaches for Optogenetic Tool Development. *Biochem. Soc. Trans.* **2021**, *49*, 2737–2748.
- (15) Seong, J.; Lin, M. Z. Optobiochemistry: Genetically Encoded Control of Protein Activity by Light. *Annu. Rev. Biochem.* **2021**, *90*, 475–501.
- (16) Zillich, O. V.; Schweiggert-Weisz, U.; Eisner, P.; Kerscher, M. Polyphenols as Active Ingredients For Cosmetic Products. *Int. J. Cosmetic Sci.* **2015**, *37*, 455–464.
- (17) Espagne, A.; Changenet-Barret, P.; Plaza, P.; Martin, M. M. Solvent Effect on the Excited-State Dynamics of Analogues of the Photoactive Yellow Protein Chromophore. *J. Phys. Chem. A* **2006**, *110*, 3393–3404.
- (18) Espagne, A.; Paik, D. H.; Changenet-Barret, P.; Martin, M. M.; Zewail, A. H. Ultrafast Photoisomerization of Photoactive Yellow Protein Chromophore Analogues in Solution: Influence of the Protonation State. *ChemPhysChem* **2006**, *7*, 1717–1726.
- (19) Espagne, A.; Paik, D. H.; Changenet-Barret, P.; Plaza, P.; Martin, M. M.; Zewail, A. H. Ultrafast Light-Induced Response of Photoactive Yellow Protein Chromophore Analogues. *Photochem. Photobiol. Sci.* **2007**, *6*, 780–787.
- (20) Espagne, A.; Changenet-Barret, P.; Baudin, J.-B.; Plaza, P.; Martin, M. M. Photoinduced Charge Shift as the Driving Force For the Excited-State Relaxation of Analogues of the Photoactive Yellow Protein Chromophore in Solution. *J. Photochem. Photobiol. A: Chem.* **2007**, *185*, 245–252.
- (21) Kuramochi, H.; Takeuchi, S.; Tahara, T. Ultrafast Structural Evolution of Photoactive Yellow Protein Chromophore Revealed by Ultraviolet Resonance Femtosecond Stimulated Raman Spectroscopy. *J. Phys. Chem. Lett.* **2012**, *3*, 2025–2029.
- (22) Wang, S.; Schatz, S.; Stuhldreier, M. C.; Böhnke, H.; Wiese, J.; Schröder, C.; Raeker, T.; Hartke, B.; Keppler, J. K.; Schwarz, K.; et al. Ultrafast Dynamics of UV-Excited *trans*- and *cis*-Ferulic Acid in Aqueous Solutions. *Phys. Chem. Chem. Phys.* **2017**, *19*, 30683–30694.
- (23) Gromov, E. V.; Burghardt, I.; Köppel, H.; Cederbaum, L. S. Photoinduced Isomerization of the Photoactive Yellow Protein (PYP) Chromophore: Interplay of Two Torsions, a HOOP Mode and Hydrogen Bonding. *J. Phys. Chem. A* **2011**, *115*, 9237–9248.
- (24) Boggio-Pasqua, M.; Groenhof, G. Controlling the Photo-reactivity of the Photoactive Yellow Protein Chromophore by Substituting at the *p*-Coumaric Acid Group. *J. Phys. Chem. B* **2011**, *115*, 7021–7028.
- (25) Isborn, C. M.; Götz, A. W.; Clark, M. A.; Walker, R. C.; Martínez, T. J. Electronic Absorption Spectra From MM and Ab Initio QM/MM Molecular Dynamics: Environmental Effects on the Absorption Spectrum of Photoactive Yellow Protein. *J. Chem. Theory Comput.* **2012**, *8*, 5092–5106.
- (26) Gromov, E. V. Unveiling the Mechanism of Photoinduced Isomerization of the Photoactive Yellow Protein (PYP) Chromophore. *J. Chem. Phys.* **2014**, *141*, 224308.
- (27) Garcia-Prieto, F. F.; Muñoz-Losa, A.; Luz Sanchez, M.; Elena Martin, M.; Aguilar, M. A. Solvent Effects on De-Excitation Channels in the *p*-Coumaric Acid Methyl Ester Anion, An Analogue of the Photoactive Yellow Protein (PYP) Chromophore. *Phys. Chem. Chem. Phys.* **2016**, *18*, 27476–27485.
- (28) Garcia-Prieto, F. F.; Muñoz-Losa, A.; Fdez. Galván, I.; Sánchez, M. L.; Aguilar, M. A.; Martín, M. E. QM/MM Study of Substituent and Solvent Effects on the Excited State Dynamics of the Photoactive Yellow Protein Chromophore. *J. Chem. Theory Comput.* **2017**, *13*, 737–748.
- (29) Nomoto, A.; Inai, N.; Yanai, T.; Okuno, Y. Substituent and Solvent Effects on the Photoisomerization of Cinnamate Derivatives: An XMS-CASPT2 Study. *J. Phys. Chem. A* **2022**, *126*, 497–505.

- (30) Adamson, B. D.; Coughlan, N. J. A.; Continetti, R. E.; Bieske, E. J. Changing the Shape of Molecular Ions: Photoisomerization Action Spectroscopy in the Gas Phase. *Phys. Chem. Chem. Phys.* **2013**, *15*, 9540–9548.
- (31) Markworth, P. B.; Coughlan, N. J. A.; Adamson, B. D.; Goerigk, L.; Bieske, E. J. Photoisomerization Action Spectroscopy: Flicking the Protonated Merocyanine-Spiropyran Switch in the Gas Phase. *Phys. Chem. Chem. Phys.* **2015**, *17*, 25676–25688.
- (32) Bull, J. N.; da Silva, G.; Scholz, M. S.; Carrascosa, E.; Bieske, E. J. Photoinduced Intramolecular Proton Transfer in Deprotonated *para*-Coumaric Acid. *J. Phys. Chem. A* **2019**, *123*, 4419–4430.
- (33) Lee, I.-R.; Lee, W.; Zewail, A. H. Primary Steps of the Photoactive Yellow Protein: Isolated Chromophore Dynamics and Protein Directed Function. *Proc. Nat. Acad. Sci.* **2006**, *103*, 258–262.
- (34) Bull, J. N.; Anstöter, C. S.; Verlet, J. R. R. Ultrafast Valence to Non-Valence Excited State Dynamics in a Common Anionic Chromophore. *Nature Comm* **2019**, *10*, 5820.
- (35) Anstöter, C. S.; Curchod, B. F. E.; Verlet, J. R. R. Geometric and Electronic Structure Probed Along the Isomerisation Coordinate of a Photoactive Yellow Protein Chromophore. *Nat. Commun.* **2020**, *11*, 2827.
- (36) Bull, J. N.; Anstöter, C. S.; Verlet, J. R. R. Fingerprinting the Excited-State Dynamics in Methyl Ester and Methyl Ether Anions of Deprotonated *para*-Coumaric Acid. *J. Phys. Chem. A* **2020**, *124*, 2140–2151.
- (37) Bull, J. N.; Buntine, J. T.; Carrascosa, E.; Stockett, M. H.; Bieske, E. J. Action Spectroscopy of Deprotomer-Selected Hydroxycinnamate Anions. *Eur. Phys. J. D* **2021**, *75*, 67.
- (38) Mustalahti, S.; Morozov, D.; Luk, H. L.; Pallerla, R. R.; Myllyperkiö, P.; Pettersson, M.; Pihko, P. M.; Groenhof, G. Photoactive Yellow Protein Chromophore Photoisomerizes around a Single Bond if the Double Bond Is Locked. *J. Phys. Chem. Lett.* **2020**, *11*, 2177–2181.
- (39) Groenhof, G.; Bouxin-Cademartory, M.; Hess, B.; de Visser, S. P.; Berendsen, H. J. C.; Olivucci, M.; Mark, A. E.; Robb, M. A. Photoactivation of the Photoactive Yellow Protein: Why Photon Absorption Triggers a Trans-to-Cis Isomerization of the Chromophore in the Protein. *J. Am. Chem. Soc.* **2004**, *126*, 4228–4233.
- (40) Groenhof, G.; Schäfer, L. V.; Boggio-Pasqua, M.; Grubmüller, H.; Robb, M. A. Arginine52 Controls the Photoisomerization Process in Photoactive Yellow Protein. *J. Am. Chem. Soc.* **2008**, *130*, 3250–3251.
- (41) Boggio-Pasqua, M.; Robb, M. A.; Groenhof, G. Hydrogen Bonding Controls Excited-State Decay of the Photoactive Yellow Protein Chromophore. *J. Am. Chem. Soc.* **2009**, *131*, 13580–13581.
- (42) Parkes, M. A.; Phillips, C.; Porter, M. J.; Fielding, H. F. Controlling Electron Emission From the Photoactive Yellow Protein Chromophore by Substitution at the Coumaric Acid Group. *Phys. Chem. Chem. Phys.* **2016**, *18*, 10329–10336.
- (43) Rocha-Rinza, T.; Christiansen, O.; Rajput, J.; Gopalan, A.; Rahbek, D. B.; Andersen, L. H.; Bochenkova, A. V.; Granovsky, A. A.; Bravaya, K. B.; Nemukhin, A. V.; et al. Gas Phase Absorption Studies of Photoactive Yellow Protein Chromophore Derivatives. *J. Phys. Chem. A* **2009**, *113*, 9442–9449.
- (44) Adamson, B. D.; Coughlan, N. J. A.; Markworth, P. B.; Continetti, R. E.; Bieske, E. J. An Ion Mobility Mass Spectrometer for Investigating Photoisomerization and Photodissociation of Molecular Ions. *Rev. Sci. Instrum.* **2014**, *85*, 123109.
- (45) Bull, J. N.; Scholz, M. S.; Carrascosa, E.; da Silva, G.; Bieske, E. J. Double Molecular Photoswitch Driven by Light and Collisions. *Phys. Rev. Lett.* **2018**, *120*, 223002.
- (46) Neese, F. The ORCA program system. *WIREs Comp. Mol. Sci.* **2012**, *2*, 73–78.
- (47) Campuzano, I.; Bush, M. F.; Robinson, C. V.; Beaumont, C.; Richardson, K.; Kim, H.; Kim, H. I. Structural Characterization of Drug-Like Compounds by Ion Mobility Mass Spectrometry: Comparison of Theoretical and Experimentally Derived Nitrogen Collision Cross Sections. *Anal. Chem.* **2012**, *84*, 1026–1033.
- (48) Mesleh, M. F.; Hunter, J. M.; Shvartsburg, A. A.; Schatz, G. C.; Jarrold, M. F. Structural Information From Ion Mobility Measurements: Effects of the Long-Range Potential. *J. Phys. Chem.* **1996**, *100*, 16082–16086.
- (49) Child, M. S. *Molecular Collision Theory*; Dover Publications Inc.: 1997.
- (50) Granovsky, A. A. Firefly Version 8.2.0. <http://classic.chem.msu.su/gran/firefly/index.html>.
- (51) Heisler, I. A.; Kondo, M.; Meech, S. R. Reactive Dynamics in Confined Liquids: Ultrafast Torsional Dynamics of Auramine O in Nanoconfined Water in Aerosol OT Reverse Micelles. *J. Phys. Chem. B* **2009**, *113*, 1623–1631.
- (52) Chosrowjan, H.; Taniguchi, S.; Tanaka, F. Ultrafast Fluorescence Upconversion Technique and its Applications to Proteins. *FEBS J.* **2015**, *282*, 3003–3015.
- (53) Jimenez, R.; Fleming, G. R.; Kumar, P. V.; Maroncelli, M. Femtosecond Solvation Dynamics of Water. *Nature* **1994**, *369*, 471–473.
- (54) Hosseinizadeh, A.; Breckwoldt, N.; Fung, R.; Sepehr, R.; Schmidt, M.; Schwander, P.; Santra, R.; Ourmazd, A. Few-fs Resolution of a Photoactive Protein Traversing a Conical Intersection. *Nature* **2021**, *599*, 697–701.
- (55) Bull, J. N.; West, C. W.; Anstöter, C. S.; da Silva, G.; Bieske, E. J.; Verlet, J. R. R. Ultrafast Photoisomerisation of an Isolated Retinoid. *Phys. Chem. Chem. Phys.* **2019**, *21*, 10567–10579.
- (56) Reichardt, C.; Welton, T. *Solvents and Solvent Effects in Organic Chemistry*; Wiley-VCH Verlag GmbH & Co. KGaA: 2010.
- (57) Anna, J. M.; Kubarych, K. J. Watching Solvent Friction Impede Ultrafast Barrier Crossings: A Direct Test of Kramers Theory. *J. Chem. Phys.* **2010**, *133*, 174506.

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