

1 **Inoculum effects of cefepime/zidebactam (WCK 5222) and ertapenem/zidebactam (WCK**
2 **6777) for Enterobacterales in relation to β -lactamase type and enhancer effect, as tested**
3 **by BSAC agar dilution**

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15 **Running head:** Inoculum effects for zidebactam combinations

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33

34 **Abstract**

35 **Introduction.** Combinations of PBP3-active β -lactams with developmental
36 diazabicyclooctanes (DBOs), e.g. zidebactam, remain active against many metallo- β -
37 lactamase (MBL) producers, via an enhancer effect. We explored how this was affected by
38 inoculum.

39 **Materials and Methods.** MICs of zidebactam and its cefepime and ertapenem combinations
40 (WCK 5222 and WCK 6777, respectively) were determined by BSAC agar dilution at inocula
41 from $3\text{-}6 \times 10^3$ to $3\text{-}6 \times 10^5$ cfu/spot. Isolates, principally *Klebsiella* spp., were chosen as
42 having previously tested resistant to zidebactam or its cefepime combination, and by β -
43 lactamase type.

44 **Results.** MICs of zidebactam, tested alone, were strongly inoculum dependent, regardless of
45 β -lactamase type; MICs of its cefepime and ertapenem combinations likewise were strongly
46 inoculum dependent – rising ≥ 32 -fold across the inoculum range tested – but only for MBL
47 producers. Combination MICs for isolates with non-metallo- β -lactamases, including those
48 with OXA-48 (where the enhancer effect remains critical for ertapenem/zidebactam), were
49 much less inoculum dependent, particularly for cefepime/zidebactam. MBL producers
50 frequently moved between putative ‘susceptible’ (MIC $\leq 8+8$ mg/L) and ‘resistant’ (MIC $> 8+8$
51 mg/L) categories according to whether the inoculum was at the high or low end of the BSAC’s
52 acceptable (1 to 4×10^4 cfu/spot) range.

53 **Conclusion.** The activity of zidebactam combinations against MBL producers – which
54 strongly depends on the enhancer effect – is inoculum dependent. Animal data suggest
55 consistent *in-vivo* activity even in high inoculum pneumonia models. Contingent on this being
56 supported by clinical experience, the combination behaviour may be best represented by the
57 MICs obtained at the lower end of BSAC’s inoculum range.

58

59 **Introduction**

60 Combinations of β -lactams with ‘triple action’ diazabicyclooctanes (DBOs) – zidebactam,
61 nacubactam or durlobactam – provide a prospective option against carbapenemase-producing
62 Gram-negatives, regardless of whether these have class A, B or D β -lactamases.¹ The furthest-
63 progressed combination, cefepime/zidebactam (WCK 5222), is entering phase III, whilst
64 nacubactam combinations are at an earlier stage, as is ertapenem/zidebactam (WCK 6777),
65 envisaged as a once-daily product suitable for OPAT.

66 The distinguishing feature of these developmental DBOs is that they not only inhibit
67 serine β -lactamases, as does avibactam (the parent DBO), but that they additionally bind to
68 PBP2, achieving a direct antibiotic activity and, when partnered with a PBP3-targetted β -
69 lactam, a β -lactamase-independent synergy dubbed the ‘enhancer effect’.²⁻⁴ These secondary
70 properties allow their combinations with PBP3-targetted β -lactams to remain active against
71 many isolates with Class B (i.e., metallo) carbapenemases, although these are not inhibited by
72 any DBO.² The enhancer effect is important also for ertapenem/zidebactam in the case of
73 isolates with OXA-48 carbapenemase, which is not inhibited by zidebactam,² though not for
74 cefepime/zidebactam, which incorporates a cephalosporin stable to OXA-48 enzyme that needs
75 only be protected against co-produced ESBLs.

76 A limitation is that high-frequency mutations, largely affecting the stringent response,
77 can compensate for inhibition of PBP2, conferring zidebactam and nacubactam resistance.⁵
78 Even then, the enhancer effect persists, and β -lactam/zidebactam or β -lactam/nacubactam
79 combinations remain active against many zidebactam- and nacubactam-resistant metallo- β -
80 lactamase (MBL) producers.²⁻⁴ Thus, multiple large surveys, variously by broth microdilution
81 or agar dilution, find that 90-95% of MBL-producing Enterobacterales are susceptible to
82 cefepime/zidebactam 8+8 mg/L;⁶⁻⁹ though, using agar dilution, we recorded a lower proportion

83 (45%) among a small collection (n=24) of *Klebsiella* spp. with both NDM and OXA-48 β -
84 lactamases, many belonging to ST14.⁹

85 What remains unclear is the extent to which the enhancer activity is affected by
86 inoculum, particularly when zidebactam itself lacks activity. To explore this aspect we
87 compared MICs of zidebactam, cefepime/zidebactam and ertapenem/zidebactam at inocula
88 from *c.* $3\text{-}6 \times 10^3$ to $3\text{-}6 \times 10^5$ cfu per spot for zidebactam-resistant MBL producers, where
89 activity depends on the enhancer effect, and zidebactam-resistant producers of zidebactam-
90 inhibited serine β -lactamases, where the enhancer effect has secondary importance.

91

92 **Method and Materials**

93 *Bacteria*

94 A panel of 93 isolates, encompassing three groups, was assembled from an earlier study,⁹
95 where testing also was performed by BSAC agar dilution with cefepime/zidebactam as a 1:1
96 gravimetric ratio. Group 1 comprised 33 isolates previously found resistant to
97 cefepime/zidebactam 8+8 mg/L; it included 32 *K. pneumoniae* and one *Escherichia coli* with
98 NDM carbapenemases alone (n=20) or together with OXA-48 (n=13). Group 2 comprised 30
99 MBL producers previously found highly resistant (MIC ≥ 32 mg/L) to zidebactam, but
100 susceptible to cefepime/zidebactam at $\leq 8+8$ mg/L. These included 28 *K. pneumoniae* and two
101 *K. oxytoca* with carbapenemases as follows: IMP (n=5), VIM (n=6), NDM (n=17) and NDM
102 plus OXA-48 (n=2). Group 3 comprised 30 MBL-negative isolates previously found highly
103 resistant to zidebactam (MIC ≥ 32 mg/L) but susceptible to cefepime/zidebactam at 8+8 mg/L.
104 They variously produced KPC (n=9), OXA-48 (n=9) or GES-5 (n=2) carbapenemases, K1
105 enzyme (n=1) and either ESBLs or AmpC enzymes alone or in combination (n=9); 25 were *K.*
106 *pneumoniae*, 3 were *K. oxytoca* and 2 *K. aerogenes*.

107

108 *MIC determinations.*

109 The UK Health Security Agency's (UKHSA, formerly Public Health England, PHE) AMRHAI
110 Reference Unit performed susceptibility tests by the BSAC agar dilution method,¹⁰ adapted as
111 below, using IsoSensitest agar from Oxoid/Thermofisher (Basingstoke, UK). Zidebactam
112 combinations were tested as 1:1 ratios, with zidebactam and ertapenem from Wockhardt
113 (Aurangabad, India) and cefepime purchased from Alfa Aesar (Heysham, UK).

114 Standard inocula were prepared by adjusting bacterial suspensions to match a 0.5
115 McFarland standard (*c.* $1-2 \times 10^8$ cfu/mL), then printing on to antibiotic-containing agars using
116 the fine-pin inoculator (Mast, Merseyside, UK) which delivers 0.3 μ l, thereby giving *c.* $3-6 \times$
117 10^4 cfu/spot inocula (see Discussion). In parallel, we performed MIC determinations with 0.3
118 (*i.e.*, 3.3-fold) and 0.1 (*i.e.*, 10-fold) dilutions of these standard suspensions, giving $1-2 \times 10^4$
119 cfu/spot and $3-6 \times 10^3$ cfu/spot, respectively, and with suspensions matched to 1.5 and 5
120 McFarlands, respectively delivering 3x and 10x standard inocula, corresponding to $1-2 \times 10^5$
121 cfu/spot and $3-6 \times 10^5$ cfu/spot.

122 Plates were read to where growth terminated: fine films and hazes were ignored, as was
123 the presence of 1 or 2 isolated colonies. Trailing was mostly an issue when zidebactam was
124 tested alone, particularly with lower inocula; on/off trailing/regrowth was ignored but
125 persistent heavy trailing was counted as growth.

126

127 **Results**

128 *Confirmation of prior results as a basis for Group assignment*

129 Most isolates behaved in accordance with previous results at the standard inoculum, supporting
130 their categorisation into Groups 1-3; disagreements largely were instances where
131 cefepime/zidebactam MICs of 8+8 mg/L were recorded for Group 1 isolates previously found
132 resistant with MICs one tube higher at 16+16 mg/L. For zidebactam, tested alone, 88/93 prior

133 and current results agreed within one doubling dilution, with larger discrepancies equally
134 distributed between cases where the present result was >1 dilution higher (n=2) or lower (n=3).
135 For cefepime/zidebactam, 77/93 isolates had prior and current results within one doubling
136 dilution, again with larger discrepancies equally distributed between instances where the
137 present result was >1 dilution higher (n=7) or lower (n=9); 90/93 of cefepime/zidebactam
138 results were within 2 doubling dilutions of previous data.

139

140 *Inoculum effects*

141 The inocula spanned a 100-fold range, from $3-6 \times 10^3$ to $3-6 \times 10^5$ cfu/spot, extending 10-fold
142 either side of PHE's $3-6 \times 10^4$ cfu/spot 'standard'. As anticipated, based upon how the panel
143 was constructed, almost all (89/93) the isolates were highly resistant to zidebactam at the
144 standard inoculum, with MICs >32 mg/L (Table 1). This proportion rose to 92/93 at 10x
145 standard inoculum; by contrast, and crucially, the proportion with zidebactam MICs >32 mg/L
146 shrank to 52/93 at 0.3x standard inoculum and to 30/93 at 0.1x standard. At this lowest
147 inoculum, zidebactam alone at 8 mg/L (i.e., as present in a 1:1 cefepime/zidebactam
148 combination with a putative 8+8 mg/L breakpoint) inhibited 18/33 Group 1 isolates, 22/30
149 Group 2 and 21/30 Group 3 (p 0.04 Chi-square test), albeit with a higher modal MIC (1 mg/L)
150 for Group 1 than for Groups 2 and 3 (both 0.25 mg/L).

151 The MBL producers of *both* Groups 1 and 2 exhibited strong inoculum effects for both
152 β -lactam/zidebactam combinations, as illustrated in Table 1 and in fig. 1, which depicts the
153 proportions susceptible to cefepime/zidebactam 8+8 mg/L and ertapenem/zidebactam 8+8
154 mg/L at different inocula. Fully 25/33 (75%) of the 'cefepime/zidebactam-resistant' Group 1
155 isolates became susceptible to cefepime/zidebactam at $\leq 8+8$ mg/L if the inoculum was lowered
156 to 0.1x standard; on the other hand, 22/30 of the 'cefepime/zidebactam-susceptible' Group 2
157 isolates became resistant if the inoculum was raised to 10x standard. We conclude that the

158 distinction between Groups 1 and 2 isolates is not fundamental; rather it is that the shift to high
159 MICs occurs at slightly lower inocula in Group 1 (fig. 1).

160 Measured across the $3\text{-}6 \times 10^3$ to $3\text{-}6 \times 10^5$ inoculum range, the MIC inoculum effects
161 for the MBL-producing Group 1 and 2 isolates typically were 32- to >128-fold (fig 2), with
162 low ratios seen only for the minority of MBL producers that remained highly resistant to
163 cefepime/zidebactam and ertapenem/zidebactam even at the lowest inocula, raising the
164 denominator for the ratio. By contrast, there were only *c.* 1-4-fold MIC effects for
165 cefepime/zidebactam in the case of the zidebactam-resistant MBL-negative Group 3 isolates.
166 Effects were more scattered for ertapenem/zidebactam, being 1- to 4-fold for 20/30 Group 3
167 isolates, including all those with GES carbapenemases, ESBLs or AmpC enzymes but 8-32-
168 fold for the remaining 10, comprising 4/9 with OXA-48 carbapenemase and 6/9 with KPC
169 carbapenemases.

170

171 **Discussion**

172 MBLs hydrolyse cefepime and ertapenem and are not inhibited by zidebactam or other
173 DBOs.^{1,2,11} Consequently, any activity of zidebactam combinations against MBL producers
174 must depend on the antibiotic activity of zidebactam and/or the enhancer effect. This enhancer
175 action must also drive the activity of ertapenem/zidebactam against isolates with OXA-48
176 carbapenemase, given that zidebactam is a poor inhibitor of this enzyme.² We explored the
177 extent to which these activities were affected by the inoculum, and the implications for
178 susceptibility testing.

179 Regardless of whether the MBL-producing isolates had previously been found resistant
180 (Group 1) or susceptible (Group 2) to cefepime/zidebactam at the standard inoculum, we
181 observed strong, mostly >32-fold, inoculum-dependent MIC rises both for zidebactam alone
182 and for its combinations (Table 1 and fig 1). By contrast, effects were absent

183 (cefepime/zidebactam) or smaller (ertapenem/zidebactam) for Group 3 isolates, which were
184 zidebactam-resistant, but lacked MBLs. The differing behaviour of the Group 3 isolates can
185 largely be explained by their having zidebactam-inhibited β -lactamases, allowing a
186 conventional mode of activity. The exception to this generalisation is the case of
187 ertapenem/zidebactam against isolates with OXA-48 enzymes, which are not inhibited by
188 zidebactam; inoculum effects for these were similarly minimal to those observed for isolates
189 with KPC carbapenemases, which are inhibited by zidebactam.² This anomaly may simply
190 reflect the fact that OXA-48 enzymes confer less *in-vitro* resistance to ertapenem than do
191 MBLs, limiting the scope for an effect: at standard inocula 6/9 isolates with OXA-48 enzymes
192 were inhibited by *unprotected* ertapenem at ≤ 8 mg/L compared with just 9/63 among MBL
193 producers (not shown).

194 β -Lactamase-related inoculum effects are well-known in Gram-negative bacteria,
195 occurring e.g. for weak-substrate oxyimino-cephalosporins in the case of ESBL producers,
196 first-generation cephalosporins for isolates with classical TEM enzymes and widely, for
197 penicillin/ β -lactamase-inhibitor combinations.¹²⁻¹⁴ In our experience, supported by the
198 published literature, such effects only become substantial above inocula of 10^5 .¹²⁻¹⁴ The effects
199 outlined here were manifest at lower inocula, particularly for the Group 1 isolates, overlapping
200 into the inoculum ranges used for susceptibility testing. Effects in this range also occur with
201 mecillinam which, like zidebactam, attacks PBP2.¹⁵ Although synergistic mecillinam/ β -lactam
202 combinations reportedly evade this issue¹⁶ they were not tested for isolates with challenging
203 combinations of resistance mechanisms, as used here.

204 Even minor inoculum variances may have a significant practical effect on results for
205 zidebactam combinations. The BSAC method,¹⁰ used here and formerly by the AMRHAI
206 Reference Unit (which has subsequently switched to EUCAST microbroth testing as its
207 standard method) specifies adjusting a bacterial suspension to the opacity of a 0.5 McFarland

208 (c. $1-2 \times 10^8$ cfu/mL), diluting this 10-fold for Enterobacterales, then using multipoint pins that
209 deposit 1-2 μ L per spot, giving an inoculum of c. $1-4 \times 10^4$ cfu/spot. Since AMRHAI used fine-
210 pin multipoint inoculators, depositing only 0.3 μ L/spot, we adapted the method to use the 0.5
211 McFarland-equivalent suspension without dilution, giving c. $3-6 \times 10^4$ cfu/spot. Taking $4.24 \times$
212 10^4 cfu/spot as the geometric mid-point of this range equates to an inoculum on the upper edge
213 of the BSAC specification or minimally above it, whilst the 0.3x dilution contains an inoculum
214 (geometric mid-point 1.41×10^4 cfu/spot) around the lower edge. Both are ‘technically
215 acceptable, yet the former would lead to more isolates being scored as resistant (Fig. 1).

216 In the absence of clinical experience, it is not possible to say which inoculum is most
217 representative of the patient, nor the extent to which this may vary between patients or infection
218 sites. Nonetheless, there are some pointers from animal testing, in particular a high-inoculum
219 ($6.69 + 0.31 \log_{10}$ cfu/lung) murine pneumonia model using humanised dosing.¹⁷ This found
220 the ertapenem/zidebactam combination effective in achieving 2-3 \log_{10} reductions in lung
221 count even for MBL producers with ertapenem/zidebactam MICs of 8+8 mg/L and zidebactam
222 MICs >128 mg/L by CLSI broth microdilution with an inoculum of c. 1×10^4 /well. The
223 reductions in bacterial load were similar to those for infections established using zidebactam-
224 resistant isolates with non-metallo-carbapenemases and lower MICs. This was despite the fact
225 that, at an MIC of 8+8 mg/L, the ertapenem concentration exceeded the MIC for only c. 15-
226 18% of the dosage interval.

227 Whilst it is difficult to translate these animal data directly to the present study, it does
228 suggest retention of enhancer activity against organisms where strong inoculum effects would
229 be predicted, and despite a heavy bacterial challenge. Accordingly, whilst further work is
230 needed, there is reason for optimism that the higher MICs observed with slightly raised inocula
231 are not predictive of clinical failure, and that results with inocula close to 1×10^4 – at the lower

232 end of the BSAC range and represented by the 0.3x standard here – may be the most
233 representative.

234

235 **Funding**

236 This work was sponsored by Wockhardt

237

238 **Transparency declarations**

239 DML: Advisory Boards or ad hoc consultancy Accelerate, Antabio, Centauri, Meiji,
240 Menarini, Mutabilis, Nordic, Paion, ParaPharm, Pfizer, QPEX, Russian Direct Investment
241 Fund, Shionogi, Sumitovant, Summit, T.A.Z., VenatoRx, Wockhardt, Zambon, Paid lectures
242 – bioMérieux, GSK, Hikma, Merck/MSD, Menarini, Nordic, Pfizer, and Shionogi. Relevant
243 shareholdings or options – Dechra, GSK, Merck and Perkin-Elmer, amounting to less than
244 10% of portfolio value. He also has nominated holdings in Arecor, Avacta, Diaceutics, Creo
245 Medical, Evgen, Genedrive, Poolbeg, Renalytics AI and Trelus (all with research/products
246 pertinent to medicines or diagnostics) through Enterprise Investment Schemes but has no
247 authority to trade these shares directly. All other authors are employees of the UKHSA's
248 Antimicrobial Resistance and Healthcare Associated Infections Reference Unit, which has
249 received financial support for conference attendance, lectures, research projects, or contracted
250 evaluations from numerous sources, including Accelerate Diagnostics, Achaogen Inc.,
251 Allegra Therapeutics, Amplex, AstraZeneca UK Ltd, AusDiagnostics, Basilea
252 Pharmaceutica, Becton Dickinson Diagnostics, bioMérieux, Bio-Rad Laboratories, BSAC,
253 Cepheid, Check-Points B.V., Cubist Pharmaceuticals, Department of Health, Enigma
254 Diagnostics, Food Standards Agency, GlaxoSmithKline Services Ltd, Helperby Therapeutics,
255 Henry Stewart Talks, IHMA Ltd, Innovate UK, Integra holdings, Kalidex Pharmaceuticals,
256 Melinta Therapeutics, Merck Sharpe & Dohme Corp, Meiji Seika Pharma Co. Ltd, Mobidiag,

257 Momentum Biosciences Ltd, Neem Biotech, Nordic Pharma Ltd, Norgine Pharmaceuticals,
258 Paratek Pharmaceuticals, Rempex Pharmaceuticals Ltd, Roche, Rokitan Ltd, Smith &
259 Nephew UK Ltd, Shionogi & Co. Ltd, Trius Therapeutics, T.A.Z., VenatoRx
260 Pharmaceuticals and Wockhardt Ltd.

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Table 1. MICs of zidebactam and its combinations for carbapenemase- and cephalosporinase- producers in relation to prior results with cefepime/zidebactam and inoculum

Drug and inoculum/spot	Inoculum	No isolates with indicated MIC (mg/L)														Total
		0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	>128	
Zidebactam																
Group 1. MBL producers, previously found resistant to cefepime/zidebactam 8+8 mg/L	0.1 x standard					1	10	4	2	1		1		4	10	33
	0.3 x standard						5	4		2		5	2		15	33
	Previous distribution at standard inocula							1				1	1		30	33
	Standard											1		1	31	33
	3 x standard											1			32	33
	10 x standard											1			32	33
Group 2. MBL producers, previously found susceptible to cefepime/zidebactam 8+8 mg/L	0.1 x standard			3	9	4	3	3				1	1	2	4	30
	0.3 x standard			1	1	4	3	2				3	2	3	11	30
	Previous distribution at standard inocula											3	1	1	25	30
	Standard					1						1	1	1	26	30
	3 x standard														30	30
	10 x standard														30	30
Group 3. Non-metallo carbapenemase and cephalosporinase producers, previously found susceptible to cefepime/zidebactam 8+8 mg/L	0.1 x standard			1	8	6	5	1							9	30
	0.3 x standard				1	1	5				1	3	5	2	12	30
	Previous distribution at standard inocula												2	1	27	30
	Standard									1				1	28	30
	3 x standard														30	30
	10 x standard														30	30

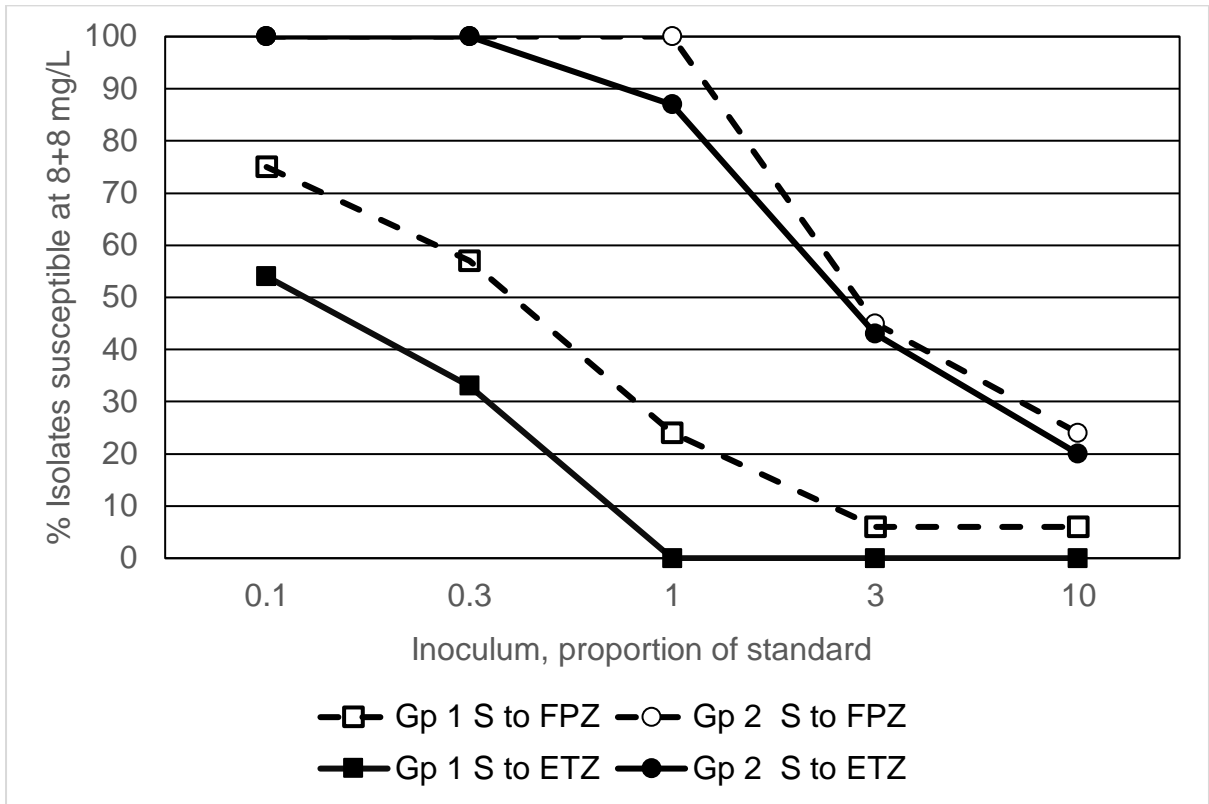
Drug and inoculum/spot	Inoculum	No isolates with indicated MIC (mg/L)														Total
		0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	128	>128	
Ertapenem/ zidebactam																
Group 1. MBL producers, previously found resistant to cefepime/zidebactam 8+8 mg/L	0.1 x standard					2	11	8	4	3	1	1	1	1	1	33
	0.3 x standard					1	5	7	4	6	1	1	4	3	1	33
	Previous distribution at standard inocula						1	1		2	2	6	13	7	1	33
	Standard						1	3		5	7	3	8		6	33
	3 x standard										2	2	4	12	13	33
	10 x standard									1	1		1	7	23	33
Group 2. MBL producers, previously found susceptible to cefepime/zidebactam 8+8 mg/L	0.1 x standard			10	2	11	3	4								30
	0.3 x standard			7	5	5	2	6	4	1						30
	Previous distribution at standard inocula			3	2	3	3	4	7	3	3	2				30
	Standard			3	3	4	2	6	2	6	3		1			30
	3 x standard				1	3		3	3	3	3	4	7	2	1	30
	10 x standard					1	2	2		1	4		4	13	3	30
Group 3. Non-metallo carbapenemase and cephalosporinase producers, previously found susceptible to cefepime/zidebactam 8+8 mg/L	0.1 x standard	2	1	3	10	8	2	4								30
	0.3 x standard		3	2	9	9	2	5								30
	Previous distribution at standard inocula	1	2	1	5	12	1	5	2	1						30
	Standard		2	2	4	8	7	4	2	1						30
	3 x standard		2	1	2	7	9	2	6		1					30
	10 x standard			3	1	4	5	8	5	3	1					30

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Values in bold – modal values

334 Figure 1
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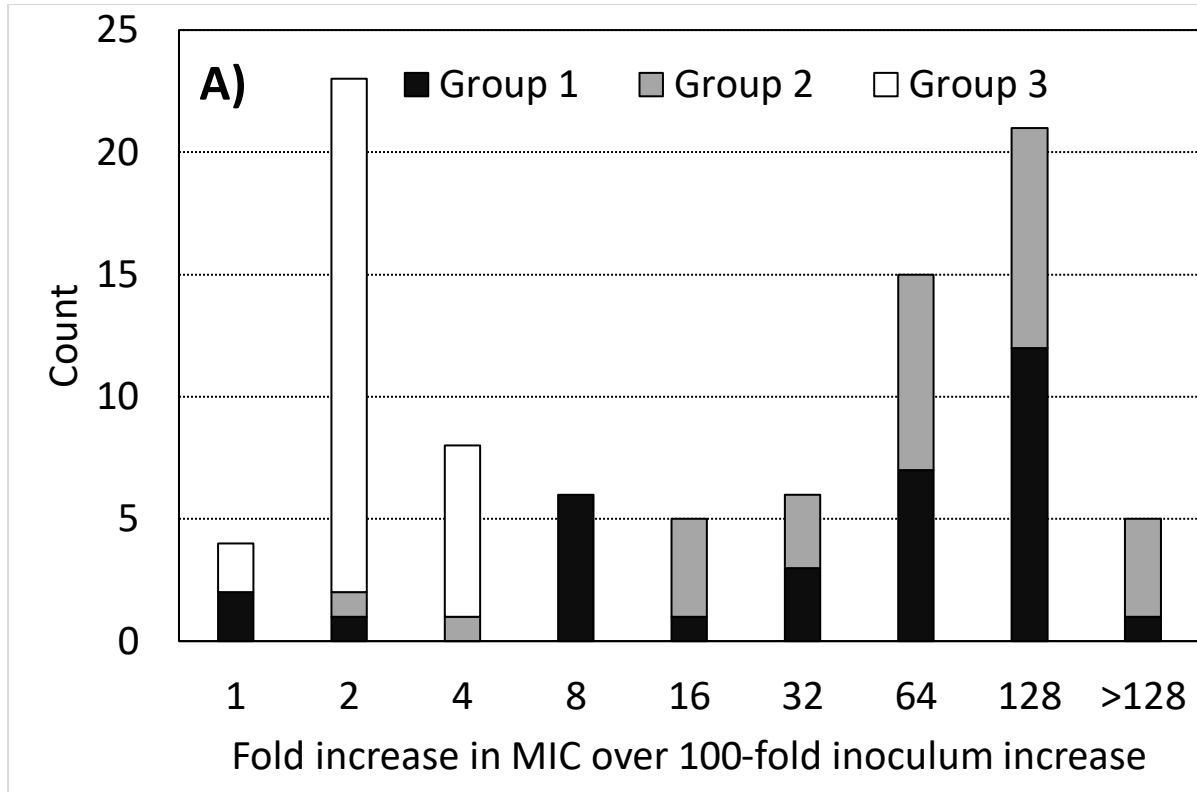


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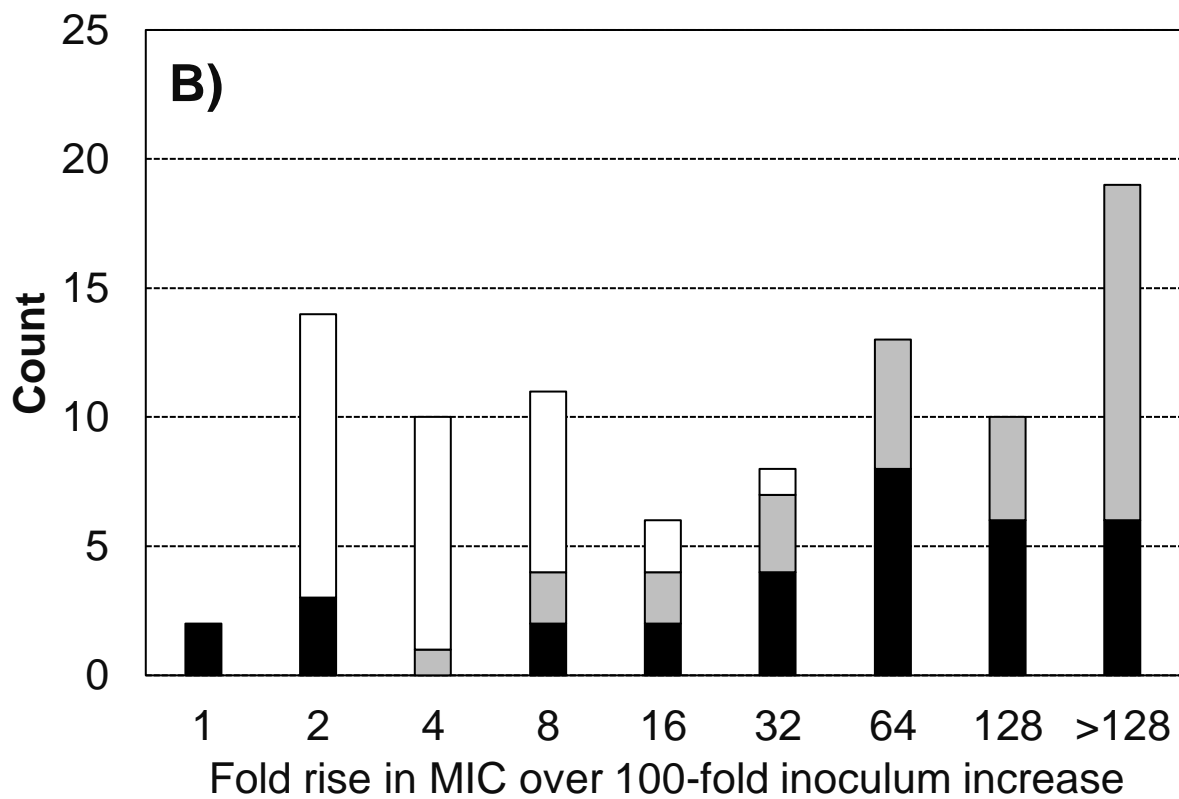
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Figure 2



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347 **Legend, Figure 1:** Percent susceptibility to cefepime/zidebactam 8+8 mg/L (FPZ) and
348 ertapenem/zidebactam 8+8 mg/L (ETZ) for Group (Gp) 1 and 2 isolates in relation to
349 inoculum.

350

351 **Legend, figure 2.** Distribution of ratios of MICs at inocula of $3-6 \times 10^5$ vs. $3-6 \times 10^3$ for (top
352 panel) cefepime/zidebactam 1:1 and (lower panel) ertapenem/zidebactam 1:1 by isolate
353 group. The large inoculum effects for most Group 1 and 2 isolates are evident; the minorities
354 of these groups with only small effects are cases where the low inoculum denominator MICs
355 remained high, typically >128 mg/L (see Table 1).

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