

1 Opinion paper

2 Should the enzyme name ‘rhodesain’ be discontinued?

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16 A B S T R A C T

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18 Rhodesain is the generic name for the cathepsin L-like peptidase of *Trypanosoma brucei*
19 *rhodesiense*. The term rhodesain was derived from the subspecies epithet *rhodesiense* which
20 itself originated from Rhodesia, a historical region in southern Africa named after the 19th
21 century British imperialist and white supremacist Cecil Rhodes. This tainting could be grounds
22 for discontinuing the name, however, there are also scientific grounds. Specifically, protein
23 sequence comparisons and frequency-based difference profiling reveal that rhodesain is
24 essentially identical (99.87-98.44%) to the cathepsin L-like peptidases of both *T. b. brucei* and
25 *T. b. gambiense*. Accordingly, and based on a previously proposed terminology for
26 kinetoplastid C1 peptidases (Caffrey and Steverding, 2009), we suggest the use of the formal
27 term, *TbrCATL*, to denote the cathepsin L-like peptidases of the *T. brucei* subspecies. The
28 earlier and informal term, ‘brucipain’, could also be used.

29

30 *Keywords:*

31 Rhodesain

32 Brucipain

33 *Trypanosoma brucei rhodesiense*

34 *Trypanosoma brucei brucei*

35 *Trypanozoon*

36

37 Trypanosomes express two Clan CA C1 cysteine peptidases that are related to mammalian
38 cathepsins B and L [1]. By tradition, the names of cathepsin L-like peptidases from various
39 parasites, including the trypanosome orthologue, have incorporated the suffix “-pain” or “-ain”
40 to the species or subspecies name based on the archetypal C1 cysteine peptidase, papain, from
41 papaya fruit. For example, brucipain, rhodesain, evansain, congopain and cruzipain/cruzain
42 (EC 3.4.22.51) are the generic names for the cathepsin L-like cysteine peptidases of
43 *Trypanosoma (brucei) brucei*, *T. (brucei) rhodesiense*, *T. evansi*, *T. congolense* and *T. cruzi*,
44 respectively [2-7]. This terminology, however, is sometimes confusing and inconsistent. For
45 instance, no such generic name exists for the cathepsin L-like peptidase of *T. equiperdum* and
46 *T. (brucei) gambiense*, and the term vivapain has been used for the respective enzymes of *T.*
47 *vivax* as well as of *Plasmodium vivax* [8,9]. Although a unifying nomenclature for kinetoplastid
48 C1 peptidases has been proposed [10], based on a system originally suggested by Clayton et
49 al. [11], informal names for the trypanosome cathepsin L-like peptidase still appear in the
50 academic literature.

51 The term rhodesain was coined to differentiate the cathepsin L-like peptidase of the *T. b.*
52 *rhodesiense* subspecies from its orthologue, brucipain, found in the subspecies *T. b. brucei*
53 [3,10]. Rhodesain is a derivation of “*rhodesiense*” itself derived from Rhodesia¹, a former
54 British protectorate in southern Africa comprising the present-day countries of Zambia and
55 Zimbabwe, and where the first case of a human infection with *T. b. rhodesiense* occurred in
56 1909 [12]. The historical region of Rhodesia² was named after the British mining magnate and
57 politician, Cecil John Rhodes (1853-1902). However, Rhodes was a passionate colonialist and
58 a believer in the supremacy of white people, and anything connected to him is contentious.
59 Since the 1950s, protesters have questioned whether it is appropriate to commemorate Cecil
60 Rhodes, and have demanded the removal of his statues and the renaming of buildings bearing
61 his name. In this context, it was recently proposed to either rename the *T. b. rhodesiense*
62 subspecies or to revoke its subspecies status, although this suggestion was also based on

¹ The Latin suffix “-ense” (singular nominative, accusative and vocative form of “-ensis”) literally means “of or from [a place]”.

² Not to be confused with the unrecognised state of Rhodesia/Republic of Rhodesia from 1965-1979 which is equivalent to the territory of present-day Zimbabwe.

63 scientific reasons [13]. By extension, the link between the name rhodesain and Cecil Rhodes
64 could be used to argue for the discontinuation of the name; again, however, there are also
65 scientific reasons.

66 The species/subspecies status of *T. b. rhodesiense* has been a constant matter of debate.
67 Molecular genetic studies (reviewed [14,15]) and phylogenetic relationship analyses [16] have
68 indicated that *T. b. rhodesiense* is only a phenotypic variant of *T. b. brucei*. In fact, the human
69 infectivity of *T. b. rhodesiense*, which distinguishes it from *T. b. brucei*, is based on just one
70 gene, the serum resistance associated (*SRA*) gene [17-19]. In addition, experimental crossing
71 and population genetics studies have provided evidence for gene flow between *T. b. brucei* and
72 *T. b. rhodesiense* [20]. Therefore, should *T. b. rhodesiense* be no longer recognised as a
73 subspecies but rather a human-infectious form of *T. b. brucei*, then there is no longer a reason
74 to use the term rhodesain. Instead, a better option would be to employ the term *TbrCATL* (for
75 *T. brucei* cathepsin L) according to the previously proposed nomenclature for kinetoplastid C1
76 peptidases, which states that the first letter of the genus should be followed by the first two
77 letters of the species (all in italics) to indicate the source organism of the peptidase [10].

78 Further support for dropping the name rhodesain comes from sequence analysis studies.
79 Comparison of the cathepsin L-like peptidase sequences of *T. b. rhodesiense* with those of *T.*
80 *b. brucei* and *T. b. gambiense* indicates that they are all essentially identical. (Fig. 1). To date,
81 two, six and two different cathepsin L-like peptidase sequences have been published for *T. b.*
82 *rhodesiense*, *T. b. brucei* and *T. b. gambiense*. The two *T. b. rhodesiense* sequences are 99.33%
83 identical while their identity with the six *T. b. brucei* and the two *T. b. gambiense* sequences
84 ranges between 99.78-98.44% and 99.78-99.33%, respectively. Thus, some *T. b. brucei* and *T.*
85 *b. gambiense* sequences show higher identity with one of the *T. b. rhodesiense* sequences than
86 the two rhodesain sequences between each other. For example, *T. b. brucei* sequence E and *T.*
87 *b. gambiense* sequence B are practically identical to *T. b. rhodesiense* sequence B apart of one
88 amino acid replacement at position 115, which is, in any case, a conservative substitution (L
89 for V; Fig. 1). Interestingly, compared to all other *T. brucei* subspecies sequences, the *T. b.*
90 *rhodesiense* sequence A is the only one that has a third N-glycosylation site motif (Fig. 1,
91 T.b.r./Seq.A: ²⁹⁵NDS). In addition, the cathepsin L-like peptidase sequences of *T. evansi* and

92 *T. equiperdum* show high identity with those of the *T. brucei* group members, with percentage
93 values ranging between 99.11-97.56% and 99.78-98.44%, respectively.

94 The close relationship between the cathepsin L-like peptidase sequences of *T. b.*
95 *rhodesiense*, *T. b. brucei*, *T. b. gambiense*, *T. evansi* and *T. equiperdum* is also evident from
96 their frequency-based difference profiles which are all very similar (Fig. 2). It should be noted
97 that *T. evansi* and *T. equiperdum* together with the three *T. brucei* subspecies belong to the sub-
98 genus *Trypanozoon*. In contrast, the cathepsin L-like peptidase sequences of *T. (Nannomona)*
99 *congolense* and *T. (Duttonella) vivax* exhibit less identity with those of the *Trypanozoon*
100 species (70.82-65.92% for *T. congolense* and 60.69-58.71% for *T. vivax*). In addition, their
101 frequency-based difference profiles are quite distinct from those of the *Trypanozoon* species
102 (Fig. 2). Based on the frequency profiles, it can be concluded that the cathepsin L-like peptidase
103 sequences of the *Trypanozoon* species represent the same protein but are clearly different from
104 the cathepsin L-like peptidase sequences of *T. congolense* and *T. vivax*. Thus, these and the
105 protein sequence alignment data, indicate that the cathepsin L-like peptidase sequences of the
106 *T. brucei* subspecies encode an identical protein with just a few, mostly irrelevant, amino acid
107 substitutions.

108 In conclusion, and apart from an association with a contentious past, we propose that the
109 name rhodesain be no longer used as the enzyme is not different from brucipain. Should the
110 subspecies status of *T. b. rhodesiense* be revoked (discussed in [13]), then the term rhodesain
111 would be automatically discontinued. In the meantime, any cathepsin L-like peptidases of the
112 *T. brucei* group should be denoted formally as *Tbr*CATL according to the previously proposed
113 unifying nomenclature system for kinetoplastid C1 peptidases, or informally as brucipain [10].

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115 **CRedit authorship contribution statement**

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117 **Dietmar Steverding:** Conceptualization, Formal Analysis, Writing – Original Draft,
118 Visualisation. **Conor R. Caffrey:** Writing – Review & Editing.

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120 **Declaration of Competing Interest**

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122 The authors declare that they have no competing interests.

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183 **Figure legends**

184

185 **Fig. 1.** Sequence alignment of cathepsin L-like peptidases from *T. b. rhodesiense* (T.b.r.), *T. b.*
186 *brucei* (T.b.b.) and *T. b. gambiense* (T.b.g.). A NCBI Protein database
187 (<https://www.ncbi.nlm.nih.gov/protein/>) search using the search terms “brucei” and “cathepsin
188 L” gave a total of 2, 35 and 6 entries for cathepsin L-like peptidases sequences for *T. b.*
189 *rhodesiense*, *T. b. brucei* and *T. b. gambiense*, respectively. However, several entries contained
190 the same sequence so that, in the end, only two, six and two specific cathepsin L-like peptidases
191 sequences for *T. b. rhodesiense*, *T. b. brucei* and *T. b. gambiense*, respectively, were obtained.
192 Species-specific sequences and their corresponding accession numbers are as follows:
193 T.b.r./Seq.A: CAC67416.1; T.b.r./Seq.B: CAA38238.1; T.b.b./Seq.A: XP_845218.1,
194 XP_845219.1, XP_845220.1, XP_845223.1, AAX80351.1, AAX80352.1, AAX80353.1,
195 AAX80356.1, AAZ11659.1, AAZ11660.1, AAZ11661.1 AAZ11664.1; T.b.b./Seq.B:
196 XP_845221.1, AAX80354.1, AAZ11662.1; T.b.b./Seq.C: XP_845222.1, XP_845227.1;
197 XP_845228.1, AAX80355.1, AAX80360.1, AAX80361.1, AAZ11663.1, AAZ11668.1,
198 AAZ11669.1; T.b.b./Seq.D: XP_845225.1, XP_845.226.1, AAX80358.1, AAX80359.1,
199 AAZ11666.1, AAZ11667.1; T.b.b./Seq.E: XP_845224.1, AAX80357.1, AAZ11665.1;
200 T.b.b./Seq.F: CAA34485.1, P14658.1; T.b.g./Seq.A: XP_011773878.1, CBH11593.1;
201 T.b.g./Seq.B: XP_011773880.1, XP_011773883.1, CBH11595.1, CBH11598.1.

202

203 **Fig. 2.** Comparison of African trypanosome cathepsin L-like peptidase sequences using the
204 NCBI COBALT Multiple Alignment Tool
205 (<https://www.ncbi.nlm.nih.gov/tools/cobalt/cobalt.cgi?CMD=Web>). The display setting shows
206 frequency-based differences which compares the residue at a position to the position consensus.
207 Darker shades of red indicate a further difference from residues in other sequences in the
208 alignment at that position. Grey indicates identical residues at that position for all sequences
209 while white indicates gaps. The following number of unique sequences were compared: *T. b.*
210 *rhodesiense*: 2; *T. b. brucei*: 6; *T. b. gambiense*: 2; *T. evansi*: 1; *T. equiperdum*: 2, *T.*
211 *congolense*: 12; *T. vivax*: 4.

212 Figure 1

213

214 T.b.r./Seq.A 1 MPRT**EMV**RFVRLPVVLLAMAAC**LA**SVALGSLHVEESLEMRFAAFKKKYGKVYKDAKEEAFRFR**AF**EENMEQAKIQAAANPYATFGVTPFS
 215 T.b.r./Seq.B 1
 216 T.b.b./Seq.A 1
 217 T.b.b./Seq.B 1I.....
 218 T.b.b./Seq.C 1
 219 T.b.b./Seq.D 1
 220 T.b.b./Seq.E 1
 221 T.b.b./Seq.F 1
 222 T.b.g./Seq.A 1
 223 T.b.g./Seq.B 1

224

225

226 T.b.r./Seq.A 91 DMTREEFRARYRNGASYFAAAQKRLRKT**V**NVTTGRAPAAVDWREKGAVTPVKDQGC**G**SCWAFSTIGNIEGQ**W**QVAGNPLVSLSEQMLVS
 227 T.b.r./Seq.B 91V.....
 228 T.b.b./Seq.A 91
 229 T.b.b./Seq.B 91
 230 T.b.b./Seq.C 91
 231 T.b.b./Seq.D 91
 232 T.b.b./Seq.E 91
 233 T.b.b./Seq.F 91V.....
 234 T.b.g./Seq.A 91
 235 T.b.g./Seq.B 91

236

237

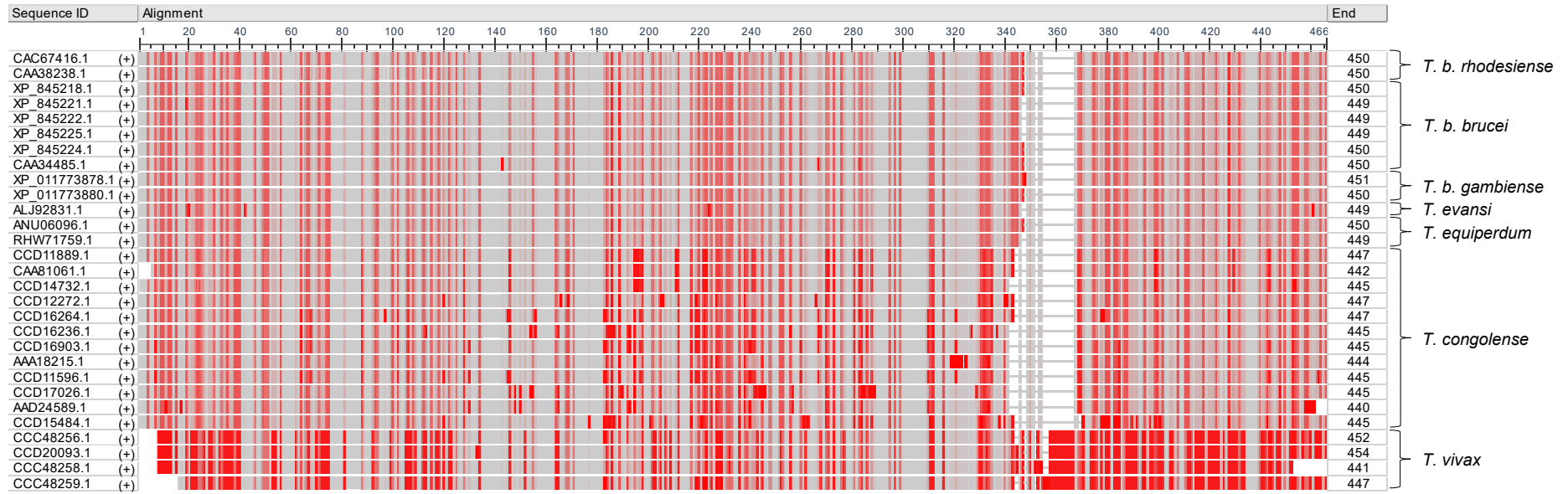
238 T.b.r./Seq.A 181 CDTIDFGCGGLMDNAFNWIVNSNGGNVFTEASYPYVSGNGEQ**Q**QC**Q**MNGHEIGAAITDHVDLPQ**DE**DAIAAYLAENGPLAIAVDAT**S**FM
 239 T.b.r./Seq.B 181
 240 T.b.b./Seq.A 181S..N.....
 241 T.b.b./Seq.B 181S..N.....
 242 T.b.b./Seq.C 181S..N.....
 243 T.b.b./Seq.D 181S..N.....
 244 T.b.b./Seq.E 181
 245 T.b.b./Seq.F 181S..N.....E.....
 246 T.b.g./Seq.A 181
 247 T.b.g./Seq.B 181

248

249				
250	T.b.r./Seq.A	271	DYNGGILTSCTSEQLDHGVLLVGYNDSSNPPYWI IKNWSNMWGEDGYIRIEKGTNQCLMNQAVSSAVVGGPTPPPP-PPPPSATFTQD	
251	T.b.r./Seq.B	271N.....-	
252	T.b.b./Seq.A	271N.....-	
253	T.b.b./Seq.D	271N.....--	
254	T.b.b./Seq.C	271N.....--	
255	T.b.b./Seq.D	271N.....--	
256	T.b.b./Seq.E	271N.....-	
257	T.b.b./Seq.F	271K.....N.....-	
258	T.b.g./Seq.A	271N.....P	
259	T.b.g./Seq.B	271N.....-	
260				
261				
262	T.b.r./Seq.A	360	FCEGKGCTKGC SHATFPTGECVQTTGVG SVIATCGASNL TQIIYPLSRSCSGLSV PITVPLDKCIPILIGSVEYHCSTNPPTKAARLVPHQ	450
263	T.b.r./Seq.B	360P.....	450
264	T.b.b./Seq.A	360P.....	450
265	T.b.b./Seq.B	359P.....	449
266	T.b.b./Seq.C	359P.....	449
267	T.b.b./Seq.D	359P.....	449
268	T.b.b./Seq.E	360P.....	450
269	T.b.b./Seq.F	360P.....	450
270	T.b.g./Seq.A	361	451
271	T.b.g./Seq.B	360	450
272				
273				

274 Figure 2

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