

**Key Points:**

- We successfully developed a multi-platform assimilative system for biogeochemistry in the North Sea
- We tested the impact of the different assimilative system components on the ecosystem reanalysis
- The multi-platform assimilation will become an essential part of future operational research

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## Towards a Multi-Platform Assimilative System for North Sea Biogeochemistry

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**Abstract** Oceanography has entered an era of new observing platforms, such as biogeochemical-Argo floats and gliders, some of which will provide three-dimensional maps of essential ecosystem variables on the North-West European (NWE) Shelf. In a foreseeable future operational centers will use multi-platform assimilation to integrate those valuable data into ecosystem reanalysis and forecast systems. Here we address some important questions related to glider biogeochemical data assimilation (DA) and introduce multi-platform DA in a preoperational model of the NWE Shelf sea ecosystem. We test the impact of the different multi-platform system components (glider vs. satellite, physical vs. biogeochemical) on the simulated biogeochemical variables. To characterize the model performance, we focus on the period around the phytoplankton spring bloom, since the bloom is a major ecosystem driver on the NWE Shelf. We found that the timing and magnitude of the phytoplankton bloom is insensitive to the physical DA, which is explained in the study. To correct the simulated phytoplankton bloom one needs to assimilate chlorophyll observations from glider or satellite Ocean Color (OC) into the model. Although outperformed by the glider chlorophyll assimilation, we show that OC assimilation has mostly desirable impact on the sub-surface chlorophyll. Since the OC assimilation updates chlorophyll only in the mixed layer, the impact on the sub-surface chlorophyll is the result of the model dynamical response to the assimilation. We demonstrate that the multi-platform assimilation combines the advantages of its components and always performs comparably to its best performing component.

**Plain Language Summary** North-West European (NWE) Shelf is a region of major importance for both European economy and climate. Observational oceanography has entered an important era of new observing biogeochemical platforms, such as Biogeochemical-Argos and gliders. Gliders are being currently deployed to measure three-dimensional distributions of some essential biogeochemical variables on the NWE Shelf. This work establishes a multi-platform assimilative system on the NWE Shelf which will be used to combine multiple different types of observing platforms (e.g., satellite, gliders) with our up to date models in order to optimize our estimate and forecast of the NWE Shelf ecosystem state. We provide an understanding for how the different components of the system interact. We demonstrate that the assimilative system is skilled to combine physical data with satellite and glider data for chlorophyll, as well as the glider data for oxygen. The work establishes the foundations of a system that is planned to be used in the future operational oceanography on the NWE Shelf.

### 1. Introduction

Understanding the state and the future of shelf sea ecosystems is essential from the point of view of economy, conservation and the global carbon cycle (Borges et al., 2006; Friedlingstein et al., 2006; Jahnke, 2010; Pauly et al., 2002). Reanalysis provide our best estimate of the ocean state by optimally combining the state of the art knowledge from models with the most up to date observations. In marine biogeochemistry the prevailing approach is to assimilate satellite products into models, either for Ocean Color (OC) derived total chlorophyll (e.g., Carmillet et al., 2001; Ciavatta, Kay, Saux-Picart, et al., 2016; Ciavatta, Torres, Saux-Picart, & Allen, 2011; Fontana et al., 2010; Ford & Barciela, 2017; Ford, Edwards, et al., 2012; Gregg, 2008; Hoteit, Triantafyllou, & Petihakis, 2005; Ishizaka, 1990; Kalaroni et al., 2016; Natvik & Evensen, 2003; Nerger & Gregg, 2007, 2008; Pradhan et al., 2019; Triantafyllou et al., 2007), phytoplankton functional

type (PFT)-specific chlorophyll (Ciavatta, Brewin, et al., 2018; Ciavatta, Kay, Brewin, et al., 2019; Skákala, Bruggeman, et al., 2020; Skákala, Ford, et al., 2018), or surface radiances (Ciavatta, Torres, Martinez-Vicente, et al., 2014; Gregg & Rousseaux, 2017; Jones et al., 2016; Shulman et al., 2013; Skákala, Bruggeman, et al., 2020). Additionally a number of studies have assimilated biogeochemical data from in situ measurements, either using single location profiles (e.g., Allen et al., 2003; Hoteit, Triantafyllou, Petihakis, & Allen, 2003; Lenartz et al., 2007; Torres et al., 2006), or using surface data from ships, floats and buoys (e.g., Anderson et al., 2000; Cossarini, Lermusiaux, & Solidoro, 2009; Song et al., 2016). The typical disadvantage of the traditionally assimilated biogeochemical data-sets is that they are either constrained to the ocean surface (e.g., in the case of satellite data), or they are typically limited to a single location (in the case of vertically measured data). Assimilating such data into the model has either only local impact, or its impact on biogeochemical fields is typically constrained to the upper oceanic layer, with uncertain impact on the vertical profiles of biomass, or nutrients.

However, the situation on the data-front is rapidly changing, with new programs (e.g., AtlantOS, Visbeck et al., 2015) aiming at revolutionizing biogeochemical oceanography with novel observing platforms covering large parts of the ocean both horizontally and vertically, such as floats deployed in the Biogeochemical-Argo program (e.g., Germaineaud et al., 2019; Johnson, 2016; Johnson & Claustre, 2016), and gliders with optical and biogeochemical sensors (Telszewski et al., 2018). Some of the Argo float oxygen data were already assimilated to constrain the biogeochemistry in the Southern Ocean (Verdy & Mazloff, 2017) and Argo-measured chlorophyll was assimilated to improve phytoplankton dynamics in the Mediterranean Sea (Cossarini, Mariotti, et al., 2019). This new observational activity quite understandably focuses on regions of high importance for fisheries, economy and climate, such as the North-West European (NWE) Shelf (e.g., Legge et al., 2020), where a number of gliders have been deployed as a part of the Alternative Framework to Assess Marine Ecosystem Functioning in Shelf Seas (AlterECO) program (<http://projects.noc.ac.uk/altereco/>). The rapid development of these new autonomous observation systems opens up an entirely new range of possibilities on how to optimally integrate multi-platform observing networks with our present oceanographic models (Bell et al., 2015; Lellouche et al., 2013). The observational work on the NWE Shelf from the AlterECO project is coupled to a sister program, the CAMPUS (Combining Autonomous observations and Models for Predicting and Understanding Shelf seas, <https://www.campus-marine.org/>) project, aiming to consistently combine the different sources of information, such as gliders, satellite OC data and models, in order to improve our capability to understand, represent and forecast the NWE Shelf biogeochemistry (e.g., spring bloom, carbon and nutrient cycle, oxygen depletion events). Future plans, based on CAMPUS and in line with the European Copernicus Marine Environment Monitoring Service (CMEMS), are to have a multi-platform assimilative system on the NWE Shelf, where the autonomous vehicles will navigate to specific locations using a combination of Artificial Intelligence (AI) and model forecast, to observe important processes such as the onset of the phytoplankton bloom, or hypoxic events.

Trying to establish glider data assimilation (DA) as part of such a multi-platform assimilative system often leads to two non-trivial problems: (1) how to consistently combine high resolution glider data with much coarser model resolution, (2) how to achieve reasonable consistency between the assimilation-corrected variables and the coupled physical-biogeochemical model dynamics. The problem of dynamical consistency needs special mention, since both physical and biogeochemical fields have typically much larger gradients in the vertical than in the horizontal dimension. The vertical correlation length scales have large spatio-temporal variability and model dynamics can be quite sensitive to spurious vertical gradients (Doney, 1999; Doney et al., 2004; Oschlies & Garçon, 1999). Such model sensitivity is often noticed when physical data (such as sea surface height, or temperature and salinity) are assimilated into the model, as the spurious vertical mixing introduced by such assimilation is known to often degrade the skill of the biogeochemical model (e.g., Berline et al., 2007; El Moussaoui et al., 2011; Holt et al., 2014; Park et al., 2018; Raghukumar et al., 2015; While et al., 2010). However, similar issues can be easily overlooked when we assimilate surface biogeochemical data (except extreme regions with substantial small-scale horizontal variability, such as the Gulf Stream, Anderson et al., 2000), since the biogeochemical fields have smaller gradients in the horizontal direction than in the vertical, which means they are more dynamically stable in the horizontal than in the vertical direction. For the gliders, it is of vital interest to understand the potentially complex interaction between the physical and the biogeochemical DA, or the interplay between the different biogeochemical variables updated by the assimilative system.

In this study we extend the operational assimilative system on the NWE Shelf to successfully produce a multi-platform reanalysis including both physical (satellite sea surface temperature, temperature and salinity from in situ platforms and an AlterEco glider) and biogeochemical (total chlorophyll *a* and oxygen from an AlterECO glider, and chlorophyll *a* from a satellite OC product) variables. The main focus of the study is to assess the impact of the different multi-platform assimilative system components (satellite vs. glider, physical vs. biogeochemical) on the simulated ecosystem processes in relation to the phytoplankton spring bloom. Being able to estimate the impact of the different system components is important, since it indicates what the assimilation impact will be on the simulated biogeochemistry in regions where only a specific type of data (e.g., satellite OC, physical variables) is available. The focus on the processes around the spring bloom is a natural choice due to (1) the availability of high-quality chlorophyll glider data, and (2) because the spring bloom is a key driver of the ecosystem dynamics on the NWE Shelf (Henson et al., 2009; Lutz et al., 2007). The results of this study should form a basis for an integrated multi-platform assimilative system, that will optimize the available information from observations and models in order to improve our understanding of the NWE Shelf biogeochemistry. The assimilated biogeochemical glider variables were selected based on the data availability, but both chlorophyll and oxygen are expected to play an important role in the future multi-platform operational assimilation: chlorophyll is a proxy for phytoplankton biomass, which forms the base of the marine food web, while oxygen needs to be monitored and forecast in order to identify oxygen depletion events (i.e., hypoxia, Vaquer-Sunyer & Duarte, 2008), which can have disastrous impacts on marine life.

## 2. Methods

The study uses a hindcast version of the operational modeling system for the NWE Shelf run by the Met Office in the framework of the CMEMS, that is, the physical model Nucleus for European Modeling of the Ocean (NEMO, Madec et al., 2015) coupled through the Framework for Aquatic Biogeochemical Models (FABM, Bruggeman & Bolding, 2014) with the biogeochemical model European Regional Seas Ecosystem Model (ERSEM, Baretta et al., 1995; Blackford, 1997; Butenschön et al., 2016). We used measurements from an AlterEco glider that operated in the central North Sea between May and August 2018 providing data for temperature, salinity, chlorophyll (derived from fluorescence), and oxygen concentrations. In multi-platform assimilation the glider data were complemented with the Ocean Color-Climate Change Initiative (OC-CCI) satellite product of the European Space Agency (ESA) for total chlorophyll (version 3.1, Sathyendranath et al., 2019), sea surface temperature (SST) data from the GCOM-W1/AMSR-2, NOAA/AVHRR, MetOp/AVHRR, MSG/SEVIRI, Sentinel-3/SLSTR, and Suomi-NPP/VIIRS satellite products, and the temperature and salinity in situ data from the EN4 data set (Good et al., 2013), which includes profiles from Argo floats, fixed moored arrays, XBTs, CTDs, gliders, and marine mammals. The physical and biogeochemical data were assimilated on a daily basis into NEMO-FABM-ERSEM using NEMOVAR (the assimilative system used operationally by the Met Office, King et al., 2018; Mogensen, Balmaseda, & Weaver 2012; Mogensen, Balmaseda, Weaver, et al., 2009; Waters et al., 2015).

The model free simulation was run from September 1, 2017 until the end of the year 2018 and was initialized from 2016–2018 run of a very similar model configuration presented in Skákala, Bruggeman, et al. (2020). The free run outputs have been analyzed for the period of the glider data availability (May 8–August 15, 2018). The assimilative runs used identical model settings as the free run, only with the added assimilation components. The different assimilative runs compared in this study are (see also Table 1): (1) physical DA (satellite SST, temperature and salinity from EN4 data and the AlterEco glider), (2) satellite OC total chlorophyll *a* assimilation, (3) AlterEco glider chlorophyll *a* assimilation, (4) AlterEco glider oxygen assimilation, and (5) multi-platform assimilation combining all the data from (1) to (4). Note that wherever we mention the assimilation of specific data (e.g., glider chlorophyll) we mean a simulation where only those data have been assimilated (as opposed to multi-platform assimilation, which assimilates all the available data). All the assimilative runs were started from the initial value conditions produced by the free simulation for May 8, 2018.

**Table 1**  
*The Observations Assimilated in the Different Data Assimilation (DA) Experiments*

Experiment	Satellite SST	EN4 T&S	Glider T&S	Satellite OC	Glider chl <i>a</i>	Glider O <sub>2</sub>
Physical DA	Yes	Yes	Yes	No	No	No
Satellite OC DA	No	No	No	Yes	No	No
Glider chl <i>a</i> DA	No	No	No	No	Yes	No
Glider O <sub>2</sub> DA	No	No	No	No	No	Yes
Multi-platform DA	Yes	Yes	Yes	Yes	Yes	Yes

Abbreviations: T, temperature; S, salinity; EN4, EN4 in situ data set.

## 2.1. The Physical Component: NEMO

The NEMO ocean physics component (OPA) is a finite difference, hydrostatic, primitive equation ocean general circulation model (Madec et al., 2015). The NEMO configuration used in this study is similar to the one used by Ford, van der Molen, et al. (2017), Skákala, Ford, et al. (2018), and almost identical to Skákala, Bruggeman, et al. (2020); we use the CO6 NEMO version, based on NEMOv3.6, a development of the CO5 configuration explained in detail by O'Dea et al. (2017). The model has 7 km spatial resolution on the Atlantic Margin Model (AMM7) domain using a terrain-following  $z^* - \sigma$  coordinate system with 51 vertical levels (Siddorn & Furner, 2013). The lateral boundary conditions for physical variables at the Atlantic boundary were taken from the outputs of the Met Office operational 1/12° North Atlantic model (NATL12, Storkey et al., 2010); the Baltic boundary values were derived from a reanalysis produced by the Danish Meteorological Institute for CMEMS. We use annually varying river discharge based on data from Lenhart et al. (2010). The model was forced at the surface by atmospheric fluxes provided by an hourly and 31 km resolution realization (HRES) of the ERA5 data set (<https://www.ecmwf.int/>).

## 2.2. The Biogeochemical Component: ERSEM

ERSEM (Baretta et al., 1995; Butenschön et al., 2016) is a lower trophic level ecosystem model for marine biogeochemistry, pelagic plankton, and benthic fauna (Blackford, 1997). The model splits phytoplankton into four functional types largely based on their size (Baretta et al., 1995): picophytoplankton, nanophytoplankton, diatoms, and dinoflagellates. ERSEM uses variable stoichiometry for the simulated plankton groups (Baretta-Bekker et al., 1997; Geider et al., 1997) and each PFT biomass is represented in terms of chlorophyll, carbon, nitrogen, and phosphorus, with diatoms also represented by silicon. ERSEM predators are composed of three zooplankton types (mesozooplankton, microzooplankton, and heterotrophic nanoflagellates), with organic material being decomposed by one functional type of heterotrophic bacteria (Butenschön et al., 2016). The ERSEM inorganic component consists of nutrients (nitrate, phosphate, silicate, ammonium, and carbon) and dissolved oxygen. The carbonate system is also included in the model (Artioli et al., 2012).

We used in this study a similar ERSEM configuration to Skákala, Bruggeman, et al. (2020), but unlike Skákala, Bruggeman, et al. (2020) we implemented an updated ERSEM version (v20.10), with a notable upgrade to the benthic code. The ERSEM parametrization is identical to the one described in Butenschön et al. (2016). The Atlantic boundary values for nitrate, phosphate, silicate and oxygen were taken from World Ocean Atlas (Garcia et al., 2013) and dissolved inorganic carbon from the GLODAP gridded data set (Key et al., 2015; Lauvset et al., 2016), while plankton and detritus variables were set to have zero fluxes at the Atlantic boundary. The ERSEM irradiance was calculated using a new bio-optical module implemented in the NEMO-FABM-ERSEM AMM7 configuration by Skákala, Bruggeman, et al. (2020). The bio-optical module resolves light spectrally and distinguishes between downwelling direct and diffuse streams. The module is forced by ERA5 atmospheric inputs (<https://www.ecmwf.int/>) for total vertically integrated ozone, water vapor, cloud cover, cloud liquid water and sea-level air pressure, as well as by a satellite product for aerosol optical thickness (Moderate Resolution Imaging Spectroradiometer, MODIS, <https://modis.gsfc.nasa.gov/data/dataprod>).



### 2.3. The Assimilative System: NEMOVAR

NEMOVAR is a variational DA system (Mogensen, Balmaseda, & Weaver 2012; Mogensen, Balmaseda, Weaver, et al., 2009; Waters et al., 2015) used for operational ocean DA at the Met Office via the assimilation of satellite OC derived (total, or PFT) chlorophyll concentrations, NEMOVAR has been demonstrated as being highly successful in improving the phytoplankton community structure (PFT chlorophyll assimilation), phytoplankton seasonal cycle, and the timing and magnitude of the spring bloom (Skákala, Bruggeman, et al., 2020; Skákala, Ford, et al., 2018). There are also indications that satellite OC assimilation can improve the carbon cycle (Skákala, Bruggeman, et al., 2020; Skákala, Ford, et al., 2018). When it comes to the non-assimilated variables, satellite OC reanalysis typically has a comparable skill to the free run (Skákala, Bruggeman, et al., 2020; Skákala, Ford, et al., 2018). The satellite OC chlorophyll assimilation using NEMOVAR on the NWE Shelf has been thoroughly validated on bi-decadal time-scales (Kay et al., 2019), showing a good overall skill and no spurious trends in biogeochemical tracer concentrations.

In this study the observations are assimilated on a daily basis. The model is first run for the day and background values are calculated in observation space by interpolating the model fields to the observation locations at the nearest model time step (300 s) to the observation time, an approach known as First Guess at Appropriate Time (FGAT). NEMOVAR is then run, calculating a set of increments for each updated variable on the model grid. After the assimilation step the model is re-run with the increments applied to the model variables gradually at each model time step using incremental analysis updates (IAU, Bloom et al., 1996). For the physical variables the increments are calculated for temperature, salinity, sea surface height and the horizontal velocity components, by accounting for their correlations by transforming those variables through a set of linear balancing equations into an independent set of variables that is assimilated separately. For biogeochemical variables, the increments are initially calculated for the observed variable. For total chlorophyll the assimilation is applied in log-space, since chlorophyll is typically log-normally distributed (Campbell, 1995). After calculating the total chlorophyll increments, we use a balancing module to split those increments into the model state variables. The applied scheme (Skákala, Bruggeman, et al., 2020; Skákala, Ford, et al., 2018) redistributes total chlorophyll increments into the 4 ERSEM PFTs based on background PFT-to-total chlorophyll ratios. The PFT chlorophyll is used to update the remaining PFT components (carbon, phosphorus, nitrogen for all PFTs, silicon for diatoms) following the background stoichiometric ratios. In the case of oxygen assimilation, the only updated variable is the simulated oxygen concentration. There were attempts to extend the currently applied balancing scheme to other ERSEM variables (e.g., nutrients), but so-far this produced sub-optimal results degrading the biogeochemical model skill (see discussion in Skákala, Ford, et al. [2018]). Any combined physical-biogeochemical assimilation in NEMOVAR is weakly coupled, which means that the physical and the biogeochemical variables are assimilated separately, with physical assimilation impacting biogeochemistry only through the model dynamics, and no feedback from biogeochemistry to physics.

The multi-platform assimilation is based on the development from Waters et al. (2015) extended to biogeochemical variables by Ford (2020), that is, the combined assimilation of satellite OC and glider chlorophyll data is performed by following a scheme previously applied to temperature by Waters et al. (2015). The satellite and in situ glider data are combined to calculate a single set of 3D increments, while allowing for different observation errors to be specified for the different data sources (for the details see Waters et al. [2015] and Ford [2020]). Since each of the physical data, chlorophyll and oxygen assimilation provides increments for different variables, the multi-platform assimilation simply aggregates the increments from the physical, chlorophyll and oxygen assimilative components.

The background covariances are represented as a product of background variances and a diffusion operator (King et al., 2018; Mirouze & Weaver, 2010). Within the diffusion operator, the same length scales are set for all the assimilated (physical, biogeochemical) variables. The horizontal correlation length scales are specified a priori, and are based on two different length scales, a longer 100 km correlation scale and a shorter length scale based on the first baroclinic Rossby radius of deformation (King et al., 2018). The vertical length scales use the scheme from Waters et al. (2015), King et al. (2018), and Ford (2020), where NEMOVAR calculates directly the set of 3D increments (we call this scheme a “3D variant”) using flow-dependent vertical length scales ( $\ell$ ), which are the following function of depth ( $d$ ):

**Table 2**

*Parts of the Multi-Platform Assimilative System With the List of the Updated Physical-Biogeochemical Variables and the Mean Values of the Background-to-Observational Error Ratio (B-O Error Ratio, With Error Understood as Standard Deviation)*

Component	Updated variables	B-O error ratio
Satellite OC chl <i>a</i>	PFT components	2.3
Glider chl <i>a</i>	PFT components	1.4
Glider O <sub>2</sub>	Oxygen	0.77
Satellite T	T, S, SSH, U,V	1.55
In situ T	T, S, SSH, U,V	1.04
In situ S	T, S, SSH, U,V	3.42

Abbreviations: T, temperature; S, salinity; SSH, sea surface height; U,V, horizontal velocity components.

$$\ell(d) = \frac{d_{ml}}{2} - \left( \frac{1}{2} - \frac{2G(d_{ml})}{d_{ml}} \right) \cdot d, \quad 0 \leq d \leq d_{ml}, \quad (1)$$

$$\ell(d) = 2G(d), \quad d > d_{ml},$$

where  $d_{ml}$  is the mixed layer depth (MLD) and  $G(d)$  is the vertical grid spacing as a function of depth. Equation 1 means the surface length scale is equal to half of the MLD, the length scale decreases linearly with depth until the MLD, while beneath MLD the length scales are two times the local vertical grid resolution. Such vertical correlation length scales are designed to minimize any spurious mixing of surface increments beneath the mixed layer (King et al., 2018). It should be noted that satellite OC DA in some previous studies (e.g., Skákala, Bruggeman, et al., 2020; Skákala, Ford, et al., 2018) used a “2D variant,” where surface chlorophyll increments were applied throughout the mixed layer. Both 2D and 3D variants were tested in this study and we have found that they produced almost identical results (not shown here). In this study we will present the outputs of the 3D variant, but these are representative of both methods.

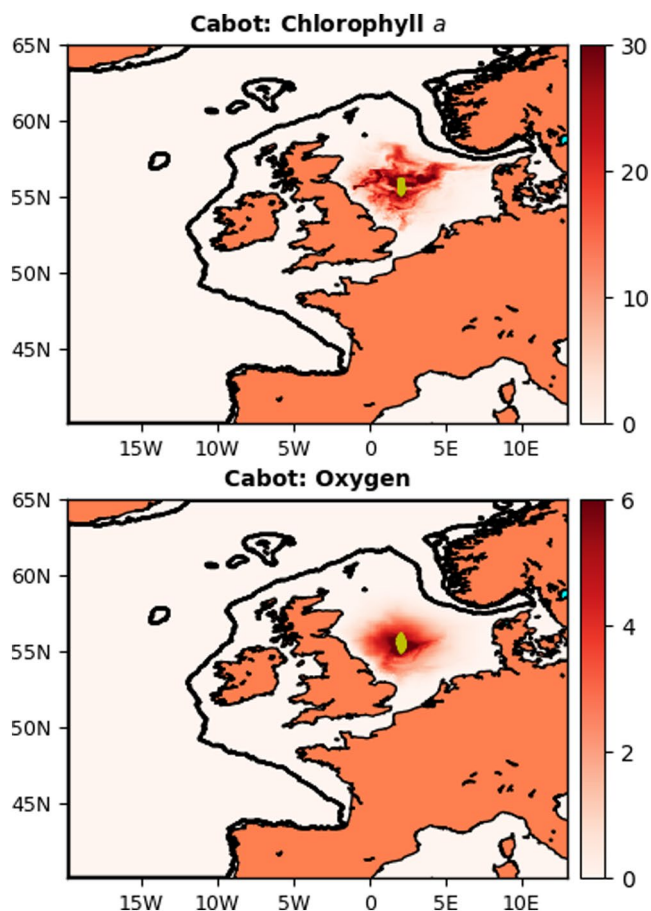
NEMOVAR has two important drawbacks: (1) the background errors (square-root of background variances) have to be specified mostly a priori and those do not always capture how the reanalysis approximates the true state, (2) it does not account for the observational error correlations. Both (1 and 2) tend to artificially increase the impact of the assimilated observations (especially when there is high density of observations) and likely contribute to the fact that biogeochemical reanalysis on the NWE Shelf are relatively insensitive to the precise value of the background-to-observational error ratio (e.g., Skákala, Ford, et al., 2018). Then, provided that the reanalysis state is sufficiently internally consistent, NEMOVAR reanalysis on the NWE Shelf tend to converge for a wide interval of background-to-observational error ratios towards the assimilated observations (Skákala, Bruggeman, et al., 2020; Skákala, Ford, et al., 2018). Improvements could be achieved by using hybrid methods (e.g., background errors calculated as a weighted combination of the parameterized component and a flow-dependent component calculated from an ensemble), or flow-dependent iterative methods based on error diagnostics, such as the scheme of Hollingsworth and Lönnberg (1986), Andersson (2003), and Desroziers et al. (2005) (e.g., Cossarini, Mariotti, et al., 2019; Mattern et al., 2018). For physical assimilation (King et al., 2018) the background errors were estimated using the innovation method of Hollingsworth and Lönnberg (1986) applied to innovations from an existing reanalysis by O'Dea et al. (2017), with background errors between 1 and 3.5 times larger than the observational errors (Table 2). For biogeochemical assimilation the background errors,  $\Sigma\{Q_{bkg}\}$ , were estimated from the observational-to-free run differences and observational errors,  $\Sigma\{Q_o\}$  ( $Q_{bkg}$ ,  $Q_m$ , and  $Q_o$  stand subsequently for the background, model free run and observed concentrations), along the scheme of Skákala, Bruggeman, et al. (2020):

$$\Sigma\{Q_{bkg}\} = \sqrt{\langle [Q_m - Q_o]^2 \rangle - \Sigma\{Q_o\}^2}, \quad (2)$$

which assumes that for a suitable spatiotemporal binning the model and observational errors are uncorrelated (Skákala, Bruggeman, et al., 2020). In the case of the glider data the total observational errors (including representation error) were estimated from the difference between variances of the observations,  $V\{Q_o\}$ , and the variances of the true state,  $V\{Q_i\}$ :

$$\Sigma\{Q_o\} = \sqrt{V\{Q_o\} - V\{Q_i\}}, \quad (3)$$

where the variances of the true state were estimated from the model outputs. This scheme assumes that the observations have zero bias and that (for the limited spatiotemporal range of glider data) the observational errors and the true state deviations from the mean are uncorrelated. After estimating the observational errors for gliders, one proceeds with Equation 2 to estimate the corresponding background errors. The methods based on Equation 2 and Equation 3 produced background and observational errors with



**Figure 1.** The panels show the NEMO-FABM-ERSEM (AMM7) domain with the Cabot glider data locations (chlorophyll data locations for the full May 8–August 15, 2018 mission, oxygen data for a shorter period of May 8–June 29, 2018) marked by yellow dots, as well as glider horizontal area of impact on the reanalysis. The color scale in the two panels shows the weekly (June 23–29, 2018) mean percentage (%) difference between reanalysis and free run in the surface chlorophyll (upper panel) and surface oxygen (bottom panel) concentrations, and reveals the horizontal extent of the glider's impact on the assimilation. The percentage difference is calculated by dividing the absolute value of the difference between reanalysis and the free run, with the free run. The black lines show the boundary of the North-West European (NWE) Shelf (<200 m bathymetry).

comparable values, with background-to-observational error ratios on average between 0.77 and 2.3 (see Table 2). For the two different chlorophyll observational products, the estimate of glider chlorophyll error (using Equation 3) turned out to be on average 22% lower than the satellite OC chlorophyll error.

## 2.4. Glider Data

The study used data from a Slocum glider (Teledyne Webb Research, Falmouth, USA) named Cabot (Unit 345, National Oceanography Center, Southampton) deployed during the AlterEco mission (deployment 454). The glider sampling transect was situated in the Central North Sea (see Figure 1), between May and August 2018, collecting data for temperature and salinity (Seabird SBE42 CTD), colored dissolved organic matter, particulate backscattering, chlorophyll *a* fluorescence (WETLabs ECOpuck), and oxygen (Aanderaa AA4831 optode). After Quality Control (QC) the quenching-corrected chlorophyll (derived from fluorescence) and oxygen concentrations were available for slightly different periods: chlorophyll for May 8–August 15, 2018 and oxygen for a shorter period of May 8–June 30, 2018. The Cabot glider was chosen because it provided high-quality data, but the period of the glider mission was also of special interest for assimilation, since it marks a known discrepancy between the timing of the spring bloom in the model and observations, with the model biased towards a late bloom (see Skákala, Bruggeman, et al., 2020). The QC glider outputs contained a substantial number of data points ( $2 \times 10^6$  for chlorophyll and  $3 \times 10^5$  for oxygen) which were mapped to the model AMM7 grid (each observation to the nearest model grid point). The observations that were mapped on the same day into the same model grid point were then averaged into a single value. The grid-averaging of glider observations is a practice adopted in the physical DA to avoid assimilating many observations at higher resolution than the model can represent. However, our tests have shown that the impact of grid-averaging on the biogeochemical reanalysis was negligible. During each day the glider typically covered three model horizontal grid-cells and for each model horizontal location the glider scanned nearly the full vertical water column.

The glider data (publicly available from [www.bodc.ac.uk](http://www.bodc.ac.uk)) were processed by the National Oceanography Center (NOC) AlterECO team using the GEOMAR glider toolbox for salinity and oxygen lag corrections (following Bittig et al. [2014]). The glider was fitted with a standard non-pumped SBE CT sensor, a WETLabs ECOpuck to measure chlorophyll fluorescence, and an Aanderaa 4,330 oxygen optode. Oxygen data were

corrected based on comparisons between Winkler samples and local crossings with the rest of the AlterEco glider fleet.

The fluorescence sensor on Cabot (454) was calibrated prior to deployment, and recovered data were converted to chlorophyll concentration from raw voltages using the manufacturer supplied calibration routine. The derived chlorophyll record was filtered such that negative values were set to zero. Multiple quenching corrections were tested, including: Hemsley et al. (2015), Swart et al. (2015), Biermann et al. (2015), and Xing et al. (2012). The former three methods rely on the use of algal particle scattering to correct for quenching. However, these approaches proved unsatisfactory for use in Case 2 waters (e.g., the North Sea). Consequently, the Xing et al. (2012) method was adopted. Under this approach the maximum value of chlorophyll concentration above the mixed layer depth (MLD) is extrapolated to the surface for daytime profiles. Night-time chlorophyll profiles are not corrected. MLD is calculated from glider CTD profiles according to the method of Holte and Talley (2009).

### 2.5. Used Metrics (Definitions)

The study uses two metrics: (1) model-to-observation bias ( $\Delta Q_{mo}$ ) defined as

$$\Delta Q_{mo} = \langle Q_m - Q_o \rangle, \quad (4)$$

where, as before,  $Q_m$  are the model free run and  $Q_o$  the observed concentrations (by the observations we will automatically mean the glider data), and (b) Bias-Corrected Root Mean Square Difference (BC RMSD,  $\Delta_{RD}Q_{mo}$ ) defined as

$$\Delta_{RD}Q_{mo} = \sqrt{\langle [Q_m - Q_o - \Delta Q_{mo}]^2 \rangle}. \quad (5)$$

The BC RMSD metric is applied in two different contexts: as a “spatial BC RMSD” and a “temporal BC RMSD.”

In the case of spatial BC RMSD, we calculate for each day ( $t_d$ ) the difference between the model and the observed daily mean, which we call model-to-observation daily bias:

$$\Delta Q_{mo}(t_d) = \langle Q_m(t_d) - Q_o(t_d) \rangle, \quad (6)$$

where  $Q_m(t_d)$  and  $Q_o(t_d)$  are the model free run and the observation data from the day  $t_d$ , and the model free run is taken only from the spatial locations visited by the glider (about 150 model grid points per day). Then we calculate “daily BC RMSD,”  $\Delta_{RD}Q_{mo}(t_d)$ , by applying Equation 5 on each day using the model and the observation daily data, as well as their daily biases:

$$\Delta_{RD}Q_{mo}(t_d) = \sqrt{\langle [Q_m(t_d) - Q_o(t_d) - \Delta Q_{mo}(t_d)]^2 \rangle}. \quad (7)$$

The spatial BC RMSD,  $\Delta_{RD}^S Q_{mo}$ , is then obtained as a time-average of the daily BC RMSD, that is, averaging  $\Delta_{RD}Q_{mo}(t_d)$  through the glider data availability period (100 days for chlorophyll and 53 days for oxygen):

$$\Delta_{RD}^S Q_{mo} = \langle \Delta_{RD}Q_{mo}(t_d) \rangle_{t_d}, \quad (8)$$

where  $\langle \rangle_{t_d}$  means averaging through the interval of  $t_d$  values. Since the glider moves on the model grid dominantly in the vertical dimension, the spatial BC RMSD mostly measures how well the model simulation represents the vertical profile of the glider observations.

The temporal BC RMSD,  $\Delta_{RD}^T Q_{mo}$ , is based on calculating a time series,  $\delta$ , of the daily mean values (for both model,  $\delta_m$ , and the observations,  $\delta_o$ ), averaged through the spatial locations visited by the glider:

$$\delta_m(t_d) = \langle Q(t_d) \rangle, \quad \delta_o(t_d) = \langle Q_o(t_d) \rangle, \quad (9)$$

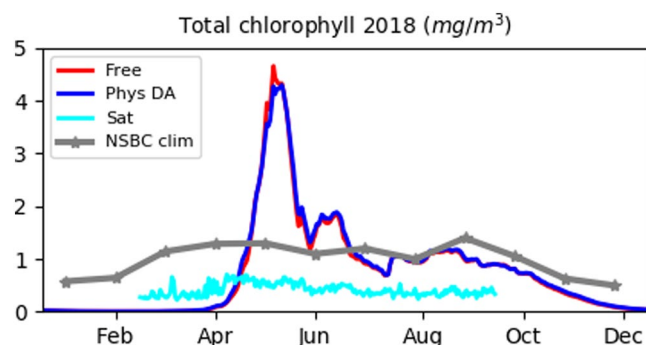
then applying Equation 5 to those time series, with bias understood as the model-to-observation difference in the temporal mean of the time series data:

$$\Delta_{RD}^T Q_{mo} = \sqrt{\langle [\delta_m(t_d) - \delta_o(t_d) - \langle \delta_m(t_d) - \delta_o(t_d) \rangle] ^2 \rangle_{t_d}}. \quad (10)$$

The temporal BC RMSD is designed to capture how the model represents the observed phytoplankton phenology.

It should be noted that the metrics discussed in this section are used to measure “the skill” of the assimilative runs by comparing the simulation outputs to the assimilated glider data, rather than to an independent validation data set. There are two reasons for this: first, to get sufficient validation data for the limited spatio-temporal region of this study is nearly impossible, however, most importantly, this study has no ambition to produce a skill-assessed reanalysis, its ambition is to test the impact of the assimilative system components on the simulated variables. Since the NEMOVAR reanalysis tend to converge under optimal conditions to





**Figure 2.** The mean daily surface chlorophyll concentrations averaged across the NWE Shelf for the year 2018. We compare a model free run used in this study with the physical data assimilation (DA) (the physical DA started on September 1, 2017 from the model free run initial values), the satellite OC and the North Sea Biogeochemical Climatology (NSBC) in situ data set (Hinrichs et al., 2017). The satellite OC chlorophyll values are masked for the October to February period when there is sparsity of data due to the extensive cloud cover and the low solar zenith angle. The model is shown to have an intense and late spring bloom: the observed bloom is much less pronounced than the bloom in the model and the timing of the observed bloom is around the early April, as opposed to the early mid-May bloom simulated by the model.

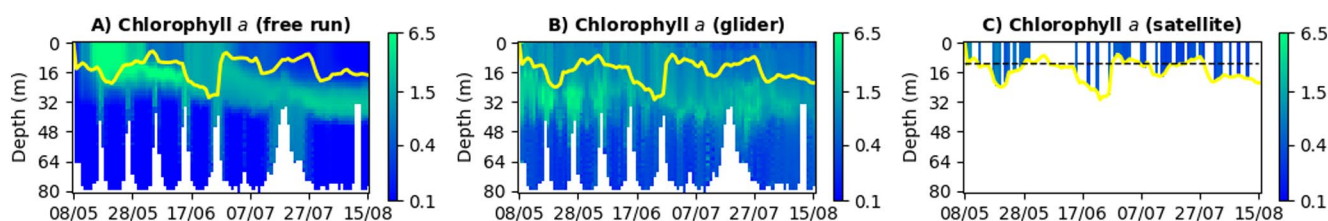
the assimilated observations (Skákala, Bruggeman, et al., 2020; Skákala, Ford, et al., 2018), the performance of the assimilative system can be measured by comparing the model to the assimilated data.

### 3. Results and Discussion

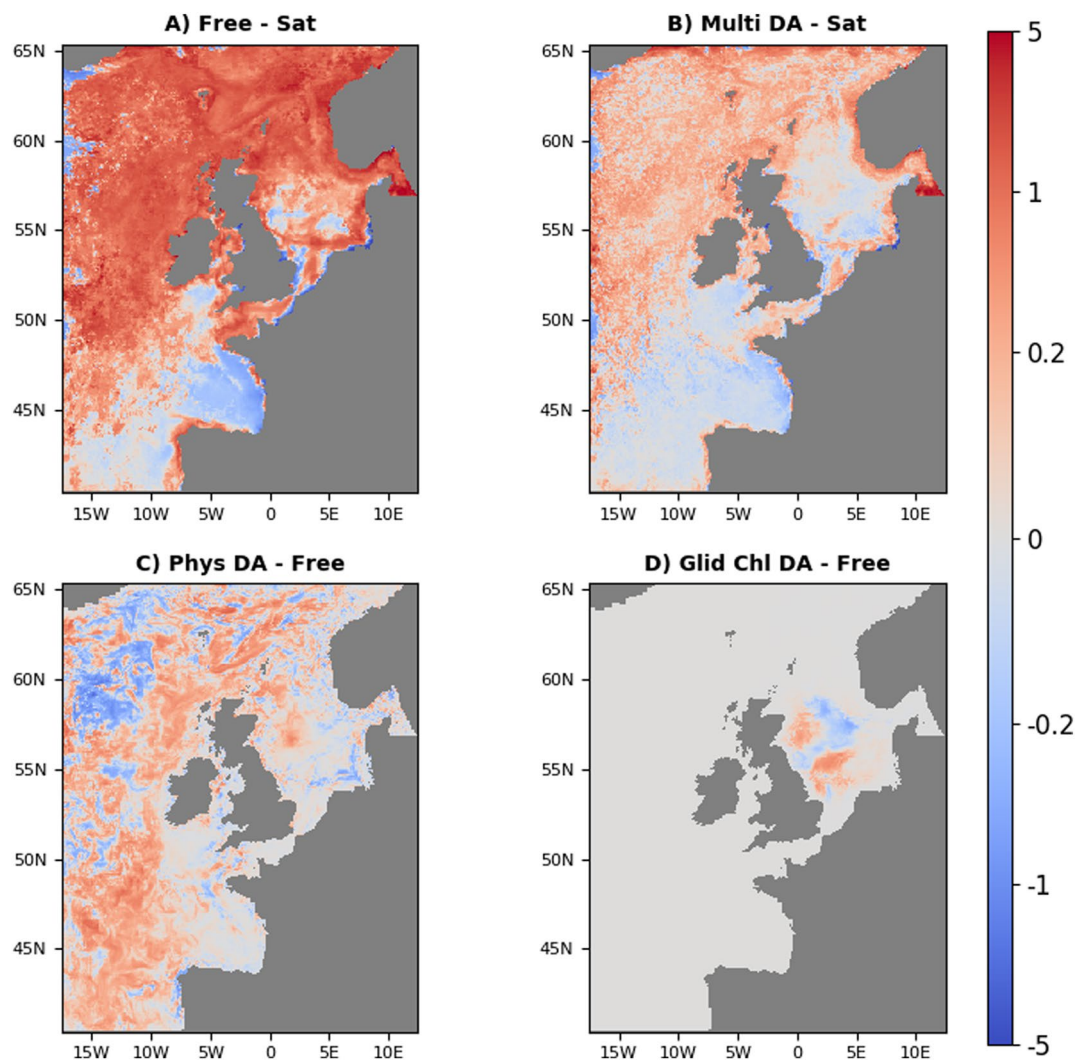
The model free run shows a late and intense spring bloom, with a timing about 1 month later than the bloom observed in the satellite OC and in situ data (Figure 2 and Skákala, Bruggeman, et al., 2020). The late timing of the model bloom is most likely influenced by the interplay between the model vertical mixing scheme and the simulated irradiance (see the discussion in Skákala, Bruggeman, et al. [2020]). The results from the study of Skákala, Bruggeman, et al. (2020) are confirmed by Figure 3, which shows the chlorophyll concentrations in the region measured by the glider between May and August 2018. When the assimilation starts in early May (Figure 3), the glider is in the post-bloom period showing some deep chlorophyll maxima, whereas the model free run has yet to see the onset of the bloom with chlorophyll concentrations predominantly in the mixed layer. Since the North Atlantic sees substantial seasonal patterns in primary productivity (e.g., Henson et al., 2009), the late and intense model bloom has a large impact on the biogeochemical model skill (Skákala, Bruggeman, et al., 2020).

The simulated surface chlorophyll on the NWE Shelf is typically corrected by the assimilation of OC satellite data (Skákala, Bruggeman, et al., 2020; Skákala, Ford, et al., 2018) and the positive impact of satellite OC assimilation on the simulated NWE Shelf surface chlorophyll is shown in Figures 4a and 4b. Around the glider locations, it is shown that both satellite OC and glider chlorophyll assimilation remove the late simulated bloom and improve the surface phytoplankton phenology (Figures 5d, 5f, 6a, and 6b). However, unlike the satellite OC component, the glider chlorophyll assimilation has a limited impact on the model domain (Figure 4d). The horizontal spatial impact of glider assimilation varies with time (Figures 7a and 7b), but any substantial impact of glider assimilation on the simulated chlorophyll (on the level of >10%) is typically constrained to a 50 km radius around the glider location (Figure 7a).

Since glider chlorophyll *a* data were assimilated across the whole water column, the glider chlorophyll assimilation is also able to substantially improve the sub-surface chlorophyll concentrations (Figure 5f). The three skill metrics (bias, spatial and temporal BC RMSD) capturing how the simulated chlorophyll *a* matches with the glider observations were all substantially improved by the glider chlorophyll assimilation: the model bias was reduced by almost 50% (Table 3 and Figure 6d), the spatial BC RMSD by 60% (Table 3) and the temporal BC RMSD by 70% (Table 3). Unlike glider chlorophyll assimilation, satellite OC assimilation

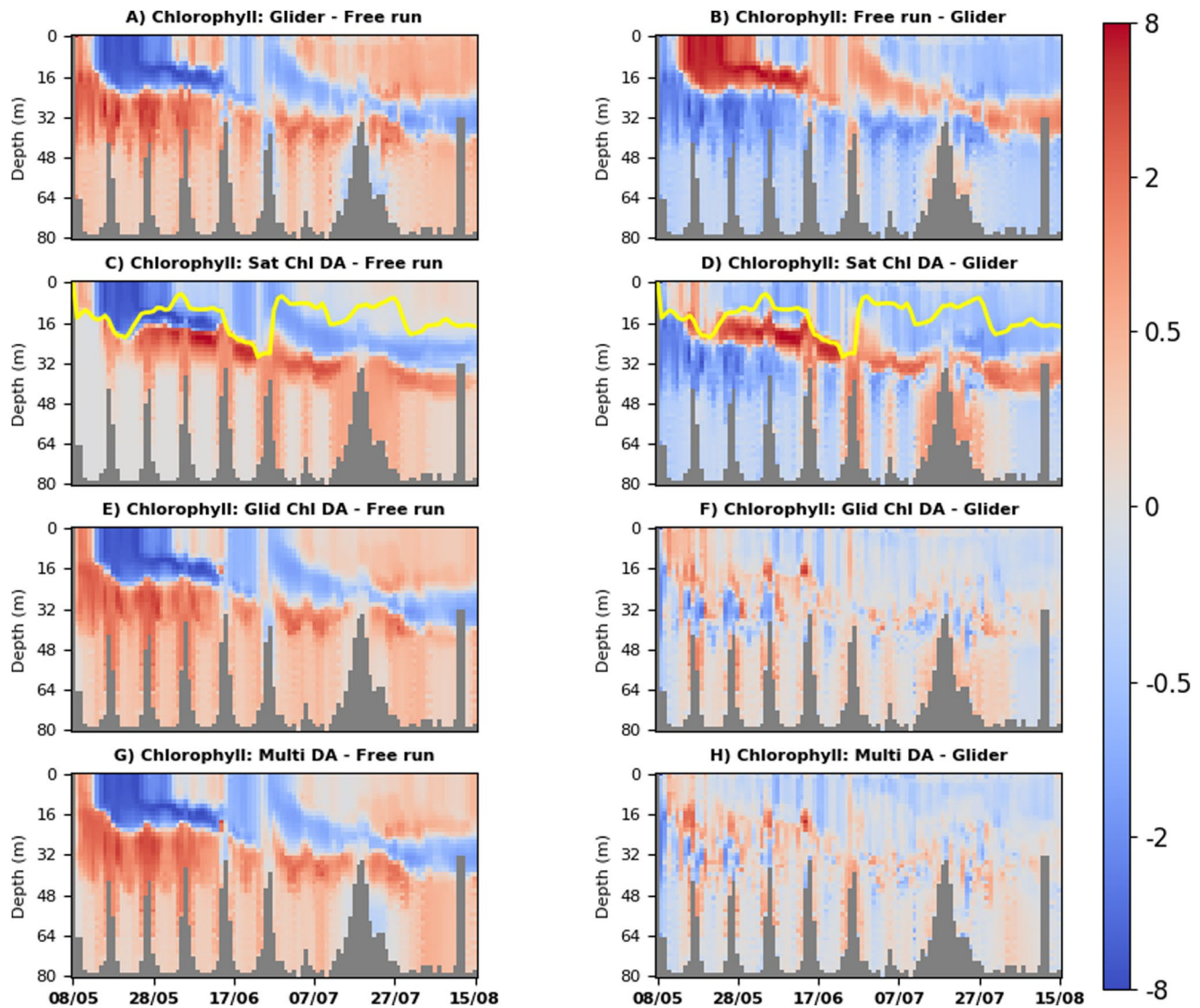


**Figure 3.** Hovmöller diagrams for the model free run and the observations. The left panel (a) shows the model free run outputs for total chlorophyll *a* ( $\text{mg m}^{-3}$ ) horizontally averaged through the area covered by the glider during each day (the plot is depth vs. time). The middle panel (b) shows the same for the glider-observed chlorophyll concentrations and the right panel (c) shows the satellite OC chlorophyll observations at the glider locations. The yellow lines mark the mixed layer depth of the model free run (left-hand panel) and of the physics-assimilative run (the middle and right-hand panels). The satellite observations are plotted in the mixed layer, with the dotted black line broadly corresponding to the average satellite optical depth (Skákala, Bruggeman, et al., 2020). The several missing data in the right-hand plot are due to the cloud cover. The missing data at the bottom of panels (a and b) are due to the varying bathymetry along the horizontal glider trajectory.



**Figure 4.** Comparison of the time median surface chlorophyll *a* distributions ( $\text{mg m}^{-3}$ ) for the simulation period (May 8–August 15, 2018) and the AMM7 domain. The upper two panels show differences in the mean concentrations between the free run (panel a), the multi-platform reanalysis (panel b) and the assimilated satellite OC product (the differences are simulated minus observed chlorophyll). The bottom two panels display the impact of the physical (panel c) and the glider chlorophyll (panel d) assimilation on the simulated surface chlorophyll *a* concentrations by showing the differences between the two reanalysis and the free run (reanalysis minus free run). The NWE Shelf-wide impact of the multi-platform assimilation on the surface chlorophyll *a* concentrations is dominated by the satellite OC assimilation component (not shown here). The multi-platform reanalysis (panel b) is therefore almost identical to satellite OC reanalysis.

updates chlorophyll concentrations only in the mixed layer, but the model dynamics propagates the updates to chlorophyll beneath the mixed layer and gradually spreads the impact of assimilation across the whole water column (Figure 5c). It is encouraging to see that the model dynamics acting on the satellite OC assimilation increments produces a qualitatively similar change to the sub-surface chlorophyll as the glider assimilation (Figures 5c and 5e). We propose a simple explanation based on chlorophyll dynamics: the satellite-only assimilative run removes the intense late model bloom in May, removing chlorophyll from the mixed layer and increasing the light penetrating into the water column. The increased irradiance combined with nutrient availability produces deep chlorophyll maxima around the pycnocline (Figure 5c). Furthermore, the removal of the late (mid-May) bloom in the satellite OC reanalysis means the assimilation also removes the gradually deepening chlorophyll maxima (the July to August period in Figures 3b and 4c), as the nutrients become confined deeper in the water column. The satellite OC assimilation improves both

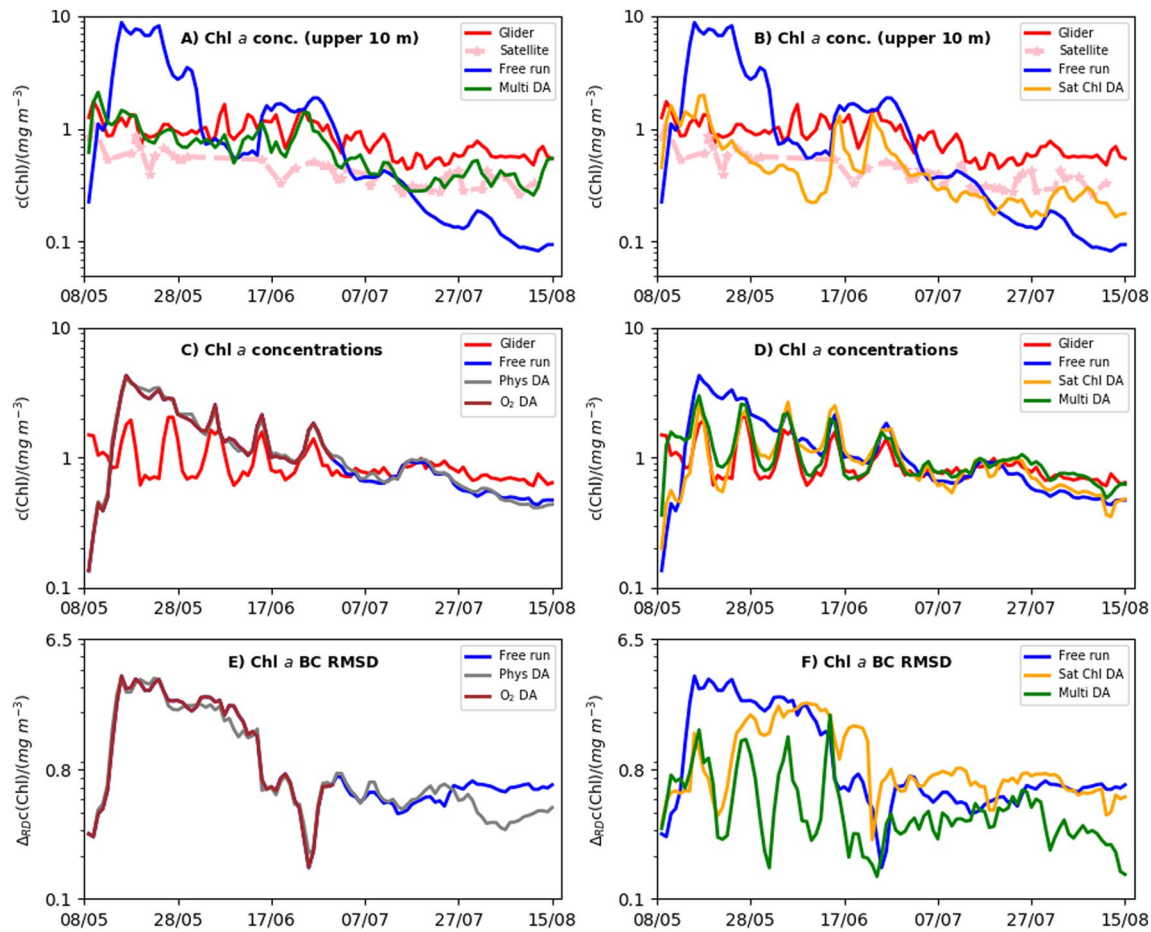


**Figure 5.** The left-hand panels (a, c, e, and g) demonstrate the spatiotemporal impact of the multi-platform system components on the simulated chlorophyll *a* concentrations ( $\text{mg m}^{-3}$ ) by comparing different simulations to the free run. One major advantage of the left-hand side panels is that they demonstrate how the changes introduced by the assimilation propagate vertically with the model dynamics, for example, for the satellite OC assimilation (panel c) that updates the model only in the mixed layer (the MLD is marked in panels c and d by a yellow line). The right-hand panels (b, d, f, and h) show the skill of each component by comparing the simulations to the glider observations. The first row shows the skill of the free run (panel b) and the required changes to the free run in order to better match the glider observations (panel a). The rows beneath the first row compare the chosen reference (free run or glider) with a range of system components: (1) the reanalysis assimilating satellite OC chlorophyll (panels c and d), (2) the reanalysis assimilating glider chlorophyll (panels e and f) and (3) the multi-platform assimilation (joint physical data, glider chlorophyll and oxygen, and satellite chlorophyll assimilation, panels g and h).

temporal BC RMSD (by 55%, Table 3) and spatial BC RMSD (by 15%, Table 3). Although the improvement of BC RMSD is in both cases outperformed by the glider chlorophyll assimilation, the substantial reduction of temporal BC RMSD by 55% in the satellite OC reanalysis is non-trivial, and it is only possible due to (1) a relative consistency between the satellite OC data and the glider surface measurements (Figures 3, 6a, and 6b), and (2) a realistic update to sub-surface chlorophyll driven by the model dynamics.

Whilst the physical DA improves the model representation of both temperature and salinity (Figure 6), it is unable to correct the late model spring bloom (Figure 2) and has a relatively modest impact on chlorophyll concentrations (Figures 3c, 5c, 5e, and 8e). This can be understood as follows: As the pycnocline is primarily controlled by temperature and salinity, we expect that assimilating the physical variables may improve vertical gradients in water density and consequently vertical mixing. However, in the well-mixed nutrient-rich



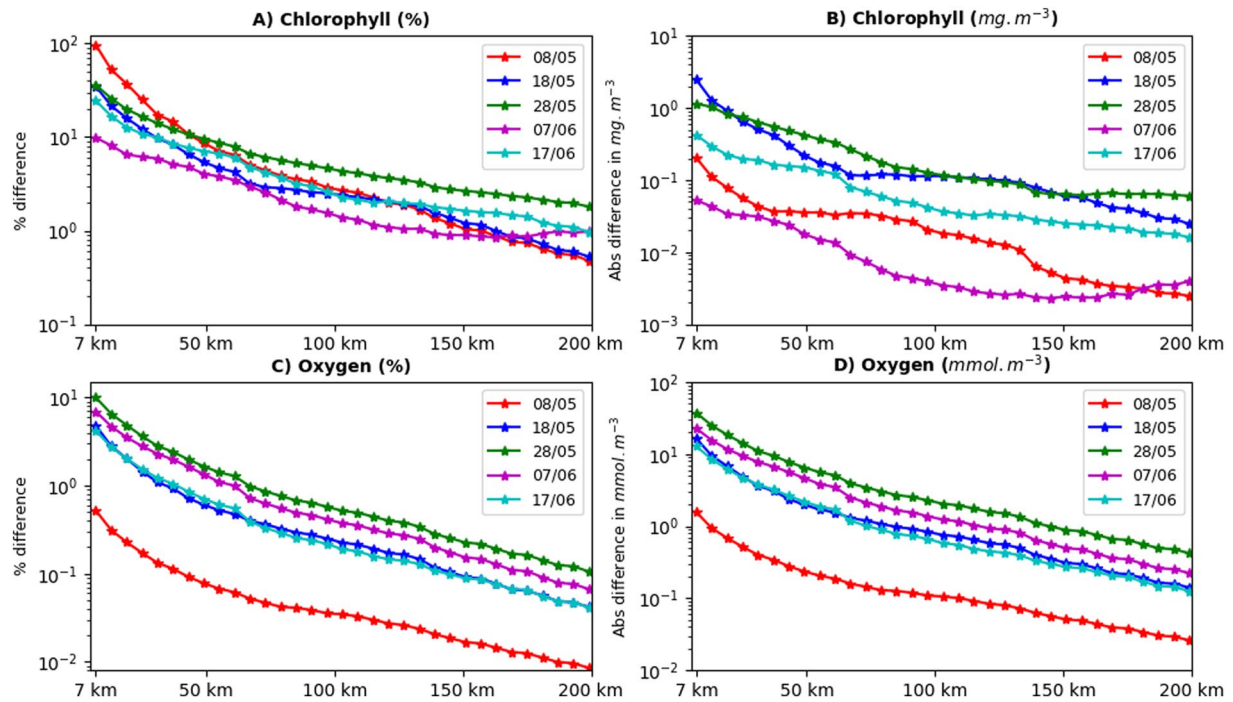


**Figure 6.** The impact of different multi-platform system components on the model chlorophyll concentrations. The panels (a and b) compare the daily chlorophyll values spatially averaged throughout the upper 10 m of the water column, within the part of the model domain visited by the glider. The panels (c and d) show the daily values spatially averaged throughout the whole water column, within the part of the model domain visited by the glider (the daily time series from Equation 9), and the remaining panels (e and f) show the daily BC RMSD (Equation 7) for the same part of the model domain as the panels (c and d). The panels display the skill of the following system components: physical DA (gray color), satellite OC chlorophyll assimilation (orange) and oxygen assimilation (brown). These components are compared with the multi-platform assimilative run (joint physical data, glider chlorophyll and oxygen, and satellite OC chlorophyll assimilation, green color), the free run (blue), the glider observations (red) and the satellite OC data (pink).

waters the onset of the spring bloom depends on the interplay between vertical mixing in the upper oceanic layer and the irradiance (e.g., Huisman et al., 1999; Smyth et al., 2014; Waniek, 2003). Such interplay is closely related to the model atmospheric forcing product for the wind stress and the net incoming short-wave radiation, but an even greater issue is the model response to the used atmospheric forcing product, which consists here mostly of the ERSEM underwater light attenuation, the phytoplankton response to specific light conditions and the model vertical mixing scheme. The ERSEM response to the atmospheric forcing is known to be sensitive to the forcing temporal resolution, leading to shifts of up to one week in the timing of the phytoplankton bloom (Powley et al., 2020). Since the assimilation does not alter the atmospheric forcing, the model mixing scheme, or the phytoplankton response to light, assimilating physical data was found to have relatively modest impact on chlorophyll bias, as well as spatial and temporal BC RMSD (between 5% and 7%, Table 3). However, the impact of physical DA on the simulated phytoplankton could become more substantial within a strongly coupled system (Goodliff et al., 2019). In such system we would mutually update the biogeochemical and the physical increments within a balancing scheme, which could be ideally defined using a two-way coupled physical-biogeochemical model (e.g., Lengaigne et al., 2007). Such development is planned in the foreseeable future.

Finally, we have observed that assimilating glider oxygen into the model has a negligible impact on the simulated chlorophyll concentrations, with a change to the skill metrics of the order  $O(10^{-2})$  percent (Table 3,





**Figure 7.** The horizontal scales for the impact of the glider chlorophyll (panels a and b) and the glider oxygen (panels c and d) assimilation. The impact of glider assimilation is shown for a range of days (between May 8 and June 17, 2018). The impact is calculated by comparing the mean absolute value of the difference in chlorophyll (panels a and b), or oxygen (panels c and d) concentration between the reanalysis and the model free run. The mean absolute difference is shown relative to the free run values (in %, panels a and c), or in the absolute values (panels b and d). The absolute difference was averaged on the circles with 7–200 km radii (the spatial scales shown on the x-axis). The circles were centered around the glider daily mean location. The mean absolute differences (y-axis) are shown on a log-scale, a straight line therefore represents an exponential decrease of the assimilation impact as a function of spatial scale.

see also Figures 5c and 5e). This is expected, as within ERSEM the oxygen variable influences phytoplankton concentrations only indirectly through a complex chain of marine chemical and biological processes (e.g., through influencing remineralization, or nitrification rates, and through the impact of hypoxia on zooplankton).

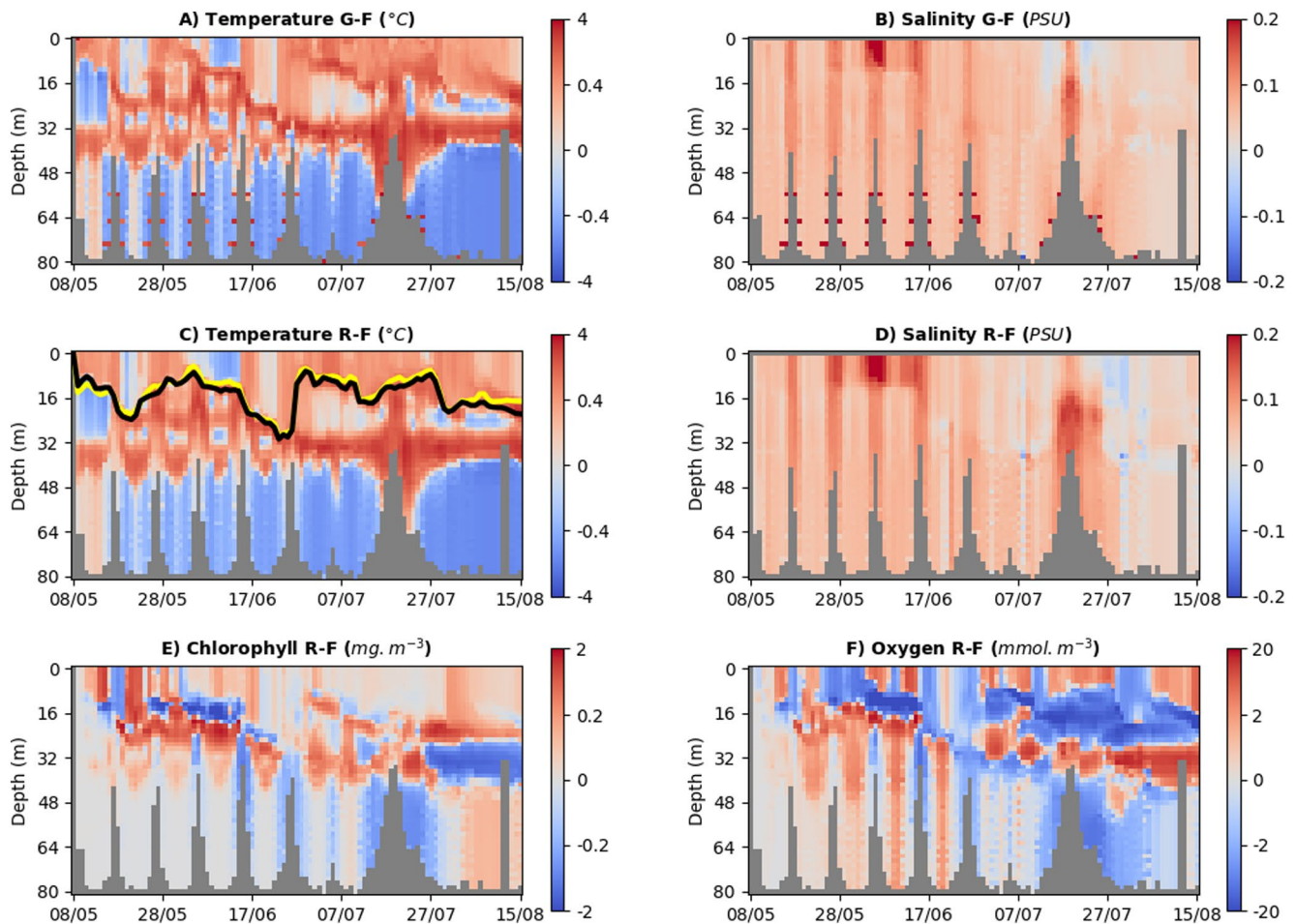
There is a clear discrepancy between the oxygen time series of the glider and the model free run (Figures 9, 10a, and 10b), with glider oxygen concentrations steadily decreasing, while the simulated oxygen peaks in late May (Figures 10a and 10b). Furthermore, simulated oxygen concentrations have a substantial positive bias (25 mmol m<sup>-3</sup>, Table 3; Figures 10a and 10b) relative to the glider observations. Figure 9a clearly shows that photosynthesis is an important driver of the simulated oxygen, producing a large oxygen surge

**Table 3**

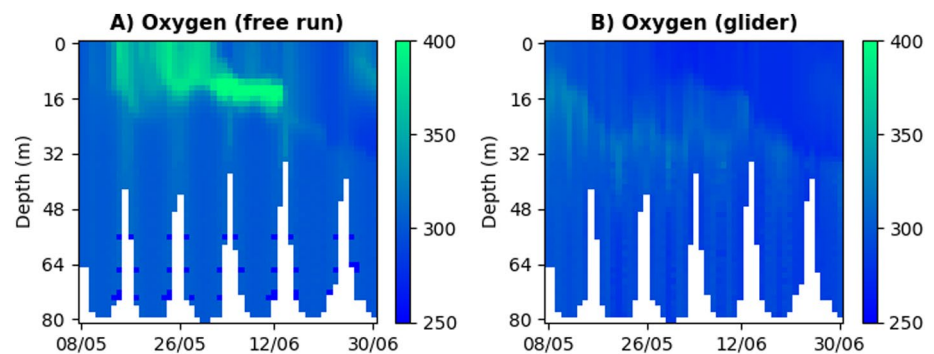
The Table Demonstrates the Skill Measured by Bias (4), Spatial BC RMSD (8) and Temporal BC RMSD (Equation 10) of the Free Run and the Relative (%) Changes to the Skill Carried by the Different Assimilative System Components

Variable	Free run	Phys DA	Sat chl a DA	Glid chl a DA	O <sub>2</sub> DA	Multi DA
Chl a bias	0.31 mg m <sup>-3</sup>	+6.8%	−80%	−46.4%	0%	−56.7%
Chl a temporal BC RMSD	0.77 mg m <sup>-3</sup>	+5.2%	−54.6%	−70.3%	0%	−65.4%
Chl a spatial BC RMSD	1.14 mg m <sup>-3</sup>	−5.5%	−15.3%	−61.9%	0%	−59%
O <sub>2</sub> bias	25 mmol m <sup>-3</sup>	−3.8%	+10.6%	+0.7%	−97%	−98%
O <sub>2</sub> temporal BC RMSD	13.5 mmol m <sup>-3</sup>	−4.3%	+10.8%	−5.4%	−83.8%	−83.7%
O <sub>2</sub> spatial BC RMSD	29.8 mmol m <sup>-3</sup>	−7%	−5.7%	−14.6%	−44.5%	−47.4%

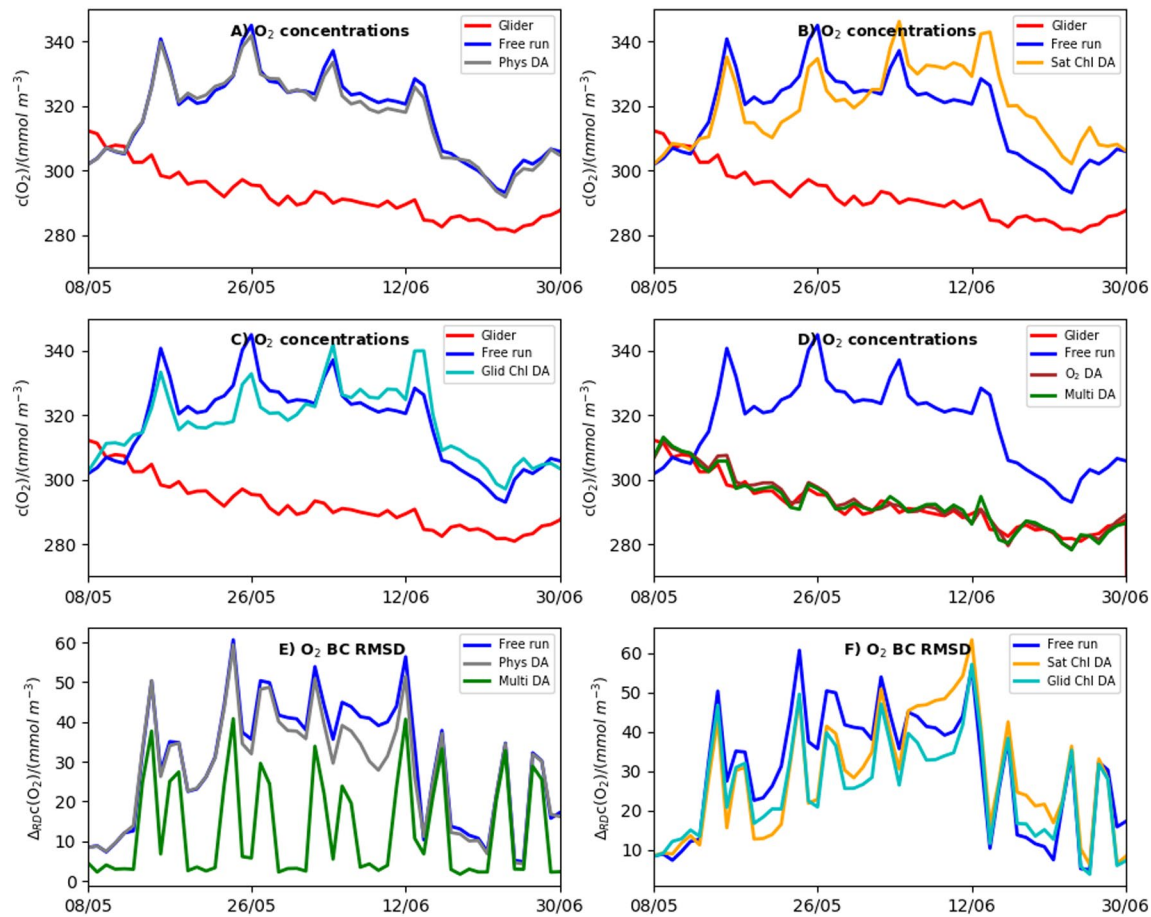
*Note.* The skill compares the model simulations with the glider data. The percentage changes in the columns for the assimilative runs are calculated relative to the free run skill. The negative percentage means that the bias, or (spatial, temporal) BC RMSD is reduced by the specific system component, whilst the positive percentages mean that bias, or (spatial, temporal) BC RMSD, increases.



**Figure 8.** Hovmöller diagrams to demonstrate the impact of physical (SST, in situ temperature and salinity, including Cabot glider data) assimilation on the model variables. The upper row (a and b) shows the difference between glider (“G” in the title) and free run (“F”) outputs for (a) temperature and (b) salinity. The middle row (c and d) shows differences for the same variables between physical reanalysis (“R”) and the free run. The bottom row (e and f) shows the same differences between physical reanalysis and the free run, but for the two biogeochemical variables addressed by this study: total chlorophyll and oxygen. The two lines in the panel c compare the mixed layer depth of the free run (yellow) and of the physical reanalysis (black). The mixed layer depth has been obtained in both cases from the model outputs.



**Figure 9.** Hovmöller diagrams for the model free run and the glider observations. The left-hand panel (a) shows the model free run outputs for oxygen ( $\text{mmol m}^{-3}$ ) horizontally averaged through the area covered by the glider during each day (the plot is depth vs. time). The right-hand panel (b) shows the same for the glider-observed oxygen.

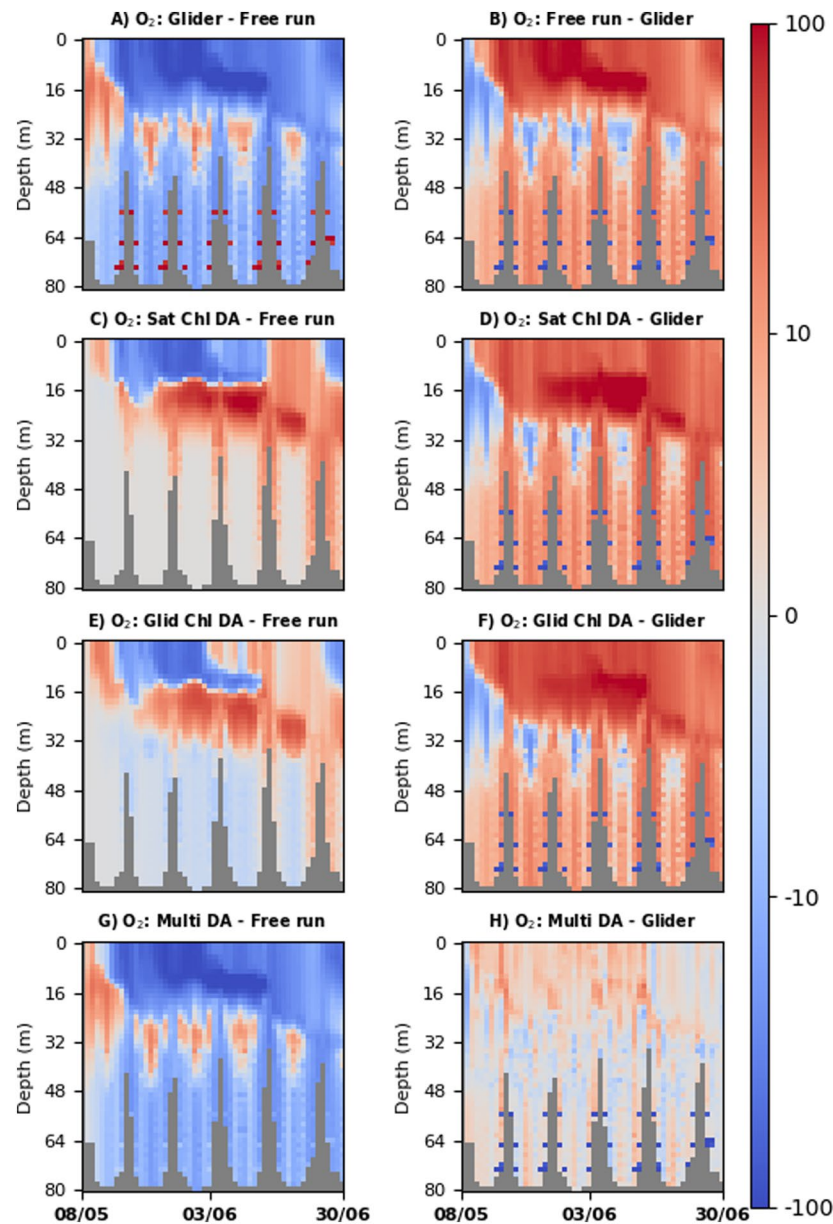


**Figure 10.** The impact of different multi-platform system components on the model oxygen. The panels (a–d) compare the daily oxygen values spatially averaged throughout the whole water column, within the part of the model domain visited by the glider (the daily time series from Equation 9), and the panels (e and f) show the daily BC RMSD (Equation 7). The panels display the skill of the following system components: physical DA (gray color), satellite OC chlorophyll assimilation (orange), glider chlorophyll assimilation (light blue) and oxygen assimilation (brown). These components are compared with the multi-platform assimilative run (joint physical data, glider chlorophyll and oxygen, and satellite chlorophyll assimilation, green color), the free run (blue) and the glider observations (red).

in the mixed layer during the simulated late spring bloom. Some connection between oxygen and chlorophyll concentrations (a proxy for primary productivity) appears also in the glider observations (Figure 9b), with the peak in oxygen concentrations located in the neighborhood of the glider deep chlorophyll maxima (Figure 3b). As for chlorophyll, a simple way to improve simulated oxygen is to assimilate the glider oxygen data into the model (Figures 10d and 11h). Assimilating glider oxygen into the model reduces the oxygen bias by 97%, temporal BC RMSD by 84% and spatial BC RMSD by 45% (Table 3). However, as in the case of chlorophyll, such assimilation has a limited spatial impact on the NWE Shelf (Figures 7c, 7d, and 12c). Unlike chlorophyll, the glider oxygen assimilation horizontal impact reduces with spatial scale at a rate largely independent of time (Figures 7c and 7d). Beyond the 50 km scale the assimilation horizontal impact decays approximately exponentially (a straight line in Figures 7c and 7d), with a halving scale of approximately 40 km, which means the impact is reduced by an order of magnitude at a 130 km scale.

Since the modeled oxygen concentrations are largely driven by the phytoplankton seasonal cycle, it is not surprising that assimilation of either satellite OC, or glider chlorophyll, has a major influence on the simulated oxygen (Figures 11c, 11e, and 12b). The assimilated chlorophyll modifies the simulated oxygen after a necessary time-lag, removing the excess oxygen from the model spring bloom and generating some deep oxygen maxima in early to mid-June (Figures 11c and 11f). The chlorophyll assimilation consistently improves oxygen in the period up to the start of June, but typically degrades oxygen in early to mid-June (Figures 10b, 10d, 10f), mostly due to the surge in oxygen concentrations around the deep oxygen maxima

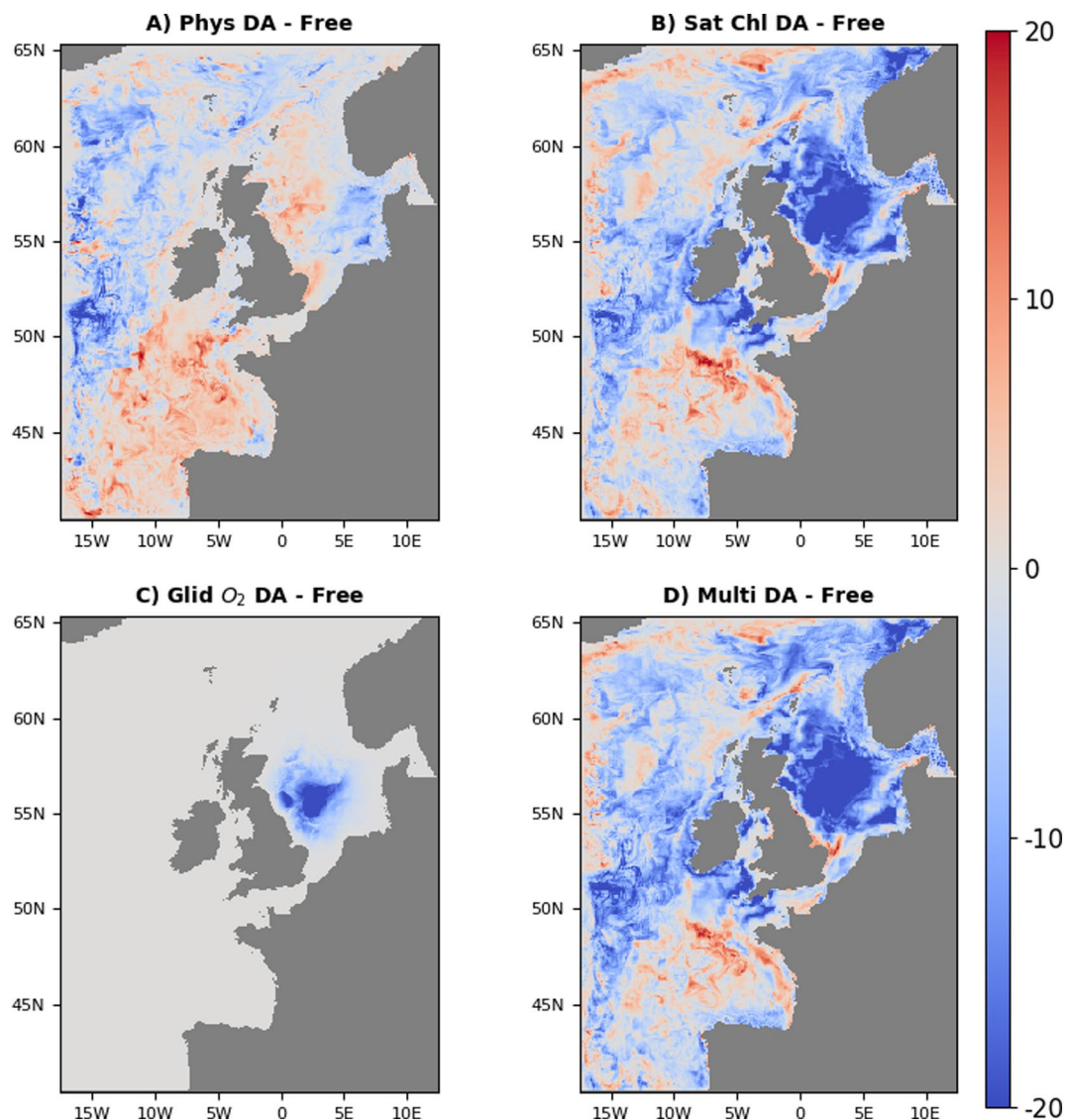




**Figure 11.** The left-hand panels (a, c, e, and g) demonstrate the impact of the multi-platform system components on the simulated oxygen concentrations ( $\text{mmol m}^{-3}$ ) by comparing different simulations to the free run. These panels are particularly well suited to see how chlorophyll assimilation dynamically influences the simulated oxygen. The right-hand panels (b, d, f, and h) show the skill of each component by comparing the simulations to the glider observations. The first row shows the skill of the free run (panel b) and the required changes to the free run in order to better match the glider observations (panel a). The rows beneath the first row compare the chosen reference (free run or glider) with a range of system components: (1) the reanalysis assimilating satellite OC chlorophyll (panels c and d), (2) the reanalysis assimilating glider chlorophyll (panels e and f), and (3) the multi-platform assimilation (joint physical data, glider chlorophyll and oxygen, and satellite chlorophyll assimilation, panels g and h).

(Figures 11c and 11e). The oxygen surge is likely to be partly driven by the deep chlorophyll maxima, for example, by the overestimated chlorophyll concentrations around the deep maxima in the satellite OC assimilation (Figure 5d). However, other drivers such as zooplankton and bacteria respiration are likely to contribute to the deep oxygen maxima. The mechanism for this is suggested by Figures 13c–13f: the chlorophyll assimilation removes phytoplankton biomass from the mixed layer, limiting the resources for the simulated zooplankton and bacteria, and reducing their concentrations. The reduced phytoplankton concentrations



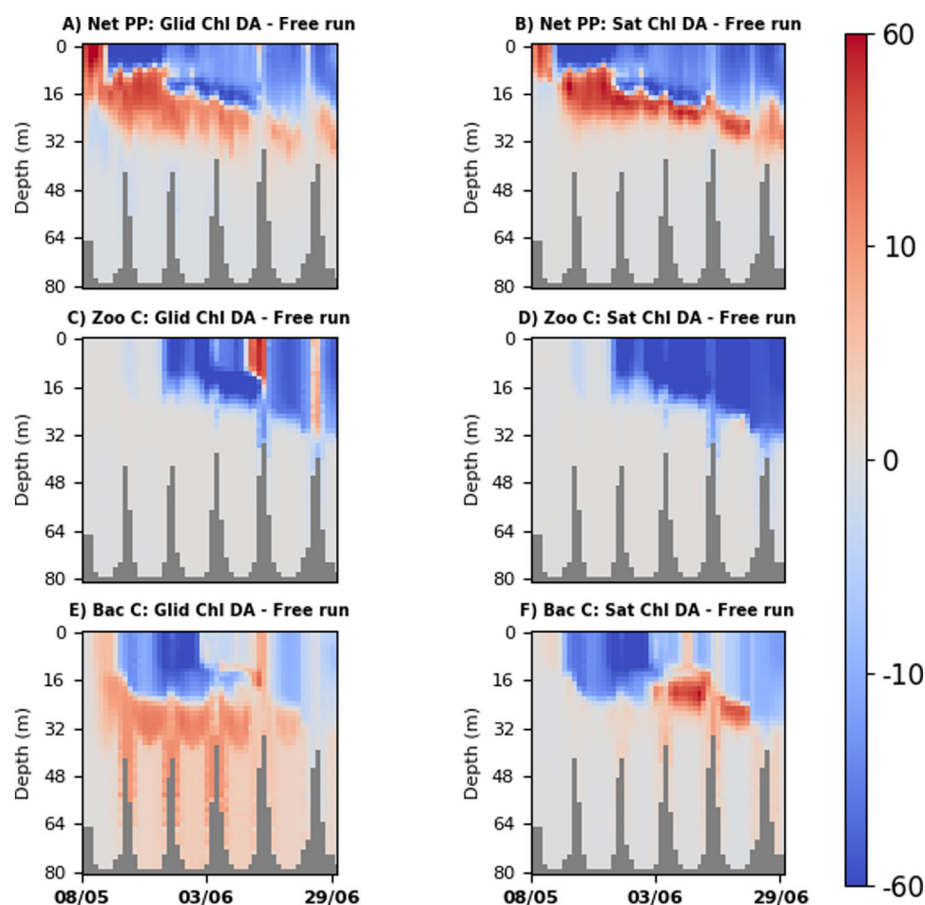


**Figure 12.** Comparison of the time median surface oxygen distributions ( $\text{mmol m}^{-3}$ ) for the oxygen glider data period (May 8, 2018–June 29, 2018). The panels show the impact of the different multi-platform system components on the modeled oxygen by comparing the differences between four reanalysis and the free run. The reanalysis presented in the panels are the physical DA (panel a), the OC satellite chlorophyll assimilation (panel b), the glider oxygen assimilation (panel c) and the multi-platform assimilation (panel d).

seem to have much larger and more consistent impact on the zooplankton concentrations than on bacteria (Figures 13c–13f) and the reduced zooplankton concentration means less oxygen is removed through respiration, which likely produces excess oxygen concentrations.

Compared to chlorophyll assimilation, the physical DA has a relatively modest impact on the simulated oxygen (Figures 8f, 12a, and 12b), but it tends to consistently improve both the oxygen bias, and the spatial and temporal BC RMSD (by 3%–7%, Table 3). The impact of physical DA on the oxygen concentrations can be explained by the lowered oxygen saturation concentrations under the increase in temperature within the reanalysis (Figure 8c).

Finally, we have combined all the assimilative system components (physical DA, satellite OC, glider chlorophyll and oxygen) into a multi-platform assimilative run and we have shown that multi-platform assimilation has the capability to optimally combine the skill of all its components (Figures 4b, 6d, 6f, 9d, and 9e; Table 3). The multi-platform chlorophyll reanalysis is dominated in the vicinity of the glider by the glider



**Figure 13.** The different panels help to interpret the impact of the simulated primary production and respiration on the modeled oxygen concentrations. We show the difference between the glider chlorophyll assimilation (left-hand side panels (a, c, and e), or OC chlorophyll assimilation (right-hand side panels (b, d, and f), and the model free run (always assimilative run minus free run). The difference is shown for (1) the total net primary production ( $\text{mg C m}^{-3} \text{ day}^{-1}$ , panels a and b), (2) total zooplankton carbon concentrations ( $\text{mg C m}^{-3}$ , panels c and d), and (3) heterotrophic bacteria carbon concentrations ( $\text{mg C m}^{-3}$ , panels e and f).

chlorophyll assimilative component (Figures 5e and 5g), whilst further away from the glider it is dominated by the satellite OC assimilation (Figure 4d). The multi-platform oxygen reanalysis is dominated near the glider locations by the glider oxygen assimilation (Figure 10d), whilst further away from the glider locations it is dominantly shaped by the satellite OC assimilation (Figures 12b and 12d).

#### 4. Summary

Present and future glider missions on the NWE Shelf will provide us with three-dimensional (3D) data on some specific biogeochemical variables (presently mostly for chlorophyll and oxygen) combined with physical measurements (e.g., temperature and salinity). These data will be, together with satellite missions, integrated into our ecosystem models by means of a multi-platform assimilative system. It is of crucial importance to understand what observed variables need to be assimilated in order to represent well a target ecosystem indicator, and what assimilation may need to be avoided because it can paradoxically degrade the model skill for the target indicator. Furthermore, different data will be available for different spatial and temporal regions on the NWE Shelf and it is essential to understand how the limitations imposed by the availability of the observational data impact on the quality of the multi-platform reanalysis. To address these questions, we explored the impact of different system components (physical data, satellite OC chlorophyll, glider chlorophyll and oxygen assimilation) on the simulated ecosystem state, using the operational

set-up currently assimilating physical variables and satellite OC chlorophyll. This study has taught us several important lessons:

- (1) Assimilating physical data (SST, in situ temperature, and salinity) has a negligible impact on the simulated phytoplankton bloom. This is because the modeled phytoplankton bloom depends in the North Sea mostly on the model response to the atmospheric forcing (wind stress and solar radiance), which remains unchanged by the temperature and salinity assimilation. Since the phytoplankton bloom is an essential driver of the ecosystem dynamics on the NWE Shelf (Henson et al., 2009), it is quite likely that physical glider DA has a relatively minor importance for the simulated ecosystem dynamics on the NWE Shelf. This is quite different from some other global regions where physical assimilation is either desirable (Anderson et al., 2000; Yu et al., 2018), or can degrade the biogeochemical model skill (Berline et al., 2007; Holt et al., 2014; Park et al., 2018; Raghukumar et al., 2015). Based on this study we would suggest that, at least around the spring bloom in the North Sea, physical assimilation can be used to improve the physical model skill, whilst its impact on the coupled biogeochemical model can be relatively ignored
- (2) In terms of chlorophyll, the glider chlorophyll assimilation is the dominant and best performing component of the multi-platform assimilative system within the 50 km horizontal proximity of the glider. Further away from the glider locations, assimilating satellite OC data substantially improves the surface chlorophyll concentrations, but it can also produce realistic updates to the sub-surface chlorophyll. Since satellite OC assimilation updates chlorophyll only within the mixed layer, the updates to the sub-surface chlorophyll are explained by the model dynamical response to the assimilation. The skill of satellite OC assimilation in sub-surface chlorophyll is important, as glider technology will be able to cover only limited parts of the NWE Shelf and future multi-platform assimilative system will have to rely heavily on satellite data
- (3) The modeled phytoplankton dynamics is impacted by the oxygen concentrations only indirectly, for example, through remineralization, or nitrification rates and the impact of hypoxia on zooplankton (Butenschön et al., 2016). It is therefore hardly surprising that univariate assimilation of oxygen has a negligible impact on the simulated phytoplankton chlorophyll concentrations. This also means that one can assimilate oxygen into ERSEM without worrying about its consequences for the modeled phytoplankton. Such an oxygen assimilation has an obvious advantage in that it outperforms any other run in the model simulation of oxygen
- (4) Two important drivers of the simulated oxygen concentrations are the primary production and respiration. Consequently, assimilating (satellite OC, or glider) chlorophyll was found to have a major impact on the modeled oxygen. The removal of the late model bloom in the reanalysis improves the modeled oxygen, however it produces spurious deep oxygen maxima, partly due to the productivity at the deep chlorophyll maxima and partly due to the reduced respiration by the ERSEM zooplankton. Physical DA has a stronger impact on the oxygen than on chlorophyll (oxygen saturation levels depend substantially on temperature), but it had substantially less impact on the simulated oxygen than the chlorophyll assimilation
- (5) The multi-platform assimilation (joint physical data, glider chlorophyll and oxygen, satellite OC chlorophyll assimilation) combines optimally the skill of its components and always performs comparably to, or better than its best performing component
- (6) Based on the results of this study we expect that the multi-platform system will provide us with improved-quality operational products on the NWE Shelf.

### Data Availability Statement

The authors thank the European Space Agency Climate Initiative “Ocean Color” (<https://esa-ocean-colour-cci.org/>) for providing the ocean color data. The glider data used in the study (<https://doi.org/10.5285/b57d215e-065f-7f81-e053-6c86abc01a82>) are publicly available on [https://www.bodc.ac.uk/data/published\\_data\\_library/catalogue/](https://www.bodc.ac.uk/data/published_data_library/catalogue/). The model was forced by atmospheric ERA5 product of The European Centre for Medium-Range Weather Forecasts (ECMWF, <https://www.ecmwf.int/>). The river forcing data used by the model were prepared by Sonja van Leeuwen and Helen Powley as part of UK Shelf Seas Biogeochemistry programme (contract no. NE/K001876/1) of the NERC and the Department for Environment Food and

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