| 1 | Mapping habitats for the suitability of jellyfish blooms around the UK and |
|----|--|
| 2 | Ireland |
| 3 | |
| 4 | Kennerley A, Lorenzoni I, Luisetti T, Wood L. E and Taylor N. G. H |
| 5 | |
| 6 | |
| 7 | A. Kennerley (ORCID: 0000-0001-5399-4000) (adam.kennerley@cefas.co.uk), N. Taylor |
| 8 | (<u>nick.taylor@cefas.co.uk</u>) and L. Wood (ORCID: 0000-0002-8649-5010) (<u>louisa.wood@cefas.co.uk</u>), Centre |
| 9 | for Environment, Fisheries and Aquaculture Science, Weymouth, Dorset, DT4 8UB, UK. I. Lorenzoni (ORCID: |
| 10 | 0000-0001-8226-5755) (i.lorenzoni@uea.ac.uk), School of Environmental Sciences, University of East Anglia, |
| 11 | Norwich Research Park, Norwich, NR4 7TJ, UK. T. Luisetti (ORCID: 0000-0001-9115-0773) |
| 12 | (tiziana.luisetti@cefas.co.uk), Centre for Environment, Fisheries and Aquaculture Science, Lowestoft, Suffolk, |
| 13 | NR33 0HT, UK. |
| 14 | |
| 15 | |
| 16 | |
| 17 | |
| 18 | |
| 19 | |
| 20 | |
| 21 | |
| 22 | |
| 23 | |
| 24 | |
| 25 | |
| 26 | |
| 27 | |
| 28 | |
| | |

29 Abstract

There is increasing focus on the frequency of jellyfish blooms in the Northeast Atlantic because of negative interactions with humans. However, uncertainty exists as to whether perceptions of increasing bloom frequencies reflect reality due to limitations within long-term population trend data. This study therefore developed and applied a semi-quantitative mapping approach to visualise bloom suitability based on the physiological tolerance of seven jellyfish to ocean temperature, salinity, and a prey index across the Northeast Atlantic. A 10% increase and a 10% decrease in the environmental parameters was then applied to the maps to assess model sensitivity and the potential influence of environmental change on bloom suitability. The study found that optimal physiological temperatures and salinities combined with peaks in prey abundance drove higher bloom suitability and determined distribution. Several locations predicted to be at high risk of bloom occurrence off British and Irish coasts were found to coincide with areas of high anthropogenic activity that could be impacted by blooms. In the absence of long-term datasets on jellyfish population dynamics, the results and methods developed in this study allow an understanding of historic bloom events and predictions of future populations that will be useful in informing monitoring and management.

44 Key words: Habitat suitability mapping, GIS, Northeast Atlantic, UK and Ireland, Jellyfish blooms,
45 Environmental change

54 Introduction

55 Several Scyphozoa, Ctenophora and Hydrozoa medusae taxa (hereafter, referred to as jellyfish) can occur in high 56 numbers causing bloom events (Purcell et al., 2007; Richardson et al., 2009). Jellyfish blooms are perceived to be 57 becoming more common globally, reflected by an increase in coverage within the media and scientific literature 58 (Condon et al., 2012). However, debate exists as to the cause of bloom events and whether the perceived increase 59 is reflective of actual conditions. Furthermore, few long-term datasets on jellyfish population abundance exist to 60 corroborate perceptions (Condon et al., 2012; Sanz-Martín et al., 2016; Pitt et al., 2018). There is a need to better 61 understand current bloom suitability and changes under future environmental scenarios. This is particularly 62 important in coastal areas where blooms coincide with anthropogenic activities such as fishing, aquaculture and 63 tourism, as well as industries that use seawater for cooling such as power generation, due to the detrimental 64 economic and social impacts associated with such events.

65 Though limited, there is evidence to suggest that bloom events are increasing in the Northeast Atlantic 66 (NEA) (Licandro et al., 2010; Palmieri et al., 2015). This is based in part, on increased detections of gelatinous 67 material in annual Continuous Plankton Recorder (CPR) surveys since the early 2000s (Licandro et al., 2010) as 68 well as a synchronous increase in the abundance of shelf and oceanic jellyfish species detected within the last 69 decade (Gibbons and Richardson, 2008). However, this relatively short-term increase could be attributed to the 70 natural long-term population cycles of certain species (Licandro et al., 2010). Within the NEA, intense blooming 71 events have recently been reported off the coasts of the UK and Ireland (Doyle et al., 2008; Licandro et al., 2010; 72 Painting et al., 2014), covering several subareas within the Food and Agriculture Organisation (FAO) major 73 fishing area 27 (see Fig. 1) (hereafter defined as area 27). For example, Pelagia noctiluca blooms have recently 74 occurred in the Porcupine and Rockall area of the NEA, which included Irish coastal shelf waters in 2007 (Doyle 75 et al., 2008), and the interannual variation of jellyfish populations that occurs across the North and Irish Seas 76 which includes blooms of Cyanea capillata, Aurelia aurita an Chrysaora hysoscella (Lynam et al., 2004, 2005). 77 Although historical blooming events occur in the area (Fraser, 1955), there is a widespread perception 78 that jellyfish numbers are increasing, which could impact anthropogenic activities if these populations bloom 79 more frequently (Palmieri et al., 2015). However, it is also possible that the perceived increase in bloom events is

a result of increases in the marine footprint of coastal industries and the subsequent increase in the likelihood of
negative interactions occurring with existing jellyfish populations (Duarte et al., 2013). More generally, the
perceived increases could also be the result of reporting bias attributed to increased attention on blooming events
as they are a significant ecosystem change. Despite the NEA being one of the most studied regions in terms of

84

85

jellyfish populations and their ecology, there is still uncertainty surrounding their abundance over time, their broad distribution patterns and how these blooms vary (Lynam et al., 2004; Doyle et al., 2007; Lucas et al., 2012).

86 One way to improve this uncertainty is to consider spatiotemporal variation in environmental factors to 87 improve understanding of bloom suitability in time and space, and how this may change in the future. As long-88 term population datasets on jellyfish species in the NEA has limitations, additional methodologies are needed to 89 further understand medusae populations and the factors that influences their distributions. Habitat suitability 90 mapping can be used to assess how environmental factors may influence spatiotemporal variation in jellyfish 91 suitability through multicriteria evaluation of biotic and abiotic factors (Store & Kangas, 2001) thought to 92 influence medusae numbers. For example, Collingridge et al., (2014) assessed the suitability of the temperature, 93 salinity, and prey availability in the North Sea for the potential establishment of the invasive Ctenophore 94 Mnemiopsis leidyi using a spatial mapping approach. Such approaches based on the suitability of study species to 95 known environmental conditions allow for predictions of the distribution of gelatinous organisms to be made 96 which could include the suitability of an area for jellyfish blooms. Spatial mapping approaches can therefore 97 support mitigation efforts of potential impacts in locations visualised to be more suitable for blooms. The influence 98 of changes to environmental parameters on a species can also be incorporated into models to estimate how overall 99 suitability could change under different climate scenarios and the potential impact on coastal communities.

100 Temperature (Holst et al., 2012; Purcell et al., 2012; Pascual et al., 2014), salinity (Holst & Jarms, 2010; 101 Purcell et al., 2007) and prey availability (Lilley et al., 2015) are known to increase survival and reproduction 102 rates of jellyfish found in the NEA during their life cycle (Purcell et al., 2012; Collingridge et al., 2014) and are 103 considered key drivers of blooming events of species that may require monitoring. Based on these environmental 104 factors, this study aims to develop and apply a spatial mapping approach to assess the spatiotemporal suitability 105 for jellyfish blooms within area 27. Identified taxa included in mapping scenarios include native and seasonal 106 species that are known to bloom. As limitations exists in long-term population trend data, it is hoped that the 107 outputs from this study will help inform future surveillance and mitigation measures of blooms in the region.

108

109 Methods

110 Mapping area

111 The extent of the present study was defined by subareas within the FAO major fishing area 27 that cover British 112 and Irish coastlines. The subareas selected for bloom assessment were the North Sea (subareas 27.4a-c), the Irish Sea and English Channel (subareas 27.7a-k) and the northwest coast of Scotland and Northern Ireland (subarea
27.6a) (Fig. 1).

115

116 Data Collection and Display

117 In the present study, only the abiotic (sea surface temperature, salinity) and biotic (prey abundance) factors 118 affecting the final medusae phase of the life cycle were considered. In the NEA, this life cycle stage generally 119 occurs during mid-year peaks (Gibbons and Richardson, 2008), so the main output displayed in this study relate 120 to the summer (June-August) for each species. Although it is acknowledged that other life cycle stages will impact 121 the formation of blooms (Holst & Jarms, 2010; Purcell, 2011; Holst, 2012), species and life cycle specific 122 thresholds are largely unknown and could not be included in this study. For example, polyp populations, 123 strobilation success and ephyrae survival rates have been linked to favourable environmental conditions for some 124 species, contributing to increased recruitment that can lead to blooms of adult medusae (Holst & Jarms, 2010; 125 Purcell, 2011; Holst, 2012).

126 NetCDF files were obtained for sea surface temperature (SST) and salinity (PPT) from the Met Office 127 Hadley centre EN4.2.0 ocean series (Good et al., 2013). The NetCDFs represented monthly (from daily averages) 128 SST in degrees centigrade (°C) (converted from kelvin) and salinity in parts per thousand (PPT) for the years 129 2000-2019. These years were chosen as a representation of a present-day time-slice of average conditions that 130 jellyfish currently experience within area 27 (Fig. 1). The NetCDFs were converted into raster data sets (1° latitude 131 X 1º longitude grid resolution) and displayed in ArcMap 10.5 using the WGS84 coordinate system. The monthly 132 data layers were then averaged and visualised according to season (winter = December to February, spring = 133 March to May, summer = June to August and autumn = September to November). The final SST and PPT data 134 within the relevant subareas of area 27 (Fig. 1) were then extracted, focussing on the summer averages (See Online 135 Resource 1, Fig. S1).

Data representing monthly average prey abundance was obtained from the Sir Alister Hardy Foundation for Ocean Science (SAHFOS) Continuous Plankton Recorder (CPR) database from the year 2000 to 2012 (SAHFOS, 2016). Although a restricted subset of years within the present-day time-slice occurred due to data availability at the time of research, the CPR data set acted as the only spatial representation of a known jellyfish prey item for the region. Total eye count zooplankton larger than 2 mm were specifically selected as a prey proxy for jellyfish medusae within the CPR data as these organisms have been reported in the stomachs of jellyfish (Lilley et al., 2009). It is acknowledged that other planktonic organisms of a smaller sizes are an important prey item to some species, particularly at different life cycle stages and that medusae are also known to consume other organisms such as tintinnids and fish larvae (Hansson et al., 2005; Javidpour et al., 2016). However, thresholds relating to additional prey items and different life cycle stages could not be sourced for each species, so the data selected within the CPR layers was defined as a prey index that gives an indication of suitability for adult medusae.

147 The point data from the database that fell within the FAO subareas (Fig. 1) were selected and plotted 148 based on sample longitude and latitude (in decimal degrees) in ArcMap. Each series of seasonal points was 149 converted to grid cell layers that estimated plankton levels across the mapping area using ordinary kriging 150 interpolations (Li & Heap, 2011), with the plankton eye counts set as the z-scores. The interpolation was based 151 on Tobler's first law of geography that states that "everything is related to everything else, but near things are 152 more related than distant things" (Sui, 2004, pg. 269). A spherical semi-variogram model was applied to the 153 kriging interpolation, using the squared differences between z-scores to identify when autocorrelation no longer 154 occurred between samples (Li & Heap, 2011). The annual seasonal raster data layers based on the interpolation 155 were averaged to produce present-day average prey index data layers, consistent with the SST and PPT NetCDFs 156 (Fig. S1).

157

158 Species Selection and Threshold Assignment

159 Study species were chosen based on known ranges and physiological tolerances to ocean temperature, salinity, 160 and the prey index available in the literature. Species included in the study are Aurelia aurita, Chrysaora 161 hysoscella, Cyanea capillata, Cyanea lamarckii, Pelagia noctiluca and Rhizostoma octopus. These are known to 162 bloom and are native to the UK and Irish waters. The siphonophore *Physalia physalis* was also included in the study because until recently it was considered to be an occasional visitor and is an example of a species that could 163 164 increase in the NEA due to climate change (Hinder et al., 2012). Recent P. physalis outbreaks in the Celtic Sea 165 Shelf have been attributed to wind acting as drift pathway into area 27 from more southernly latitudes (Headlam 166 et al., 2020). Increased storminess because of climate change (Hinder et al., 2012) could result in increased 167 numbers of *P. physalis* moving into the area. Therefore, an understanding of present-day suitability and how it 168 could change in the region is required for this economically disruptive species.

For each species, environmental thresholds in relation to each of the biotic and abiotic parameters were derived from the scientific literature, using the following google scholar searches: 'species' AND 'environmental parameter' AND 'threshold' (e.g. *"'Aurelia aurita*' AND 'ocean temperature' AND 'survival'). On occasions where no species thresholds were found, data from the Ocean Biogeographical Information System (OBIS) library of species population occurrence and associated oceanographic conditions were used.

174 Searches produced little information relating to species-specific prey requirements. Species were 175 therefore categorised according to the size of medusae (small and large) as there is evidence that the level of prey 176 consumed is correlated to medusae size (Purcell, 2003; Rosa et al., 2013; Lilley et al., 2015). This enabled 177 suitability thresholds to be assigned for the two medusae groups where species specific data was lacking. It was 178 assumed that by using this methodology, some variation in prey requirements would be represented, as greater 179 prey clearance rates within food webs have been associated with larger medusae (Graham and Kroutil, 2001; 180 Purcell, 2003). The small jellyfish group was defined as medusae with a diameter of 20 cm and under and the 181 large species were defined as having a medusae diameter larger than 20 cm. Medusae sizes stated in the Marine 182 Life Information Network (MarLIN) species list descriptions were used to place each species into the large or 183 small groups. Species that were classed as large medusa were, C. capillata, C. hysoscella and R. octopus and the 184 species classed as small medusae were A. aurita, P. noctiluca, C. lamarckii and P. physalis. However, it must be 185 acknowledged that this does not account for ontogenetic changes in the types and amount of prey consumed by 186 jellyfish species over the course of their life cycle (Graham and Kroutil, 2001; Fleming et al., 2015), which could 187 not be accounted for in this study due to the lack of life cycle specific thresholds available.

188

189 Development of Semi-Quantitative Spatial Maps

190 Semi-quantitative habitat suitability maps were developed by reclassifying the average present-day seasonal data 191 layers according to the physiological thresholds collected for each species (Fig. 2a-c), considering the range of 192 temperatures where survival, reproduction and blooms can occur (Fig. S2). Grid cells within the environmental 193 raster data layers were then given a suitability ranking based on physiological thresholds for each abiotic and 194 biotic parameter: '0', below survival threshold; '1', above survival but below reproduction thresholds; '2', above 195 reproduction but below bloom threshold; '3', above bloom threshold (Fig. 2b). This was repeated for the 'spring', 196 'summer', 'autumn' and 'winter' environmental conditions, but a focus was given to the conditions in summer 197 representing the NEA mid-year peak and this data was displayed (Fig. 3). The three corresponding

reclassifications of the SST, PPT and prey index data layers for each season were then overlaid and suitability at each raster square was assigned using minimum cell statistics (Fig. 2c-f). The lowest reclassification score within the overlay was displayed in the final map as it was assumed to be the limiting factor because of the impact on jellyfish suitability.

202

203 Validation

204 To validate the scoring methodology, environmental data layers were reclassified to assess if the methodology 205 could represent a historic and well reported bloom event and conditions in the years leading up to, during and 206 after the event. P. noctiluca bloom events occurred in 2007 and 2008 within the coordinates 45° N to 58° N and 207 1° W to 26° W, with reports indicating an increase in cnidaria occurrence in the CPR surveys in the area since 208 2002 (Licandro et al., (2010). Data layers for SST, PPT and the prev index representing the average mid-year 209 conditions from the year 2002-2006 within the stated coordinates were extracted from the average present-day 210 environmental data layers. The resultant data layers were reclassified based on the P. noctiluca physiological 211 threshold range. Reclassified layers were overlaid and average suitability scores for each raster square were 212 calculated over the time-period. The process was repeated for the average mid-year conditions for the years 2007 213 and 2008 when the outbreaks of *P. noctiluca* were reported, and also for the years 2009-2012 to represent the 214 average suitability at the end of the time period that data was available for. The average suitability scores at the 215 start of the time period were compared with the average suitability scores at the end of the time period to see if an 216 increase in suitability rankings matched the increases in occurrence reported in the CPR records. The 2007 and 217 2008 average suitability scores were then compared with the two data sets representing non bloom years to see if 218 higher bloom suitability was ranked during the known bloom years.

For the other species mapped in the study, an additional literature search (Lynam et al., 2004; Doyle et al., 2008; Painting et al., 2014; Bastian et al., 2011a; Bastian et al., 2011b) was conducted of their known presentday distributions within area 27 of the NEA (Fig. 1), including instances where blooms have been known to occur. Species occurrence records for each sub-are reported in the literature were compared with the average presentday suitability scores obtained using the mapping methodology to assess accuracy of the mapping approach in terms of jellyfish suitability and their spatial distributions.

226 Map Sensitivity

For each species, an evaluation was carried out of how varying the environmental parameters affected the sensitivity of model outputs. This was used to simulate generic changes to the environment and assess the subsequent effect on bloom suitability across the FAO subareas based on the sensitivity shown within the maps. Values within the SST, PPT and prey index data layers were increased and decreased by 10%, with data layers subsequently reclassified based on species thresholds and then overlaid. The percentage change in the highest suitability scores of grid cells was compared to the initial present-day reclassifications.

233

234 **Results**

235 Present-day Thresholds and Suitability

The physiological threshold ranges (ranges of each environmental parameter where varying levels of suitability was stated) for each species based on the initial literature review are displayed in Table 1. Generally, as the environmental parameters increased, so did the suitability rankings of cells. This was the case for both temperature and the prey index thresholds. The exception was the two *Cyanea* species that were more suited to colder temperatures than the other species. All species were able to survive, reproduce and bloom in marine waters in terms of salinity (< 35 ppt), with tolerance varying little between species.

242 Based on the literature derived species thresholds (Table 1) and the reclassifications of the present-day 243 environmental data layers (Fig. S2), blooming was predicted to be possible for five of the jellyfish species (A. 244 aurita, P. noctiluca, C. capillata, C. lamarckii and C. hysoscella), with populations of the other two species 245 capable of reproducing (R. octopus and P. physalis) at certain times within the year (Table 2). However species-246 specific physiological tolerances resulted in seasonal and spatial variation in environmental suitability for 247 medusae and the ability for populations to bloom. Four of the seven species (A. aurita, P. noctiluca, R. octopus, 248 and P. physalis) were predicted to have the highest frequency of "reproduce" and "bloom" rankings across the 249 subareas during the summer, indicating that higher mid-year temperatures were generally associated with blooms 250 (Table 2). For the two *Cyanea* species, bloom events were predicted to be more likely during spring, however, 251 greater levels of reproduction suitability were attributed to C. capillata for the summer than for C. lamarckii and 252 highest suitability for C. hysoscella was in autumn (Table 2).

253 Generally, bloom rankings occurred more frequently for the smaller scyphomedusae (e.g. A. aurita) 254 compared to the larger species (e.g. R. octopus) (Fig. 3). Waters within the northern and southern Celtic Sea and to the southwest of Ireland (subareas 27.7h, 27.7g and 27.7j respectively) were predicted to be most suited to 255 256 blooms of A. aurita, and P. noctiluca. These southwestern subareas were also predicted to be most suitable for 257 the three species that had no present-day bloom rankings (locations where "reproduce" rankings occurred for R. 258 octopus, C. hysoscella and P. physalis). In terms of the spatial distribution of C. capillata and C. lamarckii, the 259 North Sea was the most suitable region (subarea 27.4), specifically the northern North Sea (subarea 27.4a). 260 However, for some species, areas within the map were not suitable, achieving a "below survival" ranking, despite 261 being predicted as the most suitable time of the year overall. For example, subarea 27.7a achieved below survival 262 rankings for R. octopus, C. hysoscella, and C. capillata despite neighbouring subareas achieving the highest 263 suitability rankings for these species (e.g. subarea 27.g).

264 Reclassifications of the data layers representing the SST, PPT and prey index from 2002-2012 within the coordinates 45° N to 58° N and 1° W to 26° W resulted in general agreement with the patterns of P. noctiluca 265 266 samples that occurred within the CPR data that was reported by Licandro et al., (2010) (Table 3). Negligible areas 267 of below survival rankings (between 1.75 and 2.25% of the area) occurred over the 10-year averages of the data 268 layers. When comparing the earlier years within the time period (2002-2006), with the later years (2009-2012), 269 average suitability was greater during the more recent years (49.17% "reproduce" and "bloom" rankings compared 270 to 38.51%) (Table 3), indicating that the higher suitability scores do coincide with increasing medusae occurrence 271 recorded by the CPR. The average suitability score for P. noctiluca within the 2007 and 2008 average data layers 272 (years when blooms of *P. noctiluca* were reported within the coordinates) was the highest overall (62.72% of 273 raster squares achieving a reproduce or bloom ranking), which included the greatest frequency of bloom rankings 274 (Table 3). The scoring methodology was therefore deemed to be able to roughly determine when conditions within 275 area 27 were suitable for blooms to occur and capture changes in suitability over time when compared to actual 276 records of a species of medusae, despite the exact geographic distributions of suitability not matching actual 277 records.

278

279 Sensitivity Analysis

280 Sensitivity analysis was used to determine the influence that changes in environmental factors would have on

282 greatest predicted suitability increases for the larger scyphomedusae (R. octopus 23% "reproduce" and C. 283 hysoscella 16% "reproduce" increase) that had initially showed relatively low suitability across the mapping site 284 compared to the smaller scyphomedusae. Increased temperature suitability for the smaller medusae was also 285 predicted (A. aurita 5% "bloom", P. noctiluca 5% "bloom" and P. physalis 18% "reproduce" increase), but the 286 increases were proportionally lower than for the large medusae, as larger areas of highest suitability were 287 already predicted to be possible under present-day conditions (Fig. 3). The species associated with colder 288 temperatures were predicted to increase in terms of bloom suitability in response to the decreased temperatures 289 (both C. capillata and C. lamarckii 64% "reproduce" increase). The increase in PPT resulted in little predicted 290 suitability change for all species, but the decrease was predicted to result in larger decreases (between 5% and 291 11% for all species) in suitability across the mapped regions (Fig. 4b).

292 The percentage changes in maximum suitability in relation to the prey index change revealed greater 293 sensitivity within the mapping output compared to the impacts of changing the SST and PPT layers (Fig. 4c). A 294 positive association of jellyfish suitability occurred in response to changes in prey availability by the large 295 medusae and small medusae groups, but for larger medusae, proportionally greater changes were predicted. For 296 example, C. capillata, R. octopus and C. hysoscella (20%, 22% and 19% increase in maximum suitability 297 respectively) were predicted to show greater increases in highest suitability rankings than the more common and 298 smaller A. aurita and P. noctiluca medusae (5% and 17% increase respectively). Only P. physalis was more 299 sensitive in response to temperature where all other species where most sensitive to changes in the prey index 300 data layers.

301 When overlaying the SST, PPT and prey index sensitivity reclassification (Table 4), the 10% increases 302 were predicted to lead to increases in overall suitability rankings for five species. This included large increases 303 (95%+) in the number of cells being ranked with the highest suitability scores during the summer for A. aurita, P. 304 noctiluca, P. physalis (all "bloom" rankings), C. hysoscella and R. octopus ("reproduce" rankings). Overall bloom 305 rankings for the two Cyanea species were the only occasions when an overall decrease in suitability was measured 306 in responses to the 10% increases in the environmental data layers, due to the negative impact of increasing 307 temperatures (Fig. 4a). Conversely, when the 10% decreases to the environmental reclassifications were overlaid, 308 there was a 95-100% decreases in the most suitable rankings that were predicted for each species, including the 309 two Cyanea species, despite the more suitable temperatures.

311 Discussion

Jellyfish blooming events are reported to potentially be increasing in the NEA; however, corroboration of such
trends is challenging due to limitations and gaps within long-term population monitoring datasets (Condon et al.,
2012). Given the potential socio-economic impacts of bloom events, the ability to accurately predict them would
be greatly beneficial in informing monitoring and mitigation efforts.

316 In the present study, habitat suitability mapping was completed for seven bloom forming jellyfish species 317 found in UK and Irish waters, providing new insights into how key environmental factors may contribute towards 318 current and future spatiotemporal trends in bloom occurrence. Based on average environmental data layers and 319 literature derived species thresholds, habitat suitability maps were generated to reflect present-day species 320 occurrence that provide a means to understand how these populations may respond to environmental change. The 321 relatively simplistic approach was able to give a general overview of suitability and bloom risk for a greater 322 number of species and for a wider geographic area than more complex hydrodynamic suitability models of 323 gelatinous medusae that exists within area 27 (e.g. Collingridge et al., 2014).

324

325 **Present-day Jellyfish Populations**

326 Comparison of the present-day suitability maps generated in this study with reported occurrence records for 327 jellyfish medusae indicated that whilst the methodology gave a good overview of suitability for certain species, 328 for others suitability predictions did not match reported present-day distributions. For example, regions to the 329 southwest of the maps (subareas 27.7e, 27.7f and 27.7g, including parts of the Celtic Sea) where highest suitability 330 was determined are known to contain populations of A. aurita, C. hysoscella, P. noctiluca, and P. physalis 331 medusae (Doyle et al., 2008; Licandro et al., 2010; Doyle et al., 2013; OBIS, 2020). This was also the case for 332 more northern and north-eastern areas (subareas 27.4a, 27.4b and 27.6a) where populations of C. capillata, C. 333 lamarckii, P. noctiluca and A. aurita have previously been sampled in high numbers (Lynam et al., 2004; Painting 334 et al., 2014). However, there were examples where 'below survival' was projected for some species in areas where 335 medusae are known to occur (Doyle et al., 2013; Fleming et al., 2015). For example, within in subarea 27.7a there 336 are a number of coastal embayments such as Carmarthen and Tremadoc Bay, the Solway Firth and Rosslare Bay 337 where *R. octopus* is known to occur (Doyle et al., 2013) that the maps ranked as unsuitable. Also, species such as 338 R. octopus, A. aurita and C. hysoscella that are constrained to coastal environments by a benthic stage within their 339 life cycle (Doyle et al., 2007; Doyle et al., 2008) had similar (and in some cases higher) suitability projections for 340 areas further out to sea, which does not match with their known distributions. R. octopus is only recorded in coastal 341 waters (Bastian et al., 2011a), not out into the Celtic Sea where increased suitability was predicted. Both C. 342 capillata and C. lamarckii have a benthic stage, but have been recorded out into the North Sea (Painting et al., 2014) and the Irish Sea (Bastian et al., 2011a) matching the distribution of the present-day suitability projections 343 344 of medusae in this study. However, suitability was not projected in the maps across the whole of these areas (C. 345 capillata ranked as "below survival" to the southeast of subarea 27.4b and all of 27.7a) where samples of these 346 species have been recorded (Painting et al., 2014). The occasions where jellyfish suitability in this study and their 347 known distributions do not match is likely due to data gaps in the environmental data layers and species threshold 348 ranges which are discussed in the limitations

Validation indicated that the methodology could distinguish bloom years from non-bloom years within the time-slices that the maps represent using the *P. noctiluca* blooms in the Celtic Sea in 2007 and 2008 as an example. *P. noctiluca* is not constrained by a benthic phase and increased suitability was measured further out to sea, including areas where it is known to occur which included large areas projected to be suitable for blooms (Licandro et al., 2020). However, the increased bloom suitability projected for *P. noctiluca* in the Celtic Sea during this study are further south of where the species is mostly reported, which includes the bloom events in waters to the Northwest of Ireland (Doyle et al., 2008; Bastian et al., 2011b).

356

357 Potential Changes in Bloom Suitability

358 The variation in jellyfish suitability in response to the sensitivity analysis allows for some conclusions 359 to be drawn as to whether blooms could increase in the study region. The changes in ocean temperature and prey 360 abundance in the present study were predicted to significantly influence bloom suitability, with increases resulting 361 in increasing suitability overall. The predictions suggest that future spikes in jellyfish abundance, within the 362 naturally occurring long-term cyclical patterns (Condon et al., 2013), could occur in areas that are currently 363 suitable for reproduction, if temperatures were to rise (or remain suitable) and prey abundance increases, 364 especially within the seasons that are currently most suitable. The conclusions drawn from the outputs in this 365 study agree with the results of lab-based studies that highlight the importance of temperature and prey abundance 366 for the suitability of common scyphozoa linking increases to blooms (e.g. Wang & Li, 2015).

367 Such environmental conditions do occur in the NEA, which is experiencing rapid increases in water 368 temperatures (Philippart et al., 2011) and plankton blooms attributed to increasing coastal eutrophication and 369 industrial runoff (Cartensen et al., 2001; Almroth & Skogen, 2010; Condon et al., 2013). Projections for the NEA 370 suggest that in some areas 2°C increases in sea surface temperatures are possible by the end of this century 371 (Philippart et al., 2011), which is similar to the increases applied to the temperature layers in the current study 372 during the sensitivity analysis, suggesting increased bloom suitability is possible. No projections were found that 373 suggest future temperatures, salinity and prey abundance will decrease across the region, suggesting future bloom 374 suitability relating to the species studied are unlikely to reduce.

375 If bloom events increase in response to environmental change, there is potential for a rise in detrimental 376 socioeconomic impacts to occur (Graham et al., 2003; Palmieri et al., 2014) such as disruption of coastal tourism 377 activity (Graham et al., 2003; Purcell et al., 2007; Ghermandi et al., 2015) increased mortality in aquaculture 378 finfish (Purcell et al., 2007; Doyle et al., 2008) and increased interference with coastal fishery operations 379 (Knowler, 2005; Palmieri et al., 2014). For example, high numbers of marine finfish (mainly salmon) aquaculture 380 producers are situated across the Northern Irish and Scottish west coasts (particularly within subareas 27.6a and 381 27.4a), which were predicted by this study to be at high bloom suitability for stinging species such as *P. noctiluca* 382 and C. lamarckii. High economic losses and mortalities of farmed salmon were attributed to the P. noctiluca 383 blooming events of 2007 off the Irish coasts (Doyle et al., 2008), and further such events could have significant 384 implications on the industry. The bloom suitable regions for a range of species to the south west of the maps 385 (subareas sub areas 27.7h, 27.7g and 27.7j) and adjacent coasts coincide with locations associated with high levels 386 of coastal tourism, which includes Devon and Cornwall. The areas predicted to be of high bloom suitability in the 387 North Sea and to the Southwest of the UK and Ireland, also coincide with the fishing locations of large capture 388 fishing fleets (MMO, 2018). Both tourism and fisheries have been impacted by annual blooming events of the 389 study species in other geographic locations (Purcell et al., 2007), including the annual disruption to tourism by 390 P. noctiluca in the Mediterranean (Ghermandi et al., 2015) and added cost of bloom bycatch experienced by 391 fisheries in the Adriatic (Palmieri et al., 2015). The potential impact on these industries in the NEA has been 392 acknowledged, but quantification of how impacts will compare with areas that currently experience higher bloom 393 frequencies are unknown (Palmieri et al., 2015). The results of this study could therefore be used to identify 394 locations of potential socioeconomic impact of blooms due to conflict with anthrophonic activity.

396 Limitations and future development

397 Several limitations to the mapping approach taken in this study require acknowledgement. Jellyfish are a 398 relatively understudied group in terms of their environmental requirements, and it is possible that the thresholds 399 ranges do not fully represent the adaptability and plasticity of each species. For example, C. capillata is known 400 to occur further south (Painting et al., 2014) than the temperature suitability distributions suggest in this study 401 (Fig. S2a), There were also instances where it was not possible to obtain species-specific suitability thresholds, 402 particularly in relation to prey requirements and there are additional factors known to influence blooms which 403 could not be included due to the lack of data that exists. Such data gaps included dispersal of medusae by ocean 404 currents, pH, oxygenation, additional prey items as well as how each species undergo ontogenetic shifts in 405 relation to their environmental requirements throughout their life cycle (Condon et al., 2001; Richardson et al., 406 2009, Purcell 2012). This likely led to instances where suitability was underestimated, such as the below 407 survival predictions for *R. octopus* in subareas 27.7a in locations where it is known to regularly occur in coastal 408 bays.

409 There were also occasions where species showed high suitability in subareas adjacent to subareas of 410 low suitability, which is unrealistic due the lack of physical barriers within the marine environment. A possible 411 cause is data gaps within the environmental data layers. Specifically, within the CPR data, large areas within the 412 NEA are not sampled, resulting in prey index suitability rankings being based on fewer samples. Some subareas 413 (e.g. area 27.7a) therefore likely contained underestimations of suitability for certain species (Online resource 1, 414 Fig. S2c). Conversely, areas such as the Celtic sea that are sampled more frequently by the CPR tows showed 415 greater suitability for *P. noctiluca* compared to the Norwest of Ireland (where comparatively fewer CPR 416 samples are taken) where it is more widely reported (Doyle et al., 2008). If detailed data are collected in 417 relation to the prey requirements of different jellyfish (such as more spatial distribution information that also 418 captures the variety of prey items consumed and species-specific threshold ranges), improvements will be made 419 in how the methodology projects suitability distributions.

There is also an absence of polyps and ephyrae distribution data for these species in the NEA (Bastian et al., 2011a), so the relationship between the suitability for potential medusae populations and the other life cycle constraints requires further study. The methodology currently only captures suitability in relation to the medusae phase of the life cycle. Polyp recruitment, strobilation success and ephyrae survival rates have been linked to favourable temperature increases, salinity levels and prey abundances which vary from the medusae

(Holst & Jarms, 2010; Purcell, 2011; Holst, 2012). If the locations of polyp populations can be located for
species constrained to coastal areas such as *A. aurita*, the modelling could be applied to assess the contribution
of polyp populations, strobilation success and ephyrae survival towards bloom formation, particularly if life
cycle specific thresholds can be sourced and the environmental data are reclassified during the season when
each life cycle phase is known to happen in the NEA.

430 However, despite these limitations the models are still able to give a general overview of medusae 431 suitability across area 27 (Fig. 1), replicate known historical bloom events and suggests how changes in certain 432 environmental factors may influence future populations of medusae. The relatively simple methods and data 433 requirements allow much broader applications compared to more complex hydrodynamic modelling that are 434 often constrained to few species and restricted geographic ranges (e.g. Collingridge et al., 2014). The overall suitability of the NEA generated in this study can be used to identify relevant case studies of locations where 435 436 suitability and future bloom risk could be assessed in more detail using higher resolution and more region 437 species assessment of jellyfish suitability.

438

439 Conclusions

440 This study demonstrates that habitat suitability mapping is a valuable tool in helping to predict spatiotemporal 441 trends in bloom suitability under different environmental scenarios, supporting the development of monitoring 442 programmes and strategies to help mitigate socio-economic impacts. However, there is a need to improve data 443 collection and transfer between biological information and modelling approaches to overcome the challenges 444 presented by blooms of jellyfish medusae (Fleming et al., 2015). Despite the identified limitations and data gaps, 445 the methodology provides valuable insight into the distribution of jellyfish bloom suitability across a large 446 geographic area where reports of blooms are increasing that can enable potential impacts in the NEA to be 447 identified. The methods applied in this study could easily be adapted to other species, different life cycle stages 448 and locations if the correct environmental data and species threshold ranges can be sourced to identify where 449 potential bloom impact could occur. The identification of spatial patterns of the likelihood of blooming events can 450 feed into the prioritisation of monitoring and be used to develop risk-based mitigations strategies such as anti-451 jellyfish nets on beaches used for recreation that coincide with greater bloom suitability. Although effective 452 management for marine pest species poses a complex challenge due to the high level of connectivity in marine

- 453 systems (Tidbury et al., 2016), it may be possible to enable sectors at risk to implement preventative measures in
- 454 bloom-prone regions.

456 Table 1 Environmental condition threshold rangess where survival, reproduction and blooms were possible
 457 according to physiological tolerances by species as derived from the literature

| Environmental Condition Thresholds | | | | | | |
|------------------------------------|--|---|---|--|--|---|
| Species | SST (°C) | Refs | РРТ | Refs | Prey Index | Refs |
| Aurelia aurita | Survival: 5-12 Reproduce: 13-14 Bloom: 15+ | Lucas, 2001; Holst, 2012; Purcell et al., 2012; Pascual et al., 2014; Lilley et al., 2015, OBIS, 2020 | Survival: 17-29 Reproduce: 30-34 Bloom: 35+ | Purcell, 1984; Lucas, 2001; Holst & Jarms, 2012; OBIS, 2020 | Survival: 5-39 Reproduce: 40-59 Bloom: 60+ | Morand, 1987; Lilley et al., 2015 |
| Pelagia noctiluca | Survival: 5-11 Reproduce: 12-14 Bloom: 15+ | Avian, 1986; OBIS, 2020 | Survival: 30 Reproduce: 31-34 Bloom: 35+ | Doyle et al., 2008; Rosa et al., 2013; OBIS, 2020 | Survival: 5-39 Reproduce: 40-59 Bloom: 60+ | Purcell, 1984; Rosa et al., 2013; Lilley et al., 2009; Lilley et al., 2015 |
| Cyanea capillata | Survival: 16 Reproduce: 11-15 Bloom: 10 | OBIS, 2020 | Survival: 25-31 Reproduce: 32-34 Bloom: 35+ | Holst & Jarms, 2010; Holst, 2012; OBIS, 2020 | Survival: 30-59 Reproduce: 60-99 Bloom: 100+ | Fancett, 1988; Brewer, 1989; Purcell, 2003 |
| Rhizostoma octopus. | Survival: 14 Reproduce: 15 Bloom: 16-20 | Lilley et al., 2009; Fuentes et al., 2011; OBIS, 2020 | Survival: 30-33 Reproduce: 34 Bloom: 35+ | Purcell et al., 2012; Fuentes et al., 2011; OBIS, 2020 | Survival: 40-59 Reproduce: 60-99 Bloom: 100+ | Fancett, 1988; Lilley et al., 2009; Fuentes et al., 2011 |
| Chrysaora hysoscella | Survival: 13-14 Reproduce: 15 Bloom: 16+ | Sparks, 2001; Purcell et al., 2012 | Survival: 20-31 Reproduce: 32-34 Bloom: 35+ | Holst & Jarms, 2012 | Survival: 30-59 Reproduce: 60-99 Bloom: 100+ | Flynn & Gibbons, 2007; Lilley et al., 2015 |
| Cyanea lamarckii | Survival: 16 Reproduce: 11-15 Bloom: 10 | Lucas, 2001; Purcell et al., 2012 | Survival: 25-31 Reproduce: 32-34 Bloom: 35+ | Lucas, 2001; Holst & Jarms, 2010; Holst, 2012 | Survival: 15-39 Reproduce: 40-59 Bloom: 60+ | Brewer, 1989; Lilley et al., 2015 |
| Physalia physalis | Survival: 2-14 Reproduce: 15-19 Bloom: 20+ | Birsa et al., 2010; OBIS, 2020 | Survival: 30 Reproduce: 31-34 Bloom: 35+ | OBIS, 2020 | Survival: 30-39 Reproduce: 40-59 Bloom: 60+ | Fancett, 1988; Purcell, 2003 |

466 Table 2 Proportion (%) of cell rankings within the maps for each jellyfish medusae within the present-day time
 467 slice per season across the FAO subareas

| Species | Suitability | Winter | Spring | Summer | Autumn |
|----------------------|----------------|---------|---------|---------|---------|
| Aurelia aurita | Below Survival | 0.70% | 0.560/ | 0.000/ | 0.000/ |
| | Survival | 8.72% | 0.56% | 0.00% | 0.00% |
| | | 91.28% | 98.87% | 55.93% | 82.11% |
| | Reproduce | 0.00% | 0.56% | 37.29% | 14.21% |
| | Bloom | 0.000/ | 0.000/ | 6 780/ | 2 690/ |
| Pelagia noctiluca | Below Survival | 10.26% | 2.82% | 2.26% | 1.58% |
| | Survival | 88 210/ | 01 53% | 48 0.2% | 66 320% |
| | Survivar | 00.2170 | 91.5570 | 48.0270 | 00.3270 |
| | Reproduce | 1.54% | 5.65% | 42.94% | 28.42% |
| | Bloom | 0.00% | 0.00% | 6.78% | 3.68% |
| Cyanea lamarckii | Below Survival | 51.79% | 11.30% | 35.59% | 12.11% |
| | Survival | 39.49% | 41.24% | 31.64% | 61.58% |
| | Reproduce | 6.15% | 35.03% | 32.77% | 26.32% |
| | Bloom | 2.56% | 12.43% | 0.00% | 0.00% |
| Cyanea capillata | Below Survival | 51.58% | 87.69% | 51.41% | 35.59% |
| | Survival | 40.00% | 3.59% | 25.99% | 49.15% |
| | Reproduce | 8.42% | 6.15% | 22.60% | 14.69% |
| | Bloom | 0.00% | 2.56% | 0.00% | 0.56% |
| Rhizostoma octopus | Below Survival | 100.00% | 99.44% | 71.19% | 96.07% |
| | Survival | 0.00% | 0.56% | 21.47% | 3.93% |
| | Reproduce | 0.00% | 0.00% | 7.34% | 0.00% |
| | Bloom | 0.00% | 0.00% | 0.00% | 0.00% |
| Chrysaora hysoscella | Below Survival | 100.00% | 99.44% | 39.52% | 29.55% |
| | Survival | 0.00% | 0.56% | 52.10% | 41.48% |
| | Reproduce | 0.00% | 0.00% | 8.38% | 20.45% |
| | Bloom | 0.00% | 0.00% | 0% | 8.52% |
| Physalia physalis | Below Survival | 89.01% | 32.09% | 31.92% | 49.47% |
| | Survival | 10.99% | 65.78% | 45.74% | 41.06% |
| | Reproduce | 0.00% | 1.60% | 21.28% | 9.47% |
| | Bloom | 0.00% | 0.53% | 1.06% | 0.00% |

- **Table 3** Average mid-year suitability scores for *P. noctiluca* within the coordinates 45° N to 58° N and 1° W to
- 471 26° W during time periods when increasing occurrences where being recorded (2002-12) and a period when 472 outbreaks were reported (2007-8)

| Time slice | Below Survival | Survival | Reproduction | Bloom |
|------------|----------------|----------|--------------|--------|
| 2002-2006 | 1.72% | 59.77% | 36.78% | 1.74% |
| 2007-2008 | 2.25% | 35.03% | 49.15% | 13.56% |
| 2009-2012 | 2.25% | 48.58% | 42.93% | 6.21% |

473

474 Table 4 Overall proportional (%) change in the highest cell suitability rankings for each species during the mid 475 year peak in response to the 10% sensitivities applied to the environmental data layers

| 476 | Species | Percentage change in highest overall suitability | | | |
|-----|----------------------|--|-----------------------|--|--|
| | | Plus 10% Sensitivity | Minus 10% Sensitivity | | |
| 477 | Aurelia aurita | 242% | -100% | | |
| | Pelagia noctiluca | 242% | -100% | | |
| 478 | Cyanea lamarckii | -48% | -95% | | |
| | Cyanea capillata | -40% | -100% | | |
| 479 | Rhizostoma octopus | 200% | -100% | | |
| | Chrysaora hysoscella | 178% | -100% | | |
| 480 | Physalia physalis | 95% | -100% | | |

481

482 Figure legends

483 Fig. 1 Subareas off the British and Irish coasts within the FAO major fishing area 27 where jellyfish bloom
484 suitability was assessed (subareas 27.4a-c, 27.6a and 27.7a-k)

485

Fig 2 Pictorial overview of the mapping methodology from the initial reclassification of environmental data
 layers to the final cell overlay. a) the initial environmental parameter to be reclassified, b) the physiological
 threshold ranges and suitability scores used to reclassify the environmental parameters, c) the reclassification of
 the environmental parameter, d) the temperature reclassification, e) the salinity reclassification, f) the prey index
 reclassification, g) the minimum cells statistics overlay of the 3 reclassified environmental parameters

491

492 Fig. 3 Predicted suitability rankings of UK and Irish waters to resident jellyfish species within the mid-year in
 493 terms of SST, PPT and the prey index layers

494

495 Fig. 4 Proportional (%) change in the highest suitability rankings within the maps in response to the +/-10%
 496 SST, PPT and the prey index changes for each species. Grey bars: 10% increase, Black bars: 10% decrease

497 **References**

- 498 Almroth, E. & Skogen. M. D. 2010. A North Sea and Baltic Sea model ensemble eutrophication assessment.
 499 Ambio 39: 59-69.
- Avian, M. 1986, Temperature influence on in vitro reproduction and development of *Pelagia noctiluca*(Forskål). Italian Journal of Zoology 53: 385-391.
- Bastian, T., Haberlin, D., Purcell, J.E., Hays, G.C., Davenport, J., McAllen, R. & Doyle, T.K. 2011a. Largescale sampling reveals the spatio-temporal distributions of the jellyfish Aurelia aurita and Cyanea
 capillata in the Irish Sea. Marine biology 158(12): 2639-2652.
- Bastian, T., Stokes, D., Kelleher, J.E., Hays, G.C., Davenport, J. & Doyle, T.K. 2011b. Fisheries bycatch data
 provide insights into the distribution of the mauve stinger (Pelagia noctiluca) around Ireland. ICES
 Journal of Marine Science 68(3): 436-443.
- Birsa, L. M. Verity, P. G. & Lee R. F. 2010. Evaluation of the effects of various chemicals on discharge of
 and pain caused by jellyfish nematocysts. Comparative Biochemistry and Physiology Part C:
 Toxicology & Pharmacology 151: 426–30.
- 511 Brewer, R. H. 1989. The annual pattern of feeding, growth, and sexual reproduction in Cyanea (Cnidaria:
 512 Scyphozoa) in the Niantic River estuary, Connecticut. The Biological Bulletin 176(3): 272-281.
- 513 Carstensen, J., Dahl, K., Hansen, J., Nygaard, K., Rygg, B., Severinsen, G., Casartelli, S., Schrimpf, W.,
 514 Schiller, C. & Druon, J. N. 2001. Eutrophication in Europe's coastal waters. EEA topic report 7.
- Collingridge, K., van der Molen, J. & Pitois, S. 2014. Modelling risk areas in the North Sea for blooms of
 the invasive comb jelly *Mnemiopsis leidyi* A. Agassiz, 1865. Aquatic Invasions 9: 21–36.
- 517 Condon R. H., Decker M. B. & Purcell J. E. 2001. Effects of low dissolved oxygen on survival and asexual
 518 reproduction of scyphozoan polyps (*Chrysaora quinquecirrha*). In: Purcell J. E., Graham W. M.
 519 & Dumont H. J. (eds) Jellyfish Blooms: Ecological and Societal Importance. Developments in
 520 Hydrobiology, Dordrecht: 89-95.
- 521 Condon, R. H., Graham, W. M., Duarte, C. M., Pitt, K. A., Lucas, C. H., Haddock, S. H., Sutherland, K. R.,
 522 Robinson, K. L., Dawson, M. N., Decker, M. B. & Mills, C. E. 2012. Questioning the rise of
 523 gelatinous zooplankton in the world's oceans. BioScienc 62: 60-169.

| 524 | Condon, R. H. Duarte, C. M., Pitt, K. A., Robinson, K. L., Lucas, C. H., Sutherland, K. R., Mianzan, H. W., |
|-----|--|
| 525 | Bogeberg, M., Purcell, J. E., Decker, M. B. & Uye, S. I. 2013. Recurrent jellyfish blooms are a |
| 526 | consequence of global oscillations. Proceedings of the National Academy of Sciences 110: 1000- |
| 527 | 1005. |
| 528 | Doyle, T. K., De Haas, H., Cotton, D., Dorschel, B., Cummins, V., Houghton, J. D., Davenport, J. & Hays, |
| 529 | G. C. 2008. Widespread occurrence of the jellyfish Pelagia noctiluca in Irish coastal and shelf |
| 530 | waters. Journal of Plankton Research 30: 963–968. |
| 531 | Doyle, T.K., Georges, J.Y. & Houghton, J.D. 2013. A leatherback turtle's guide to jellyfish in the North East |
| 532 | Atlantic. Munibe Monographs. Nature Series 1: 15-21. |
| 533 | Doyle, T. K., Houghton, J. D., Buckley, S. M., Hays, G. C. & Davenport, J. 2007. The broad-scale distribution |
| 534 | of five jellyfish species across a temperate coastal environment. Hydrobiologia 579: 29–39. |
| 535 | Duarte, C. M., Pitt, K. A., Lucas, C. H., Purcell, J. E., Uye, S. I., Robinson, K., Brotz, L., Decker, M. B., |
| 536 | Sutherland, K. R., Malej, A. & Madin, L. 2013. Is global ocean sprawl a cause of jellyfish |
| 537 | blooms? Frontiers in Ecology and the Environment 11(2): 91-97. |
| 538 | Fancett, M. S. 1988. Diet and prey selectivity of scyphomedusae from Port Phillip Bay, Australia. Marine |
| 539 | Biology 98: 503-509. |
| 540 | Fleming, N.E., Harrod, C., Newton, J. & Houghton, J.D. 2015. Not all jellyfish are equal: isotopic evidence |
| 541 | for inter-and intraspecific variation in jellyfish trophic ecology. PeerJ 3: 1110. |
| 542 | Flynn, B. A. & Gibbons, M. J. 2007. A note on the diet and feeding of Chrysaora hysoscella in Walvis Bay |
| 543 | Lagoon, Namibia, during September 2003. African Journal of Marine Science 29: 303–307. |
| 544 | Fraser, J. H. 1955. The plankton of the waters approaching the British Isles in 1953. HM Stationery Office. |
| 545 | Fuentes, V., Straehler-Pohl, I., Atienza, D., Franco, I., Tilves, U., Gentile, M., Acevedo, M., Olariaga, A. and |
| 546 | Gili, J. M. 2011. Life cycle of the jellyfish Rhizostoma pulmo (Scyphozoa: Rhizostomeae) and its |
| 547 | distribution, seasonality and inter-annual variability along the Catalan coast and the Mar Menor |
| 548 | (Spain, NW Mediterranean). Marie Biology 158: 2247-2266. |
| | |

- 549 Ghermandi, A., Galil, B., Gowdy, J. & Nunes, P. A. 2015. Jellyfish outbreak impacts on recreation in the
 550 Mediterranean Sea: welfare estimates from a socioeconomic pilot survey in Israel. Ecosystem
 551 Services 11: 140-147.
- Gibbons, M. J. & Richardson, A. J. 2008. Patterns of jellyfish abundance in the North Atlantic. In Jellyfish
 Blooms: Causes, Consequences, and Recent *Advances*. Springer, Dordrecht: 51-65.
- Good, S. A., Martin, M. J. & Rayner, N. A. 2013. EN4: quality-controlled ocean temperature and salinity
 profiles and monthly objective analyses with uncertainty estimates. Journal of Geophysical Research
 118: 66704-6716.
- Graham, W. M., Martin, D. L., Felder, D. L., Asper, V. L. & Perry, H. M. 2003. Ecological and economic
 implications of a tropical jellyfish invader in the Gulf of Mexico. Biological Invasions 5: 53-69.
- Graham, W.M. & Kroutil, R.M. 2001. Size-based prey selectivity and dietary shifts in the jellyfish, Aurelia
 aurita. Journal of Plankton Research, 23(1): 67-74.
- Hansson, L.J., Moeslund, O., Kiørboe, T. & Riisgård, H.U. 2005. Clearance rates of jellyfish and their
 potential predation impact on zooplankton and fish larvae in a neritic ecosystem (Limfjorden,
 Denmark). Marine Ecology Progress Series 304: 117-131.
- Headlam, J.L., Lyons, K., Kenny, J., Lenihan, E.S., Quigley, D.T., Helps, W., Dugon, M.M. & Doyle, T.K.
 2020. Insights on the origin and drift trajectories of Portuguese man of war (Physalia physalis) over
 the Celtic Sea shelf area. Estuarine, Coastal and Shelf Science: 107033.
- 567 Hinder, S.L., Hays, G.C., Edwards, M., Roberts, E.C., Walne, A.W. & Gravenor, M.B. 2012. Changes in
 568 marine dinoflagellate and diatom abundance under climate change. Nature Climate Change 2(4):
 569 271-275.
- Holst, S. 2012. Effects of climate warming on strobilation and ephyra production of North Sea scyphozoan
 jellyfish. Hydrobiologia 690: 127–140.
- Holst, S. & Jarms, G. 2010. Effects of low salinity on settlement and strobilation of Scyphozoa (Cnidaria): Is
 the lion's mane *Cyanea capillata* (L.) able to reproduce in the brackish Baltic Sea? Hydrobiologia
 645: 53–68.

- 575 Javidpour, J., Cipriano-Maack, A.N., Mittermayr, A. & Dierking, J. 2016. Temporal dietary shift in jellyfish
 576 revealed by stable isotope analysis. Marine biology 163(5): 112.
- 577 Knowler, D. 2005. Reassessing the costs of biological invasion: *Mnemiopsis leidyi* in the Black Sea.
 578 Ecological Economics 52: 187-199.
- 579 Li, J. & Heap, A. D. 2011. A review of comparative studies of spatial interpolation methods in environmental
 580 sciences: performance and impact factors. Ecological Informatics 6: 228-241.
- Licandro, P., Conway, D. V. P., Daly Yahia, M. N., Fernandez de Puelles, M. L., Gasparini, S., Hecq, J.H.,
 Tranter, P. & Kirby, R. R. 2010. A blooming jellyfish in the Northeast Atlantic and Mediterranean.
 Biology Letters 6: 688-691.
- Lilley, M. K., Houghton, J. D. R. & Hays, G. C. 2009. Distribution, extent of inter-annual variability and diet
 of the bloom-forming jellyfish Rhizostoma in European waters. Journals of Marine Biological
 Association of the United Kingdom 89: 39–48.
- 587 Lilley, M. K., Ferraris, M., Elineau, A., Berline, L., Cuvilliers, P., Gilletta, L., Thiéry, A., Gorsky, G. &
 588 Lombard, F. 2014. Culture and growth of the jellyfish *Pelagia noctiluca* in the laboratory. Marine
 589 Ecology Progress Series 510: 265-273.
- Lucas, C. H. 2001. Reproduction and life history strategies of the common jellyfish , *Aurelia aurita* , in
 relation to its ambient environment. Hydrobiologia 451: 229–246.
- Lucas, C.H., Graham, W.M. & Widmer, C. 2012. Jellyfish life histories: role of polyps in forming and
 maintaining scyphomedusa populations. Advances in marine biology 63, 133-196.
- Lynam, C.P., Hay, S.J. & Brierley, A.S. 2004. Interannual variability in abundance of North Sea jellyfish and
 links to the North Atlantic Oscillation. Limnology and Oceanography, 49(3): 637-643.
- Lynam, C.P., Hay, S.J. & Brierley, A.S. 2005. Jellyfish abundance and climatic variation: contrasting
 responses in oceanographically distinct regions of the North Sea, and possible implications for
 fisheries. Journal of the Marine Biological Association of the United Kingdom, 85(3): 435-450.
- 599 MMO, 2019. UK SEA FISHERIES STATISTICS 2018. Dandy Booksellers Limited.
- Morand, P., Carré, C. & Biggs, D. C. 1987. Feeding and metabolism of the jellyfish *Pelagia noctiluca*(scyphomedusae, semaeostomae). Journal of Plankton Research 9: 651–665.

- 602 OBIS, 2020. Species Distribution records. [Dataset] Available: Ocean Biogeographic Information System.
 603 Intergovernmental Oceanographic Commission of UNESCO. Available at: <u>www.iobis.org.</u>
- Painting, S., Lynam, C., Barry, J. & Stephens D. 2014. IFMA Module 1h Emerging Threats: Jellyfish
 outbreaks and climate change. Cefas contract report C6185.
- Palmieri, M. G., Barausse, A., Luisetti, T. & Turner, K. 2014. Jellyfish blooms in the Northern Adriatic Sea:
 Fishermen's perceptions and economic impacts on fisheries. Fisheries Research 155: 51-58.
- Palmieri, M. G., Schaafsma, M., Luisetti, T., Barausse, A., Harwood, A., Sen, A. and Turner, R.K. 2015.
 Jellyfish blooms and their impacts on welfare benefits: recreation in the UK and fisheries in Italy.
 In: Coastal Zones Ecosystem Services. Springer Cham: 219-240.
- Pascual, M., Fuentes, V., Canepa, A., Atienza, D., Gili, J. M. & Purcell, J. E. 2015. Temperature effects on
 asexual reproduction of the scyphozoan *Aurelia aurita*: differences between exotic (Baltic and Red
 seas) and native (Mediterranean Sea) populations. Marine Ecology 36: 994-1002.
- Philippart, C. J. M. Anadón, R., Danovaro, R., Dippner, J. W., Drinkwater, K. F., Hawkins, S. J., Oguz, T.,
 O'Sullivan, G. & Reid, P.C. 2011. Impacts of climate change on marine ecosystems: observations,
 expectations, and indicators. Journal of Experiential Marine Biology and Ecology 400: 52-69.
- 617 Pitt, K. A., Lucas, C. H., Condon, R. H., Duarte, C. M. & Stewart-Koster, B. 2018. Claims that anthropogenic
 618 stressors facilitate jellyfish blooms have been amplified beyond the available evidence: a systematic
 619 review. Frontiers in Marine Science 5, 451.
- 620 Purcell, J. E. 1984. Predation on fish larvae by *Physalia physalis*, the Portuguese man of war. Mar Ecol Prog
 621 Ser. 19: 189-191.
- 622 Purcell, J. E. 2003. Predation on zooplankton by large jellyfish (*Aurelia labiata, Cyanea capillata, Aequorea*623 *aequorea*) in Prince William Sound, Alaska. Marine Ecology Progress Series 246: 137–152.
- 624 Purcell, J.E. 2011. Jellyfish and ctenophore blooms coincide with human proliferations and environmental625 perturbations.
- Purcell, J. E., Uye, S. I. & Lo, W. T. 2007. Anthropogenic causes of jellyfish blooms and their direct
 consequences for humans: a review. Marine Ecology Progress Series 350: 153-174.

- Purcell, J. E., Atienza, D., Fuentes, V., Olariaga, A., Tilves, U., Colahan, C. & Gili, J. M. 2012. Temperature
 effects on asexual reproduction rates of scyphozoan species from the northwest Mediterranean Sea.
 Hydrobiologia 690: 169–180.
- Richardson, A. J., Bakun, A., Hays, G. C. & Gibbons, M. J. 2009. The jellyfish joyride: causes, consequences,
 and management responses to a more gelatinous future. Trends in Ecology and Evolution 24: 312322.
- Rosa, S., Pansera, M., Granata, A. & Guglielmo, L. 2013. Interannual variability, growth, reproduction and
 feeding of *Pelagia noctiluca* (Cnidaria: Scyphozoa) in the Straits of Messina (Central Mediterranean
 Sea): Linkages with temperature and diet. Journal of Marine Systems 111: 97-107.
- 637 SAHFOS, 2016. Records of plankton samples with the North East Atlantic from 2000-12. DOI:
 638 <u>https://doi.org/10.7487/2014.170.1.31</u>
- 639 Sanz-Martín, M., Pitt, K. A., Condon, R. H., Lucas, C. H., Novaes de Santana, C. & Duarte, C. M. 2015.
 640 Flawed citation practices facilitate the unsubstantiated perception of a global trend toward increased
 641 jellyfish blooms. Global Ecology and Biogeography. 25: 1039-1049.
- Sparks, C., Buecher, E., Brierley, A. S., Axelsen, B. E., Boyer, H. & Gibbons, M.J. 2001. Observations on
 the distribution and relative abundance of the scyphomedusan *Chrysaora hysoscella* (Linné, 1766)
 and the hydrozoan *Aequorea aequorea* (Forskål, 1775) in the northern Benguela ecosystem.
 Hydrobiologia 451: 275-286.
- 646 Store, R. & Kangas, J. 2001. Integrating spatial multi-criteria evaluation and expert knowledge for GIS-based
 647 habitat suitability modelling. Landscape and urban planning, 55(2): 79-93.
- Sui, D. Z. 2004. Tobler's first law of geography: A big idea for a small world? Annals of the Association of
 American Geographers 94: 269-277.
- Tidbury, H. J., Taylor, N. G., Copp, G. H., Garnacho, E. & Stebbing, P. D. 2016. Predicting and mapping
 the risk of introduction of marine non-indigenous species into Great Britain and Ireland. Biological
 Invasions 18: 3277-3292.
- Wang, N. & Li, C. 2015. The effect of temperature and food supply on the growth and ontogeny of Aurelia
 sp. ephyrae. Hydrobiologia 754: 157-167.

Figure 1





Initial reclassification (repeated for SST, PPT and Prey)



Reclassification overlay (minimum cell statistics)



a)EnvironmentalParameter

b) Threshold ranges and suitability scores

c) Reclassification





d) Temperature Reclassification

e) Salinity Reclassification

f) Prey Reclassification

g) Cell Overlay

658 Figure 3



Fig 4a



676 Fig 4b



692 Fig 4c

