

Functional Structure/Activity Relationships

**Differential effects of quercetin and its two derivatives
(isorhamnetin and isorhamnetin-3- glucuronide) in
inhibiting proliferation of human breast cancer MCF-7 cells**

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1 **Differential effects of quercetin and its two derivatives (isorhamnetin and**
2 **isorhamnetin-3- glucuronide) in inhibiting proliferation of human breast cancer MCF-7**
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17 **ABSTRACT:** Quercetin (Que) has consistently been reported to be useful cytotoxic compound
18 *in vivo and in vitro*, but little is known on its metabolites. Here we examined and compared
19 cytotoxic effect of Que and its water-soluble metabolites, isorhamnetin (IS) and
20 isorhamnetin-3-glucuronide (I3G) in human breast cancer MCF-7 cells to uncover their
21 tumor-inhibitory mechanism and structure-function relationship. The results showed that Que,
22 IS and I3G could dose-dependently inhibit the growth of MCF-7 cells, and the cytotoxic effect
23 was ranked as Que > IS > I3G. Furthermore, Que, IS and I3G mediated the cell-cycle arrest
24 principally in S phase, followed by the decrease in the number of G0/G1 and G2/M, and 70.8%,
25 68.9% and 49.8% MCF-7 tumor cells entered early phase apoptosis when treated with 100 μ M
26 Que, IS and I3G for 48 h, respectively. Moreover, induction of apoptosis by Que, IS and I3G
27 were accompanied with the marginal generation of intracellular ROS. Given these results, Que,
28 IS and I3G possess strong cytotoxic effect through a ROS-dependent apoptosis pathway in
29 MCF-7 cells.

30 **KEYWORDS:** *Quercetin, isorhamnetin, cytotoxicity, cell circle, apoptosis*

31 INTRODUCTION

32 Breast cancer is the leading cause of tumor death among women,^{1,2} and more and more
33 reports consistently show that regular consumption of fruits and vegetables is strongly
34 associated with reduced risk of tumor.^{3,4} Furthermore, many benefits of fruits and vegetables
35 are shown to be due to the ingestion of vast flavonoids, a type of functional compounds with a
36 common phenylbenzopyrone structure (C6-C3-C6).^{5,6} As one of the primary flavonoids,
37 quercetin (3,3',4',5,7-pentahydroxyflavone) has been reported to have anti-tumor effect on
38 many tumor cells, which may be related to catechol moiety in B ring and free hydroxyl groups
39 in the quercetin structure.⁷⁻¹¹

40 Recent studies have showed that quercetin can be metabolized into various sulphated,
41 glucuronidated and methylated forms in different organs, such as liver, kidney, colon and small
42 intestine, and its metabolites may still act as antioxidants with higher hydrophily.¹² Our
43 previous studies have also showed that QS (quercetin-5',8-disulfonate) can possess remarkably
44 high anti-tumor activity in human breast cancer MCF-7 cells,¹¹ indicating that sulfated
45 metabolites of quercetin may play an important role in cytotoxic effects. For this reason,
46 whether the methylated-, methylated- and glucuronidated- metabolites of quercetin also play a
47 crucial role in quercetin-induced biological effects remains poorly understood, and few studies
48 reported the cytotoxic effect of them. Accordingly, it is necessary to further study the
49 anti-tumor effect and their molecular mechanism of methylated quercetin and other metabolites.
50 Significantly, it is interesting to note that the difference in cytotoxic activities between
51 quercetin and its metabolites may also help understand the structure-activity relationship of the
52 tested compounds.¹³

53 With this in mind, in present study we chose the water-solubility metabolites Isorhamnetin
54 (IS) and Isorhamnetin-3-glucuronide (I3G) (Fig. 1), which were synthesized by Paul W.
55 Needs,¹⁴ to evaluate the cytotoxic effect and make clear the structure-activity relationship of
56 them by investigating and comparing the cytotoxicity, cell cycle distribution, apoptosis,
57 cellular morphology and intracellular ROS generation in human breast cancer MCF-7 cell line,
58 and ultimately purify the molecular mechanism. These findings can help understand the
59 structure-activity relationship in tumor-inhibitory effects and have important implications for
60 the potential use of the quercetin and its metabolites in the treatment of human breast cancer.

61

62 **MATERIALS AND METHODS**

63 **Chemicals and reagents**

64 The quercetin was the product of National Institute for the Control of Pharmaceutical and
65 Biological Products (Beijing, China) and its purity (>98%) was verified by UPLC. IS and I3G
66 (Fig. 1) were synthesized and presented as solid sodium salts form by Paul W. Needs.¹⁴ EDTA
67 and Triton X-100 were the products of Sinopharm Chemical Reagents Co., Ltd (Shanghai,
68 China). 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT), dimethyl
69 sulfoxide (DMSO), Rnase-A and propidium iodide (PI) were purchased from Sigma-Aldrich
70 (St. Louis, Mo, USA). Dihydroethidium (DHE) and dichlorofluorescein diacetate (DCFH-DA)
71 were obtained from BestBio Co. (Shanghai, China). Millipore Milli Q-plus System (Millipore,
72 Bedford, MA, USA) was used to prepare deionized water. The other reagents were all
73 analytical reagents.

74 **Cell lines and culture**

75 Human breast carcinoma MCF-7 cells were products of Cell Bank of Institute of
76 Biochemistry and Cell Biology, Chinese Academy of Sciences (Shanghai, China). Cells were
77 grown in RPMI-1640 medium in a humidified incubator at 37°C with 5% CO₂, and the
78 medium consists of 100 U/mL penicillin, 10% heat-inactivated fetal bovine serum (FBS) and
79 100 µg/mL streptomycin.¹¹ To improve the reliability of the data, we repeated all experiments
80 for three times at least per experimental point.

81 **MTT assay**

82 Determination of live cell numbers is often used to assess the rate of cell proliferation
83 caused by drugs and cytotoxic agents. Among all non-radioactive viability assays, MTT assay
84 developed by Mossman is one of the most versatile and popular assays. MTT is a tetrazolium
85 salt that is turned into a purple formazan product after reduction by mitochondrial enzymes
86 that are only present in metabolically active live cells, not in dead cells.¹¹ The cells were
87 seeded and grew in 96-well plates at concentration of 3×10⁵ cells/well in 100 µL medium for
88 24 h (the cells were grew to 70% confluence). Then we used medium consists of different
89 concentrations (0, 25, 50 and 100 µM) of Que, IS, I3G or 5-fluorouracil (5-Fu, 100 µM) to
90 treat the cells, respectively. The 5-Fu is used as a positive control in cell experiments. In this
91 study, we added 10 µL of MTT (5 mg/mL) in PBS solution to each well. After blending them,
92 we further incubated the plate. 100 µL of solution containing 0.01 M HCl, 5% isobutyl
93 alcohol and 10% SDS (pH 4.8) was added to each well in 4 h, mixed and put in incubator for
94 one night. The absorbancy was observed at 570 nm using a microplate spectrophotometer
95 (RT6000, Guangdong, China). The viability of cells was calculated with the following
96 formula: cell viability (%) = OD_{test}/OD_{control} × 100%, and the compounds IC₅₀ (50% inhibition

97 concentration) values were counted using the Origin 7.0 software.

98 **LDH assay for cytotoxicity**

99 Lactate dehydrogenase (LDH) is a cytosolic enzyme present in many different types of cells.
100 When the plasma membrane is damaged, LDH is released into cell culture media. The released
101 LDH can be quantified by a coupled enzymatic reaction.¹¹ The cytotoxic effects of Que, IS and
102 I3G on human breast MCF-7 cells were investigated by LDH assay. In this study, LDH kit
103 (Jiancheng BioEngineering, Nanjing, China) was used to test cellular membrane damage of
104 MCF-7 cells in response to Que, IS and I3G (0, 25, 50 and 100 μ M) treatments as outlined by
105 manufacture with minor modifications. After incubation at room temperature for 30 minutes,
106 reactions are stopped and 20 μ L of culture supernatant was taken out for the activity analysis of
107 extracellular LDH, which could catalyze the lactate turn into pyruvate. Then the culture
108 supernatant was reacted with 2,4-dinitrophenylhydrazine to make the basic solution present
109 brownish red color, and LDH activity was determined by spectrophotometric absorbance at 450
110 nm.

111 **Morphological study**

112 Regularly examining the morphology (shape and appearance) of the cells in culture is
113 essential for successful cell culture experiments.¹⁵ In this study, the morphological study of
114 MCF-7 cells treated with Que, IS and I3G was investigated using an Inverted Fluorescence
115 Microscope. Human breast cancer MCF-7 cells were seeded onto a glass slide and treated with
116 Que, IS and I3G for 48 h. After washing at least two times with ice-cold PBS, they were
117 blended with 4% (V/V) formaldehyde in PBS and then washed PBS. After the cells were
118 stained with 1 mg/mL Hoechst 33258 in PBS at 37°C for 15 min, we examined the morphology

119 using a fluorescence microscope (Leica DMIL LED, Leica, Germany) with an excitation
120 wavelength of 345 nm through the filter of 420 nm.

121 **Assessment of cell apoptosis**

122 Apoptosis is a distinct form of cell death controlled by an internally encoded suicide
123 program.¹⁶ The extent of apoptosis was investigated by Annexin V-FITC/PI double staining
124 assay. In this study, an Annexin V-FITC/PI Apoptosis Detection Kit (BestBio, Shanghai,
125 China) was used to determine early and late apoptotic changes in MCF-7 cells. MCF-7 cells
126 (3×10^5) were collected, washed with PBS for two times and then suspended in 400 μ L of
127 binding buffer (adding 5 μ L of annexin V-FITC and 10 μ L of PI). After incubating for 10 min
128 at 2-8 °C in the dark, we used a GUAVA[®] easy Cyte[™] 8HT flow cytometry (Millipore
129 Corporation, Billerica, MA, USA) to analyze the samples. Then we counted the number of
130 annexin V-FITC-positive and PI-positive of cells in each field. In order to ensure the
131 reliability of the data, we independently did the whole experiments for three times at least.

132 **Cell circle analysis by flow cytometry**

133 DNA content assay for cell circle is a classical method that frequently employs flow
134 cytometry to distinguish cells in different phases of the cell circle.¹⁷ For the purpose of
135 studying the relationship between growth inhibitory effect of Que, IS and I3G and cell circle
136 arrest, we treated MCF-7 cells with Que, IS and I3G for 48 h and then examined cell circle
137 phase distribution of PI-stained by using flow cytometry. MCF-7 cells (3×10^5) were seeded in
138 6-well flat-bottomed plates and grown overnight until they reached 80% concentration, and the
139 medium was changed after 24 h. After treatment with Que, IS and I3G (0, 50, 100 μ M) for 48 h,
140 we collected the detached cells in culture, and then combined them with the remaining

141 adherent cells that were detached by brief trypsinization (0.25% trypsin-EDTA,
142 Sigma-Aldrich). After mixing and washing the cell pellets in 75% ethanol with PBS, we
143 resuspended them in PBS (1 mL) containing 1 mg/mL RNase (Sigma-Aldrich) and 50 µg/mL
144 PI (Sigma-Aldrich). The cells were incubated in the dark for 30 min at 26°C, and then
145 investigated by the GUAVA® easy Cyte™ 8HT flow cytometry (Millipore Corporation,
146 Billerica, MA, USA).

147 **Measurement of ROS**

148 Generation of intracellular ROS was assessed using dichlorofluorescein diacetate probes.¹⁶
149 MCF-7 cells were seeded in 12-well plates, and then incubated with Que, IS and I3G for 24 h.
150 After detaching with trypsin-EDTA, we washed the cells with PBS for twice and then incubated
151 them with 5 µM DCFH-DA for 30 min at 37°C. Then flow cytometry was used to determine the
152 fluorescence intensity of MCF 7 cells.

153 **Statistical analysis**

154 All data are expressed as mean ± SD of three independent experiments. The significant
155 difference from the respective control for each experimental group was examined by one-way
156 analysis of variance (ANOVA) using SPSS 19.0 software. A value of $p < 0.05$ is considered
157 statistically significant and a value of $p < 0.01$ means extremely significant difference.

158

159 **RESULTS**

160 **Growth-inhibitory effects of quercetin, IS and I3G on MCF-7 cells**

161 To identify the growth-inhibitory effects of quercetin and its metabolites, we cultured MCF-7
162 cells treated with the indicated concentrations of Que, IS and I3G at 25, 50, 100 µM for 48 h.

163 As shown in Fig. 2A, a significant growth-inhibitory effect induced by 25 μ M of Que, IS and
164 I3G was observed as compared to the untreated control cells ($p<0.01$), and a further decrease in
165 the percentage of MCF-7 living cells was observed as the concentrations of Que, IS and I3G
166 increased to 100 μ M, indicating that the inhibition was in a dose-dependent manner. After the
167 MCF-7 cells were treated with Que, IS and I3G at the high dose of 100 μ M for 48 h, MCF-7
168 cells viability was markedly decreased to 33.1%, 34.2% and 40.7% in comparison with the
169 control group, respectively ($p<0.01$). More interestingly, the highest concentration of Que (100
170 μ M) and IS (100 μ M) exhibited similar effect with the same concentration of 5-Fu, suggesting
171 that Que, IS and I3G all could exhibit high tumor-inhibitory effect in human breast cancer
172 MCF-7 cells, and this effect was sort as follows: Que > IS > I3G, which might be related to the
173 different structure of metabolites.

174 **Cytotoxicity of quercetin and its metabolites on MCF-7 cells**

175 The release of LDH can be regarded as an index of the integrity of cell membrane necrosis in
176 response to cytotoxic efficiency, and it can be detected by colorimetric assay.¹⁸ Herein, we
177 evaluated LDH release to evaluate the cytotoxicity of Que, IS and I3G (0, 25, 50 and 100 μ M)
178 on MCF-7 cells after 48 h of incubation. As can be seen in Fig. 2B, when treated with Que, IS
179 and I3G at the concentration of 25 μ M for 48 h, the LDH release of MCF-7 cells was 957 U/L,
180 942 U/L and 880 U/L, which were 8-10 times higher than that of control group (100 U/L,
181 $p<0.01$). Along with the increase of quercetin and metabolites concentration, the LDH release
182 of MCF-7 cells represented a significant improvement, indicating that the cytotoxicity effects of
183 Que, IS and I3G in MCF-7 cells were in a dose-dependent manner. When treated with Que, IS
184 and I3G at the high dose of 100 μ M for 48 h, LDH release of MCF-7 cells was markedly

185 increased to 1390 U/L, 1359 U/L and 1279 U/L, which was 12-14 times higher than that of
186 control group (100 U/L, $p < 0.01$), respectively and this effect was similar to that of the same
187 concentration of 5-Fu (1599 U/L, $p > 0.05$), suggesting the induction of cell membrane injury.
188 Similarly, the cytotoxicity of these compounds on MCF-7 cells could be ranked as Que > IS >
189 I3G, indicating that methylation in 3'-position of Que could decrease the cytotoxicity of Que in
190 MCF-7 cells, and glucuronidation could further decrease the tumor-inhibitory effect of IS
191 (methylation of quercetin in 3'-position). This sensitivity of MCF-7 tumor cells to quercetin and
192 its metabolites led to further examination on the mechanism of antiproliferative effects of them.

193 **Morphological study**

194 After treated with Que, IS and I3G (100 μ M), MCF-7 cells were incubated for 48 h and
195 observed the morphological characteristics with an Inverted Fluorescence. As seen in [Fig. 2C](#),
196 untreated control cells grew well, but the tumour cells treated with Que, IS and I3G were
197 gradually reduced, and cells fusion, shrinkage, nuclear condensation, apoptotic body and lysis
198 appeared, which were similar to that of 5-Fu treated MCF-7 cells. The morphology assay results
199 indicated that similar with quercetin, IS and I3G were anti-tumor compounds as well, and their
200 anti-tumor effect can be ranked as Que > IS > I3G, which were consistent with that of the MTT
201 assay and the LDH assay.

202 **Effects of quercetin and its metabolites on cell apoptosis in MCF-7 cells**

203 Apoptosis is a process of programmed cell death that occurs in multicellular organisms, and
204 always be considered as the preferred way to eliminate tumor cells.¹⁶ MCF-7 cells apoptosis
205 was measured by flow cytometry using annexin V-FITC and PI labeling.¹⁸ As displayed in [Fig.](#)
206 [3A and B](#), in untreated control groups, 95.8% of MCF-7 cells were in normal state and almost

207 no apoptotic nuclei were observed. When treated the MCF-7 cells with 25 μ M Que, IS and I3G
208 for 48 h, 36.6%, 35.3% and 16.8% of MCF-7 cells transformed into apoptotic state, which were
209 significant higher than that of control group (1.3%), respectively ($p < 0.01$). Furthermore, with
210 the increase of concentrations, Que, IS and I3G dose-dependently induced 70.8%, 68.9% and
211 49.8% of MCF-7 cells to transform into apoptotic state. Meanwhile, the cytotoxic effects of IS
212 and I3G were similar to that of 5-Fu (75.2%, $p > 0.05$). Consistent with MTT, LDH and
213 morphology analysis, Que, IS and I3G were proved to induce cell apoptosis in a
214 dose-dependent manner, and the effects could be ranked as Que > IS > I3G, suggesting that
215 methylated and methylated-glucuronidated complex metabolites of quercetin could decrease its
216 pro-apoptosis effect in MCF-7 cells in varying degree.

217 **Quercetin and its metabolites induced cell cycle arrest in MCF-7 tumor cells**

218 For the purpose of studying the relationship between growth inhibitory effects of the tested
219 flavonoids and cell cycle arrest, MCF-7 cells were treated with Que, IS and I3G for 48 h. The
220 cell cycle phase distribution of PI-stained cells was examined by using flow cytometry. As is
221 shown in [Fig. 4A](#) and [Fig. 4B](#), after treated with Que, IS and I3G at 50 μ M, a significant
222 amount number of MCF-7 cells accumulated at the S-phase, corresponding to DNA synthesis,
223 from 5.7% to 38.1%, 34.5% and 25.2% ($p < 0.01$), accompanied by a decrease in the G0/G1 and
224 G2/M cells, respectively. After 48 h of treatment with 100 μ M of Que, IS and I3G, a further
225 increasing arrest in the S-phase of MCF-7 cell cycle was observed ($p < 0.01$), which was similar
226 with that of the same concentration of 5-Fu ($p > 0.05$).

227 **ROS was involved in quercetin- or its metabolites-induced apoptosis in MCF-7 cells**

228 Mitochondrial ROS production is a crucial early driver of cell injury, and has been

229 considered to have a big relationship with the induction of apoptosis.^{19,20} To investigate
230 whether intracellular ROS is related to the apoptosis induced by Que, IS and I3G, we
231 determined the ROS level by using flow cytometry. As shown in Fig. 5A and B, after treated
232 with Que, IS and I3G at 25 μM for 12 h, the accumulation of O_2^- in MCF-7 cells was 58.8%,
233 50.0% and 44.7%, which was significantly higher than that of control cells (6.03%, $p < 0.01$),
234 respectively. With an increase in Que, IS and I3G concentrations, a dose-dependent effect was
235 further observed. The accumulation of O_2^- in a high concentration (100 μM) of Que, IS and
236 I3G treated cells was significantly increased to 84.1%, 77.6% and 60.7%, which was similar to
237 that of the same concentration of 5-Fu ($p > 0.05$), respectively. Similar results could be observed
238 from the accumulation of H_2O_2 , compared with untreated MCF-7 cells, and the accumulation
239 of H_2O_2 in Que, IS and I3G groups at 25 μM was significantly elevated to 48.9%, 36.0% and
240 20.0% (Fig. 5C and D). When concentrations increased to 100 μM , the accumulation of H_2O_2
241 was dose-dependently increased to 68.1%, 55.8% and 43.1%, respectively ($p < 0.01$). These
242 results indicated that Que, IS and I3G could induce apoptosis through the increasing of
243 intracellular oxidative stress of MCF-7 tumor cells.

244 **Growth-inhibitory effects and cytotoxicity of Que, IS and I3G on normal mammary** 245 **epithelial cell H184B5F5/M10 cells**

246 Similar with MCF-7 cells, the cytotoxic effects of Que, IS and I3G on normal mammary
247 epithelial cell H184B5F5/M10 cells were also determined using the MTT assay and LDH assay,
248 respectively. Just as shown in Fig.6 (A) and (B), cytotoxic effects of Que, IS and I3G on
249 H184B5F5/M10 normal mammary epithelial cells were not observed in the same test
250 concentrations with MCF-7 cells, indicating that Que, IS and I3G had no growth-inhibitory

251 effects and cytotoxicity in the normal mammary epithelial H184B5F5/M10 cells.

252

253 **DISCUSSION**

254 Quercetin (3,3',4',5,7-pentahydroxyflavone) is one of the most widespread flavonoids and
255 was regarded as a promising compound in tumor prevention.^{21,22} *In vivo*, absorbed Que is
256 rapidly metabolized to various methylated, glucuronidated and sulfated forms in different
257 organs, such as liver, kidney, colon and small intestine. The metabolites include Que,
258 Isorhamnetin (IS), Isorhamnetin-3-glucoside(I3G), Quercetin aglycone(QA), quercetin
259 3-glucoside(Q3G), 3'-O-methylated quercetin, 4'-O-methylated quercetin, quercetin-3'-
260 sulfate(Q3'S) and so on.^{23, 24} Among these metabolites, IS and I3G (Fig. 1) are very important
261 metabolites for quercetin and they are likely to possess biological properties different from
262 parent quercetin, making it significant to examine their anti-tumor activities and investigate the
263 relationship between structure and function.²³ Nevertheless, there is a paucity of research on
264 the issue with anti-tumor effects of quercetin metabolites. Consequently, in this study we
265 mainly tested and compared the anti-tumor effects of Que, IS and I3G, to figure out the
266 mechanism and try to reveal the structure-activity relationship of them by testing
267 growth-inhibitory effects, cytotoxicity, cell cycle effects and ROS level in human breast cancer
268 MCF-7 cells.

269 IS and I3G both inhibited the growth of MCF-7 cell, and the effect can be ranked as Que >
270 IS > I3G (Fig. 2). Notably, we firstly demonstrated an interesting phenomenon that similar with
271 parent quercetin, structurally related metabolites IS and I3G possessed strong inhibitory effects
272 on human breast cancer MCF-7 cells in a dose-dependent manner, and their inhibitory

273 activities were similar with positive 5-Fu. Apoptosis is a process of programmed cell death that
274 occurs in multicellular organisms.²⁵ The results of annexin V/PI co-staining assays in this study
275 clearly showed that Que, IS and I3G could induce MCF-7 cells apoptosis and necrosis, and the
276 activities can be ranked as Que > IS > I3G (Fig. 3). And these results were consist with that of
277 biochemical and morphological assay, which showed that in Que, IS and I3G treatment groups,
278 cell shrinkage, chromatin condensation, inter nucleosomal DNA fragmentation, and formation
279 of “apoptotic bodies” appeared in MCF-7 cells (Fig. 2C). Mounting evidence suggests that
280 apoptosis have a big relationship with cell circle and apoptosis may be induced by cell circle
281 disruption.^{26,27} Similar with some previous reports,²⁸ our cell circle essay indicated that Que, IS
282 and I3G exhibited effective cell growth inhibition by accumulating cells in S-phase, decreasing
283 the MCF-7 cells number of G2/M and G0/G (Fig. 4). To conclusion, this finding suggests that
284 quercetin and its metabolites IS and I3G inhibit hyperplasia of tumor cells mainly by arresting
285 the cells in the S-phase and decreasing the number of G0/G1 and G2/M cells in the cell circle.

286 Extensive literatures have indicated that ROS plays a crucial role in cell apoptosis and
287 participates in multiple signaling pathways which can mediate high anti-proliferation effect.²⁹
288 In order to determine the pathway by which Que, IS and I3G induced apoptosis, we examined
289 ROS generation in MCF-7 cells.^{30,31} Our results showed that Que, IS and I3G led to a
290 significant dose-dependent increase of intracellular ROS in MCF-7 cells, and the antioxidant
291 effect order is as follows: Que > IS > I3G, indicating that ROS production led to apoptotic
292 cell-death through the mitochondrial pathway (Fig. 5). In agreement with some previews report,
293 Que, IS and I3G possess strong antioxidation activities *in vitro*.^{32,33} The result of “Que > IS” is
294 in agreement with a previous study which revealed that radical scavenging activity decreased

295 in the order Que > IS.³⁴ The result of “IS > I3G” has never been compared before, and this may
296 related to the fact that in quercetin and its derivatives, 3-OH is an important activity position,
297 glucuronidation at the 3-position had a marked decrease in their antioxidant activity.³⁵ The assay
298 results of H₂O₂ and O₂⁻ indicated that ROS played a crucial role in cell apoptosis, and ROS
299 might participate in the Que-, IS- and I3G-elicited MCF-7 cell death. In the subsequent study,
300 we will explore the relationship of ROS and Que, IS and I3G induced MCF-7 cell death by
301 using the antioxidant such as Acetylcysteine (NAC), DPI and so on. Overall, we firstly reveals
302 the reality that IS and I3G, which are main metabolites of quercetin *in vivo*, possess strong
303 tumor-inhibitory activities in MCF-7 cells via cell cycle arrest at S phase and apoptosis by
304 ROS-dependent mitochondrial pathway, indicating that quercetin metabolites may still possess
305 strong activities *in vivo*.

306 In addition, the anti-tumor mechanism of Que, IS and I3G may also be related to the
307 cell-permeability of them. The extensive reports have indicated that quercetin has the high
308 cell-permeability,⁹ which may contribute to its cytotoxic activity. An extensive literature has
309 indicated that quercetin has a significantly high anti-proliferation effect, which may also
310 contribute to its antitumor activity *in vitro*.³⁶ As to the “Que > IS > I3G”, we think that it may
311 be related to the different molecular weight and polarity of them. Small molecules were easier to
312 cross cell membrane and the molecular weight of these compounds was sorted as Que
313 (302.24) > IS (316.2623) > I3G (492.39), which was consistent with the cytotoxic efficiency
314 order of them.

315 Meanwhile, we investigated the growth-inhibitory effects and cytotoxicity of Que, IS and
316 I3G on normal mammary epithelial cell H184B5F5/M10 cell. As shown in Fig. 6, Que, IS and

317 I3G has no cytotoxicity against the normal mammary epithelial cells, indicating that Que, IS
318 and I3G have a good selectivity on the tested tumor cells. Our previous studies have showed
319 that Que and QS (5', 8-disulfonate substituted metabolite of quercetin) possess high anti-tumor
320 activity in human colon cancer LoVo cells and breast cancer MCF-7 cells,¹¹ which can further
321 provide evidence for the application of the tested Que, IS and I3G. Furthermore, some other
322 studies have showed that Que has strong anticancer effect on a wide range of cancer cells such
323 as acute lymphoid, myeloid leukemia cells, human gastric and colon cancer cells.⁷⁻¹¹ It is also
324 reported that Que, IS and I3G have high antioxidant ability,³⁴ which further provide evidence
325 for the application of the three compounds. In general, after Que ingestion, its metabolites
326 quercetin, isorhamnetin and isorhamnetin-3-glucuronide are mainly present in the
327 physiological fluids, and concentrations of animal tissues are in the range of 0.015-0.125 μM ,
328 0.53-0.65 μM or 0.03-0.18 μM .^{23, 37} It must be noted that the concentrations (25, 50 and 100
329 μM) we used in this study may generally be not physiological and achievable *in vivo* because
330 of the low bioavailability of these compounds. In this study we mainly investigated the
331 anticancer-structure relationships between Que, IS and I3G *in vitro* and try to provide the
332 foundation for high bioavailability and water solubility metabolites of quercetin *in vivo*
333 evaluation, and to evaluate the potential clinical use of this study.

334 In conclusion, we investigated and compared the cytotoxic activities of Que, IS and I3G in
335 human breast cancer MCF-7 cells for the first time, and found that they possess strong
336 cytotoxic effect through a ROS-dependent apoptosis pathway in MCF-7 cells. Significantly, we
337 firstly point out the fact that 3'-methylation can decrease the cytotoxic properties of quercetin,
338 and 3-glucuronidation may further decrease the cytotoxic activity of IS (3'-methylation of

339 quercetin), and display the structure-anti tumor activity relationship of them (Que > IS > I3G).
340 In our previous studies, we have investigated and compared the cytotoxic effect of Que and its
341 another metabolite Q3G (glucuronidation metabolite of quercetin in 3 position) on human
342 breast cancer MCF-7 cells, the results showed that Que and Q3G possess cytotoxic effect in
343 breast cancer MCF-7 cells and it was ordered “Que > Q3G”, indicating that glucuronidation
344 may decreased the cytotoxic effect of Que *in vitro*.^[38] Our studies *in vitro* provide a new insight
345 into the cytotoxic effect of quercetin metabolites, and further studies should be carried out in
346 animal studies and ultimately in clinical trials.

347 In the subsequent study, we will further develop the synthesis method and do some
348 metabonomics experiments to further investigate the cytotoxic mechanism of Que, IS and I3G
349 in MCF-7 cells. Metabolomics is an integral part of the systems biology and is rapidly
350 advancing with the aims of detecting many metabolites with low molecular weight in single
351 cell, bio-fluids, and tissue extracts.^[39] Therefore, the application of metabolomics method may
352 contribute to the further understanding of the cytotoxic mechanism of Que, IS and I3G.

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359 **CONFLICT OF INTEREST STATEMENT**

360 The authors declare no competing financial interest.

361 **REFERENCES**

- 362 (1) Fan, L.; Strasser-Weippl, K.; Li, J.; St Louis, J.; Finkelstein, D. M.; Yu, K.; Chen, W.; Shao,
363 Z.; Goss, P. E. Breast cancer in China. *Lancet Oncol.* **2014**, 15(14), e 279-289.
- 364 (2) Zhu, J.; Sanidad, K. Z.; Sukamtoh, E.; Zhang, G. Potential roles of chemical degradation in
365 the biological activities of curcumin. *Food Funct.* **2017**, 8, 907-914.
- 366 (3) Chen, J.; Duan, Y.; Zhang, X.; Ye, Y.; Ge, B.; Chen, J. Genistein induces apoptosis by the
367 inactivation of the igf-1r/p-akt signaling pathway in MCF-7 human breast cancer cells.
368 *Food Funct.* **2015**, 6, 995-1000.
- 369 (4) Liu, R. H. Potential Synergy of phytochemicals in cancer prevention: mechanism of action.
370 *J. Nutr.* **2004**, 134(12), 3479S-3485S.
- 371 (5) Havsteen, B.H. The biochemistry and medical significance of the flavonoids. *Pharmacol.*
372 *Therapeut.* **2002**, 96, 67-202.
- 373 (6) Lila, M. A.; Burton-Freeman, B.; Grace, M.; Kalt, W. Unraveling Anthocyanin
374 Bioavailability for Human Health. *Annu. Rev. Food Sci. Technol.* **2016**, 7, 375-393.
- 375 (7) Muthurajan, T.; Rammanohar, P.; Rajendran, N. P.; Sethuraman, S.; Krishnan, U. M.; Soc,
376 R. Evaluation of a quercetin–gadolinium complex as an efficient positive contrast enhancer for
377 magnetic resonance imaging. *RSC Adv.* **2015**, 5(106), 86967–86979.
- 378 (8) Nile, S. H; Nile, A. S.; Keum Y. S.; Sharma, K. Utilization of quercetin and quercetin
379 glycosides from onion (*Allium cepa* L.) solid waste as an antioxidant, urease and xanthine
380 oxidase inhibitors. *Food Chem.* **2017**, 235, 119-126.

- 381 (9) Atala, E.; Fuentes, J.; Wehrhahn, M. J.; Speisky, H. Quercetin and related flavonoids
382 conserve their antioxidant properties despite undergoing chemical or enzymatic oxidation.
383 *Food Chem.* **2017**, 234, 479-485.
- 384 (10) Kang, T. B.; Liang, N. C. Studies on the inhibitory effects of quercetin on the growth of
385 HL460 leukemia cells. *Biochem. Pharmacol.* **1997**, 54(9), 1013-1018.
- 386 (11) Zhang, H. S.; Zhang, M.; Yu, L. H.; Zhao, Y; He. N. W.; Yang, X. B. Antitumor activities
387 of quercetin and quercetin-5',8-disulfonate in human colon and breast cancer cell lines.
388 *Food Chem. Toxicol.* **2012**, 50(5), 1589-1599.
- 389 (12) Nabavi, S. F.; Russo, G. L.; Daglia, M. Seyed Mohammad Nabavi, Role of quercetin as an
390 alternative for obesity treatment: You are what you eat. *Food Chem.* **2015**, 179, 305-310.
- 391 (13) Benavente-García, O.; Castillo, J. Update on uses and properties of citrus flavonoids: new
392 findings in anticancer, cardiovascular, and anti-inflammatory activity. *J. Agric. Food*
393 *Chem.* **2008**, 56, 6185-6205.
- 394 (14) Needs, P. W.; Kroon, P. A. Convenient syntheses of metabolically important quercetin
395 glucuronides and sulfates. *Tetrahedron* **2006**, 62, 6862–6868.
- 396 (15) Ma, Y. S.; Weng, S. W.; Lin, M. W.; Lu, C. C.; Chiang, J. H.; Yang, J. S. Antitumor effects
397 of emodin on LS1034 human colon cancer cells in vitro and in vivo: roles of apoptotic
398 cell death and LS1034 tumor xenografts model. *Food Chem. Toxicol.* **2012**,
399 50(5):1271-1278.
- 400 (16) Hanahan, D.; Weinberg, R. A. The hallmarks of cancer. *Cell* **2000**, 100(1):57-70.
- 401 (17) Tero, A.; Takagi, S.; Saigusa, T.; Ito K.; Bebber, D. P.; Fricker, M. D.; Yumiki, K.;
402 Kobayashi, R.; Nakagaki, T. Rules for biologically inspired adaptive network design.

- 403 *Science* **2010**, 327(5964):439-42.
- 404 (18) Lemieszek, M. K.; Cardoso, C.; Fh, F. M. N.; Barros, A. I. R. N. A.; Marques, G.;
405 Pozarowski, P.; Rzeski, W. Boletus edulis biologically active biopolymers induce cell
406 cycle arrest in human colon adenocarcinoma cells. *Food Func.* **2013**, 4(4):575-585.
- 407 (19) Chouchani, E. T.; Pell, V. R.; Gaude, E.; Aksentijević, D.; Sundier, S. Y.; Robb, E. L.
408 Ischaemic accumulation of succinate controls reperfusion injury through mitochondrial
409 ROS. *Nature* **2014**, 515(7527), 431-445.
- 410 (20) Chen, A. Y.; Chen, Y. C. A review of the dietary flavonoid, kaempferol on human health
411 and cancer chemoprevention. *Food Chem.* **2013**, 138(4), 2099-2107.
- 412 (21) Pool, H.; Mendoza, S.; Xiao, H.; Mcclements, D. J. Encapsulation and release of
413 hydrophobic bioactive components in nanoemulsion-based delivery systems: impact of
414 physical form on quercetin bioaccessibility. *Food Func.* **2013**, 4(1), 162-174.
- 415 (22) Kobori, M.; Takahashi, Y.; Sakurai, M.; Akimoto, Y.; Yukari, A.; Tsushida, T.; Oike H.
416 Quercetin suppresses immune cell accumulation and improves mitochondrial gene
417 expression in adipose tissue of diet-induced obese mice. *Mol. Nutr. Food Res.* **2016**, 60(2),
418 300-312.
- 419 (23) Yeh, S. L.; Lin, Y. C.; Lin, Y. L.; Li, C. C.; Chuang, C. H. Comparing the metabolism of
420 quercetin in rats, mice and gerbils. *Eur. J. Nutr.* **2016**, 55(1), 413-422.
- 421 (24) Pilařová, V.; Plachká, K.; Chrenková, L.; Najmanová, I.; Mladěnka, P.; Švec, F.; Novák,
422 O.; Nováková, L. Simultaneous determination of quercetin and its metabolites in rat plasma
423 by using ultra-high performance liquid chromatography tandem mass spectrometry. *Talanta*
424 **2018**, 185, 71-79.

- 425 (25) Hengartner, M. O. The biochemistry of apoptosis. *Nature* **2000**, 407(6805):770-776.
- 426 (26) Lawrence, M. S.; Stojanov, P.; Mermel, C. H.; Garraway, L. A.; Golub, T. R.; Meyerson,
427 M.; Gabriel, S. B.; Lander, E. S.; Getz, G. Discovery and saturation analysis of cancer
428 genes across 21 tumor types. *Nature* **2014**, 505(7484), 495-500.
- 429 (27) Strowig, T.; Henao-Mejia, J.; Elinav, E.; Flavell R. Inflammasomes in health and disease.
430 *Nature* **2012**, 481(7381), 278-286.
- 431 (28) Richter, M.; Ebermann, R.; Marian, B. Quercetin-induced apoptosis in colorectal tumor
432 cells: possible role of egf receptor signaling. *Nutr. Cancer* **1999**, 34(1), 88-99.
- 433 (29) Roca, F. J.; Ramakrishnan, L. TNF dually mediates resistance and susceptibility to
434 mycobacteria via mitochondrial reactive oxygen species. *Cell* **2013**, 153(3), 521-534.
- 435 (30) Kim, J.; Tchernyshyov, I.; Semenza, G. L.; Dang, C. V. HIF-1-mediated expression of
436 pyruvate dehydrogenase kinase: a metabolic switch required for cellular adaptation to
437 hypoxia. *Cell Metab.* **2006**, 3(3), 177-185.
- 438 (31) Xie, J. H.; Liu, X.; Shen, M, Y. Purification, physicochemical characterisation and
439 anticancer activity of a polysaccharide from *Cyclocarya paliurus* leaves. *Food Chem.*
440 **2013**, 136(3-4):1453-1460.
- 441 (32) Jaramillo, S.; Lopez, S.; Varela, L. M.; Rodriguezarcos, R.; Jimenez, A.; Abia, R. The
442 flavonol isorhamnetin exhibits cytotoxic effects on human colon cancer cells. *J. Agr. Food*
443 *Chem.* **2010**, 58(20), 10869-10875.
- 444 (33) Kong, C.; Kim, J.; Qian, Z.; Kim, Y.; Lee, J.; Kim, S.; Nam, T. J.; Seo, Y. Protective effect
445 of isorhamnetin 3-o- β -d-glucopyranoside from *salicornia herbacea* against
446 oxidation-induced cell damage. *Food Chem. Toxicol.* **2009**, 47(8), 1914-1920.

- 447 (34) Yamamoto, N.; Moon, J. H.; Tsushida, T.; Nagao, A.; Terao, J. Inhibitory effect of
448 quercetin metabolites and their related derivatives on copper ion-induced lipid
449 peroxidation in human low-density lipoprotein. *Arch. Biochem. Biophys.* **1999**, 372(2),
450 347-354.
- 451 (35) Day, A. J.; Bao, Y.; Morgan, M. R.; Williamson, G. Conjugation position of quercetin
452 glucuronides and effect on biological activity. *Free Radical Bio. Med.* **2000**, 29(12),
453 1234-1243.
- 454 (36) Lee, S. H.; Lee, E. J.; Min, K. H.; Hur, G. Y.; Lee, S. H.; Lee, S. Y.; Kim, J. H.; Shin, H.;
455 Shim, J. J.; In, K. H.; Kang, K. H.; Lee, S. Y. Quercetin enhances chemosensitivity to
456 gemcitabine in lung cancer cells by inhibiting heat shock protein 70 expression. *Clin.*
457 *Lung Cancer* **2015**, 16(6), e235-e243.
- 458 (37) Lee, J.; Ebeler, S. E.; Zweigenbaum, J. A.; Mitchell, A. E. UHPLC-(ESI)QTOF MS/MS
459 profiling of quercetin metabolites in human plasma postconsumption of applesauce
460 enriched with apple peel and onion. *J. Agric. Food Chem.* **2012**, 60(34):8510-8520.
- 461 (38) Wu, Q.; Paul, W. N.; Lu, Y.; Paul, A. K.; Ren, D.; Yang, X. Different antitumor effects of
462 quercetin, quercetin-3'-sulfate and quercetin-3-glucuronide in human breast cancer mcf-7
463 cells. *Food Func.* **2018**, 9(3), 1736-1746.
- 464 (39) Sreekumar, A.; Poisson, L.M.; Rajendiran, T.M.; Khan, A.P.; Cao, Q.; et al. Metabolomic
465 profiles delineate potential role for sarcosine in prostate cancer progression. *Nature* **2009**,
466 457: 910-914.

467 **Figure Captions**

468 **Fig.1.** Chemical structure of quercetin (Que), Isorhamnetin (IS) and Isorhamnetin-3-
469 glucuronide (I3G)

470
471 **Fig.2.** The MTT assay results of Que, IS and I3G on MCF-7 cells was assayed (A), and
472 cytotoxic effect was measured by LDH assay (B) and the change of cell morphology was
473 measured by optic microscopic observation (C). In Fig 2(C), red arrow means normal MCF-7
474 cells, yellow arrow means cell shrinkage, green arrow means cell fusion and bronzing arrow
475 means cell lysis. Data are expressed as the mean \pm SD (n = 3). $p < 0.05$ (*) or $p < 0.01$ (**) indicates a significant difference versus control.

476
477
478 **Fig.3.** Quantitative analysis of apoptotic cells induced by Que, IS and I3G using annexin V/PI
479 double staining assay. (A) Representative dot plots of Annexin V/PI staining. (B) Column bar
480 graph of apoptotic cells. Cells were treated with Que, IS and I3G at 25 and 100 μ M for 48 h,
481 respectively. 3000 cells were analyzed by flow cytometry. The results are expressed as mean \pm
482 SD of three independent experiments. $p < 0.01$ (**), as compared to the control.

483
484 **Fig.4.** Effects of Que, IS and I3G on cell cycle phase distribution of MCF-7 cells. (A)
485 Representative histograms of DNA content in the cells incubated with Que, IS and I3G at 50
486 and 100 μ M for 48 h. Horizontal and vertical axes indicate the relative nuclear DNA content
487 and number of cells, respectively. (B) Percentage of cell populations in G0/G1, S and G2/M
488 phases. All values are expressed as mean \pm SD of three independent experiments. Significant

489 difference from the control at the same phase is indicated at $p < 0.05$ (*) or $p < 0.01$ (**).

490

491 **Fig.5.** Effects of Que, IS and I3G on ROS (H_2O_2 and O_2^-) generation of MCF-7 cells. (A)
492 Representative flow cytometric images for O_2^- generation in MCF-7 cells. (B) Levels of O_2^- (%)
493 in MCF-7 cells when treated with Que, IS and I3G. (C) Fluorescence intensity analysis for
494 H_2O_2 generation in MCF-7 cells. (D) Levels of H_2O_2 (%) in MCF-7 cells when treated with
495 Que, IS and I3G. About 100 μM H_2O_2 were used as positive control. The results represent the
496 mean \pm SD of three independent experiments. $p < 0.05$ (*) or $p < 0.01$ (**) indicate statistically
497 significant difference with control, which was considered to be 100%.

498

499

500 **Fig.6.** Growth-inhibitory effects and cytotoxicity of Que, IS and I3G on normal mammary
501 epithelial H184B5F5/M10 cells. MTT assay (A) and LDH assay (B). The results represent the
502 mean \pm SD of three independent experiments. $p < 0.05$ (*) or $p < 0.01$ (**) indicate statistically
503 significant difference with control, which was considered to be 100%.

Figure 1

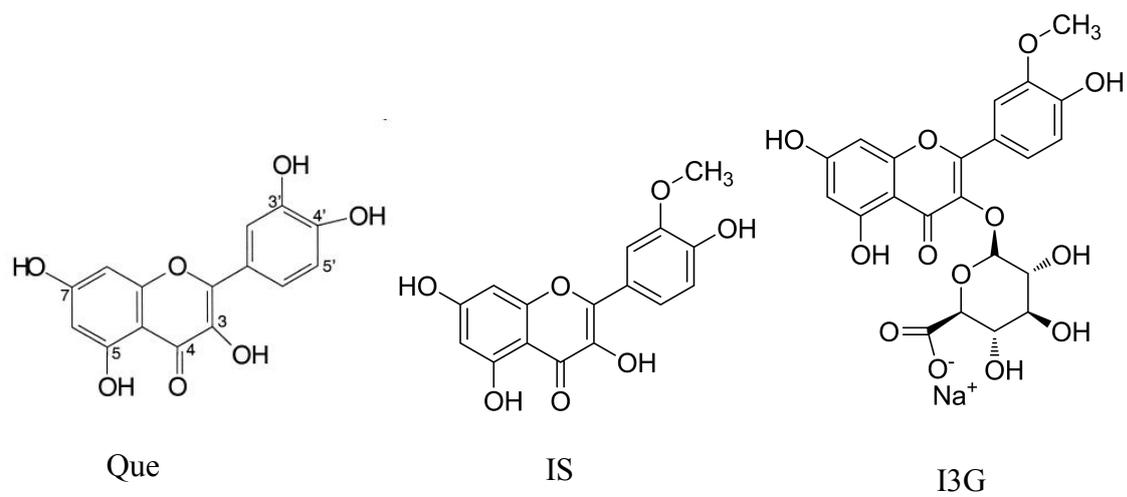


Figure 2

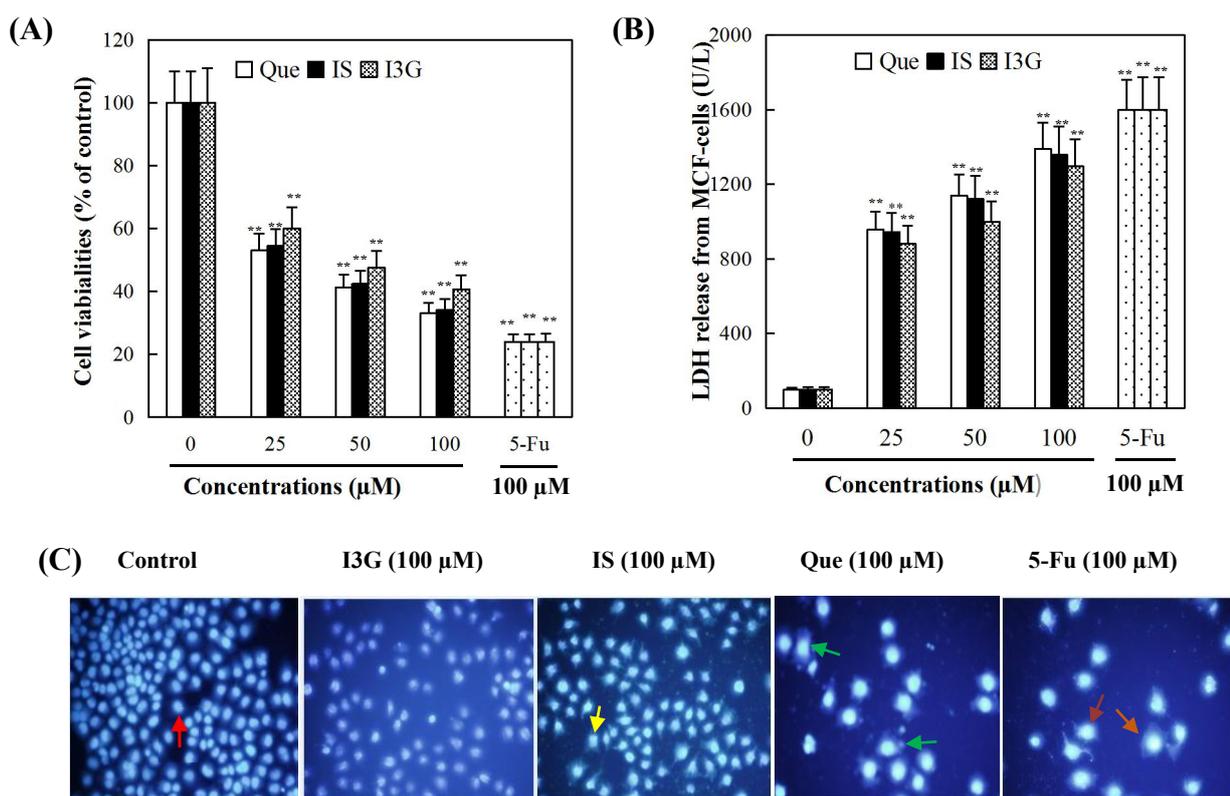


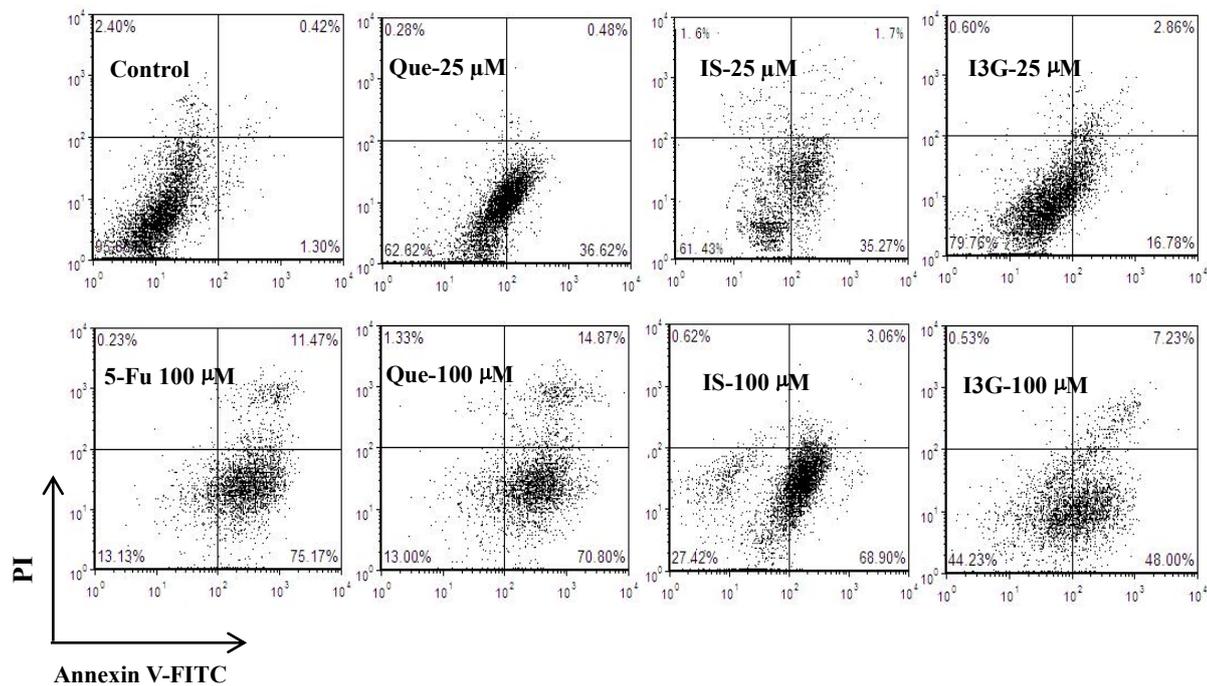
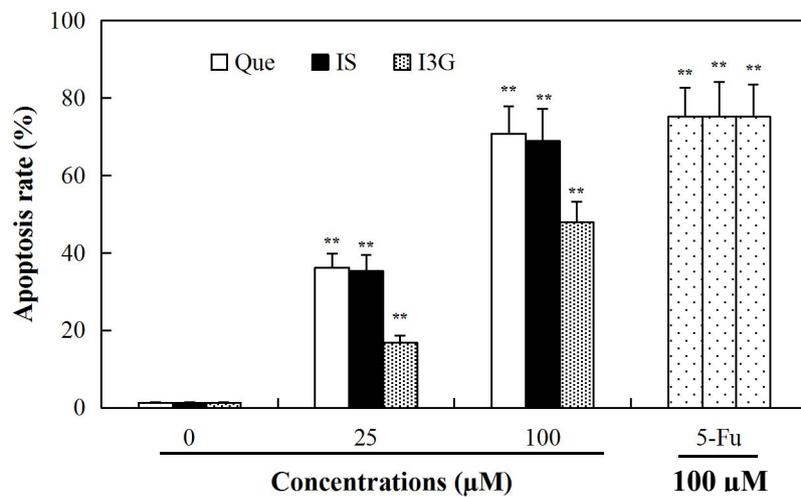
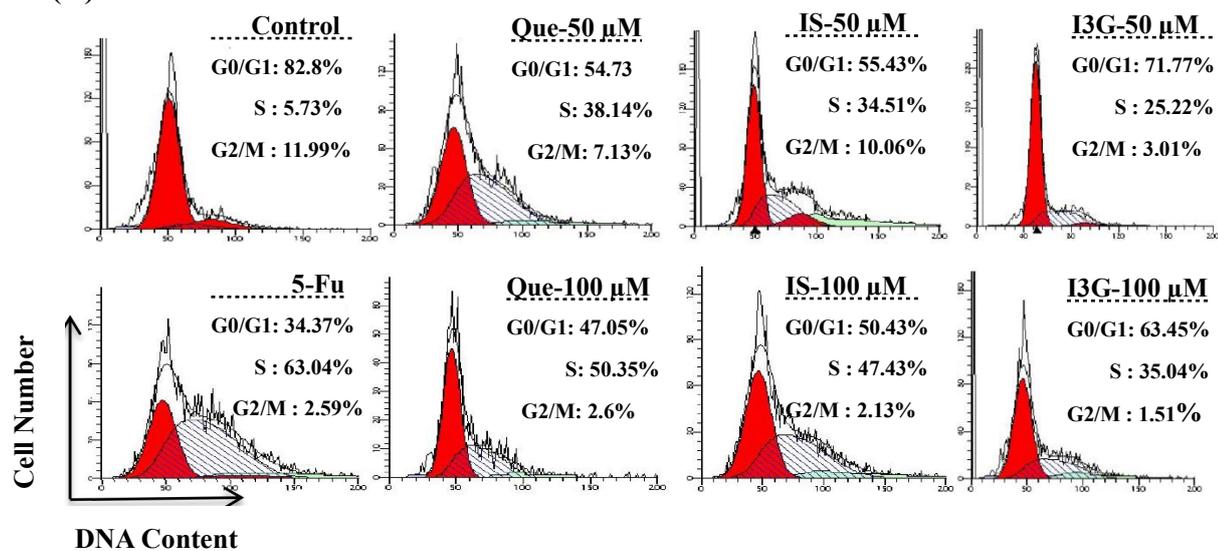
Figure 3**(A)****(B)**

Figure 4

(A)



(B)

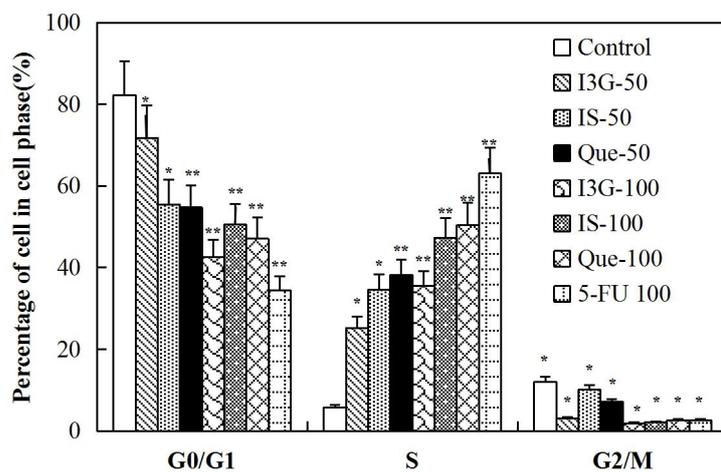


Figure 5

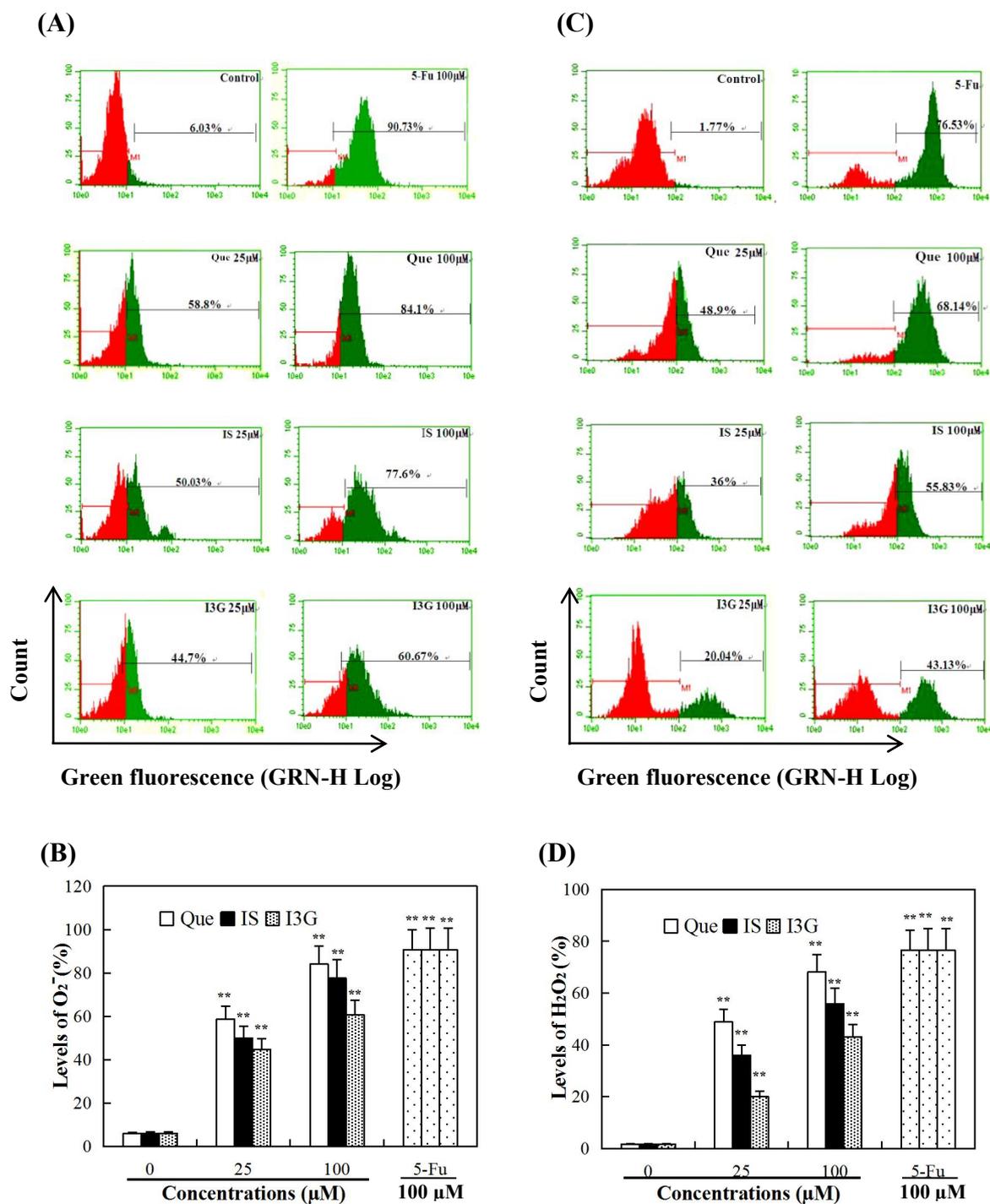
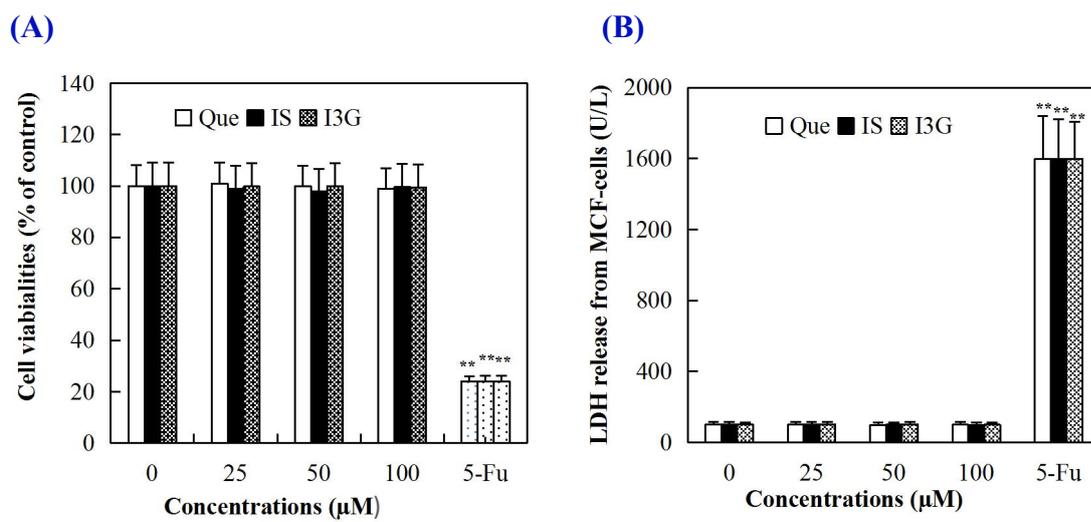
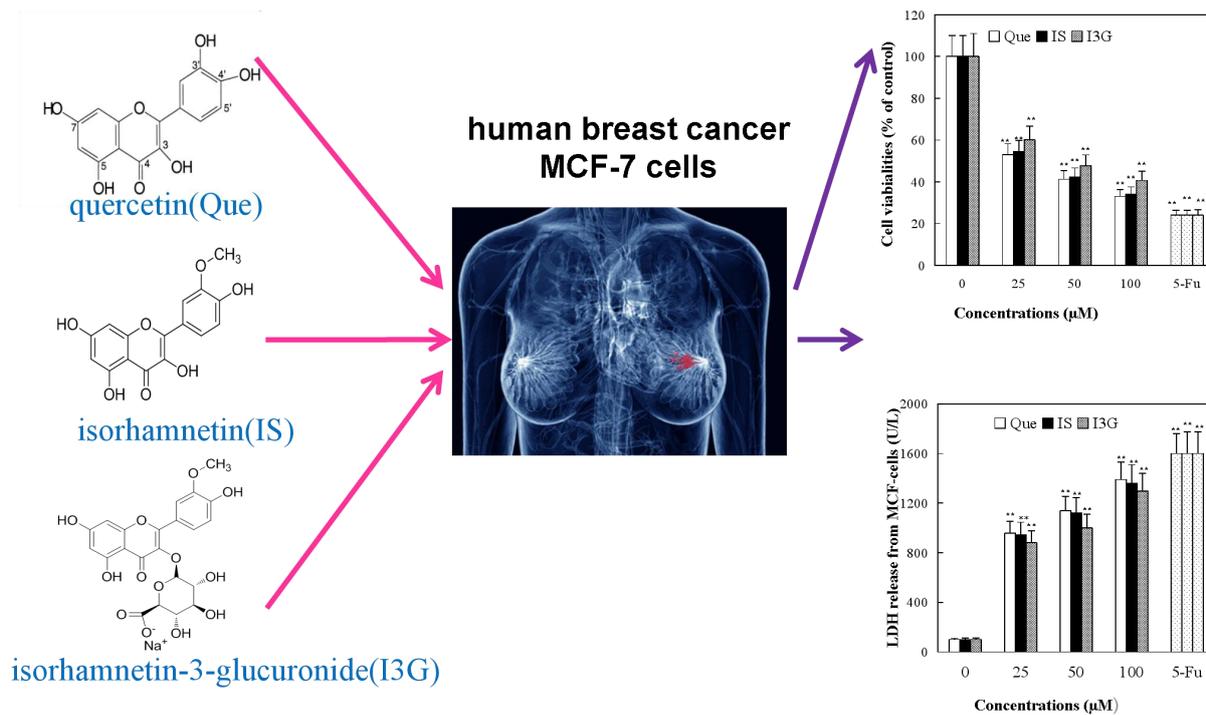
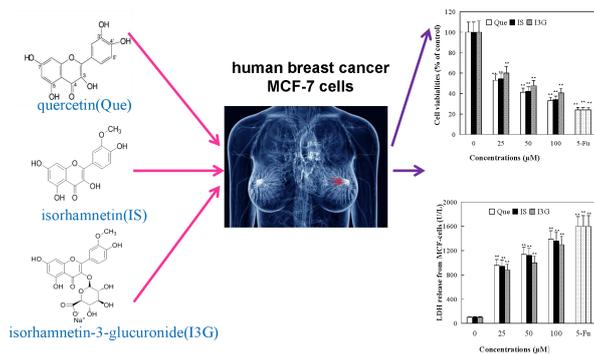


Figure 6





TOC Graphic (original drawing)



TOC Graphic(shrunken drawing)