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Pseudomonas aeruginosa ST357 with VEB ESBLs in the UK: relatedness and

ESBLs are uncommon in *Pseudomonas aeruginosa* in the United Kingdom (UK), but may be frequent e.g. in parts of the Middle East [1]. The types of ESBL predominantly found in the species are unusual, with VEB and PER often encountered, rather than the CTX-M, SHV, and TEM variants that dominate in Enterobacteriaceae [1,2]

From 2006-16, the Antimicrobial Resistance and Healthcare Associated Infections (AMRHAI) Reference Unit confirmed 30 VEB-positive *P. aeruginosa* isolates by PCR. Seven of these were from an outbreak in North West (NW) England, the early stages of which were described previously [2]; the remaining 23 were reference service submissions selected for *blaveb* PCR tests based on either: (i) a ceftazidime MIC  $\geq$ 256 mg/L reduced to  $\leq$ 32 mg/L by 2 mg/L clavulanate; or (ii) being from the same hospital as known positives and being resistant to ceftazidime at  $\geq$ 256 mg/L, irrespective of ceftazidime/clavulanate potentiation.

Despite their coming from 20 different hospitals we were struck that all 30 isolates 32 had the VNTR profile 13, 2, 1, 5, 2, 3, 6, 5, x, determined as previously described [3], where 33 'x' is variable. This profile corresponds to sequence type (ST) 357, a recognised 'high-risk P. 34 aeruginosa clone' for metallo  $\beta$ -lactamases (MBLs) [4]. These findings led us to screen 38 35 36 further ST357 isolates - referred for typing but not susceptibility testing - identifying an 37 additional 12 blaveb positives and giving a final working collection of 42 VEB-positive ST357 isolates from 36 patients at 26 hospitals (2006: one isolate, 2008: one isolate, 2010: four 38 39 isolates, 2012: seven isolates, 2013: ten isolates, 2014: seven isolates, 2015: eleven isolates, 2016: one isolate). Although we receive incomplete information on patients' origins and prior 40 41 travels, many isolates were likely imports to the UK: among 15 isolates from 13 patients hospitalized in the London area 11 were from private hospitals with international clienteles, 42 including many admissions from the Middle East. Seven isolates originated from patients 43 44 involved in a fire in Bucharest and transferred to different UK hospitals following prior 45 hospitalisation in Romania.

46 Fourteen isolates also had MBLs, as predicted from phenotypes and confirmed by PCR; 13 of these, including seven from the NW outbreak, had *blavim*; one had *blavim*. 47 48 Irrespective of MBL co-production, the VEB-positive isolates were extremely resistant to antipseudomonal agents, as determined by BSAC agar dilution with EUCAST breakpoints 49 50 (<u>http://www.eucast.org</u>). Ceftazidime MICs all were ≥256 mg/L, carbenicillin ≥1024 mg/L, cefepime  $\geq 64$  mg/L and piperacillin/tazobactam  $\geq 32$ mg/L, indicating consistent high-level 51 resistance. All but one isolate were resistant to impenem (41/42; 97.7%) whilst resistance to 52 53 meropenem was observed in 40/42 (95.3%). Resistance to ciprofloxacin (MICs  $\geq$  4 mg/L) was 54 universal and that to aminogly cosides was near universal (tobramycin MICs all  $\geq$ 16 mg/L. 100% non-susceptible; amikacin MICs  $8 \ge 64 \text{ mg/L}$ , 41/42 (97.7%) non-susceptible; 55 gentamicin MICs 4-≥32 mg/L, 41/42 (97.7%) non-susceptible). All remained susceptible to 56 colistin (MICs  $\leq 2$  mg/L). Where MBLs were absent (28/42 isolates), carbapenem resistance 57 58 likely depended on loss of porin OprD, which is frequent in *P. aeruginosa*, although this was 59 not examined directly.

Despite their frequent ceftazidime/clavulanate synergy, all of the isolates were 60 61 resistant to ceftolozane/ tazobactam, with MICs >16 mg/L, regardless of MBL co-production; it remains unclear whether this reflects poor penetration of *P. aeruginosa* by tazobactam or 62 tazobactam having little capacity to inactivate VEB enzymes. The behaviour of 63 ceftazidime/avibactam was more interesting. Avibactam evidently penetrates P. aeruginosa 64 as it potentiates ceftazidime against AmpC-derepressed strains, and reportedly can inhibit 65 66 extracted VEB β-lactamases [5]. Nevertheless, and despite avibactam being used at double 67 the concentration of clavulanate, resistance to ceftazidime/avibactam was observed in 95.3% of the isolates (40/42), with MICs above the 8+4 mg/L breakpoint and mostly remained above 68 69 those of ceftazidime/clavulanate (Table 1).

From our experience, potentiation of ceftazidime by clavulanate coupled with resistance to ceftolozane/tazobactam (MIC >16 mg/L) have good specificity as indicators of VEB ESBLs in *P. aeruginosa*, but sensitivity is poor, with ceftazidime/clavulanate MICs for
11.9% of producers identified here remaining above our top concentration of 32+2 mg/L.

74 To date, the genome sequences of three ST357 P. aeruginosa isolates have been 75 published, including one with a VEB-1 enzyme from a bloodstream infection in India. More 76 generally, ST357 is well known as a 'high-risk clone' also for VIM and IMP MBLs [4]. These carbapenemases, like VEB enzymes, are integron-borne, and ST357 may have a particular 77 78 ability to host these elements, We did not examine *blaveb* gene type or location in this study: 79 previous characterization of isolates from the NW outbreak mostly found *blaveb-1*, but with blaveb.9 in one representative [2]. The blaveb integron was chromosomally located in these 80 outbreak isolates but has also been recorded on plasmids from P. aeruginosa in other studies. 81

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Although VEB-positive *P. aeruginosa* remain uncommon in the UK, the present data show that they are repeatedly being introduced, commonly belong to an internationally successful lineage, and are extremely drug resistant - often remaining susceptible only to colistin. At a time when international emphasis is on carbapenemase producers, they should not be underestimated.

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## 89 Funding

90 This study was carried out as part of our routine work.

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## 92 Transparency declaration

DML: Advisory Boards or ad-hoc consultancy Accelerate, Achaogen, Adenium, Allecra,
AstraZeneca, Auspherix, Basilea, BioVersys, Centauri, Discuva, Meiji, Nordic, Pfizer,
Roche, Shionogi, Tetraphase, The Medicines Company, VenatoRx, Wockhardt, Zambon,
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97 Relevant shareholdings in – Dechra, GSK, Merck, Perkin Elmer, Pfizer amounting to <10% of</li>
98 portfolio value.

All other authors: none to declare. However, PHE's AMRHAI Reference Unit has received 99 100 financial support for conference attendance, lectures, research projects or contracted 101 evaluations from numerous sources, including: Accelerate Diagnostics, Achaogen Inc, Allecra Therapeutics, Amplex, AstraZeneca UK Ltd, AusDiagnostics UK Ltd, Basilea Pharmaceutica, 102 Becton Dickinson Diagnostics, bioMérieux, Bio-Rad Laboratories, The BSAC, Cepheid, 103 Check-Points B.V., Cubist Pharmaceuticals, Department of Health, Enigma Diagnostics, Food 104 Standards Agency, GlaxoSmithKline Services Ltd, Henry Stewart Talks, IHMA Ltd, Kalidex 105 106 Pharmaceuticals, Melinta Therapeutics, Merck Sharpe & Dohme Corp, Meiji Seika Pharmo Co., Ltd, Mobidiag, Momentum Biosciences Ltd, Nordic Pharma Ltd, Norgine 107 108 Pharmaceuticals, Rempex Pharmaceuticals Ltd, Roche, Rokitan Ltd, Smith & Nephew UK Ltd, Trius Therapeutics, VenatoRx Pharmaceuticals and Wockhardt Ltd. 109

110 Ethical approval: Not required

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## 112 **References**

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[1] Al-Agamy MH, Jeannot K, El-Mahdy TS, Samaha HA, Shibl AM, Plésiat P *et al.* Diversity of
 molecular mechanisms conferring carbapenem resistance to *Pseudomonas aeruginosa* isolates from Saudi Arabia. Can J Infect Dis Med Microbiol 2016;2016: 4379686.

117

[2] Woodford N, Zhang J, Kaufmann ME, Yarde S, Tomas M, Faris C *et al.* Detection of
 *Pseudomonas aeruginosa* isolates producing VEB-type extended-spectrum beta-lactamases
 in the United Kingdom. J Antimicrob Chemother 2008;62: 1265-8.

121

[3] Turton JF, Turton SE, Yearwood L, Yarde S, Kaufmann ME, Pitt TL. Evaluation of a ninelocus variable-number tandem-repeat scheme for typing of *Pseudomonas aeruginosa*. Clin
Microbiol Infect 2010;16: 1111-6.

125

- 126 [4] Wright LL, Turton JF, Livermore DM, Hopkins KL, Woodford N. Dominance of international
- 127 'high-risk clones' among metallo- $\beta$ -lactamase-producing *Pseudomonas aeruginosa* in the UK.
- 128 J Antimicrob Chemother 2015;70: 103-10.
- 129
- 130 [5] Lahiri SD, Alm RA. Identification of novel VEB  $\beta$ -Lactamase enzymes and their impact on
- avibactam inhibition. Antimicrob Agents Chemother 2016;60: 3183-6.

**Table 1**. Relationship of ceftazidime/clavulanate and ceftazidime/avibactam MICs for *P*.

Ceftazidime/clavulanate	n	Ceftazidime/avibactam (MIC mg/L)	n	MBL enzymes (n)
(MIC mg/L)*	n		n	MBL enzymes (II)
4	1	2	1	
		16	4	
8	9	32	3	
		>32	2	
		16	3	VIM (1)
16	14	32	5	
		>32	6	VIM (2)
		8	1	
32	13	16	2	VIM (1)
52	10	32	5	VIM (3)
		>32	5	VIM (3)
>32	5	32	1	VIM(1)
20Z	5	>32	4	NDM(1), VIM (2)

## 133 aeruginosa with blaveb

135 \*MICs of unprotected ceftazidime were  $\geq$ 256 mg/L in all cases