

1 **miR-16 is highly expressed in Paget's associated osteosarcoma**

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28 Dear Editor,

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30 Bone is an endocrine organ that produces key hormones and cytokines (Green et al., 2015b).
31 Paget's Disease of Bone (PDB) is a polygenic disorder of bone turnover first described by Sir
32 James Paget in 1876. PDB is characterised by hyper differentiation and hyper activity of
33 osteoclast cells which induces increased bone remodelling by osteoblasts. The resultant
34 mosaic of bone is structurally weaker, larger, more vascular and porous with an increased
35 susceptibility to fracture. On histology the osteoclasts are increased in size, population and
36 number of nuclei, expressing a "pagetic phenotype" that distinguishes them from normal
37 osteoclasts. Malignant transformation is a rare complication of PDB reported to arise in <1%
38 of PDB patients. Paget's associated osteosarcoma (PDB-OS) consistently arises in sites of
39 pagetic bone and may present with multifocal lesions (Hansen et al., 2006). On histology the
40 lesions are osteoblastic and characterised as an exaggerated form of the accelerated bone
41 remodelling that manifests in PDB. Median survival at diagnosis is 21 months for those treated
42 with curative intent and 7 months for those treated palliatively (Shaylor et al., 1999).

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44 PDB has a strong genetic component. A number of loci have been linked to the disorder with
45 sequestosome 1 (*SQSTM1*) variants associated with more severe symptoms, polyostotic foci
46 and heritable transmission (Hansen et al., 2006). *SQSTM1* is multifunctional protein which
47 serves as a signalling hub for diverse cellular events including activation of nuclear factor
48 kappa B (NFkB) and tumour necrosis factor superfamily member 11 (TNFSF11). *SQSTM1*
49 also serves as an autophagy receptor for degradation of ubiquitinated molecules via its
50 ubiquitin binding domain (Katsuragi et al., 2015). *SQSTM1* variants associated with PDB are
51 typically located within the coding region of the ubiquitin binding domain. Impairment of
52 autophagy is accompanied by massive accumulation of *SQSTM1* and formation of *SQSTM1*-
53 positive aggregate structures (Katsuragi et al., 2015). As the role of *SQSTM1* has not been
54 fully elucidated in transformation and there is no transcriptomic analysis of this cancer, we

55 took a next generation sequencing approach to evaluate the expression of small RNAs such
56 as microRNAs (miRNAs) in PDB and PDB-OS.

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58 MiRNAs are key regulators of gene expression through gene silencing. MiRNAs can also be
59 used as biomarkers to classify poorly differentiated cancers and cancer tissue origin (Green
60 et al., 2015a). We extracted RNA using the miRCURY RNA isolation kit (Exiqon) from two
61 tissue specimens of PDB-OS with a proven histological diagnosis of osteosarcoma in Paget's
62 affected bone (ages 73 and 81, 2 men). We extracted RNA from four **SQSTM1-positive** PDB
63 tissue specimens taken from **affected trabecular bone** (ages 79-87, 2 women and 2 men). We
64 extracted RNA from five control bone tissue specimens taken from the femoral heads of
65 trauma patients (ages 68-86, 3 women and 2 men). **Tissue samples were collected and**
66 **preserved at -20 °C. RNA was stored at -80 °C.** We generated small RNA libraries using high
67 definition (HD) adapters as previously described (Xu et al., 2015). HD adapters increase the
68 annealing efficiency between small RNAs and adapters. An increased annealing efficiency
69 significantly reduces the RNA ligase-dependent ligation bias in next generation sequencing
70 studies (Xu et al., 2015). We performed sequencing on the HiSeq 2500 Ultra-High-Throughput
71 Sequencing System (Illumina) at the Earlham Institute, Norwich Research Park.

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73 Raw fastq files were converted to fasta format. The HD signatures of the sequencing reads
74 were trimmed. Quality checking was performed using The UEA Small RNA Workbench
75 (www.srna-workbench.cmp.uea.ac.uk). Reads were mapped with no gaps allowed to the
76 human genome v38 using PatMaN. Small RNA expression levels were normalised using a
77 scaling approach, reads per total, to a fixed total of 10 million reads (Mohorianu et al., 2011).
78 Comparison of the samples was conducted using scatter plots, size-split boxplot of the
79 replicate-to-replicate differential expression, intersection and Jaccard similarity analyses
80 (Mohorianu et al., 2011). Differentially expressed reads between the control, PDB and PDB-
81 OS samples were identified using both an expression interval approach and pairwise
82 comparison using offset fold change (Mohorianu et al., 2011; Mohorianu et al., 2013). Analysis

83 was conducted using custom made Perl (5.24.0.1) and R (3.2.2) scripts. We observed a high
84 proportion of reads matching to the reference genome (Table 1). The size class distributions
85 were bimodal with peaks at 22 and 32 nt (Figure 1). These peaks correspond to miRNAs and
86 tRNA fragments which was confirmed using annotations. The small number of unique
87 sequences with high abundance is also visible in the complexity distributions (Figure 1). The
88 lower complexities correspond to 22 and 32 nt which indicate a small number of highly
89 abundant sequences (Figure 1). As with all human studies there was variability in the number
90 of reads assigned to either miRNAs or tRNA fragments in the control samples (when
91 compared to inbred genetic models such as mice and fruit flies where variability is minimal).
92 We identified a low expression of miR-16 in PDB compared to controls and a high expression
93 of miR-16 in PDB-OS compared to controls and PDB. We also identified a downregulation of
94 miR-144 and upregulation of miR-21 in PDB-OS compared to controls. Downregulation of
95 miR-144 is in line with previous data in osteosarcoma cells. Upregulation of miR-21 is
96 associated with many types of cancer. In this study we explored miR-16 further.

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98 Total RNA used for next generation sequencing was quantified by density measurement after
99 separation by agarose gel electrophoresis with ethidium bromide staining. Equal amounts of
100 RNA across the sample pools were reverse transcribed using the TaqMan advanced miRNA
101 cDNA synthesis kit (Thermo Fisher Scientific). Differential expression of miR-16 was validated
102 three times in triplicate using TaqMan miRNA advanced assays (Thermo Fisher Scientific).
103 Digital PCR was performed on the QuantStudio 3D Digital PCR System using the GeneAmp
104 PCR System 9700 (Thermo Fisher Scientific). After PCR the chips were imaged on the
105 QuantStudio 3D Instrument which assesses raw data and calculates the concentration of the
106 cDNA sequence targeted by FAM and VIC labelled probes by Poisson distribution (Fazekas
107 de St, 1982). For more in depth analysis the QuantStudio 3D AnalysisSuite was used to report
108 the data as copies/ μ L. Probe sequence used was miR-16-5p (5'-
109 UAGCAGCACGUAAAUAUUGGCG-3'). We confirmed the downregulation of miR-16 in PDB

110 compared to controls and upregulation of miR-16 in PDB-OS compared to controls and PDB
111 (Figure 1).

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113 To elucidate a mechanistic role of miR-16 in PDB-OS we used TargetScan v7.1 to search for
114 its messenger RNA targets. *SQSTM1* was identified as a candidate which was supported by
115 performing a second search and identification on miRSearch v3. Target identification of
116 miRNAs is computationally difficult due to the relatively low homology between miRNAs and
117 their targets. We searched the literature and identified *SQSTM1* as an experimentally
118 confirmed target of miR-16, i.e. levels of *SQSTM1* are reduced by miR-16 overexpression
119 (Selbach et al., 2008). We find that miR-16 is downregulated in PDB which means there is a
120 loss of *SQSTM1* negative regulation. In line with previous data *SQSTM1* variants in PDB are
121 activating mutations. *SQSTM1* is a signalling hub for the activation of TNFSF11. TNFSF11 is
122 the primary ligand for stimulating the differentiation of monocyte progenitors to osteoclasts
123 through interaction with tumour necrosis factor receptor superfamily member 11a
124 (TNFRSF11A). TNFSF11 is produced by osteoblasts and downstream repercussions of the
125 TNFSF11-TNFRSF11 interaction is the increase of bone resorption. Bone resorption achieved
126 by various osteoclast-produced cytokines is set within a positive feedback loop which
127 encourages osteoblast proliferation and activity. Increased proliferation of osteoblast cells
128 containing *SQSTM1* activating variants repeats the bone remodelling cycle each time
129 increasing the number of *SQSTM1* positive osteoblasts. PDB does not typically present until
130 patients are >55-years-old. In patients harbouring *SQSTM1* variants, presentation can appear
131 earlier. Presentation may be due to the speed of the bone remodelling cycle or the gradual
132 loss of miR-16 mediated silencing of *SQSTM1*.

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134 We find a high expression of miR-16 in PDB-OS. The effect of upregulated miR-16 means
135 there is increased negative regulation of genes including *SQSTM1*. Loss of *SQSTM1* has little
136 effect on autophagy (Katsuragi et al., 2015). The lack of phenotypic impact might be explained
137 by the presence of other autophagy regulators including optineurin (OPTN) (Katsuragi et al.,

138 2015). As a signalling hub SQSTM1 is a partner of the Kelch like ECH associated protein 1
139 (KEAP1)-nuclear factor erythroid 2 like 2 (NFE2L2) pathway, a major cellular defence
140 mechanism against oxidative stress (Katsuragi et al., 2015). In normal conditions NFE2L2 is
141 constitutively degraded by the ubiquitin-proteasome system because its binding partner
142 KEAP1 is an adapter of ubiquitin ligases. Upon exposure to oxidative stress KEAP1 is
143 structurally modified which releases NFE2L2 to translocate to the nucleus. NFE2L2 is a
144 transcription factor which then induces a multitude of antioxidant and anti-inflammatory genes.
145 SQSTM1 is able to abrogate the interaction between KEAP1 and NFE2L2 leading to protection
146 against oxidative stress (Katsuragi et al., 2015). We propose miR-16 mediated silencing of
147 *SQSTM1* incapacitates the cell's ability to protect itself against oxidative stress-induced
148 oncogenesis. This proposal warrants further molecular investigation **to show an inverse**
149 **correlation between the expression of miR-16 and *SQSTM1*.**

150

151 Molecular testing may help provide a robust diagnosis and is particularly useful in rare
152 cancers. This is especially true in PDB where transformation to osteosarcoma is often missed
153 until late stage. Radiograph morphology is subtly different between PDB and PDB-OS.
154 Symptoms of PDB-OS can be similar to the day-to-day symptoms experienced by a PDB
155 patient. Biomarkers such as serum total alkaline phosphatase used in the diagnosis of PDB
156 are the same biomarkers used to aid the diagnosis of PDB-OS. These features combined
157 make it difficult to distinguish between a chronic non-life threatening disorder and a fatal
158 disease which require very different treatment strategies. Quantitative PCR or next generation
159 sequencing performed on RNA extracted from a biopsy of a presenting lesion is precise for
160 detecting a reduced expression of miR-16 in 'normal PDB' and high expression of miR-16 in
161 PDB-OS. The switch in miR-16 expression could alert physicians to the change to an
162 osteosarcoma phenotype.

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164 A limitation of this study is the size of the cohorts studied. PDB-OS arises in <1% of PDB
165 patients and donation to tissue banks is scarce. We are in the process of collecting a larger

166 cohort of tissue specimens to validate the findings from this study. Our data highlights the
167 value of being able to provide a robust tissue diagnosis in addition to identifying regulatory
168 transcriptomic molecules that could be exploited for targeted therapy. Expression of miR-16
169 could also be investigated in paediatric osteosarcoma to evaluate similarities and/or
170 differences between the two age-related incidence peaks of osteosarcoma.

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172 **DATA AVAILABILITY**

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174 The data presented in this study is publicly available on Gene Expression Omnibus
175 under the accession numbers GSE85809 (GSM2284729 to GSM228473 are control samples;
176 GSM2284736 and GSM2284736 are PDB-OS samples) and GSE87018 (GSM2318966 to
177 GSM2318970 are PDB samples).

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