

# Multifunctional quantum dots and liposome complexes in drug delivery

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## Abstract:

Incorporate both diagnostic and therapeutic functions into a single nanoscale system is an effective modern drug delivery strategy. Combine liposomes with Semiconductor quantum dots (QDs) have great potential to achieve such dual functions, referred to in this review as a liposomal QD hybrid system (L-QD). Here we review the recent literature dealing with the design and application of L-QD for advanced in bio-imaging and drug delivery. After a summary of L-QD synthesis process and evaluation of their properties, we will focus on their multifunctional applications, ranging from *in vitro* cell imaging to theranostic drug delivery approaches.

**Keywords:** liposomes, nanomedicine, drug delivery

## Introduction of liposomal quantum dot Hybrid delivery system

The use of nanoparticles has increased in the areas of drug delivery<sup>[1, 2]</sup>, cancer detection<sup>[3]</sup>, and therapeutics<sup>[1, 4-7]</sup>. Quantum dots (QDs) are one of the most interesting examples due to unique optical characteristics that enable them to overcome the limitations of previously used

organic fluorophores<sup>[8, 9]</sup>. On the other hand, liposomes are nanoscale spherical vesicles composed of phospholipid bilayers<sup>[10]</sup>. The more recent proposition to combine liposomes with QDs not only increases biocompatibility of QDs, but also opens opportunities for creating theranostic nanoscale delivery systems, which can combine simultaneous therapeutic and imaging functions<sup>[11]</sup>. Combining QD with therapeutic agent in a liposomal delivery system allows the bio-distribution of the payload can be monitored *in vivo*, reducing the potential for unintended side effects of drug toxicity in healthy tissues. In addition, the use of such hybrid systems potentially allows clinicians to monitor the progress and efficacy of a therapy throughout the course of treatment<sup>[11]</sup>. In this review, we have focused on the liposomal QD hybrid system (L-QD), where imaging agent and therapeutic agent and can be co-delivered.

### **Liposome based drug delivery system**

Liposomes were discovered by Alec D Bangham in the 1960s at the Babraham Institute, University of Cambridge<sup>[12, 13]</sup>. Since their discovery, liposomes have been widely investigated, and are considered to be one of the most successful drug delivery systems. The first liposome formulation was composed solely of natural lipids; now liposomes can be created from cholesterol and natural or synthetic phospholipids with additional modification to prolong their *in vivo* circulation time<sup>[10]</sup>. In aqueous solutions, due to their amphipathic nature phospholipids have a strong tendency to form membranes<sup>[14]</sup>. Hydrophobic drugs/QDs can be enclosed in-between membranes, while hydrophilic drugs/QDs can be encapsulated inside central compartment. To improve the delivery of therapeutic molecules, surface modification of liposomes by the inclusion of hydrophilic carbohydrates, glycolipids or polymers, such as polyethylene glycol (PEG) can be used<sup>[15, 16]</sup>. The PEGylation of the liposomes would prolong the blood-circulation time while suppressing the uptake by the reticuloendothelial system (RES)<sup>[17, 18]</sup>. These long-circulating liposomes can passively accumulate in tumour site through the porous endothelium present in tumours—the enhanced permeability and retention (EPR) effect<sup>[16]</sup>. Or active targeting can be achieved by further modify the PEG terminus with functional molecules e.g. specific ligands or monoclonal antibodies<sup>[19-21]</sup>.

Liposomes can vary from a few nanometres to several micrometres and may have one or more lipid bilayers. The sizing of liposome is a critical parameter which determines their therapeutic efficiency<sup>[22]</sup> and helps characterise liposomes. On the basis of their size and lamellarity, liposomes can be classified into one of three categories: multilamellar vesicles

(MLV), large unilamellar vesicles (LUV) and small unilamellar vesicles (SUV)<sup>[23]</sup>. MLV have an onion like structure with diameters of 1-5 $\mu$ m. SUV are typically 50nm in diameter while LUV range from 100-250nm or even larger<sup>[24]</sup>. There are several methods for the preparation of liposomes. One of the most widely used techniques for liposome synthesis is the thin-film hydration <sup>[10]</sup>, where dry lipid film is hydrated by adding an aqueous buffer solution under agitation at temperature above the lipid transition temperature<sup>[25]</sup>. During this process hydrophilic drugs are entrapped by using the aqueous solution of these materials as hydrating fluid, while lipophilic drugs are solubilised in the organic solution of the constitutive lipid. However, using this method, large and nonhomogeneous MLVs have formed and further steps are needed to produce homogeneous SUV. The second method is solvent solvent-injection using either ether or ethanol. This method involves the dissolution of the lipids into either or ethanol, followed by the injection of the lipid solution into aqueous solution <sup>[26, 27]</sup>. The ether injection method has one advantage over ethanol injection method as ether is immiscible with water so can be removed directly during injection process by using warmed aqueous phases above the boiling point of the ether<sup>[25]</sup>. The third method is reverse-phase evaporation, which is based on the creation of inverted micelles. Briefly, the water-in-oil emulsion is formed by brief sonication of organic solvent containing phospholipids and aqueous buffer. Then the liposomes are shaped when the organic solvent is evaporated during continued rotary evaporation under reduced pressure<sup>[28]</sup>. Other methods for liposomes preparation includes detergent removal<sup>[29]</sup>, Spray-drying<sup>[30]</sup>, freeze drying<sup>[31]</sup>, and microfluidisation<sup>[32]</sup>.

Drug loading into liposome can be attained either passively (i.e., the drug is encapsulated during liposome formation) or actively (i.e., after liposome formation). Hydrophobic drugs can be directly combined into liposomes during the formation process. The encapsulation efficiency is dependent on the properties of the lipid such as length and packing density as well as drug-lipid interactions. Passive loading of hydrophilic drugs depends on the ability of liposomes to trap aqueous solution containing a dissolved drug during vesicle formation. Only a small percentage of a hydrophilic drug can be encapsulated by passive loading using the most common thin film hydration method<sup>[33]</sup>. The highest encapsulation efficiency by means of passive loading (up to 65%) is achieved by the reverse-phase evaporation method. However, this technique uses organic solvents, which will leave probably a small amount of residual in the liposome suspension, and thus is not feasible for commercial production<sup>[23, 33]</sup>. Compared to passive loading, active loading employing pH gradient methods could achieve near 100% encapsulation efficiencies <sup>[34, 35]</sup>.

Due to their biocompatibility and biodegradability, liposomes are considered safer drug delivery systems. Both diagnostic and therapeutic agents can be encapsulated into liposomes. The encapsulation of drugs into the liposomes protects them against enzymatic degradation and immunologic inactivation, thus improves their therapeutic activity. Meanwhile the encapsulation minimises exposure of healthy tissue to the drugs during their circulation in the blood. Therefore, the systemic toxicity will be largely reduced compared with the free drugs<sup>[36]</sup>. Currently, several liposome-based drugs are approved for clinical practice such as Doxyl®<sup>[37, 38]</sup> and AmBisome®<sup>[39]</sup>; many others are in the various stages of clinical trials<sup>[40-43]</sup>.

## **Quantum dots**

Amongst the various types of nanomaterials, *i.e.* metals, metal oxides, organic materials or biomaterials, semiconductor nanoparticles (NPs), also referred to as quantum dots (QDs), has become a major interdisciplinary area of science due to their unique properties<sup>[44]</sup>. QDs are generally considered to be particles of material which have diameters in the range of 1 to 10nm<sup>[45]</sup>. The small dimensions of QDs result in properties differing from those seen in the corresponding bulk material. These unique physical properties give rise to many potential applications in areas such as luminescence, electronics, catalysis, and optoelectronics<sup>[46-49]</sup>. The size of the particle and surface to volume ratio are two fundamental factors of individual QD, which are responsible for these unique properties. As a particle becomes smaller the band gap gradually becomes larger because of quantum confinement effect. Also, as the size of the semiconductor material becomes smaller the ratio of the number of surface atoms to those in the interior increases, which leads to the surface playing an important role in the properties of the material<sup>[50]</sup>. Thus, both particle size and the synthetic method determine the physical and electronic properties of the QD produced, which gives scientists the unique ability to change the electronic and chemical properties of a QD.

QDs have quite a few advantages over traditional fluorescent organic molecules. They have size- and composition-tuneable fluorescence emission from ultra violet to infrared wavelengths, narrow spectral line widths, high luminescence quantum efficiency, broad absorption profiles, and stability against photo-bleaching<sup>[9, 51-53]</sup>. The broad absorption and narrow emission spectra of QDs allow the simultaneous detection of multiple colours of QDs upon illumination with a single light source which is very useful for fluorescent multiplexed analysis in biological system<sup>[51, 54]</sup>. Due to their photophysical characteristics, QDs are thought to have potential as novel fluorescent probes for diagnostic purposes<sup>[9, 52]</sup>. However, their biocompatibility and potential toxicity remain critical issues for use in humans and thus

limit their application<sup>[55, 56]</sup>. The combination of liposome and QDs to create stable and multifunctional assemblies could overcome the shortfall of QDs and pave the ways for the development of novel drug delivery system.

### **Liposome and QD Hybridisation**

QD is an ideal candidate for fluorescent imaging in both the biomedical and pharmaceutical fields. However, QD are mostly synthesised in non-polar organic solvents, their surface hydrophobicity or poor colloidal stability at physiological conditions frequently renders them inappropriate for biomedical and clinical applications. Many strategies have been developed to overcome this limitation, such as functionalisation of QD with peptides, antibodies, and polymers<sup>[57, 58]</sup>. However, evidence shows that the presence of these ligands could be disadvantageous for their application. Surface modification often quenches QD fluorescence and decreases their photostability<sup>[59, 60]</sup>, which for in vivo applications requires higher doses of administered QD and therefore increases potential toxicity. Encapsulation of QD within liposomes may avoid the chemical functionalisation of QDs, which can lead to enhanced stability in plasma, better control of the pharmacological fate, and an overall improvement in their biocompatibility. Moreover, therapeutic agent e.g. anticancer drug could also be added to the QD liposome hybrid to create a device with both therapeutic and diagnostic functions. For example, hydrophobic QDs could be embedded in the lipid bilayer, whereas hydrophilic therapeutic agents can be encapsulated within the internal liposome aqueous core<sup>[61, 62]</sup>. The hybridisation of the QDs with liposome generates a novel, interesting class of nano-range delivery devices, which combines the unique optic properties of QD and the lipid functional moiety, thus offering new opportunities in multifunctional drug delivery system development. A summary of the types of hybrid delivery systems that could be used for simultaneous diagnostics and therapeutics of cancer is presented in Table 1.

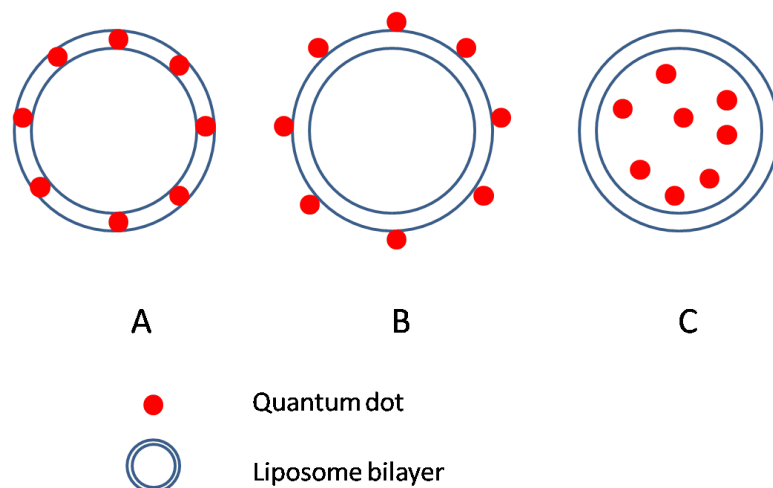
*Table 1 Representative liposome QD hybrid systems for simultaneous imaging and therapy of cancer*

<b>Types of QD used</b>	<b>Incorporated therapeutic agent</b>	<b>Structure of hybrid systems</b>	<b>Target (tumour) cells or xenografts [targeting mechanism/species employed]</b>	<b>Ref.</b>
<b>Trioctylphosphine oxide (TOPO)-coated CdSe QDs</b>	N/A	QD inside hydrophobic interior of lipid bilayer	HEK293 human embryonic kidney cells [non-specific internalization]	[63]
<b>TOPO-coated CdSe/ZnS</b>	N/A	QD inside hydrophobic interior of lipid bilayer	A549 human epithelial lung cells and CD-1 nude mice 7 inoculated with C33a human cervical carcinoma cells [non-specific internalization]	[64, 65]
<b>TOPO-coated CdSe/ZnS and Carboxyl-functionalised QD</b>	N/A	TOPO-QD inside hydrophobic interior of lipid bilayer and Carboxyl-QD inside interior aqueous compartment	HuH-7 human hepatocellular carcinoma cells [non-specific internalization]	[66]
<b>Cetrimonium Bromide (CTAB) capped CdTe or CdHgTe QDs</b>	N/A	QD inside hydrophobic interior of lipid bilayer	MCF7 human breast carcinoma cells and Kunming mice [non-specific internalization]	[67]
<b>TOPO-coated CdSe/ZnS</b>	Doxorubicin (Dox)	TOPO-QD inside hydrophobic interior of lipid bilayer and Dox inside interior aqueous compartment	N/A	[61, 62]
<b>CdSe and CdSe/ZnS QDs (Lumidot®)</b>	Cisplatin	QD inside hydrophobic interior of lipid bilayer and Cisplatin inside interior aqueous compartment	Melanoma cells and nude mice [non-specific internalization]	[68]

<b>Dihydroxyloipoic acid (DHHA)-capped CdSe/ZnS QDs</b>	N/A	QD on the surface of liposome	B16F10 cells and C57BL/6 mice [tracking extravasation with QD labelled cells]	[69]
<b>Carboxyl CdSe/ZnS QDs</b>	Dox	QD on the surface of liposome and Dox inside interior aqueous compartment	MCF-7/HER2 cells and xenografts [anti-HER2 single chain Fv fragments]	[70]
<b>Streptavidin conjugates CdSe/ZnS QDs and ITK-carboxyl CdSe/ZnS QDs</b>	N/A	streptavidin-QDs on the surface and carboxyl-QD inside interior aqueous compartment	A431 human epidermoid carcinoma and CHO Chinese hamster ovary cell lines [epidermal growth factor (EGF) ligand]	[71]
<b>3-mercaptopropionic acid (MPA)-capped CdTe QDs</b>	N/A	QD inside interior aqueous compartment	Xenografts [non-specific internalization]	[72]
<b>thiol-capped CdTe QDs</b>	N/A	QD inside interior aqueous compartment	U2OS human osteosarcoma cell, Hela human cervical carcinoma cell and 293T human embryonic kidney cell [non-specific internalization]	[73]

## Formulation of liposomal QD Hybrid system

Developing liposome-based drug delivery system with QD-based fluorescence imaging requires the hybridisation of QD with liposomes. Figure 1 depicts three different strategies for QD incorporation. Hydrophobic QDs have been inserted into the hydrophobic interior of the liposomal membrane<sup>[61, 63, 64]</sup>, and hydrophilic QDs have been encapsulated in the interior aqueous compartment<sup>[66, 71, 74, 75]</sup> or on the outer membrane of liposomes<sup>[69, 70]</sup>. To achieve these, different methods have been used, depend on properties of QD.



*Figure 1 Quantum dots incorporation models of hybrid delivery systems. A-hydrophobic quantum dots encapsulated shell model, B-hydrophilic quantum dots linked shell model, C-hydrophilic quantum dots encapsulated core model.*

Thin film hydration method is one of the most common techniques used in liposome production. Both hydrophobic and hydrophilic molecules can be encapsulated into liposomes during the synthesis process. Hydrophobic QDs could be easily incorporated into the lipid bilayer when added to the thin lipid film. lipid Vogel and co-workers have incorporated hydrophobic TOPO functionalised CdSe QDs into the bilayer membrane of liposome using thin film hydration<sup>[63]</sup>. A high yield of LUV along with some MLV was obtained using this approach, which sizes ranged from 50nm to 50µm. The researchers also used electric field aided hydration to shorten process time and produced more homogeneous unilamellar structures. However, they didn't show if there was any influence on vesicle size by such process and there weren't any further steps used to reduce the liposome size. In another study, Al-Jamal, Al-Jamal, Tian, Lacerda, Bornans, Frederik and Kostarelos<sup>[64]</sup> encapsulated TOPO capped CdSe/ZnS QDs into zwitterionic and cationic lipid bilayer of SUV using thin film hydration plus sonication. The incorporation of hydrophobic QDs in the lipid bilayer led to



significant enhancement of their optical stability during storage and exposure to UV irradiation compared to that of QD alone in toluene. This method was adapted by Tian, Al-Jamal, Al-Jamal and Kostarelos<sup>[61]</sup>, who not only incorporated QDs into the lipid bilayer, but also encapsulated doxorubicin (Dox) into the core of the liposome. The loading of Dox into L-QD was done by the osmotic gradient technique and achieved more than 97% loading efficiency.

Unlike encapsulating hydrophobic QD, where thin film hydration is the dominant method<sup>[61, 62, 64, 76-78]</sup>, there are several different methods to incorporate hydrophilic QD into liposomes. First, thin film hydration could also be used for incorporating hydrophilic QD into the aqueous core of liposome<sup>[72, 74, 75]</sup>. Al-Jamal, Al-Jamal, Bomans, Frederik and Kostarelos<sup>[74]</sup>,<sup>[75]</sup> synthesised COOH-PEG-lipid coated L-QDs by thin film hydration followed by sonication as they did for the hydrophobic CdSe/ZnS QD. The thin lipid film was prepared and was hydrated with QD suspension. SUV were then prepared by further bath sonication. Ye, Wang, Li, Yu and Hu<sup>[67]</sup> have also incorporated hydrophilic CdTe QDs into the vesicle lipid bilayer using the classic thin film hydration method. CTAB was used to make negatively charged hydrophilic QDs soluble in lipid mixture. Further sonication step was also used to form SUV loaded with QDs. Second, Bothun, Rabideau and Stoner<sup>[66]</sup> used a single step reverse phase evaporation method to create a liposomal system that deliver both green hydrophobic and red hydrophilic CdSe/ZnS QDs into carcinoma cells. In the reverse phase evaporation process, an organic solvent phase containing lipids and the hydrophobic QDs was mixed with an aqueous phase containing the hydrophilic QDs to form an emulsion. The organic solvent was then removed by rotary evaporation to collapse the emulsion phase and drive QD incorporation within the liposome. Hansen, van Emmerik, van Gaal, Storm, van Hest and Lowik<sup>[79]</sup> also used reversed phase evaporation approach and further purified their QD-loaded liposome by Ion-exchange method. Third, the detergent dialysis technique was also used to encapsulate carboxyl CdSe/ZnS QD into core of the PEGylated liposome<sup>[71]</sup>. To do this, the lipid film was hydrated in Hepes Buffer Saline (HBS) containing both 1-O-N-Octyl- $\beta$ -D-glucopyranoside (OGP) detergent and carboxyl CdSe/ZnS QDs. The result solution was immediately dialysed against HBS to form hybrid liposome.

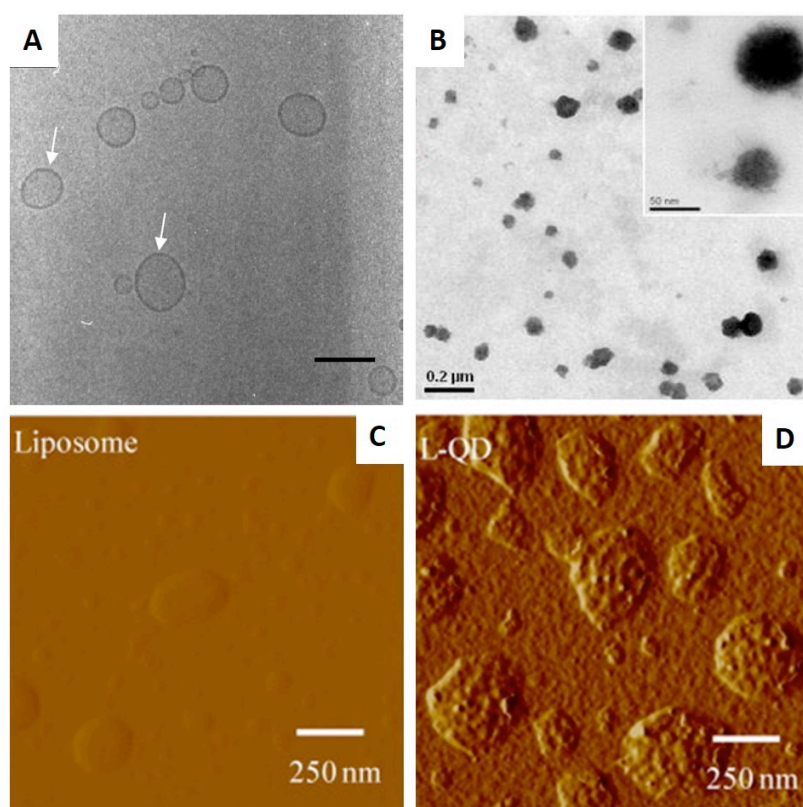
An alternative approach to hybridise hydrophilic QD with liposome is to conjugate the particle onto the surface of liposome. Voura, Jaiswal, Mattoussi and Simon<sup>[69]</sup> combined DHLA-capped QDs with the commercially available Lipofectamine 2000. Because the liposome consisted of cationic lipids are positive charged and the DHLA-QDs are negatively charged, they joined together by electrostatic force. The L-QD was simply formed by

incubation of QD with liposome solution. In another report, Weng, Noble, Papahadjopoulos-Sternberg, Chen, Drummond, Kirpotin, Wang, Hom, Hann and Park <sup>[70]</sup> have developed a multifunctional L-QD with imaging, targeting, and therapeutic modalities. The liposomes were simply prepared by extrusion. The carboxyl CdSe/ZnS QDs were then covalently linked to liposomes through a cross-linker. Furthermore, Dox was also encapsulated into the L-QD by a gradient loading technique either before or after covalent linking of the QDs to liposomes. It is worth to note that lower drug loading efficacy (~30%) was showed when drug loaded into liposomes after QD conjugation, which was probably due to the shielding of QD on liposome surface.

## Characterisation of liposomal QD hybrid system

### *Morphology of liposomal QD hybrid system*

Morphological characterisation of liposomal QD hybrid system helps in understanding the relative structure, position of the QD molecules and assembly of the L-QDs. These characteristics can influence the stability of the L-QD in vitro as well as in vivo and also can impact the bio-distribution. Morphology of QD and liposome complexes can be studied using techniques such as, cryogenic transmission electron microscopy (cryo-TEM), freeze fracture electron microscopy (ff-EM) and Atomic force microscopy (AFM).



*Figure 2 (A) Cryo-TEM images of Liposome-QD hybrid vesicles, scale bar 100nm. Reproduced with permission from ref. 64. Copyright 2008 American Chemical Society; (B) TEM images of the liposome-coated QDs. Reproduced with permission from ref. 72. Copyright 2010 Springer; (C) AFM images of empty liposomes; (D) AFM images of (DSPC) Liposome-QD hybrid vesicles. Reproduced with permission from ref. 61. Copyright 2011 Elsevier.*

Using Cryo-TEM Al-Jamal, Al-Jamal, Bomans, Frederik and Kostarelos<sup>[74]</sup> showed clearly that vesicular structures of liposome were formed and that hydrophilic QDs were incorporated into the core of vesicle. It was also consistently observed that the QDs were interacting with the lipid bilayer rather than being simply encapsulated into the vesicle inner aqueous compartment, suggesting some degree of mixing between the bilayer lipids and the functional ligands on the QD surface. In a later study, the same group showed several Cryo-TEM images which indicated that incorporating hydrophobic QD into lipid bilayer with low lipid:QD ratios (1000:1) resulted in elongated and deformed vesicular structures. Moreover, the lipid bilayer thickened at specific locations, which suggested that the incorporation of QD was taking place in “pockets” rather than being evenly distributed throughout the bilayer. According to their results, the ideal lipid to QD ratio for L-QD formation is 10000 to 1. At this ratio, perfect spherical liposomes were formed, and there was no noticeable physical deformation of the liposomes by hydrophobic QDs<sup>[64]</sup>. In general, hydrophobic QD containing liposome showed darker rim than the interior of the vesicles because of the presence of the electron-dense QD in the lipid bilayer<sup>[64, 77]</sup> (Figure 2A). Whereas, in case of hydrophilic QD (third model, Figure 1C), TEM images showed a pale edge and dark centre of the obtained L-QD, which suggested the QDs were incorporated into the core of liposome<sup>[72]</sup> (Figure 2B). The technique of AFM was used by Tian, Al-Jamal, Al-Jamal and Kostarelos<sup>[61]</sup> to investigate the structural elucidation of the hybrid vesicles. Interestingly, in contrast to the liposome control which showed a smooth surface, the image of L-QD showed that the incorporation of QD into the lipid bilayers resulted in a rough surface (Figure 2C&D). 3D image analysis indicated that QD associated with the lipid bilayers and distributed throughout the vesicle surface. Furthermore, the cross-section analysis suggested that QD incorporation increased the height of the liposome from 8 nm to almost 20 nm. Weng, Noble, Papahadjopoulos-Sternberg, Chen, Drummond, Kirpotin, Wang, Hom, Hann and Park<sup>[70]</sup> used ff-EM to characterise the nanoscale structures of the both free carboxyl QDs and when conjugated to liposomes outer layer. QDs appeared as small, mostly spherical particles with

shadowing behind the structure, which was typical for hard-core particles. In contrast to QDs, liposomes displayed convex and concave fracture planes, which was typical for membrane-bound structures. The ff-EM images also showed that a small number of QDs appeared on liposomal fracture planes, which reflected imprints of surface-attached QDs showing through the semifluid, liquid-crystalline bilayers.

### ***Particle size and surface charge***

Size and size distribution measurements are formulation parameters that indicate homogeneity of the particles in liposomal formulations and are very important for formulation and process optimisation. The poly-dispersity index (PI) of liposome reflects the range of liposome species present around the target average liposome size. In general, uniformly sized liposomes with a lower PI ( $<0.2$ ) are preferred for drug delivery. Moreover, changes in the average particle size and PI can be used as indicators of long-term stability. Dynamic light scattering (DLS) and transmission electron microscopy (TEM) are the most common techniques to determine the size of liposomes. Al-Jamal, Al-Jamal, Bomans, Frederik and Kostarelos <sup>[74]</sup> demonstrated that incorporating of hydrophilic QD into the core of liposome largely increased liposome size as determined by both DLS and cryo-TEM. The size distribution was wide, as expected from thin film hydration preparation method. The surface charge characteristics of the L-QDs were in accordance with the characteristics of the lipid molecules used to form the bilayers. Sigot, Arndt-Jovin and Jovin <sup>[71]</sup> showed a similar result from TEM measurement that hydrophilic QD and liposome hybrid was larger than empty liposome, and the vesicle size increased with increase number of encapsulated QD. In addition, Chu, Zhuo, Xu, Sheng, Hou and Wang <sup>[72]</sup> noted that hydrophilic QD and liposome hybrids in water retain their spherical structures, as well as the most of QDs remain trapped within the vesicles even after storing at 4°C for 609 days. This suggested that incorporation of QD to the core of liposome not only prevent the QDs being released from the vesicles but also reinforce the vesicle structure. Furthermore, Weng, Noble, Papahadjopoulos-Sternberg, Chen, Drummond, Kirpotin, Wang, Hom, Hann and Park <sup>[70]</sup> conjugated hydrophilic QD to the surface of liposome, which also resulted in adversely increase of both vesicle size and size distribution. Interestingly, for hydrophobic QD and liposome hybrids, Al-Jamal, Al-Jamal, Tian, Lacerda, Bornans, Frederik and Kostarelos <sup>[64]</sup> have shown that incorporating QDs did not significantly change the size and surface charge of hybrids. The mean L-QD diameter and surface charge were in the range of that empty liposomes. Subsequent studies carried out in this group have showed that both the size and the surface charge of

hydrophobic QD and liposome hybrid vesicles remain almost the same even after Dox loading, which is consistent with liposome control. This suggested that the process of hydrophilic drug loading does not affect the physicochemical properties of L-QD<sup>[62]</sup>. The same group also demonstrated that such L-QD had pronounced colloidal stability and no mean vesicle diameter increase at both 4°C and 25°C over three weeks<sup>[61]</sup>. However, Kethineedi, Crivat, Tarr and Rosenzweig<sup>[77]</sup> have found that incorporating hydrophobic QD slightly increase the size of L-QD from a mean diameter of 75 nm to 100 nm. Zhang, Wen, Al-Suwayeh, Yen and Fang<sup>[68]</sup> has also showed an increase of liposome size with QD and cisplatin loading. Such changes also happened on surface charge when incorporating cisplatin and QDs into the liposome. The loading of cisplatin or CdSe QDs significantly reduced the negative zeta potential, and dual loading further shielded the electric surface potential. Such differences may be resulted from different formulations and production procedures used among studies.

#### *Photostability of QDs in the hybrid system*

Photostability of QD after incorporating to the L-QD could be quantitatively studied by photoluminescence (PL) spectroscopy<sup>[61, 64, 67, 72, 78]</sup>. Interestingly, incorporation of QDs within the lipid bilayer led to enhanced photostability compared to that of bare QDs<sup>[64, 67]</sup>. Al-Jamal, Al-Jamal, Tian, Lacerda, Bornans, Frederik and Kostarelos<sup>[64]</sup> showed that the spectral characteristics of TOPO-capped CdSe/ZnS QD in obtained MLV and SUV were similar to those QDs in toluene. Only 30–40% reduction of the initial fluorescence intensity was observed when the QD were embedded within the MLV and SUV lipid bilayers, compared to more than 70% loss of fluorescence intensity in the case of QD in toluene suspensions. Furthermore, QD in toluene were photochemically unstable when exposed to UV light, witnessed as a sharp reduction in fluorescence intensity and a marked blue shift at 7 days, and a complete loss of fluorescence after 14 days of UV exposure. On the other hand, the hybrid exhibited improved photostability after both 7 and 14 days of UV exposure. Similar results were obtained by Ye, Wang, Li, Yu and Hu<sup>[67]</sup>. When incorporating CTAB-coated CdTe or CdHgTe QD into the lipid bilayer, fluorescence of QD was preserved and the photostability was enhanced. After 10 hours exposed to UV light, fluorescent intensity of the bare QDs reduced sharply, whereas, only 20–30% reduction was observed in the case of encapsulated QD. These results indicated that the L-QD improved the photostability of QD on storage and against UV light exposure, which can be attributed to the tight packing of the QD within the lipid bilayer. The increased photostability of liposome-encapsulated QDs

might be expected to improve their performance as fluorescence markers. In another report, photostability of CdSe QD encapsulated in lipid bilayers with different physical state was investigated<sup>[78]</sup>. Three phospholipids with different melting temperature (T<sub>m</sub>) were used. The PL of CdSe QDs changed in a phospholipid-dependent manner when stored under ambient conditions. Their results suggested that the T<sub>m</sub> of the lipid membrane controls optical and chemical properties of embedded QDs and QDs encapsulated within gel-phase lipid bilayer were the most stable. Furthermore, Tian, Al-Jamal, Al-Jamal and Kostarelos<sup>[61]</sup> and Zhang, Wen, Al-Suwayeh, Yen and Fang<sup>[68]</sup> demonstrated the addition of model drugs to the liposomes did not significantly alter the PL of QD which is quite important for such multifunctional drug delivery system.

### *Encapsulation efficiency*

Encapsulation efficiency (EE) determines the loading of QDs in liposomal hybrid formulations. EE is critical as it can be used to optimise the formulation composition as well as the manufacturing process. EE is calculated using the following formula:

$$\%EE = \frac{\text{Encapsulated QD concentration}}{\text{Initial QD concentration}} \times 100$$

The initial QD concentration is usually a known concentration determined in the formulation. There are two techniques have been used to determine EE namely fluorescence spectroscopy (FS) and Thermal lens microscopy (TLM). In the first technique, encapsulated QD concentration is estimated using photoluminescence. Percentage EE is then calculating using the formula. For example, Wang, Zhao, Wang, Yang and Chen<sup>[73]</sup> encapsulated water soluble thiol-capped CdTe QDs into liposome vesicles by agglomeration. The PL intensities of obtained L-QD in solutions were measured with a fluorescence spectrophotometer, and compared with that of the initial QDs solution. The PL in L-QD solution was totally come from the loaded QDs and no fluorescence self-quenching of the QDs inside the liposomes was observed, so that the comparison of PL intensities between the L-QD solution and the initial QD solution could estimate the loading content of QDs in L-QD. In this study, 95% EE was achieved with a lipid/QDs molar ratio of 0.15:1. The second technique, TLM, was developed by Batalla, Cabrera, San Martin-Martinez, Korte, Calderon and Marin<sup>[80]</sup> which is an useful tool for determining EE when quenching of the fluorescence happens after liposome encapsulation of the QDs. Self-quenching of QD is quite rare in hydrophobic loaded QD liposome hybrid, however it may happen when hydrophilic QD incorporating to the liposomes. The methodology of TLM consists in measuring the TL signal amplitude as a

function of known QDs concentrations in water, *i.e.* the calibration curve. Next, the unknown concentration of encapsulated QDs is determined using the calibration curve. Then, the encapsulation efficiency can be calculated as the ratio of the encapsulated QDs concentration versus the initial concentration of QDs mixed with the empty liposomes solution. It was found that the optimal EE for encapsulating carboxyl CdSe/ZnS inside aqueous compartment of the classic soy lecithin/cholesterol liposomes by thin film hydration method was 36% with an initial QDs concentration ranged between 1.25 and 10 nM. The authors suggested that TLM not only determine the encapsulation efficiency but also the optimum quantity of the initial concentration of QDs to be mixed with liposomes.

#### *In vitro and in vivo biodistribution of liposomal QD hybrid system*

*In vitro* and *in vivo* biodistribution of L-QD could be studied using flow cytometry and fluorescence microscopy. Flow cytometry studies help determine the average QD fluorescence intensity inside the cells, thus can be used to quantify cellular uptake of QD liposome hybrids. For example, cellular bonding and uptake of free QDs and various liposomal QD constructs were examined in MCF cells with HER2 overexpression 60 minutes post incubation at 37 °C<sup>[70]</sup>. Flow cytometry studies revealed that the uptake of anti-HER2 L-QDs in HER2-overexpressing MCF cells was significantly higher than in normal MCF cells. In contrast, free carboxyl QDs bound non-specifically to all tested cells. Furthermore, anti-HER2 L-QDs showed markedly greater uptake vs free QDs or vs non-targeted L-QDs in HER2-overexpressing MCF7 cells at matching concentrations. Therefore, reduced amounts of QD could be used with anti-HER2 L-QDs to reduce the potential cytotoxicity of QDs. The biodistribution of L-QDs was also visualised using confocal microscopy. Free carboxyl QDs were observed in close association with the cell surface, while anti-HER2 L-QDs accumulated mostly in the perinuclear region of the cells (Figure 3A&B). In addition, intravenous administration of L-QDs resulted fluorescence signals were readily detected at the tumour site 24 hours post-injection. Interestingly, tumour accumulation of both targeted and non-targeted L-QD showed similar results, which suggested that long circulating L-QDs localise in tumours predominantly via the EPR effect rather than via antibody-mediated targeting<sup>[70]</sup>. In another study, Kostarelos and co-workers studied intracellular trafficking of cationic and zwitterionic L-QD by confocal microscopy<sup>[64, 65, 74, 75]</sup>. The L-QD intracellular signal was found to be time and dose dependent for both L-QD types<sup>[64, 74]</sup>. It is evident from the microscopy images obtained that the cationic L-QD were uptake more efficiently by A549 cells, compared to weak negative or neutral surface charge L-QD<sup>[64, 74]</sup>. Cationic L-QDs

were bound to the cell membrane within 1h and internalised throughout the cell volume and close to the nucleus after 3h incubation presumably through endosomal uptake<sup>[74]</sup>. Similar results were demonstrated by Bothun, Rabideau and Stoner<sup>[66]</sup>, where fluorescence microscopy images showed that cationic L-QDs by HuH-7 was higher than Zwitterionic L-QD. These studies suggested that the positive charge is a key parameter responsible for increased cellular uptake of the liposomal QD hybrid system. *In vivo* transportation of L-QD were investigated using a three-dimensional multicellular tumour spheroids (MCS) cultures from melanoma cells (B16F10)<sup>[74]</sup>. Confocal microscopy images depict strong interaction between cationic L-QDs and the MCS. The L-QD localised 30–50-mm deep within the MCS mass. However, zwitterionic L-QD with a weak negative surface charge were able to diffuse deeper into the spheroids. *In vivo* uptake and retention of L-QD in Tumour Xenografts were also investigated<sup>[64, 65, 74, 75]</sup>. Intratumorally injection of cationic L-QD gave much stronger fluorescence signals than zwitterionic L-QD after both 5 minutes (Figure 3C&D) and 24 hours<sup>[64, 74]</sup>. It is suggested that zwitterionic vesicles around 100nm in diameter were leaking from the tumour immediately after intratumoral administration, while, cationic delivery vesicles of similar size significantly increased tumour retention<sup>[81]</sup>. This is agreed with the liposomal QD hybrid system, where cationic L-QD vesicles were uptaken by tumour cells and retained within tumour xenografts 24 hours post-injection, in contrast to zwitterionic L-QD vesicles that were drained out of the xenografts within 5 min following administration<sup>[64, 74]</sup>. *In vivo* behaviour of L-QD were further investigated following intravenous administration<sup>[65, 75]</sup>. Sharp differences were obtained between the tissue biodistribution of the various L-QD types dependent on their lipid composition. Cationic L-QD exhibited rapid clearance from blood circulation due to transient lung accumulation. After 24 hours post-administration, redistribution of the cationic L-QD led to their localisation mainly in the liver and spleen is thought to be due to adsorption of negatively charged plasma proteins on the surface of the cationic L-QDs hinders the nonspecific interaction with the pulmonary endothelium<sup>[65]</sup>. PEGylated L-QD exhibited improved blood circulation compared to cationic L-QD. The gel phase PEGylated L-QD accumulated in the solid tumour very rapidly and could be retained at the tumour site for at least 24 hours<sup>[75]</sup>. Overall, L-QD offers great potential for tumour imaging applications.



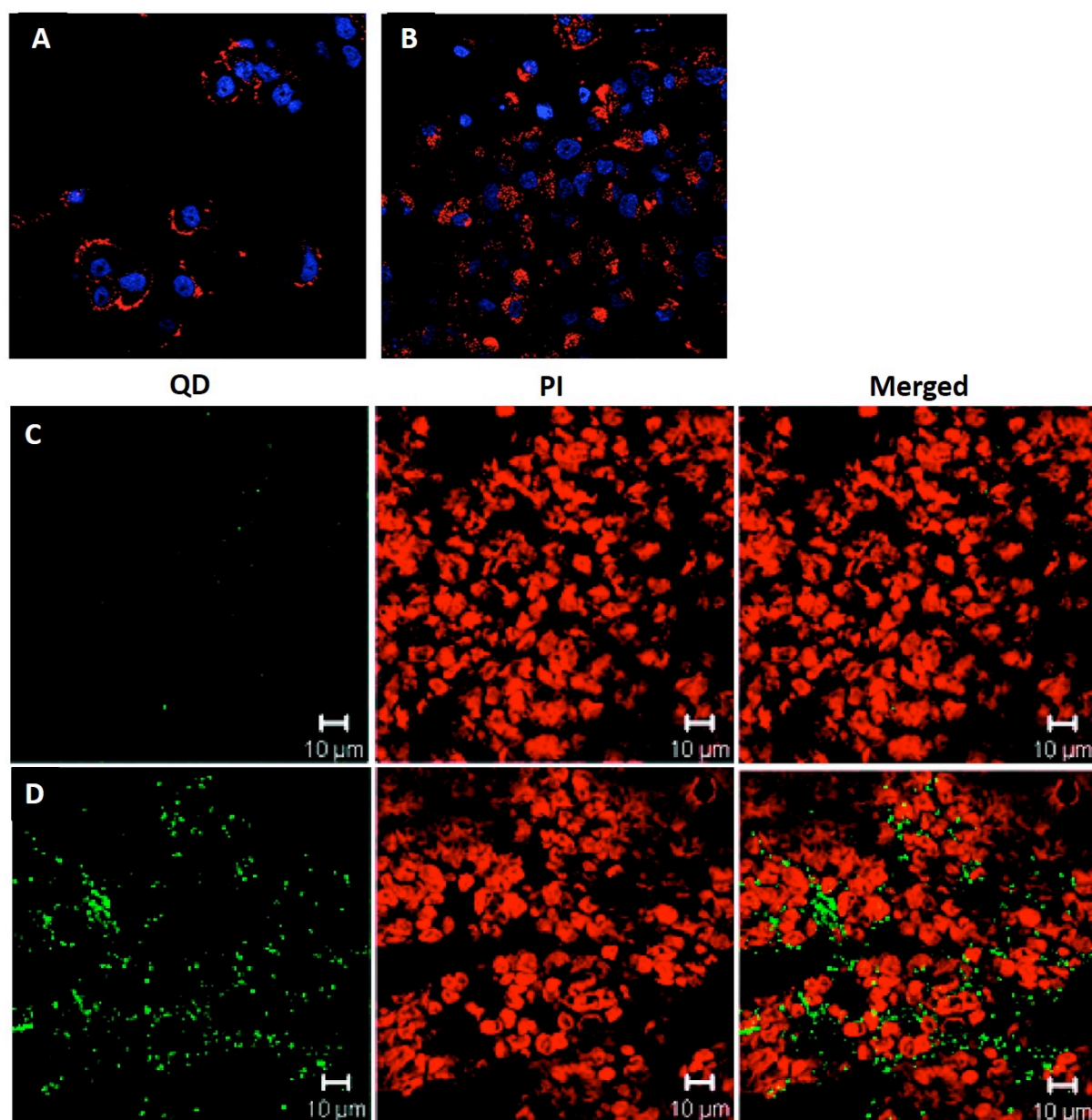


Figure 3 (A) SK-BR-3 cells were treated with free QDs, showing nonspecific association on the cell surface (B) SK-BR-3 cells were treated with QD-conjugated anti-HER2 liposomes (red fluorescence) for 60 min at 37 °C. QD-conjugated liposomes were internalised in SK-BR-3 cells. Cell nuclei were stained by DAPI (blue fluorescence). Reproduced with permission from ref. 70. Copyright 2008 American Chemical Society. Panels C-D represent in vivo tumour xenograft uptake and retention of L-QD hybrid vesicles. Confocal microscopy images of human cervical carcinoma (C33a) tumours dissected 5 min after intratumoral injection with (C) zwitterionic L-QD and (D) cationic L-QD. Left panels, L-QD fluorescence; middle panels, PI-stained nuclei; and right panels, the merged green

*and red channels. Scale bars are 10  $\mu$ m. Reproduced with permission from ref. 64. Copyright 2008 American Chemical Society.*

### ***In vivo degeneration of liposomal QD hybrid system***

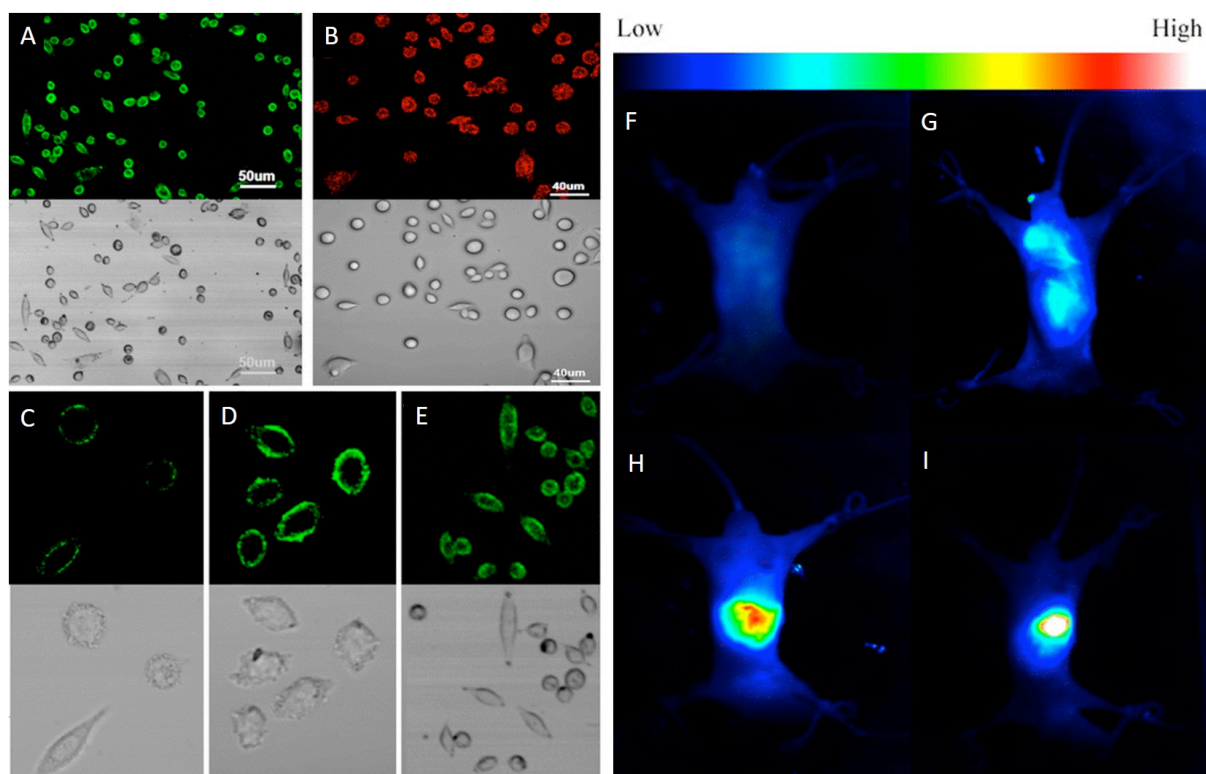
Though the *in vivo* biodistribution of L-QDs is relatively well investigated, there is still a lack of knowledge about the degeneration of such delivery system. Biomedical applications mainly focus on intravenous injection or oral application routes. Upon injection or ingestion, L-QDs come into contact with various complex physiological environments, leading to formation of a corona made of various biomolecules<sup>[82]</sup> which covering the L-QDs<sup>[83-86]</sup>. So for the degeneration process, L-QD should be considered as a system including corona, liposome envelope and QDs. So far the protein corona (PC) has mostly been studied, which can either has a stabilising effect or has a destabilising impact<sup>[84, 87]</sup>. The corona complexes are quite stable in plasma, however once they have been undertaken into the liver, corona and L-QD degradation could be triggered<sup>[88]</sup>. Then the elimination of liposome envelope takes place in different ways, such as metabolised by Kupffer cells, splenic macrophages or eliminated by the target tissues after their accumulation<sup>[10]</sup>. Eventually, innermost part of QDs may also be degraded, which will be highly dependent on the composition of the QD. For example, QDs made of CdSe are known to corrode, and thus release metal ions<sup>[89]</sup>, while Silica QDs can be, completely dissolved by hydrolysis<sup>[90]</sup>.

### **Application of liposomal QD hybrid system**

#### ***In vitro and in vivo bio-imaging***

QDs have been used as imaging agents to overcome many of the limitations of conventional contrast agents (i.e. organic dyes). The liposomal QD hybrid system provides further improvement of photostability and biocompatibility of QD, which could be used in bio-imaging applications *in vitro* and *in vivo*. Quite a few reports have appeared describing the use of such hybrid liposomal devices. For example, Voura, Jaiswal, Mattoussi and Simon<sup>[69]</sup> injected the tail veins of C57BL/6 mice with L-QD labelled B16F10 cells. The author demonstrated that QD labelling by L-QD has no detectable toxicity to the labelled cells or the host animal. The QD signal was observed in B16F10 cells seeded organs, which suggested that metastatic tumour cell extravasation could be tracked using L-QD. In addition, the use of a set of L-QDs with difference emission spectra in conjunction with multiphoton and emission-scanning microscopy provides the opportunity to simultaneously identify and study the interactions of different populations of tumour cells and normal tissue cells within the

same animal. In another study, Chu, Zhuo, Xu, Sheng, Hou and Wang <sup>[72]</sup> used three different size of L-QDs for sentinel lymph node (SLN) mapping. The encapsulated CdTe QD had near-infrared (NIR) emission band which was ideal for *in vivo* imaging. The NIR light offers great advantages for deep tissue imaging applications, because the NIR light could penetrate deeply in living tissues, as well as autofluorescence and absorbance from tissue-intrinsic chromophores reach their minima in this range. After intradermally injection of L-QDs, a bright red fluorescent spot appeared rapidly in the axillary location of a nude mouse. The fluorescent signal was retained for 24 hours post-injection. Ye, Wang, Li, Yu and Hu <sup>[67]</sup> also labelled MCF7 cells with CdTe or CdHgTe QDs encapsulated in liposomes (Figure 3A&B). It was found that L-QDs were efficiently internalised by MCF7 cells in a time-dependent manner (Figure 3C-E). Meanwhile, no cytotoxic effects were observed when cells exposed to the L-QD during the whole incubation period. All these findings suggest that L-QD increased biocompatibility and stability of QDs, thus improving the imaging effects for cancer cell labelling. Their *in vivo* imaging study using L-QD containing CdHgTe QDs also demonstrated the capability of L-QD for imaging in living animals (Figure 3F-I). In the imaging process, strong fluorescence signal throughout the entire body was observed immediately after L-QD injection. Within 20 minutes of injection, fluorescence signal increased strongly in liver and spleen and decreased sharply from the rest of the body, which suggested that L-QDs were taken up mainly in the liver and spleen due to RES effect. In addition, the authors showed that the injection of L-QD did not appear to induce significant toxicity in living animal. All these findings suggested that the L-QD is a promising tool for *in vitro* and *in vivo* bio-imaging study.



*Figure 4 Confocal laser scanning microscopy images of MCF-7 cells incubation with lipid-QD after different time, top panel-fluorescence channel, bottom panel-bright field, (A): lipid-CdTe 120 minutes, (B): lipid-CdHgTe 120 minutes, (C): lipid-CdTe 30 minutes, (D): lipid-CdTe 60 minutes, (E): lipid-CdTe 120 minutes. (F)to(I) represent NIR images of the denuded mouse after lipid-CdHgTe was injected via tail vein for 10 seconds, 5, 10 and 20 minutes, respectively. Reproduced with permission from ref. 67. Copyright 2013 Springer.*

### **Multifunctional (therapeutic and diagnostic) delivery devices**

One of the key advantages offered by the QD liposome hybrid system is the versatility of potential structural and surface characteristics by selection of different lipid components with minimal manipulation of the QD. Also, they offer the possibility to simultaneously encapsulate therapeutic agents for the construction of multifunctional (therapeutic and diagnostic) delivery devices (Table 1). The first QD liposome theranostic systems was developed by Kostarelos and co-authors, who loaded L-QD with Dox using the osmotic gradient technique<sup>[61]</sup>. The PL spectra of Dox-loaded L-QD showed that both Dox and QD could be simultaneously detected, indicating the coexistence of both QD and Dox in the hybrid vesicle population<sup>[62]</sup>. The process of Dox loading into hybrid vesicles is further investigated at various loading concentrations compared with liposome control. The highest loading efficiency in hybrid vesicles was 97% compared with liposome control (99%), which

indicated that QD incorporation into the lipid bilayer does not affect Dox loading through the lipid membrane of liposomes using the pH-gradient technique. It was also found that high Dox loading efficiency was achieved at a certain Dox concentration ( $>0.5$  mM) when Dox crystal-like structures were formed. Moreover, the following release study showed that fastest Dox release was observed from Egg phosphocholine (EPC)-QD vesicles in serum with 50% of Dox released over 6 hours compared with that from DSPC-QD vesicles ( $< 10\%$ ), which indicated that Dox release profile from L-QD could be modulated simply by changing lipid compositions. In another attempt, Weng *et. al.* <sup>[70]</sup> have covalently conjugated hydrophilic QD at the outer surface of HER2-targeted liposomes (monoclonal antibody fragments of HER2 conjugated to liposomes) and then loaded Dox into the aqueous core of these vesicles through ammonium sulfate gradient method. While drug loading into liposomes prior to QD conjugation was highly efficient ( $>90\%$ ), drug loading after QD conjugation was largely reduced with  $\sim 30\%$  loading efficiency. The obtained L-QDs were relatively stable with 15-30% Dox lost after cold storage in buffer solution for 2 months. The anticancer activity of the Dox-loaded hybrids was evaluated in HER2-overexpressing SK-BR-3 cells. The Dox-loaded hybrids showed potent cytotoxicity against SK-BR-3 cells ( $IC_{50} \sim 0.5$   $\mu\text{g/mL}$ ), which was comparable to Dox-loaded liposome without QD ( $IC_{50} \sim 0.7$   $\mu\text{g/mL}$ ). This result suggested that intracellular Dox delivery of the hybrid system was as efficient as the direct permeation of free Dox in vitro and also indicated that the HER-2 targeting effect was not compromised by QD conjugation. Furthermore, L-QD without dox showed minimal cytotoxicity, confirming that anticancer activity was due to Dox and not from the QD liposome carrier system. In addition, in vivo properties of L-QD delivery system were studied in nude mice. The L-QD exhibited moderately prolonged circulation time, which was greatly prolonged over that of free QDs. No weight loss or obvious signs of toxicity were observed on mice during study period (3 months). After 24 hours, following intravenous injection (IV), in nude mice bearing HER2-overexpressing MCF-7/HER2 xenografts, fluorescence signals were readily detected at the tumour site as well as in mononuclear phagocytic system (MPS) organs known to mediate liposome clearance (Figure 4A). Tumour fluorescence reached a plateau after 24 h, with up to  $18 \pm 5\%$  of total body fluorescence localising to the tumour region. Tissue sections of tumours collected 48 h post-injection showed that L-QD extensive accumulation within tumour tissue and intracellularly within tumour cells (Figure 4B). In another report, Zheng *et. al.* <sup>[68]</sup> have encapsulated both Cisplatin and CdSe/ZnS into liposomes using thin film hydration method. The loading of cisplatin is approximately 50%



with or without QDs. This suggested incorporation of QDs did not affect the drug entrapment efficiency, which is agreed with previous studies. *In vitro* cytotoxicity assay on melanoma cells demonstrated that the L-QD without the drug had no cytotoxic effect on melanoma cells, while cisplatin-loaded L-QD showed greater cytotoxicity (22% viability). *In vivo* bio-imaging study showed the possibility to track such theranostic systems by IVIS imaging system after intravenous injection. The QD activity was primarily found in the brain and skin, which was also confirmed by *ex vivo* imaging of organs (Figure 4C-E). The biodistribution of cisplatin also showed that the drug accumulation in brain and skin was significantly increased after loading into QD liposome vesicles as compared to that in the free control. Meanwhile, liver and spleen uptake of both QD and cisplatin was significantly reduced after encapsulation. These results suggested that entrapment of QDs and the drug into liposomes could protect them against degradation, reduce the RES uptake and enhance their delivery to the specific tissue site. All these studies demonstrated the potential of the QD liposome hybrid delivery system could be used as a platform for synchronous therapeutic and diagnostic modalities.

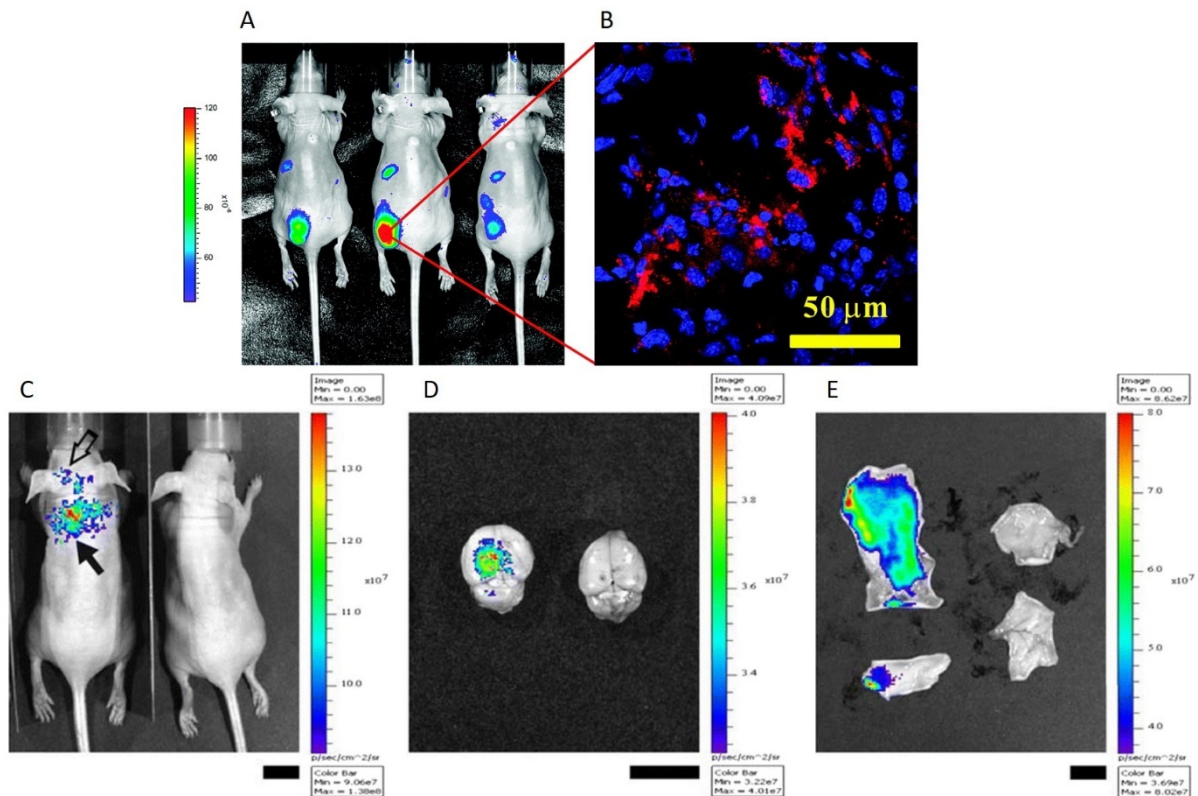


Figure 5 (A) *In vivo* fluorescence imaging of three nude mice bearing MCF-7/HER2 xenografts implanted in the lower back 30 hours after i.v. injection with anti-HER2 QD-ILs. Imaging showed that QD-ILs had localized prominently in tumours as well as in MPS organs.

*Units: efficiency (the fractional ratio of fluorescence emitted per incident photon). (B) A 5  $\mu$ m section cut from frozen tumour tissues harvested at 48 hours postinjection and examined by confocal microscopy. The tumour section was examined in two-colour scanning mode for nuclei stained by DAPI (blue) and QD-ILs (red). Reproduced with permission from ref. 70. Copyright 2008 American Chemical Society. (C-E) represent fluorescence imaging of representative nude mice detected with the IVIS system following an intravenous injection of CdSe/ZnS QD-loaded liposomes for 2 hours. The negative control (normal saline treatment) mice are represented by the animals placed on the right side of each image. (C) whole-body imaging in vivo (open arrow indicates the possible site of the brain; close arrow indicates the possible site of the skin), (D) ex vivo imaging of the brain, (E) ex vivo imaging of the skin. The scale bar is 1 cm. Reproduced with permission from ref. 68. Copyright 2012 Springer.*

## **Conclusion and Perspectives**

Liposomal QD hybrid systems have great potential for future clinical use since they are designed to integrate the well-established physicochemical and pharmacodynamic properties of liposomes with unique photochemical properties of QDs<sup>[11]</sup>. QDs can be loaded into the inner space of a liposome, insertion into the liposomal membrane, or attached onto the liposome surface. Careful attention should be paid to stability of such complex systems since incorporation of QDs and/or therapeutic agents in a liposome may cause payload leakage or a decrease in vivo stability of the liposomal nanostructure. Physical deformation of the liposomes could be eliminated by optimisation lipids and QD ratio in preparation formulation. In addition, high drug loading could be achieved by gradient method after liposome formation (with hydrophobic QD) or before hydrophilic QD attached onto liposome surface. For *in vitro* bio-imaging, the L-QD were shown to be efficiently uptaken by living cells in the absence of cell death. For *in vivo* applications, surface charge and lipid composition are important factors for blood circulation and tumour accumulation/retention of L-QDs. For example, cationic L-QD were retained much longer than zwitterionic L-QD when administrated intratumorally. Whereas, PEGylated gel phase L-QD showed prolonged blood circulation than cationic L-QD when administrated intravenously. Much more effort should be focused on the *in vivo* behaviour of such hybrid delivery systems. Further investigations are needed to further prolong the L-QD blood circulation time, increase their accumulation in tumours and reduce their uptake in liver and spleen. Overall, L-QD offers great potential for

tumour imaging applications and could be easily adapted to construct theranostic device which achieve both diagnostic and therapeutic functions.

**Disclosure:**

The authors report no conflicts of interest in this work.



## Reference:

- [1] Mandal B, Bhattacharjee H, Mittal N, et al. Core-shell-type lipid-polymer hybrid nanoparticles as a drug delivery platform[J]. *Nanomedicine-Nanotechnology Biology and Medicine* 2013,9(4):474-491.
- [2] Tan SW, Li X, Guo YJ, et al. Lipid-enveloped hybrid nanoparticles for drug delivery[J]. *Nanoscale* 2013,5(3):860-872.
- [3] Key J, Leary JF. Nanoparticles for multimodal in vivo imaging in nanomedicine[J]. *International Journal of Nanomedicine* 2014,9:711-726.
- [4] Jeong SH, Kim JH, Yi SM, et al. Assessment of penetration of quantum dots through in vitro and in vivo human skin using the human skin equivalent model and the tape stripping method[J]. *Biochemical and Biophysical Research Communications* 2010,394(3):612-615.
- [5] Schutz CA, Juillerat-Jeanneret L, Mueller H, et al. Therapeutic nanoparticles in clinics and under clinical evaluation[J]. *Nanomedicine* 2013,8(3):449-467.
- [6] Shao K, Singha S, Clemente-Casares X, et al. Nanoparticle-Based Immunotherapy for Cancer[J]. *Acs Nano* 2015,9(1):16-30.
- [7] Kim CS, Duncan B, Czeran B, et al. Triggered nanoparticles as therapeutics[J]. *Nano Today* 2013,8(4):439-447.
- [8] Zrazhevskiy P, Sena M, Gao XH. Designing multifunctional quantum dots for bioimaging, detection, and drug delivery[J]. *Chemical Society Reviews* 2010,39(11):4326-4354.
- [9] Medintz IL, Uyeda HT, Goldman ER, et al. Quantum dot bioconjugates for imaging, labelling and sensing[J]. *Nature Materials* 2005,4(6):435-446.
- [10] Bozzuto G, Molinari A. Liposomes as nanomedical devices[J]. *International Journal of Nanomedicine* 2015,10:975-999.
- [11] Sailor MJ, Park JH. Hybrid Nanoparticles for Detection and Treatment of Cancer[J]. *Advanced Materials* 2012,24(28):3779-3802.
- [12] Bangham AD, Horne RW. Negative staining of phospholipids+their structural modification by-surface active agents as observed in electron microscope[J]. *Journal of Molecular Biology* 1964,8(5):660-&.
- [13] Bangham AD, Hill MW, Miller NGA. Preparation and Use of Liposomes as Models of Biological Membranes. In: Korn E, editor. *Methods in Membrane Biology Springer US*, 1974:1-68.
- [14] Papahadjopoulos D, Kimelberg HK. Phospholipid vesicles (liposomes) as models for biological membranes: Their properties and interactions with cholesterol and proteins[J]. *Progress in Surface Science* 1974,4:141-232.
- [15] Allen TM, Hansen CB, Demenezes DEL. Pharmacokinetics of long-circulating liposomes[J]. *Advanced Drug Delivery Reviews (in English)* 1995,16(2-3):267-284.
- [16] Milla P, Dosio F, Cattel L. PEGylation of Proteins and Liposomes: a Powerful and Flexible Strategy to Improve the Drug Delivery[J]. *Current Drug Metabolism* 2012,13(1):105-119.
- [17] Harris JM, Martin NE, Modi M. Pegylation - A novel process for modifying pharmacokinetics[J]. *Clinical Pharmacokinetics* 2001,40(7):539-551.
- [18] Howard MD, Jay M, Dziublal TD, et al. PEGylation of nanocarrier drug delivery systems: State of the art[J]. *Journal of Biomedical Nanotechnology* 2008,4(2):133-148.
- [19] Kirpotin D, Park JW, Hong K, et al. Sterically stabilized Anti-HER2 immunoliposomes: Design and targeting to human breast cancer cells in vitro[J]. *Biochemistry* 1997,36(1):66-75.
- [20] Mamot C, Drummond DC, Greiser U, et al. Epidermal growth factor receptor (EGFR)-targeted immunoliposomes mediate specific and efficient drug delivery to EGFR- and EGFRvIII-overexpressing tumor cells[J]. *Cancer Research* 2003,63(12):3154-3161.

- [21] Hansen CB, Kao GY, Moase EH, et al. ATTACHMENT OF ANTIBODIES TO STERICALLY STABILIZED LIPOSOMES - EVALUATION, COMPARISON AND OPTIMIZATION OF COUPLING PROCEDURES[J]. *Biochimica Et Biophysica Acta-Biomembranes* 1995,1239(2):133-144.
- [22] Nagayasu A, Uchiyama K, Kiwada H. The size of liposomes: a factor which affects their targeting efficiency to tumors and therapeutic activity of liposomal antitumor drugs[J]. *Advanced Drug Delivery Reviews* 1999,40(1-2):75-87.
- [23] Akbarzadeh A, Rezaei-Sadabady R, Davaran S, et al. Liposome: classification, preparation, and applications[J]. *Nanoscale Research Letters* 2013,8(1):102.
- [24] Fan Y, Zhang Q. Development of liposomal formulations: From concept to clinical investigations[J]. *Asian Journal of Pharmaceutical Sciences* 2013,8(2):81-87.
- [25] Laouini A, Jaafar-Maalej C, Limayem-Blouza I, et al. Preparation, Characterization and Applications of Liposomes: State of the Art[J]. *Journal of Colloid Science and Biotechnology* 2012,1(2):147-168.
- [26] Batzri S, Korn ED. Single bilayer liposomes prepared without sonication[J]. *Biochimica et Biophysica Acta (BBA) - Biomembranes* 1973,298(4):1015-1019.
- [27] Deamer D, Bangham AD. Large volume liposomes by an ether vaporization method[J]. *Biochimica Et Biophysica Acta* 1976,443(3):629-634.
- [28] Szoka F, Papahadjopoulos D. Procedure for preparation of liposomes with large internal aqueous space and high capture by reverse-phase evaporation[J]. *Proceedings of the National Academy of Sciences of the United States of America* 1978,75(9):4194-4198.
- [29] Alpes H, Allmann K, Plattner H, et al. Formation of large unilamellar vesicles using alkyl maltoside detergents[J]. *Biochimica et Biophysica Acta (BBA) - Biomembranes* 1986,862(2):294-302.
- [30] Skalko-Basnet N, Pavelic Z, Becirevic-Lacan M. Liposomes Containing Drug and Cyclodextrin Prepared by the One-Step Spray-Drying Method[J]. *Drug Development and Industrial Pharmacy* 2000,26(12):1279-1284.
- [31] Li CL, Deng YJ. A novel method for the preparation of liposomes: Freeze drying of monophasic solutions[J]. *Journal of Pharmaceutical Sciences* 2004,93(6):1403-1414.
- [32] Jahn A, Vreeland WN, Gaitan M, et al. Controlled Vesicle Self-Assembly in Microfluidic Channels with Hydrodynamic Focusing[J]. *Journal of the American Chemical Society* 2004,126(9):2674-2675.
- [33] Gubernator J. Active methods of drug loading into liposomes: recent strategies for stable drug entrapment and increased in vivo activity[J]. *Expert Opinion on Drug Delivery* 2011,8(5):565-580.
- [34] Fritze A, Hens F, Kimpfler A, et al. Remote loading of doxorubicin into liposomes driven by a transmembrane phosphate gradient[J]. *Biochimica et Biophysica Acta (BBA) - Biomembranes* 2006,1758(10):1633-1640.
- [35] Allen TM, Cullis PR. Liposomal drug delivery systems: From concept to clinical applications[J]. *Advanced Drug Delivery Reviews* 2013,65(1):36-48.
- [36] Zamboni WC. Concept and clinical evaluation of carrier-mediated anticancer agents[J]. *Oncologist* 2008,13(3):248-260.
- [37] Bladé J, Sonneveld P, Miguel JFS, et al. Efficacy and Safety of Pegylated Liposomal Doxorubicin in Combination With Bortezomib for Multiple Myeloma: Effects of Adverse Prognostic Factors on Outcome[J]. *Clinical Lymphoma Myeloma and Leukemia* 2011,11(1):44-49.
- [38] Campos SM, Matulonis UA, Penson RT, et al. Phase II study of liposomal doxorubicin and weekly paclitaxel for recurrent Mullerian tumors[J]. *Gynecologic Oncology* 2003,90(3):610-618.

- [39] Hann IM, Prentice HG. Lipid-based amphotericin B: A review of the last 10 years of use[J]. *International Journal of Antimicrobial Agents* 2001,17(3):161-169.
- [40] Chang H-I, Yeh M-K. Clinical development of liposome-based drugs: formulation, characterization, and therapeutic efficacy[J]. *International Journal of Nanomedicine* 2012,7:49-60.
- [41] Fasol U, Frost A, Büchert M, et al. Vascular and pharmacokinetic effects of EndoTAG-1 in patients with advanced cancer and liver metastasis[J]. *Annals of Oncology* 2012,23(4):1030-1036.
- [42] Poon RTP, Borys N. Lyso-thermosensitive liposomal doxorubicin: an adjuvant to increase the cure rate of radiofrequency ablation in liver cancer[J]. *Future Oncology* 2011,7(8):937-945.
- [43] McDonagh CF, Huhlov A, Harms BD, et al. Antitumor Activity of a Novel Bispecific Antibody That Targets the ErbB2/ErbB3 Oncogenic Unit and Inhibits Heregulin-Induced Activation of ErbB3[J]. *Molecular Cancer Therapeutics* 2012,11(3):582-593.
- [44] Bangal M, Ashtaputre S, Marathe S, et al. Semiconductor Nanoparticles[J]. *Hyperfine Interactions* 2005,160(1):81-94.
- [45] Alivisatos AP. Perspectives on the physical chemistry of semiconductor nanocrystals[J]. *Journal of Physical Chemistry* 1996,100(31):13226-13239.
- [46] Weller H. Colloidal Semiconductor Q-Particles: Chemistry in the Transition Region Between Solid State and Molecules[J]. *Angewandte Chemie International Edition in English* 1993,32(1):41-53.
- [47] Weller H. Quantized Semiconductor Particles: A novel state of matter for materials science[J]. *Advanced Materials* 1993,5(2):88-95.
- [48] Talapin DV, Lee J-S, Kovalenko MV, et al. Prospects of Colloidal Nanocrystals for Electronic and Optoelectronic Applications[J]. *Chemical Reviews* 2010,110(1):389-458.
- [49] Chan WCW, Maxwell DJ, Gao XH, et al. Luminescent quantum dots for multiplexed biological detection and imaging[J]. *Current Opinion in Biotechnology* 2002,13(1):40-46.
- [50] Pickett NL, O'Brien P. Syntheses of semiconductor nanoparticles using single-molecular precursors[J]. *The Chemical Record* 2001,1(6):467-479.
- [51] Chan WCW, Maxwell DJ, Gao X, et al. Luminescent quantum dots for multiplexed biological detection and imaging[J]. *Current Opinion in Biotechnology* 2002,13(1):40-46.
- [52] Michalet X, Pinaud FF, Bentolila LA, et al. Quantum dots for live cells, in vivo imaging, and diagnostics[J]. *Science* 2005,307(5709):538-544.
- [53] Resch-Genger U, Grabolle M, Cavaliere-Jaricot S, et al. Quantum dots versus organic dyes as fluorescent labels[J]. *Nature Methods* 2008,5(9):763-775.
- [54] Chen C-S, Yao J, Durst R. Liposome encapsulation of fluorescent nanoparticles: Quantum dots and silica nanoparticles[J]. *Journal of Nanoparticle Research (in English)* 2006,8(6):1033-1038.
- [55] Oberdorster G, Oberdorster E, Oberdorster J. Nanotoxicology: An emerging discipline evolving from studies of ultrafine particles[J]. *Environmental Health Perspectives* 2005,113(7):823-839.
- [56] Lewinski N, Colvin V, Drezek R. Cytotoxicity of nanoparticles[J]. *Small* 2008,4(1):26-49.
- [57] Karakoti AS, Shukla R, Shanker R, et al. Surface functionalization of quantum dots for biological applications[J]. *Advances in Colloid and Interface Science* 2015,215:28-45.
- [58] Wegner KD, Hildebrandt N. Quantum dots: bright and versatile in vitro and in vivo fluorescence imaging biosensors[J]. *Chemical Society Reviews* 2015,44(14):4792-4834.
- [59] Clarke SJ, Hollmann CA, Zhang Z, et al. Photophysics of dopamine-modified quantum dots and effects on biological systems[J]. *Nat Mater* 2006,5(5):409-417.

- [60] Smith AM, Duan H, Rhyner MN, et al. A systematic examination of surface coatings on the optical and chemical properties of semiconductor quantum dots[J]. *Physical Chemistry Chemical Physics* 2006,8(33):3895-3903.
- [61] Tian B, Al-Jamal WT, Al-Jamal KT, et al. Doxorubicin-loaded lipid-quantum dot hybrids: Surface topography and release properties[J]. *International Journal of Pharmaceutics* 2011,416(2):443-447.
- [62] Tian BW, Al-Jamal WT, Kostarelos K. The engineering of doxorubicin-loaded liposome-quantum dot hybrids for cancer theranostics[J]. *Chinese Physics B* 2014,23(8).
- [63] Gopalakrishnan G, Danelon C, Izewska P, et al. Multifunctional lipid/quantum dot hybrid nanocontainers for controlled targeting of live cells[J]. *Angewandte Chemie-International Edition* 2006,45(33):5478-5483.
- [64] Al-Jamal WT, Al-Jamal KT, Tian B, et al. Lipid-quantum dot bilayer vesicles enhance tumor cell uptake and retention in vitro and in vivo[J]. *Acs Nano* 2008,2(3):408-418.
- [65] Al-Jamal WT, Al-Jamal KT, Cakebread A, et al. Blood Circulation and Tissue Biodistribution of Lipid-Quantum Dot (L-QD) Hybrid Vesicles Intravenously Administered in Mice[J]. *Bioconjugate Chemistry* 2009,20(9):1696-1702.
- [66] Bothun GD, Rabideau AE, Stoner MA. Hepatoma Cell Uptake of Cationic Multifluorescent Quantum Dot Liposomes[J]. *Journal of Physical Chemistry B* 2009,113(22):7725-7728.
- [67] Ye C, Wang YQ, Li CG, et al. Preparation of liposomes loaded with quantum dots, fluorescence resonance energy transfer studies, and near-infrared in-vivo imaging of mouse tissue[J]. *Microchimica Acta* 2013,180(1-2):117-125.
- [68] Zhang L-W, Wen C-J, Al-Suwayeh SA, et al. Cisplatin and quantum dots encapsulated in liposomes as multifunctional nanocarriers for theranostic use in brain and skin[J]. *Journal of Nanoparticle Research* 2012,14(7).
- [69] Voura EB, Jaiswal JK, Mattoussi H, et al. Tracking metastatic tumor cell extravasation with quantum dot nanocrystals and fluorescence emission-scanning microscopy[J]. *Nat Med* 2004,10(9):993-998.
- [70] Weng KC, Noble CO, Papahadjopoulos-Sternberg B, et al. Targeted tumor cell internalization and imaging of multifunctional quantum dot-conjugated immunoliposomes in vitro and in vivo[J]. *Nano Letters* 2008,8(9):2851-2857.
- [71] Sigot V, Arndt-Jovin DJ, Jovin TM. Targeted Cellular Delivery of Quantum Dots Loaded on and in Biotinylated Liposomes[J]. *Bioconjugate Chemistry* 2010,21(8):1465-1472.
- [72] Chu M, Zhuo S, Xu J, et al. Liposome-coated quantum dots targeting the sentinel lymph node[J]. *Journal of Nanoparticle Research* 2010,12(1):187-197.
- [73] Wang J-Y, Zhao J-F, Wang P-N, et al. Liposome Encapsulation of Thiol-Capped CdTe Quantum Dots for Enhancing the Intracellular Delivery[J]. *Journal of Fluorescence* 2011,21(4):1635-1642.
- [74] Al-Jamal WT, Al-Jamal KT, Bomans PH, et al. Functionalized-quantum-dot-liposome hybrids as multimodal nanoparticles for cancer[J]. *Small* 2008,4(9):1406-1415.
- [75] Al-Jamal WT, Al-Jamal KT, Tian B, et al. Tumor targeting of functionalized quantum dot-liposome hybrids by intravenous administration[J]. *Molecular pharmaceutics* 2009,6(2):520-30.
- [76] Wi HS, Kim SJ, Lee K, et al. Incorporation of quantum dots into the lipid bilayer of giant unilamellar vesicles and its stability[J]. *Colloids and Surfaces B-Biointerfaces* 2012,97:37-42.
- [77] Kethineedi VR, Crivat G, Tarr MA, et al. Quantum dot-NBD-liposome luminescent probes for monitoring phospholipase A(2) activity[J]. *Analytical and Bioanalytical Chemistry* 2013,405(30):9729-9737.

- [78] Zheng WW, Liu Y, West A, et al. Quantum Dots Encapsulated within Phospholipid Membranes: Phase-Dependent Structure, Photostability, and Site-Selective Functionalization[J]. *Journal of the American Chemical Society* 2014,136(5):1992-1999.
- [79] Hansen MB, van Emmerik C, van Gaal E, et al. Quick-and-easy preparation and purification of quantum dot-loaded liposomes[J]. *Journal of Nanoparticle Research* 2013,15(1).
- [80] Batalla J, Cabrera H, San Martin-Martinez E, et al. Encapsulation efficiency of CdSe/ZnS quantum dots by liposomes determined by thermal lens microscopy[J]. *Biomedical Optics Express* 2015,6(10):3898-3906.
- [81] Nomura T, Koreeda N, Yamashita F, et al. Effect of Particle Size and Charge on the Disposition of Lipid Carriers After Intratumoral Injection into Tissue-isolated Tumors[J]. *Pharmaceutical Research (in English)* 1998,15(1):128-132.
- [82] Cedervall T, Lynch I, Lindman S, et al. Understanding the nanoparticle–protein corona using methods to quantify exchange rates and affinities of proteins for nanoparticles[J]. *Proceedings of the National Academy of Sciences* 2007,104(7):2050-2055.
- [83] Nel AE, Madler L, Velegol D, et al. Understanding biophysicochemical interactions at the nano-bio interface[J]. *Nat Mater* 2009,8(7):543-557.
- [84] Monopoli MP, Aberg C, Salvati A, et al. Biomolecular coronas provide the biological identity of nanosized materials[J]. *Nat Nano* 2012,7(12):779-786.
- [85] Tenzer S, Docter D, Kuharev J, et al. Rapid formation of plasma protein corona critically affects nanoparticle pathophysiology[J]. *Nat Nano* 2013,8(10):772-781.
- [86] Xia X-R, Monteiro-Riviere NA, Riviere JE. An index for characterization of nanomaterials in biological systems[J]. *Nat Nano* 2010,5(9):671-675.
- [87] Bargheer D, Nielsen J, Gébel G, et al. The fate of a designed protein corona on nanoparticles in vitro and in vivo[J]. *Beilstein Journal of Nanotechnology* 2015,6:36-46.
- [88] Feliu N, Docter D, Heine M, et al. In vivo degeneration and the fate of inorganic nanoparticles[J]. *Chemical Society Reviews* 2016,45(9):2440-2457.
- [89] Soenen SJ, Parak WJ, Rejman J, et al. (Intra)Cellular Stability of Inorganic Nanoparticles: Effects on Cytotoxicity, Particle Functionality, and Biomedical Applications[J]. *Chemical Reviews* 2015,115(5):2109-2135.
- [90] Mahon E, Hristov DR, Dawson KA. Stabilising fluorescent silica nanoparticles against dissolution effects for biological studies[J]. *Chemical Communications* 2012,48(64):7970-7972.