Impact of a single oral dose of 100,000 IU vitamin D3 on profiles of serum 25(OH)D3 and its metabolites 24,25(OH)₂D3, 3-epi-25(OH)D3, and 1,25(OH)₂D3 in adults with vitamin D insufficiency

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Abstract

Background: We investigate the effect of a high dose of vitamin D3 on circulating concentrations of 25(OH)D3 and its metabolites $24,25(OH)_2D3$, 3-epi-25(OH)D3, and $1,25(OH)_2D3$ in healthy individuals with self-perceived fatigue and vitamin D insufficiency (25(OH)D3 < 50 nmol/L).

Methods: 107 study participants (age 20–50 years) were randomized to receive a single 100,000 IU dose of vitamin D3 (n= 52) or placebo (n= 55). Vitamin D metabolite concentrations in serum were measured before, and 4 weeks after, supplementation.

Results: Overall, 52% of participants receiving vitamin D3 attained a serum 25(OH)D3 level >75 nmol/L. Among individuals who received vitamin D3, there were significant increases in serum concentrations of 25(OH)D3 and its metabolites 24,25(OH)₂D3, 3-epi-25(OH)D3, and 1,25(OH)₂D3 at 4 weeks; however, inter-individual variability in these changes was substantial. Positive correlations between serum 25(OH)D3 and 24,25(OH)₂D3 and 3-epi-25(OH)D3, and a significant negative correlation between serum 1,25(OH)₂D3 and 3-epi-25(OH)D3, were found 4 weeks after supplementation. The 24,25(OH)₂D3/25(OH)D3 and 24,25(OH)D3 and 25(OH)D3 and 25(OH)D3

Conclusions: Administration of a single high dose of vitamin D3 leads to a significant increase in concentrations of 25(OH)D3, 24,25(OH)₂D3, 3-epi-25(OH)D3 and 1,25(OH)₂D3; induction of the catabolic pathway predominates over the production of 1,25(OH)₂D3. Due to the high inter-individual variation in the 25(OH)D3 response to supplementation, any given dose of vitamin D is unlikely to achieve optimal vitamin D status in all treated individuals

Keywords

Vitamin D, 25(OH)D3, 24,25(OH)2D3, 3-epi-25(OH)D3, 1,25(OH)2D3, Supplementation

Introduction

Vitamin D plays a key role in the regulation of calcium and phosphate homeostasis, and deficiency of this vitamin is associated with secondary hyperparathyroidism, an increase in bone turnover and bone loss an increased risk of several diseases, including osteoporosis, cardiovascular disease, diabetes and cancer [1]. Vitamin D synthesized in the skin (vitamin D3 [cholecalciferol]) or orally ingested (either vitamin D3 or vitamin D2 [ergocalciferol]) is metabolized in the liver by the enzyme 25-hydroxylase (CYP2R1) to form 25-hydroxy vitamin D3 (25(OH)D3), which is then further metabolized primarily in the kidney by 1α -hydroxylase (CYP27B1) to form the active vitamin D metabolite, 1,25-dihydroxy vitamin D3 (1,25(OH)₂D3). Both 25(OH)D3 and 1,25(OH)₂D3 undergo further metabolism, predominantly by renal 24-hydroxylase (CYP24A1), to generate 24,25-dihydroxy vitamin D3 (24,25(OH)₂ D3) and 1,24,25-trihydroxyvitamin D3 (1α,24,25(OH)₃D3), respectively [2–4]. Mutations in the CYP24A1 gene are associated with partial or total loss of 24-hydroxylase activity, which in turn leads to hypercalcaemic conditions [5-7]. The production of 24,25(OH)₂D3 has been shown to be 25(OH)D3-dependent, and is moderately affected by vitamin D supplementation [5,6]; the physiological role of this metabolite remains to be established, although it is known to be involved in embryogenesis, cartilage development and fracture repair [8–10].

Measurement of total 25(OH)D (comprising both 25(OH)D3 and 25(OH)D2) in serum is widely accepted as a marker of vitamin D status; however, the optimum threshold concentration of 25(OH)D continues to be debated. The Institute of Medicine (IOM) recommends a threshold of 50 nmol/L for bone health [11], whereas the Endocrine Society recommend a threshold of 75 nmol/L for optimal reductions in fall or fracture risk [1]. with concentrations below 50 nmol/L being regarded as indicative of vitamin D insufficiency [11]. However, tThe 25(OH)D3 response to vitamin D supplementation varies markedly between individuals, and a significant proportion of patients may have persistent suboptimal levels despite supplementation [12–17]. Furthermore, the relationship between circulating 25(OH)D3 concentrations and clinical outcomes such as osteoporosis and fracture risk may differ between racial groups, raising the question of whether 25(OH)D3 provides a reliable estimate of vitamin D status in all populations [18,19]. For these reasons, increasing attention is being paid to the measurement of 24,25(OH)2D3 (the major circulating catabolite of vitamin D), and the ratio of 24,25(OH)2D3 to 25(OH)D3, as potential markers of vitamin D catabolism and predictors of the serum 25(OH)D response to vitamin D supplementation [5,6,12,18,20].

Measurement of vitamin D metabolites as biomarkers of vitamin D status has been further complicated in recent years by the identification of C3 epimeric forms of 25(OH)D3 and 1,25(OH)₂D3 [21]. These epimers were originally identified in infants and neonates, in whom they account for approximately 21% of total 25(OH)D3 concentrations [21], but were subsequently shown to be present in lower concentrations in adults, in whom they account for approximately 6% of total 25(OH)D3 [21–23]. The 3-epi-25(OH)D3 metabolite is produced endogenously, and circulating concentrations increase following vitamin D supplementation [22]; however, the physiological significance of these epimers remains to be established [20,21].

In view of the continuing uncertainty surrounding the clinical utility of different vitamin D metabolites as markers of vitamin D status, and to better understand the vitamin D metabolism pathway in response to supplementation, the present study was performed to investigate the effect of a single high dose (100,000 IU) of vitamin D3 on profiles of circulating 25(OH)D3 and its metabolites 24,25(OH)₂D3, 3-epi25(OH) D3, and 1,25(OH)₂D3 in healthy individuals with self-perceived fatigue and vitamin D insufficiency (25(OH)D3 <50 nmol/L), and to assess the inter-individual variability in the response to vitamin D supplementation. A further objective was to investigate the hypothesis that the baseline 24,25(OH)₂D3/25(OH)D3 ratio is a predictor of the response to supplementation.

Material and Methods

Clinical samples

Frozen serum samples (n=214) were obtained from a prospective randomized, double-blind, placebo-controlled clinical trial conducted at the University Hospital of Zurich, Switzerland (latitude 47° 22' N) (ClinicalTrials.gov Registry number NCT02022475). The trial was conducted in accordance with the declaration of Helsinki and Good Clinical Practice guidelines; the study protocol and its amendment were approved by the Zurich Cantonal Ethical Committee and Swissmedic, and informed consent was obtained from all participants prior to enrolment. The primary aim of the trial was to determine the effects of a single high dose of vitamin D3, compared with placebo, on serum 25(OH)D3 concentrations and clinical outcomes

such as fatigue at 4 weeks after treatment. Full details of this trial has been described elsewhere [23].

The trial involved 107 participants (age 20–50 years, body mass index [BMI] 18–25 kg/m²) who had serum 25(OH)D3 concentrations below 50 nmol/L. The 50 nmol/L threshold for vitamin D insufficiency was used in accordance with the recommendation of the Institute of Medicine (IOM) [11]. Participants were randomized to receive either a single 100,000 IU dose of vitamin D3 (n= 52) or placebo (n= 55).

Blood samples were obtained at a screening visit immediately before treatment and at a second visit 4 weeks after supplementation. Serum was separated by centrifugation at 2000*g* for 10 minutes, and aliquots were stored at -80°C prior to analysis. Serum concentrations of 25(OH)D3, 3-epi 25(OH)D3, 24,25(OH)₂D3 and 25-hydroxy vitamin D2 (25(OH)D2) were measured by a validated NIST traceable LC-MS/MS assay using a Micromass Quattro Ultima Pt mass spectrometer (Waters Corp., Milford, MA, USA) at Bioanalytical Facility, University of East Anglia, Norwich, UK; details of the assay are provided in the online supplementary material. For all analytes, the assay showed good linearity ($r^2 \ge 0.98$) and low intra-assay and inter-assay variability (see supplementary table S1).

Measurements of total 1,25 (OH)₂D3 were performed using a commercial immunoextraction enzyme immunoassay kit (IDS, Bolden, UK). The inter- and intra assay imprecision, as expressed by the coefficient of variation (CV), was less than 12.5%. Serum concentrations of calcium, phosphate, parathyroid hormone (PTH), C-reactive protein (CRP), and creatinine were measured using a Cobas 8000 analyzer (Roche Diagnostics, Mannheim, Germany) at the Institute of Clinical Chemistry, University Hospital of Zurich. All analyses were carried out according to the manufacturer's instructions. For all analytes, intra-assay and inter-assay variability, as expressed by the coefficient of variation (CV), were $\leq 1.7\%$ and 3.1%, respectively.

Statistical analyses

Demographic data and serum concentrations of vitamin D metabolites at baseline and followup were summarized using descriptive statistics (means, SDs, medians and interquartile ranges). Differences between baseline and post-supplementation values were analysed by means of paired t tests for vitamin D metabolites, unpaired t tests for normally distributed demographic variables, Mann-Whitney rank tests for non-normally distributed variables, and χ^2 tests for categorical variables. All comparisons were two-sided. Associations between vitamin D3 metabolites, and other clinical variables (age, BMI, serum calcium, serum phosphate, and serum PTH), at baseline and at 4 weeks after supplementation were investigated using Spearman rank correlation analysis.

Simple and multiple regression analyses were used to build prediction models for the 25(OH)D3 response to vitamin D3 supplementation. Four different models were used: model 1 included only baseline 25(OH)D3 concentrations as covariate; model 2 included baseline 25(OH)D3, $24,25(OH)_2D3$ and 3-epi-25(OH)D3 concentrations as covariates; model 3 included the same covariates as model 2 in addition to age, gender and body mass index (BMI), while model 4 included baseline $1,25(OH)_2D3$ concentrations in addition to the same covariates as model 2. All analyses were performed using IBM SPSS Statistics 22 software (SPSS Inc., Chicago, IL), and *P* values below 0.05 were considered significant.

Results

Baseline demographic and clinical characteristics of study participants are summarized in Table 1. No statistically significant differences between the vitamin D supplemented and placebo groups were observed. At baseline, 3-epi-25(OH)D3 was present in 88% of study participants, at a mean concentration equivalent to 3.9% of serum 25(OH)D3 concentrations. (Table 1).

Changes in vitamin D metabolites following vitamin D supplementation

Serum concentrations of vitamin D metabolites at baseline are summarized in Table 1, and changes in these concentrations 4 weeks after a single oral dose of 100,000 IU vitamin D3 are presented in Fig. 1. At 4 weeks, participants receiving vitamin D3 showed significant absolute increases in serum 25(OH)D3, $24,25(OH)_2D3$, 3-epi-25(OH)D3 and $1,25(OH)_2D3$ concentrations (all *P*<0.001 versus baseline), whereas no such changes were seen in placebotreated participants.

Interestingly, the ratios of $24,25(OH)_2D3$ to 25(OH)D3 and 24,25(OH)2D3 to 1,25(OH)3D3 were significantly increased, compared with baseline, in study participants receiving vitamin

D3 supplementation (Fig. 1). The mean 24,25(OH)D3/25(OH)D3 ratio at baseline was 0.076 \pm 0.02, and this had increased to 0.086 \pm 0.02 (*P*=0.006) at 4 weeks after supplementation. Similarly, the ratio of 24,25(OH)₂D3 to 1,25(OH)₂D3 increased 2.4-fold after vitamin D3 supplementation, from 0.023 \pm 0.01 at baseline to 0.056 \pm 0.025 (*P*<0.0001) at 4 weeks. In participants receiving placebo, both ratios remained unchanged following supplementation (*P*=0. 36 and *P*=0.92, respectively, versus baseline), as shown in Fig. 1(E and F).

At 4 weeks after dosing, all participants in the vitamin D3 group had attained a serum 25(OH)D3 concentration ≥ 50 nmol/L, except for one patient in whom the 25(OH)D3 concentration increased from a baseline value of 17.5 nmol/L to 35.6 nmol/L. Overall, 52% of participants receiving vitamin D3 supplementation attained a serum 25(OH)D3 concentration of >75 nmol/L, while 46% attained a serum 25(OH)D3 concentration between 50-75 nmol/L. No significant differences were observed in vitamin D metabolite concentration changes from baseline in study subjects who attained 25(OH)D3 concentration between 50-75 nmol/L, as compared to those who attained a serum 25(OH)D3 concentration >75 nmol/L (Table 2).

Substantial inter-individual variability in changes in serum 25(OH)D3, 3-epi-25(OH)D3, 24,25(OH)₂D3 and 1,25(OH)₂D3 was observed following administration of 100,000 IU vitamin D3. This variability was not dependent on baseline serum levels of the respective analytes, as shown in Fig. 2.

Overall, 25(OH)D3 accounted for approximately 89–90% of circulating vitamin D metabolites at baseline, 24,25(OH)₂D3 accounted for 7%, and 3-epi-25(OH)D3 for approximately 3–4%. These proportions did not change after vitamin D3 supplementation (Fig. 3).

Correlations between vitamin D3 metabolites before and after vitamin D3 supplementation In the overall study population (n=107), there were significant correlations at baseline between serum concentrations of 25(OH)D3 and 1,25(OH)₂D3, 24,25(OH)₂D3 or 3-epi-25(OH)D3 (ρ =0.39, 0.86 and 0.36, respectively; *P*<0.001 for all) as shown in Fig. 4. Serum concentrations of 24,25(OH)₂D3 at baseline correlated significantly with 3-epi-25(OH)D3 (ρ =0.37, *P*<0.001), but there were no other significant correlations between the other metabolites. There were also weak but significant correlations at baseline between serum 25(OH)D3 and calcium concentrations (ρ =0.24, *P*=0.013), and between serum 24,25(OH)₂D3 and PTH concentrations (ρ =0.20, *P*=0.043).

Among participants who received vitamin D3 supplementation (n=52), there were significant positive correlations at 4 weeks between serum 25(OH)D3 concentrations and 24,25(OH)₂D3 (ρ =0.47, *P*<0.001) and 3-epi-25(OH)D3 (ρ =0.35, *P*=0.011), and a significant negative correlation between serum 1,25(OH)₂D3 and 3-epi-25(OH)D3 (ρ =-0.46, *P*<0.001). The change in serum 25(OH)D3 concentrations from baseline to 4 weeks after supplementation was significantly correlated with the change in 24,25(OH)₂D3 concentrations (ρ =0.49, *P*<0.0001), but not with changes in 1,25(OH)D₂D3 concentrations (ρ =0.05, *P*=0.71).

Predictors of 25(OH)D3 response to vitamin D3 supplementation

Multiple regression analyses were performed to identify predictors of the 25(OH)D3 response to vitamin D3 supplementation. The results of these analyses are summarized in Table 23. The variance in the 25(OH)D3 level after supplementation explained by a simple regression model that included only 25(OH)D3 at baseline was 15% (R²=0.17, F(1,50)=10.2, *P*=0.002) Adjustment for other vitamin D3 metabolites (1,25(OH)₂D3, 24,25(OH)₂D3 or 3-epi-25(OH)D3), age, sex or BMI did not further improve the prediction of 25(OH)D3 levels after supplementation. Similarly, other putative markers of vitamin D3 status, including the $24,25(OH)_2D3/25(OH)D3$ ratio alone or in combination with age, sex, and BMI were not predictive of 25(OH)D3 concentrations after vitamin D3 supplementation. None of the regression models could predict the variance in the 25(OH)D3 change after supplementation (Table 23).

Changes in other circulating biomarkers of calcium homeostasis

Participants receiving vitamin D supplementation showed a significant decrease in PTH concentrations at 4 weeks, whereas PTH concentrations were increased in placebo-treated participants (mean change -2.6 ± 13 versus 3.9 ± 18 ng/L, respectively; *P*=0.03). Calcium and phosphate concentrations remained unchanged in both groups.

Discussion

This study has shown that serum concentrations of 25(OH)D3, $24,25(OH)_2D3$, 3-epi-25(OH)D3 and $1,25(OH)_2D3$ all increase significantly 4 weeks after a single high oral dose of 100,000 IU vitamin D3, whereas no such changes are seen in placebo-treated participants. The increase in 25(OH)D3 concentrations after supplementation was significantly associated with the increase in $24,25(OH)_2D3$ concentrations after supplementation.

Taking the 24,25(OH)₂D values and the ratio of 24,25(OH)₂D/25(OH)D3 and 24,25(OH)₂D3/1,25(OH)₂D3 as markers of vitamin D catabolism, we found significant increases in these variables following supplementation with a high dose of vitamin D3, which indicates induction of the vitamin D catabolic pathway. This suggests that, when adequate amounts of biologically active vitamin D are available, the production of the vitamin D catabolite 24,25(OH)D3, due to increased activity of 24-hydroxylase (CYP24A1), thereby avoiding excessive production of 1,25(OH)D3 and associated toxicity. Interestingly, a previous study from our group, which analysed vitamin D metabolite profiles in three supplementation studies, showed that the production of 24,25(OH)₂D3 is favoured over 1,25(OH)₂D3 following administration of high doses of vitamin D3, compared with lower doses [20].

The majority of participants receiving vitamin D3 attained serum 25(OH)D3 concentrations above 50 nmol/L, a widely accepted threshold for vitamin D insufficiency [11], but only 52% of subjects attained serum 25(OH)D3 concentrations above 75 nmol/L. This indicates that the use of a single high dose of vitamin D is not sufficient to ensure that adequate vitamin D levels are attained in all study participants. This would be consistent with the finding by Binkley et al [14] that suboptimal 25(OH)D3 levels persisted in approximately 20% of individuals despite dosing with vitamin D3, 50,000 IU monthly, for 1 year. Furthermore, our results demonstrate large inter-individual variations in the increase in 25(OH)D3 and 24,25(OH)₂D concentrations following administration of 100,000 IU vitamin D3. In addition, we provide the first evidence that the increase in the 3-epimer 25(OH)D metabolite following vitamin D supplementation also shows large inter-individual variation in adults, probably due to modifying factors, as has previously been described for 25(OH)D3 and 24,25(OH)₂D3 [5,12,14,16]. This inter-individual variation in the response to vitamin D3 supplementation. For example, looking

at Fig.2, it can be seen that participants 45 and 47 in the vitamin D supplementation group had similar baseline concentrations of 25(OH)D3, but the increases in 24,25(OH)₂D3 and 3-epi25(OH)D3 following supplementation differed markedly between the two participants. These large individual variations in the response to supplementation should be taken into account when giving recommendations for vitamin D supplementation. Clearly, a single fixed dose of vitamin D will not suffice to ensure adequate 25(OH)D levels in all patients unless the dose is very large, thereby increasing the risk of toxicity [16]. It is therefore desirable to tailor the dose of vitamin D in order to achieve pre-specified 25(OH)D3 targets in individual patients [16].

Several factors may contribute to the inter-individual variability in the response to vitamin D supplementation, including BMI, baseline 25(OH)D3 concentrations and genetic factors. Single nucleotide polymorphisms (SNPs) involved in the synthesis (DHCR7 and CYP2R1), binding and transportation (DBP/GC) and degradation (CYP24A1) of vitamin D and its metabolites have been shown to contribute to differences in the vitamin D response to supplementation [15,25–27]. In contrast to findings from other studies [12], the change in 25(OH)D3 concentrations after therapy in our study was not dependent on the age and BMI of the study participants at baseline. This could be due to the narrow age and BMI ranges of the participants in our study (age: 29 ± 6 years; BMI: 22 ± 2 kg/m²).

The well accepted negative correlation between baseline levels of 25(OH)D3, and the increase in this metabolite following supplementation [12,28], was not seen in this study. Similar negative findings have been reported by Binkley et al [16]. This lack of correlation in our study may be due to the short time period over which concentrations were measured, and the fact that only a single dose was used. In our regression model including only 25(OH)D3 at baseline, the baseline value explained 15% of the variance in the 25(OH)D3 concentration after supplementation. The inclusion of other vitamin D3 metabolites in the regression models did not improve the predictive power of baseline 25(OH)D3, and the 24,25(OH)D₂D3/25(OH)D3 ratio was not predictive of the 25(OH)D3 response.

The epimeric metabolite 3-epi-25(OH)D3 was present in 88% of participants at baseline in this study, at a mean concentration equivalent to 3.5% of serum 25(OH)D3 concentrations. This finding is consistent with previous studies that found vitamin D3 epimers to be present in adults, albeit in lower concentrations than in infants [17,21,28,29]. However, the physiological

significance of these metabolites is unknown [21,22]. Due to the low concentrations of vitamin D epimers in adults, the inclusion of 3-epi-25(OH)D3 has only a marginal effect on the classification of vitamin D status [24]. In the present study, 3-epi-25(OH)D3 concentrations were not predictive of the increase in 25(OH)D3 following supplementation.

To our knowledge, this is the first study to report the concentrations of key vitamin D metabolites following the administration of a high oral dose of vitamin D3 in young healthy adults with vitamin D deficiency/insufficiency. It is possible that changes in vitamin D metabolites after vitamin D administration might be different in the elderly as compared to young adults. Further studies are required to address the impact of vitamin D supplementation on key vitamin D metabolite concentration changes in elderly as vitamin D deficiency/insufficiency is more common in elderly subjects. Limitations of the study include the small sample size, the narrow age and BMI ranges of the participants and the short and noncomprehensive follow-up after supplementation. As described by Binkely et al [14], following administration of 50,000 IU vitamin D3, 25(OH)D3 concentrations rise rapidly and reach a peak after 3 days, whereas in our study blood collection was only performed 4 weeks after dosing. An analysis of the kinetics of vitamin D catabolism by measuring changes in 24,25(OH)₂D concentrations over time following supplementation would be of great interest. We did not analyse the activities of enzymes involved in the enzymatic conversion of vitamin D metabolites (CYP27B1, CYP2R1, and CYP24A1), or polymorphisms of these enzymes. Moreover, we did not assess the genetic variants of vitamin D binding protein, which is well known to affect the response to vitamin D3 supplementation [31].

In conclusion, this study has shown that administration of a single high oral dose of vitamin D3 leads to a significant increase in concentrations of 25(OH)D3 and its metabolites 24,25(OH)2D3, 3-epi (OH)D3 and 1,25(OH)2D3, with induction of the catabolic pathway predominating over the production of the active metabolite 1,25(OH)D3. The study has also highlighted the substantial heterogeneity in the 25(OH)D response to supplementation, which means that any given dose of vitamin D is unlikely to achieve optimal vitamin D status in all treated individuals. New cost-effective screening strategies are urgently needed to avoid the current trend toward universal supplementation on sight, and to help identify individuals requiring lower- or higher-dose vitamin D supplements: it should be emphasised that high doses of vitamin D are often counter-productive as they may not achieve an adequate increase in 25(OH)D.

References

- Holick MF, Binkley NC, Bischoff-Ferrari HA, Gordon CM, Hanley DA, Heaney RP, et al (2011) Evaluation, treatment, and prevention of vitamin D deficiency: an Endocrine Society clinical practice guideline. J Clin Endocrinol Metab 96:1911–1930
- Herrmann M, Farrell CL, Pusceddu I, Fabregat-Cabello N, Cavalier E (2016) Assessment of vitamin d status - a changing landscape. Clinical chemistry and laboratory medicine: CCLM / FESCC 2016
- Beckman MJ, Tadikonda P, Werner E, Prahl J, Yamada S, DeLuca HF (1996) Human 25-hydroxyvitamin D3-24-hydroxylase, a multicatalytic enzyme. Biochemistry 35:8465–84
- Prosser DE, Jones G (2004) Enzymes involved in the activation and inactivation of vitamin D. Trends Biochem Sci 29:664–673
- Wagner D, Hanwell HE, Schnabl K, Yazdanpanah M, Kimball S, Fu L, Sidhom G, Rousseau D, Cole DE, Vieth R (2011) The ratio of serum 24,25-dihydroxyvitamin D(3) to 25-hydroxyvitamin D(3) is predictive of 25-hydroxyvitamin D(3) response to vitamin D(3) supplementation. J Steroid Biochem Mol Biol 126:72–77
- Cashman KD, Hayes A, Galvin K, Merkel J, Jones G, Kaufmann M, Hoofnagle AN, Carter GD, Durazo-Arvizu RA, Sempos CT (2015) Significance of serum 24,25dihydroxyvitamin D in the assessment of vitamin D status: a double-edged sword? Clin Chem 61:636–645
- Jones G, Prosser DE, Kaufmann M (2012) 25-Hydroxyvitamin D-24-hydroxylase (CYP24A1): its important role in the degradation of vitamin D. Arch Biochem Biophys 523:9–18
- Gal-Moscovici A1, Gal M, Popovtzer MM (2005) Treatment of osteoporotic ovariectomized rats with 24,25(OH)2D3. Eur J Clin Invest 35:375–379
- 9. Henry HL, Norman AW (1978) Vitamin D: two dihydroxylated metabolites are required for normal chicken egg hatchability. Science 201:835–837
- Norman AW, Okamura WH, Bishop JE, Henry HL (2002) Update on biological actions of 1alpha,25(OH)2-vitamin D3 (rapid effects) and 24R,25(OH)2-vitamin D3. Mol Cell Endocrinol 197(1-2):1–13
- Ross AC, Manson JE, Abrams SA, Aloia JF, Brannon PM, Clinton SK, Durazo-Arvizu RA et al (2011) The 2011 report on dietary reference intakes for calcium and

vitamin D from the Institute of Medicine: what clinicians need to know. J Clin Endocrinol Metab 96:53–58

- Lehmann U, Riedel A, Hirche F, Brandsch C, Girndt M, Ulrich C, Seibert E, Henning C, Glomb MA, Dierkes J, Stangl GI (2015) Vitamin D3 supplementation: response and predictors of vitamin D3 metabolites - a randomized controlled trial. Clin Nutr 35:351–358
- Cashman KD, Hill TR, Lucey AJ, Taylor N, Seamans KM, Muldowney S, Fitzgerald AP, Flynn A, Barnes MS, Horigan G, Bonham MP, Duffy EM, Strain JJ, Wallace JM, Kiely M (2008) Estimation of the dietary requirement for vitamin D in healthy adults. Am J Clin Nutr 88:1535–1542
- Binkley N, Gemar D, Engelke J, Gangnon R, Ramamurthy R, Krueger D, Drezner MK (2011) Evaluation of ergocalciferol or cholecalciferol dosing, 1,600 IU daily or 50,000 IU monthly in older adults. J Clin Endocrinol Metab 96:981–988
- 15. Sollid ST, Hutchinson MY, Fuskevåg OM, Joakimsen RM, Jorde R (2016) Large individual differences in serum 25-hydroxyvitamin D response to vitamin D supplementation: effects of genetic factors, body mass index, and baseline concentration. Results from a randomized controlled trial. Horm Metab Res 48:27–34
- Binkley N, Lappe J, Singh RJ, Khosla S, Krueger D, Drezner MK, Blank RD (2015) Can vitamin D metabolite measurements facilitate a "treat-to-target" paradigm to guide vitamin D supplementation? Osteoporos Int 26:1655–1660
- 17. Kaufmann M, Gallagher JC, Peacock M, Schlingmann KP, Konrad M, DeLuca HF, Sigueiro R, Lopez B, Mourino A, Maestro M, St-Arnaud R, Finkelstein JS, Cooper DP, Jones G (2014) Clinical utility of simultaneous quantitation of 25hydroxyvitamin D and 24,25-dihydroxyvitamin D by LC-MS/MS involving derivatization with DMEQ-TAD. J Clin Endocrinol Metab 99:2567–2574
- Berg AH, Powe CE, Evans MK, Wenger J, Ortiz G, Zonderman AB, Suntharalingam P, Lucchesi K, Powe NR, Karumanchi SA, Thadhani RI (2015) 24,25dihydroxyvitamin D3 and vitamin D status of community-dwelling black and white Americans. Clin Chem 61:877–884
- Carter GD, Phinney KW (2014) Assessing vitamin D status: time for a rethink? Clin Chem 60:809–811
- Tang J, Nicholls H, Dutton J, Piec I, Washbourne C, Saleh L, Novak A, Macdonald H, Jackson S, Greeves J, Fraser W (2016) Profiles of 25 hydroxyvitamin D and its

metabolites 24,25-dihydroxyvitamin D and 1,25-dihydroxyvitamin D in vitamin D₃ supplementation studies. Bone Abstracts 5:P21. Doi:10.1539/boneabs.5.P21

- 21. Bailey D, Veljkovic K, Yazdanpanah M, Adeli K (2013) Analytical measurement and clinical relevance of vitamin D(3) C3-epimer. Clin Biochem 46:190–196
- 22. Cashman KD, Kinsella M, Walton J, Flynn A, Hayes A, Lucey AJ, Seamans KM, Kiely M (2014) The 3 epimer of 25-hydroxycholecalciferol is present in the circulation of the majority of adults in a nationally representative sample and has endogenous origins. J Nutr 144:1050–1057
- 23. Nowak A, Boesch L, Andres E, Battegay E, Hornemann T, Schmid C, Bischoff-Ferrari HA, et al (2016) Effect of vitamin D3 on self-perceived fatigue: a double-blind randomized placebo-controlled trial. Medicine (Baltimore) 95(52):e5353 Albina Nowak LB, Erik Andres, Edouard Battegay, Thorsten Hornemann, Christoph Schmid, Heike A. Bischoff-Ferrari, Paolo M. Suter, Pierre-Alexandre Krayenbuehl. Medicine. 2016;in press
- 24. Lutsey PL, Eckfeldt JH, Ogagarue ER, Folsom AR, Michos ED, Gross M (2015) The 25-hydroxyvitamin D3 C-3 epimer: distribution, correlates, and reclassification of 25hydroxyvitamin D status in the population-based Atherosclerosis Risk in Communities Study (ARIC). Clin Chim Acta 442:75–81
- 25. Nimitphong H, Saetung S, Chanprasertyotin S, Chailurkit LO, Ongphiphadhanakul B (2013) Changes in circulating 25-hydroxyvitamin D according to vitamin D binding protein genotypes after vitamin D₃ or D₂ supplementation. Nutr J;12:39
- 26. Didriksen A, Grimnes G, Hutchinson MS, Kjærgaard M, Svartberg J, Joakimsen RM, Jorde R (2013) The serum 25-hydroxyvitamin D response to vitamin D supplementation is related to genetic factors, BMI, and baseline levels. Eur J Endocrinol 169:559–567
- 27. Fu L, Yun F, Oczak M, Wong BY, Vieth R, Cole DE (2009) Common genetic variants of the vitamin D binding protein (DBP) predict differences in response of serum 25-hydroxyvitamin D [25(OH)D] to vitamin D supplementation. Clin Biochem 42:1174–1177
- Schwartz JB, Kane L, Bikle D (2016) Response of Vitamin D Concentration to Vitamin D3 Administration in Older Adults without Sun Exposure: A Randomized Double-Blind Trial. J Am Geriatr Soc 64:65–72
- Lensmeyer G, Poquette M, Wiebe D, Binkley N (2012) The C-3 epimer of 25 hydroxyvitamin D(3) is present in adult serum. J Clin Endocrinol Metab 97:163–168

30. Strathmann FG, Sadilkova K, Laha TJ, LeSourd SE, Bornhorst JA, Hoofnagle AN, Jack R (2012) 3-epi-25 hydroxyvitamin D concentrations are not correlated with age in a cohort of infants and adults. Clin Chim Acta 413:203–206

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Table 1 Baseline demographic and clinical characteristics

Data are shown as mean (SD), and groups were compared using unpaired two-sided t tests, unless indicated otherwise.

	Therapy	Placebo	P value
	n=52	n=55	
Age (years)	29 (6)	28 (6)	0.30
Gender (females/males)	27/25	26/29	0.15 ^a
	(52%/48%)	(47%/53%)	
BMI (kg/m ²)	22 (2)	22 (2)	0.54
Arterial blood pressure (mmHg)			
Systolic	123 (11)	126 (11)	0.16
Diastolic	78 (9)	77 (8)	0.44
Parathyroid hormone (ng/L)	44 (16)	46 (18)	0.59
Calcium (mmol/L) ^b	2.23 (0.07)	2.22 (0.07)	0.97
Phosphate (mmol/L)	0.99 (0.18)	1.00 (0.15)	0.69
Creatinine (µmol/L)	71 (14)	75 (13)	0.13
C-reactive protein (mg/L) ^c	0.5 (0.0–1.2)	0.6 (0.3–1.8)	0.27
24,25(OH)2D3 (nmol/L)	2.2 (0.9)	2.5 (1.0)	0.08
25(OH)D3 (nmol/L)	28 (9)	32 (11)	0.06
1,25(OH)2D3 (pmol/L)	100 (29)	94 (25)	0.23
3-epi-25(OH)D3 (nmol/L)	1.0 (0.9)	1.3 (0.93)	0.08
25(OH)D2 (nmol/L) ^c	1.8 (1.1–2.2)	2 (1.4–2.6)	0.06

 $a\chi^2$ -test; ^bAdjusted for serum albumin concentrations; ^cmedian (interquartile range)

Table 2 Mean (\pm SD) vitamin D metabolite concentration changes from baseline in supplemented subjects who attained serum 25(OH)D3 concentrations between 50-75 nmol/L versus those who attained a serum 25(OH)D3 concentration >75 nmol/L, four weeks after a single oral dose of 100.000 IU vitamin D3 administration.

Vitamin D metabolites	50-75 nmol/L	>75 nmol/L	
Vitamin D metadontes	[n=24]	[n=27]	p-value
25(OH)D3 [nmol/L]	39.2 ± 10.3	59.6 ± 18.9	< 0.001
24,25(OH)2D3 [nmol/L]	3.9 ± 1.2	4.9 ± 2.8	0.13
1,25(OH) ₂ D3 [pmol/L]	20.9 [-29.4 - 78.0]*	32.3 [-45.8 - 83-5]*	0.20*
3-epi-25(OH)D3 [nmol/L]	1.7 ± 1.6	2.4 ± 1.9	0.17
24,25(OH)2D3/25(OH)D3	0.091 ± 0.016	0.081 ± 0.026	0.10
24,25(OH)2D3/1,25(OH)2D3	0.053 ± 0.017	0.060 ± 0.031	0.36

*median [5th-95th percentile], Mann-Whitney test.

Model	Covariate	Beta coefficient (95% CI)	<i>P</i> value
Model 1	25(OH)D3	0.41 (0.36 - 1.58)	0.002
Model 2	25(OH)D3	0.71 (0.58 - 2.79)	0.004
	24,25(OH)2D3	-0.34 (-19.17 - 3.17)	0.156
	3-epi-(OH)2D3	-0.06 (-8.86 - 5.56)	0.648
Model 3	25(OH)D3	0.74 (0.57 - 2.93)	0.005
	24,25(OH)2D3	-0.38 (-20.61 - 2.60)	0.125
	3-epi-(OH)2D3	-0.01 (-7.78 - 7.21)	0.939
	age	-0.18 (-1.58 - 0.34)	0.203
	sex	0.005 (-11.02 - 11.40)	0.973
	BMI	-0.11 (-4.05 - 1.79)	0.440
Model 4	25(OH)D3	0.84 (0.73 - 3.21)	0.002
	24,25(OH)2D3	-0.41 (-21.3 - 1.96)	0.101
	3-epi-(OH)2D3	-0.05 (-8.61 - 5.84)	0.702
	1,25(OH)2D3	-0.15 (-0.33 - 0.11)	0.308

Table 3 Regression models for the 25(OH)D3 response to vitamin D3 supplementation

Model summaries: Model 1: $R^2=0.17$, adjusted $R^2=0.15$, F(1,50)=10.2, P=0.002; Model 2: $R^2=0.21$, adjusted $R^2=0.16$, F(3,48)=4.3, P=0.009; Model 3: $R^2=0.27$, adjusted $R^2=0.17$, F(6,45)=4.3, P=0.023; Model 4: $R^2=0.23$, adjusted $R^2=0.16$, F(3,47)=4.3, P=0.014

Figure captions



Fig. 1 Absolute changes in vitamin D metabolites from baseline (dark shading) to 4 weeks after (light shading) a single 100,000 IU oral dose of vitamin D3
(A) 25(OH)D3; (B) 24,25(OH)D3; (C) 1,25 (OH)₂ D3; (D) 3-epi-25(OH)D3: (E) 24,25(OH)₂D3/(25(OH)D3; (F) 24,25(OH)₂D3/1.25(OH)₂D3/



Fig. 2 Changes in serum 25(OH)D3 [nmol/L] (**A**, **B**), 1,25(OH)2D3 [pmol/L] (**C**, **D**), (**B**) 24,25(OH)2D3 [nmol/L] (**E**, **F**), (**C**) 3-epi-25(OH)D3 [nmol/L] (**G**, **H**) concentrations from baseline to 4 weeks after vitamin D supplementation in individual participants Open circles: baseline, black-filled circles: post-supplementation; asterisks indicate participants specifically referred to in the discussion.



Fig. 3 Relative proportions of vitamin D3 metabolites in serum at baseline (visit A) and 4 weeks after a 100,000 IU single oral dose of vitamin D3 or placebo (visit B)



Fig. 4 Correlations between baseline concentrations of vitamin D metabolites.