Assessment of Vitamin D status using Mitra® volumetric absorptive microsampling (VAMS) device

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Introduction

- > Since the introduction of Guthrie cards in the 70s, filter paper method for collection of dried blood spots (DBS) has gained popularity as an alternative form of sampling technique to venepuncture.
- > DBS sampling is less invasive than venepuncture; whilst the low sample volume requirement (typically 10-50 μL) is ideally suited for use in paediatric practice and in the elderly population.
- > The reduced storage and shipping requirements allow patients to send their samples via the post, which could streamline the process of transporting samples to the laboratory and improve efficiency.
- > However, analysts face many challenges with paper-based methods; concerns over volumetric inaccuracy, variability in spot sizes, analyte stability and reproducibility of measurements in subpunches, have prohibited the wider use of DBS sampling techniques. Volumetric microsampling devices are able to accurately collect a fixed amount of samples, and because sub-punching is not required, it overcomes many drawbacks associated with filter paper collection method.

Aims and Objectives

- > To describe the use of Mitra[®] volumetric absorptive microsampler (VAMS) (Torrance, CA, USA) for LC-MS/MS measurement of $250HD_3$ and interpretation of vitamin D status.
- > To compare assay performance of Mitra VAMS against paper-based dried blood spot techniques.

Assay Characteristics

Intra-assay precision

wDBS	SpDBS	VAMS

Inter-assay precision

wDBS	SpDBS	VAMS
Mean 250H	ID ₃ nmol/L (%	CV) (n = 6)



Method of analysis

- > Whole blood K3EDTA samples 157 patients were from
- selected at random, following routine analysis.
- \geq 10 µL of blood was pipetted into Whatman[®] 903 Protein saver cards or sampled using Mitra[®] VAMS (10 µL fixed product number volume: 10006), then left to dry for 18 hours at room temperature prior to storage at -20°.
- Prior to analysis, DBS samples were extracted by 1) cutting the whole spot (wDBS), or by 2) making two 3 mm subpunches (spDBS).



Sample extraction procedure





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1	7.5 (16.1)	7.6 (13.6)	7.3 (8.2)		
2	23.4 (8.9)	23.2 (9.0)	20.5 (6.7)		
3	43.6 (9.1)	45.0 (12.0)	40.3 (6.4)		
4	64.2 (6.5)	71.1 (9.8)	59.5 (7.7)		

Assay recovery

Expected	Mean recovery (%)			
25OHD ₃ nmol/L	wDBS	SpDBS	VAMS	
8.5	102.4	110.2	98.4	
21.0	90.0	108.6	104.0	
37.5	93.3	97.1	101.7	
62.5	89.2	96.7	100.7	
87.5	90.3	104.6	95.4	
106.3	91.8	102.5	99.6	
Overall	92.8	103.3	100.0	
average	(78.9-	(88.3-	(93.4-	
(range)	114.1)	120)	114.1)	

Low QC	10.6 (16.6)	12.2 (15.1)	10.9 (7.4)
Medium QC	57.7 (6.9)	62.7 (7.5)	64.8 (7.1)
High QC	82.0 (3.6)	93.3 (8.3)	91.3 (7.0)

 \blacktriangleright Linearity from 0-125 nmol/L (Typical r² value \leq 0.98) Lower limit of quantification: wDBS 1.6 nmol/L, spDBS 2.5 nmol/L, Mitra[®] VAMS 1.5 nmol/L.



A 209-day stability plot showing percentage change of 250HD₃ concentration from day one in samples collected by wDBS and VAMS stored at -20°C.

Comparison of dried blood-to-plasma equivalency values (PEV)					
1) Whole	e spot (wDBS)	against plas	ma concentra	tion	
ن ¹⁴⁰	Identity line Y = X	180 160	Identity line Y = X	120 100	n = 70

Effects of Haematocrit (Hct)

displacement on 250HD₃ concentrations

1) Removal of plasma volume



- The plasma layer was taken off until completely in steps removed.
 - Decrease in $[250HD_3]$ were proportional to the decrease in plasma volume.
 - \succ This shows 250HD₃ in blood is present primarily in the fluid compartment, the intracellular space contained <1.7% of total 250HD₃.
- > When plasma-free packed cells were added to a whole blood full sample until saturation, [250HD₃] decreases as Hct level increased.
- Despite the constant plasma volume in the sample, the increasing level of Hct prevented the uptake of 250HD₃ into the microsampling devices.
- > Findings from the above studies indicated the concentration of 250HD₃ in whole blood is dependent upon the level of Hct present, and that measurements using microsampling devices must be corrected for the level of Hct.

 \succ Fig a)-c) Raw 250HD₃ values produced from DBS and VAMS (n=97) were correlated with plasma concentration, but showed an average negative bias of -39.3%.

into a clinically-relevant PEVs (n=70). Analysis of concordance correlation coefficient (CCC) and correctional bias (Cb) showed good agreement with plasma concentrations.

showed the assay bias was negatively associated with the increase in Hct levels.

Mitra VAMS showed the least deviation across the Hct range.

Conclusions

- \succ VAMS demonstrated benefits over the conventional paper-based method; the consistency in sampling volume, ease of use without the need for sub punches, and preservation of sample constituency.
- > Our study provides validation of microsampling methodologies for measurement of $250HD_3$ and interpretation of vitamin D status.
- \succ VAMS produced more precise measurements of 250HD₃, and the most accurate reflection of vitamin D status compared to wDBS and spDBS.
- \succ Although the recovery of the analyte remains Hct-dependent, the use of an empirically-derived model to transform DBS values into clinically-relevant equivalency improves the interpretability of results.

Use of dried blood-to-plasma equivalency value for interpretation of vitamin D status

Vitamin D status definitions	Plasma (no. of cases, % in cohort)	PEV _{wDBS} (n, Δ%)	PEV _{spDBS} (n, Δ%)	PEV _{vams} (n, Δ%)
<30 nmol/L, Deficiency	27 (38.6%)	20 (↓10%)	15 (↓17.1%)	24 (↓4.3%)
30-50 nmol/L, Insufficiency	24 (34.3%)	29 (个7.1%)	31 (↑10%)	26 (↑2.9%)
>50 nmol/L, sufficiency	19 (27.1%)	21 (↑2.9%)	24 (↑7.1%)	20 (↑1.4%)

- Following interpretation guidelines from the IOM, we classified the vitamin D status in our patient cohort (n = 70).
- Using PEVs resulted in an small underestimation of individuals with deficiency status.
- three Between the microsampling techniques, results produced from VAMS were most representative of plasma.