

## pyisotopomer: A Python package for obtaining intramolecular isotope ratio differences from mass spectrometric analysis of nitrous oxide isotopocules

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Keywords:	Nitrous oxide, isotopomers, isotopocules, scrambling, Python
Abstract:	RATIONALE Obtaining nitrous oxide isotopocule measurements with isotope ratio mass spectrometry (IRMS) involves analyzing the ion current ratios of the nitrous oxide parent ion $(N_2O^+)$ as well as those of the NO <sup>+</sup> fragment ion. The data analysis requires correcting for "scrambling" in the ion source, whereby the NO <sup>+</sup> fragment ion obtains the outer N atom from the N <sub>2</sub> O <sup>+</sup> molecule. While descriptions exist for this correction, and interlaboratory intercalibration efforts have been made, there has yet to be published a package of code for implementing isotopomer calibrations. METHODS We developed a user-friendly Python package (pyisotopomer) to determine two coefficients ( $\gamma$ and $\kappa$ ) that describe scrambling in the IRMS ion source, and then to use this calibration to obtain intramolecular isotope deltas in N <sub>2</sub> O samples. RESULTS With two appropriate reference materials, $\gamma$ and $\kappa$ can be determined robustly and accurately for a given IRMS. An additional third reference material is needed to define the zero-point of the delta scale. We show that IRMS scrambling behavior can vary with time, necessitating regular calibrations. Finally, we present an intercalibration between two IRMS laboratories, using pyisotopomer to calculate $\gamma$ and $\kappa$ and to obtain intramolecular N <sub>2</sub> O isotope deltas in lake water unknowns. CONCLUSIONS Given these considerations, we discuss how to use pyisotopomer to obtain high-quality N <sub>2</sub> O isotopocule data from IRMS

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Figure 2. Linearity relations for reference materials used to normalize measured isotope ratios to a peak area of 20 Vs (10 nmol N<sub>2</sub>O), using the dummy variable method.  ${}^{31}\delta$ +1 (a,d,g),  ${}^{45}\delta$ +1 (b,e,h), and  ${}^{46}\delta$ +1 (c, f, i) are plotted against m/z 44 peak area. Linearity relations are shown for reference materials prepared and run in Lab 1 (a-c), reference materials prepared in Lab 2 but run in Lab 1 (d-f), and reference materials run in Lab 2 (g-i). A common slope (black line) calculated from the dummy variable method for each molecular ion ratio is overlain on each data series (colored circles). The estimated isotope ratio corresponding to a peak area of 20 Vs is also shown for each series (colored diamonds, error bars correspond to the standard error of the predicted y-value).

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Figure 3.  $\gamma - \kappa$  for the Lab 1 IRMS from June 2018 to March 2021. Daily mean  $\gamma - \kappa$  (black line) values are plotted with a 5-day rolling average (dots).

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3	1	In preparation for Rapid Communications in Mass Spectrometry
4	2	pyisotopomer: A Python package for obtaining intramolecular isotope ratio differences
5	3	from mass spectrometric analysis of nitrous oxide isotopocules
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19 20	14	
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50	39 40	high-quality N <sub>2</sub> O isotopocule data from IRMS systems, including the use of appropriate
51	40	reference materials and frequency of calibration.
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### **1. Introduction**

Nitrous oxide (N<sub>2</sub>O) is a potent greenhouse gas, with a global warming potential 265 times that of carbon dioxide over a 100 year time horizon<sup>1,2</sup>. N<sub>2</sub>O is also likely to be the most emitted ozone depletion agent in the 21st century, due to production of NO radicals in the stratosphere that interact destructively with ozone<sup>3-6</sup>. Historically, the bulk stable isotopes of nitrogen and oxygen in N<sub>2</sub>O have been used to quantify its microbial cycling in soils<sup>7,8</sup> and in the ocean<sup>9-12</sup>, its destruction by photolysis and  $O(^{1}D)$ , and its cycling in the atmosphere<sup>13,14</sup>. This approach often fails at disentangling different N<sub>2</sub>O production and consumption mechanisms, because the bulk nitrogen and oxygen isotope ratios of N<sub>2</sub>O depend on the isotopic composition of the substrate, as well as the isotope effects of production and consumption processes<sup>12</sup>. Furthermore, in the context of microbial N<sub>2</sub>O cycling in soils and the ocean, bacterial nitrification and denitrification produce N<sub>2</sub>O with similar bulk  $\delta$ <sup>(15</sup>N)<sup>1</sup> values, preventing partitioning between these processes on the basis of bulk  $\delta$ <sup>(15</sup>N) alone<sup>15,16</sup>. 

The site-specific nitrogen isotope ratios of N<sub>2</sub>O provide a more nuanced constraint on the biogeochemical cycling of N<sub>2</sub>O than its bulk composition alone. N<sub>2</sub>O isotopomers have been used extensively to quantify its biogeochemical cycling in soils<sup>17-20</sup>, the atmosphere<sup>14,21-23</sup>, and the ocean<sup>24–34</sup>. The individual isotopic compositions of each nitrogen atom were first measured by Friedman and Bigeleisen, who quantified the yields of isotopomers <sup>14</sup>N<sup>15</sup>N<sup>16</sup>O and <sup>15</sup>N<sup>14</sup>N<sup>16</sup>O from enriched ammonium nitrate by measuring the NO<sup>+</sup> fragment ion signal in an isotope ratio mass spectrometer (IRMS)<sup>35</sup>. 50 years later, these N<sub>2</sub>O isotopomers were quantified at natural abundance from the  $N_2O^+$  species with mass numbers 44, 45, and 46 and the mass 30 and 31 NO<sup>+</sup> fragment ion<sup>36,37</sup>. The central nitrogen atom in the N<sub>2</sub>O molecule has been designated with locants  $\alpha$ ,  $\mu$ , or 2; the terminal atom, with locants  $\beta$ ,  $\tau$ , or  $1^{38,39}$ . Here, we use the definitions from Toyoda and Yoshida (1999) for the site-specific isotope number (N) ratios of the central ( $\alpha$ ) nitrogen atom and terminal ( $\beta$ ) nitrogen atom<sup>36</sup>: 

$${}^{15}R^{\alpha} = \frac{N({}^{14}N^{15}N0)}{N({}^{14}N^{14}N0)}$$
(1)

 ${}^{15}R^{\beta} = \frac{N({}^{15}N{}^{14}NO)}{N({}^{14}N{}^{14}NO)}$ (2)

The N<sub>2</sub>O isotopomer measurement was initially performed with two sequential measurements of the same sample on an isotope ratio mass spectrometer, one at m/z 44, 45, and 46. and the other at m/z 30 and 31<sup>36</sup>. Use of dedicated cup-configurations on lower-dispersion IRMS instruments allowed simultaneous analysis of all five masses together<sup>40</sup>.

The slight difference in absorption cross sections between the isotopocules of N<sub>2</sub>O result in different isotopic fractionations during photolysis and photo-oxidation in the stratosphere<sup>41</sup>, making the isotopomers of N<sub>2</sub>O a powerful tool for understanding its atmospheric cycling<sup>21,42-45</sup>. Likewise, N<sub>2</sub>O site preference, defined as  $\delta(^{15}N^{sp}) = \delta(^{15}N^{\alpha}) - \delta(^{15}N^{\beta})$ , was shown in microbial culture experiments to be largely a function of reaction mechanism, independent of source composition<sup>24,46–50</sup>. This allowed for the differentiation between N<sub>2</sub>O from bacterial nitrification  $(\delta(^{15}N^{sp}) \approx 28-38 \%)$  and denitrification  $(\delta(^{15}N^{sp}) \approx 0)^{24,46-50}$ , although more studies are needed

<sup>&</sup>lt;sup>1</sup> We write  $\delta$  values with parentheses, e.g.,  $\delta^{(15N)}$ , because  $\delta$  is the quantity symbol and "<sup>15</sup>N" is the label. See SI Brochure: https://www.bipm.org/en/publications/si-brochure/

to better constrain the SPs for diverse fungal, bacterial, and archaeal strains in both terrestrial and marine environments<sup>49,51</sup>. During N<sub>2</sub>O consumption,  $\delta(^{15}N^{\alpha})$  and  $\delta(^{18}O)$  were shown in microbial culture<sup>52</sup> and soil mesocosm<sup>19</sup> experiments to exhibit a characteristic relationship, allowing subsequent studies to use this relationship to distinguish between oxidative and reductive regimes of N<sub>2</sub>O cycling<sup>30,33</sup>.

Site-specific nitrogen isotope ratio measurements based on mass spectrometry need to be corrected for a phenomenon called "scrambling," whereby the NO<sup>+</sup> fragment ion contains the terminal N atom, rather than the central N attached to the O atom (as in the original molecule). A number of approaches have been taken to calibrate an IRMS system for this effect: 1) the use of a single "rearrangement factor" to describe scrambling $^{36,53}$ , 2) the use of nine coefficients to describe the different fragmentation behaviors of the different isotopocules of N<sub>2</sub>O<sup>54</sup>, and finally 3) the use of two coefficients to describe scrambling in the ion source<sup>50</sup>. While descriptions exist for each of these approaches, and interlaboratory intercalibration efforts have been made<sup>55,56</sup>. there has yet to be published a package of code for implementing any of the above isotopomer calibrations.

We developed a Python software package ("pyisotopomer") that implements the two-coefficient approach described by Frame and Casciotti<sup>32</sup> to calibrate an IRMS for scrambling and use that calibration to obtain high-quality N<sub>2</sub>O isotopocule data. This software solves a set of equations, either analytically or with an optimization routine, to quantify the scrambling behavior of an IRMS. To quantify the performance of the software, we tested the sensitivity of the analytical and optimization-based solutions to their input conditions and assessed when each method is most appropriate. To quantify the variability of the fragmentation behavior of an instrument over time, we examined the scrambling behavior of one IRMS over the course of four years of measurements. We derived a simplified equation and used a Monte Carlo simulation approach to quantify the effect of uncertainty in the scrambling coefficients on the final isotope deltas. Finally, we performed an intercalibration using this software across two labs, at Stanford University ('Lab 1') and the University of Basel ('Lab 2'). This paper introduces the theory, practical applications, and testing of pyisotopomer; instructions on how to use pyisotopomer are available in the documentation on the Python Package Index<sup>57</sup>. 

## **2. Mathematical framework**

110 The molecular ion number ratios  $45/44 ({}^{45}R)$  and  $46/44 ({}^{46}R)$  can be written in terms of atomic isotope ratios as  ${}^{36,53}$ :

$${}^{45}R = {}^{15}R^{\alpha} + {}^{15}R^{\beta} + {}^{17}R \tag{3}$$

$${}^{46}R = \left({}^{15}R^{\alpha} + {}^{15}R^{\beta}\right){}^{17}R + {}^{18}R + {}^{15}R^{\alpha}{}^{15}R^{\beta}$$
(4)

<sup>46</sup> <sup>47</sup> <sup>48</sup><sup>47</sup><sup>48</sup><sup>113</sup><sup>14</sup>N<sub>2</sub><sup>18</sup>O, respectively, to <sup>14</sup>N<sub>2</sub><sup>16</sup>O, assuming a stochastic isotope distribution between mono- and <sup>14</sup>N<sub>2</sub><sup>18</sup>O, respectively. to <sup>14</sup>N<sub>2</sub><sup>16</sup>O, assuming a stochastic isotope distribution between mono- and poly-substituted isotopocules.

For many N<sub>2</sub>O samples,  ${}^{17}R$  covaries with  ${}^{18}R$  according to the oxygen isotope ratios of Vienna Standard Mean Ocean Water (VSMOW)<sup>58,59</sup> and a mass-dependent relationship between  $^{17}R$  and  $^{18}R$  with coefficient  $\beta = 0.516^{60}$ . Deviations from this relationship are expressed by the oxygen triple isotope excess  $\Delta$ (<sup>17</sup>O)<sup>60–62</sup>, which provides additional information about the sources and sinks of N<sub>2</sub>O<sup>60,63</sup>: 

 ${}^{17}R/{}^{17}R_{\rm VSMOW} = ({}^{18}R/0.0020052)^{\beta}[\Delta({}^{17}O) + 1]$ (5)  $\Delta$ (<sup>17</sup>O) is sometimes assumed to be equal to zero but should be measured separately for samples with a significant  $\Delta$ (<sup>17</sup>O) anomaly, such as atmospheric nitrate<sup>60,62,63</sup>. The simplest formulation for the NO<sup>+</sup> fragment ion number ratio 31/30 (<sup>31</sup>*R*) is given as<sup>36</sup>:  ${}^{31}R = {}^{15}R^{\alpha} + {}^{17}R$ (6)This equation would represent the  ${}^{31}R$  measured by IRMS if no scrambling occurred. To describe instead the scrambled  ${}^{31}R$ , Toyoda and Yoshida ${}^{36}$  define the rearrangement factor y (which was later given the symbol y) as "the fraction of NO<sup>+</sup> bearing the  $\beta$  nitrogen of the initial N<sub>2</sub>O to the total NO<sup>+</sup> formed," to yield:  ${}^{31}R = (1 - \gamma)^{15}R^{\alpha} + \gamma^{15}R^{\beta} + {}^{17}R$ (7)where  ${}^{15}R^{\alpha}$  and  ${}^{15}R^{\beta}$  represent atomic isotope ratios of the sample. In other words,  $\gamma$  relates the scrambled NO<sup>+</sup> fragment ratio to the unscrambled  ${}^{15}R^{\alpha}$  and  ${}^{15}R^{\beta}$  of the sample. Kaiser et al.<sup>53</sup> introduced a more complete representation of  ${}^{31}R$ , adding terms for  $^{15}N^{16}O$ ,  $^{14}N^{15}N^{17}O$ , and  $^{15}N^{14}N^{17}O$  to m/z 31, and terms for  $^{15}N^{14}N^{16}O$  and  $^{14}N^{15}N^{16}O$  to m/z30:  ${}^{31}R = (1-\gamma)^{15}R^{\alpha} + \gamma^{15}R^{\beta} + {}^{17}R - \frac{\gamma(1-\gamma)({}^{15}R^{\alpha} - {}^{15}R^{\beta})^2}{1+\gamma^{15}R^{\alpha} + (1-\gamma)^{15}R^{\beta}}$ (8)  $=\frac{(1-\gamma)^{15}R^{\alpha}+\gamma^{15}R^{\beta}+{}^{15}R^{\alpha 15}R^{\beta}+{}^{17}R[1+\gamma^{15}R^{\alpha}+(1-\gamma)^{15}R^{\beta}]}{15}$  $= \frac{1 + \gamma^{15}R^{\alpha} + (1 - \gamma)^{15}R^{\beta}}{1 + \gamma^{15}R^{\alpha} + (1 - \gamma)^{15}R^{\beta}}$ Note that Kaiser et al.<sup>53</sup> use the symbol "s" for  $\gamma$ , <sup>15</sup> $R_1$  for <sup>15</sup> $R^{\beta}$ , and <sup>15</sup> $R_2$  for <sup>15</sup> $R^{\alpha}$ . To account for different fragmentation rates of different N<sub>2</sub>O isotopocules, Westley et al.<sup>54</sup> replaced the rearrangement factor  $\gamma$  with nine separate coefficients:  ${}^{31}R = \frac{a_{31}{}^{15}R^{\alpha} + b_{31}{}^{15}R^{\beta} + c_{31}{}^{15}R^{\alpha 15}R^{\beta} + {}^{17}R[d_{31} + e_{31}{}^{15}R^{\alpha} + f_{31}{}^{15}R^{\beta}]}{1 + a_{30}{}^{15}R^{\alpha} + b_{30}{}^{15}R^{\beta} + c_{30}{}^{15}R^{\alpha 15}R^{\beta}}$ (9) While this approach considers the possibility of different rearrangement factors for every  $N_2O$  isotopocule as well as  ${}^{15}N_2^+$  formation, it also requires solving for three to nine coefficients, depending on whether  $a_{30}$ ,  $b_{30}$  and  $c_{30}$ , as well as  $d_{31}$ ,  $e_{31}$  and  $f_{31}$ , are considered separately from coefficients  $a_{31}$ ,  $b_{31}$  and  $c_{31}$ . Frame and Casciotti<sup>50</sup> simplify this equation by reducing the number of rearrangement factors to two coefficients,  $\gamma$  and  $\kappa$ , which represent the yield of <sup>14</sup>NO<sup>+</sup> from <sup>14</sup>N<sup>15</sup>N<sup>16</sup>O and <sup>14</sup>N<sup>15</sup>N<sup>17</sup>O, and the yield of <sup>15</sup>NO<sup>+</sup> from <sup>15</sup>N<sup>14</sup>N<sup>16</sup>O, respectively. This produces the equation:  ${}^{31}R = \frac{(1-\gamma)^{15}R^{\alpha} + \kappa^{15}R^{\beta} + {}^{15}R^{\alpha 15}R^{\beta} + {}^{17}R[1+\gamma^{15}R^{\alpha} + (1-\kappa)^{15}R^{\beta}]}{1+\gamma^{15}R^{\alpha} + (1-\kappa)^{15}R^{\beta}}$ (10)The important pieces of information contained within the two scrambling factors are the unequal rates of fragmentation for the isotopomers <sup>14</sup>N<sup>15</sup>NO and <sup>15</sup>N<sup>14</sup>NO, which eqns. (7) and (8) assume are equal. Eqn. (10) is formulated by assuming that the <sup>17</sup>O-isotopocules have the same scrambling behavior as the <sup>16</sup>O-isotopocules, i.e.,  $e_{31} = 1 - a_{31}$  and  $f_{31} = 1 - b_{31}$ , in terms of the coefficients in eqn. (9). It is also assumed that  $c_{31} = 1$ , i.e., the yield of  ${}^{15}N^{16}O^+$  from  ${}^{15}N_2{}^{16}O$ is equal to the yield of  ${}^{14}N{}^{16}O{}^{+}$  from  ${}^{14}N{}_{2}{}^{16}O{}$ . Given that naturally occurring N<sub>2</sub>O contains very 

little  ${}^{15}N_{2}{}^{16}O_{2}$ , a small difference in this yield would not significantly alter  ${}^{31}R^{64}$ . Finally, it is assumed that  $d_{31} = 1$ , or that the yield of <sup>14</sup>N<sup>17</sup>O<sup>+</sup> from <sup>14</sup>N<sub>2</sub><sup>17</sup>O is equal to the yield of <sup>14</sup>N<sup>16</sup>O<sup>+</sup> from  ${}^{14}N_2{}^{16}O$ ; again, an assumption yielding little error in  ${}^{31}R$ , given the low natural abundance of 17O in N<sub>2</sub>O<sup>60</sup>. Eqn. (10) can be rearranged to give an equation for  $\gamma$  as a function of  $\kappa$  (the full derivation is presented in Supplementary text S1):  $\gamma = \frac{{}^{15}R^{\alpha} + \kappa^{15}R^{\beta} + {}^{15}R^{\alpha 15}R^{\beta} - ({}^{31}R - {}^{17}R)[1 + (1 - \kappa)^{15}R^{\beta}]}{{}^{15}R^{\alpha}(1 + {}^{31}R - {}^{17}R)}$ (11)For two reference materials, we can write two such equations and solve for two unknowns,  $\gamma$  and  $\kappa$ . <sup>15</sup> $R^{\alpha}$  and <sup>15</sup> $R^{\beta}$  represent *known* values for each reference material, and <sup>31</sup>R is the observed quantity. Essentially, we are asking what values of y and  $\kappa$  for a pair of known  ${}^{15}R^{\alpha}$ and  ${}^{15}R^{\beta}$  values gives the observed  ${}^{31}R$  for each reference gas. Setting the two solutions for  $\gamma$ equal allows us to determine  $\kappa$  and  $\gamma$  algebraically from the assigned <sup>15</sup>R values of reference materials 1 and 2 ( ${}^{15}R_{1}^{\alpha}$ ,  ${}^{15}R_{1}^{\beta}$ ,  ${}^{15}R_{2}^{\alpha}$ ,  ${}^{15}R_{2}^{\beta}$ ), their observed  ${}^{31}R$  values ( ${}^{31}R_{1}$ ,  ${}^{31}R_{2}$ ), and the  ${}^{17}R_{1}^{\beta}$ values  $({}^{17}R_1, {}^{17}R_2)$ :  $\kappa = \frac{\left(\frac{{^{15}R_1^{\alpha} - {^{31}R_1} + {^{17}R_1}}\right)\left(1 + {^{15}R_1^{\beta}}\right)}{{^{15}R_1^{\alpha}(1 + {^{31}R_1} - {^{17}R_1})}} - \frac{\left(\frac{{^{15}R_2^{\alpha} - {^{31}R_2} + {^{17}R_2}}\right)\left(1 + {^{15}R_2^{\beta}}\right)}{{^{15}R_2^{\alpha}(1 + {^{31}R_2} - {^{17}R_2})}}$   $\kappa = \frac{{^{15}R_2^{\beta}}}{{^{15}R_2^{\beta}}} - \frac{{^{15}R_2^{\beta}}}{{^{15}R_1^{\beta}}} - \frac{{^{15}R_2^{\beta}}}{{^{15}R_1^{\beta}}} - \frac{{^{15}R_2^{\beta}}}{{^{15}R_1^{\beta}}}$ (12a) $\frac{15}{15}R_{2}^{\alpha}$   $\frac{15}{15}R_{1}^{\alpha}$  $=\frac{\frac{\binom{15}{R_{1}^{\alpha}}-31}{R_{1}^{\alpha}+17}R_{1}\left(1+\frac{15}{R_{1}^{\beta}}\right)}{\frac{15}{R_{2}^{\beta}}}\left(\frac{15}{R_{2}^{\beta}}R_{2}^{\beta}\right)}-\frac{\binom{15}{R_{2}^{\alpha}}-31}{R_{2}^{\alpha}+17}R_{2}\left(1+\frac{15}{R_{2}^{\beta}}\right)}\left(1+\frac{15}{R_{2}^{\beta}}R_{2}^{\beta}\right)}{\frac{15}{R_{2}^{\alpha}}\left(1+\frac{31}{R_{2}^{\alpha}}-\frac{17}{R_{2}^{\beta}}\right)}{\frac{15}{R_{2}^{\beta}}}-\frac{\frac{15}{R_{1}^{\beta}}}{\frac{15}{R_{2}^{\alpha}}}-\frac{15}{R_{1}^{\beta}}}{\frac{15}{R_{1}^{\alpha}}}$ (12b)After substituting  ${}^{45}R - {}^{15}R^{\alpha} - {}^{15}R^{\beta}$  for  ${}^{17}R$ , the equations for  $\gamma$  and  $\kappa$  can also be written as follows:

$$\kappa = \frac{\frac{\binom{45}{R_1} - \frac{31}{R_1} - \frac{15}{R_1^{\beta}} \left(1 + \frac{15}{R_1^{\beta}} + \frac{31}{R_1} - \frac{45}{R_1} R_1\right)}{\frac{15}{R_1^{\alpha}} \left(1 + \frac{15}{R_1^{\alpha}} + \frac{15}{R_1^{\beta}} + \frac{31}{R_1} - \frac{45}{R_1} R_1\right)} - \frac{\binom{45}{R_2} - \frac{31}{R_2} - \frac{15}{R_2^{\beta}} \left(1 + \frac{15}{R_2^{\beta}} + \frac{31}{R_2} - \frac{45}{R_2} R_2\right)}{\frac{15}{R_2^{\beta}} - \frac{15}{R_1^{\beta}} - \frac{15}{R_1^{\beta}} - \frac{15}{R_1^{\beta}} \frac{15}{R_1^{\beta}} -$$

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$$\begin{array}{l} & \left(\frac{4^{5}R_{1} - 3^{1}R_{1} - 1^{5}R_{1}^{0}\right)\left(1 + 1^{5}R_{1}^{0}\right)}{1^{5}R_{1}^{2}(1 + 1^{5}R_{2}^{0} + 1^{5}R_{2$$

pyisotopomer

## **3.1 Preparation and analysis of dissolved N<sub>2</sub>O reference materials**

A series of dissolved N<sub>2</sub>O reference materials (Table 1) were prepared and analyzed in both Lab 1 and Lab 2. Reference materials were prepared by filling 160-mL glass serum bottles (Wheaton) with de-ionized water and removing a 4-mL headspace (Lab 1) or 10 to 20-mL headspace (Lab 2), then capped with a gray butyl rubber septum (National Scientific) and sealed with an aluminum crimp seal. These bottles were purged with helium for 90 minutes at yields a minimum flow rate of 100 mL/min to remove all background N<sub>2</sub>O. The purged bottles were then injected with 2 to 43 nmol N<sub>2</sub>O to give N<sub>2</sub>O concentrations of 13 to 275 nM (Lab 1) or 1 to 60 nmol N<sub>2</sub>O to give N<sub>2</sub>O concentrations of 6 to 427 nM (Lab 2) in a matrix of He or synthetic air (Table 1) using a gas-tight syringe. Reference materials prepared in Lab 1 were preserved with 100 µL saturated mercuric chloride (HgCl<sub>2</sub>) solution; those prepared in Lab 2 contained no added preservative. For Lab 1, atmosphere-equilibrated seawater was prepared by filtering surface seawater (collected in Half Moon Bay, CA) through a 0.22 mm Sterivex filter, allowing it to undergo static equilibration with outdoor air for three days, then re-filtering into 160-mL serum bottles, removing a 1-mL headspace, and preserving with 100 µL saturated mercuric chloride solution. For Lab 2, atmosphere-equilibrated reference materials were prepared by purging either de-ionized water or a sodium chloride solution with helium, allowing it to undergo static equilibration with outdoor air for three days, filling into 160-mL serum bottles, and removing a 10-mL headspace. While we were able to correct for these differences in reference material preparation, future intercalibration efforts should aim to prepare reference materials the same way in participating laboratories. In addition, the linearity relationships should be determined from analyzing different amounts of gaseous reference materials, to separate any artifacts due to preparation and extraction of dissolved N<sub>2</sub>O reference materials from the abundance linearity of the isotope ratio mass spectrometer itself. 

Reference materials were run in the same format as samples to account for any potential fractionation associated with the purge-and-trap system. The magnitude of such fractionation was quantified for Lab 1 by running aliquots of the pure N<sub>2</sub>O reference tank in sample format; this test yielded offsets of  $(0.22\pm0.52)$  % for  $\delta(^{15}N^{\text{bulk}})$  and  $(0.16\pm0.62)$  % for  $\delta(^{18}O)$  vs. the reference tank injection (see Supplementary text S3 for a full discussion of potential fractionation effects in the purge-and-trap system). 

The reference gases were calibrated independently by J. Mohn (EMPA; mini-QCLAS aerodyne) or S. Toyoda (Tokyo Tech; IRMS), except for one internal standard used by Lab 1 (B6; Table 1). The  $\delta(^{17}\text{O})$  values for each gas were calculated assuming a mass-dependent relationship between  $^{17}R$  and  $^{18}R$  (eqn. 5).

Reference gases and samples were measured on Thermo Finnigan DELTA V Plus isotope ratio mass spectrometers (IRMS; Thermo Fisher Scientific, Waltham, MA) in Labs 1 and 2. Each IRMS had Faraday cups configured to simultaneously measure m/z 30, 31, 44, 45, and 46. The measurements from the Lab 1 DELTA V Plus were made under normal operating conditions, using an ionization energy of 124 eV, accelerating voltage of 3 kV, emission current of 1.50 mA, and box and trap currents of 0.68 and 0.82 mA, respectively. The measurements from the Lab 2 DELTA V Plus were made under normal operating conditions, using an ionization energy of 110 eV, accelerating voltage of 3 kV, emission current of 1.00 mA, and box and trap currents of 0.45 and 0.55 mA, respectively. Reference materials and samples were analyzed on custom purge-and-trap systems coupled to each IRMS, which was run in continuous flow mode<sup>66</sup> (Table 1). The two systems had slight differences in the purge-and-trap method: in Lab 1, liquid from each sample bottle was transferred under helium pressure to a sparging column to extract the 

dissolved gases<sup>67</sup>; in Lab 2, each sample was extracted by purging directly from the bottle. The effects of these differences are discussed further in Results and Discussion. 

#### **3.2 Data corrections**

### 3.2.1 Linearity relation

The measured ion current ratios 31/30, 45/44, and 46/44 of each sample peak were divided by those of the working reference peak. This produced three molecular isotope delta values  ${}^{31}\delta+1$ ,  ${}^{45}\delta+1$ , and  ${}^{46}\delta+1$ , where  $\delta = R_s/R_{wr} - 1$ , with the subscripts "s" and "wr" denoting sample and working reference, respectively (Figure 1, Step 5). 

The  $\delta$  values were corrected for the effect of peak size<sup>33</sup>. For Lab 1, this was accomplished by running six reference materials (reference gases S2, B6, A01, CA06261, 90454, and 94321; Table 1) in size series ranging from 2-43 nmol N<sub>2</sub>O. For Lab 2, three reference materials (CA06261, 53504, and CA08214) were run in size series ranging from 1-60 nmol N<sub>2</sub>O (Figure 1, Step 6).

To obtain a single size correction slope from multiple size series, we used the dummy-variable method of combining regressions<sup>68</sup>. The dummy variable method is an improvement over simply averaging each individually calculated slope because it implicitly weighs each size series by its informativeness, producing a slope that is more likely to reflect the overall linearity behavior of the instrument<sup>68</sup>. For a given material, each measured  $\delta$ +1 is a linear function of its peak area (A) plus an intercept  $(y_1 + y_2D_2 + y_3D_3)$ : 

$$\delta + 1 = \hat{\beta}A + \gamma_1 + \gamma_2 D_2 + \gamma_3 D_3$$
(16)

where  $\hat{\beta}$  represents the regression coefficient for a particular peak area (for m/z 31,45, or 46), obtained by multiple linear regression. The intercept for reference material 1 is  $\gamma_1$ .  $D_2$  and  $D_3$  are 'dummy variables' to adjust  $\gamma_1$  by an appropriate intercept for reference material 2 ( $\gamma_1 + \gamma_2$ ) and reference material 3 ( $\gamma_1 + \gamma_3$ ). Thus, for reference material 1,  $D_2 = D_3 = 0$ ; for reference material 2,  $D_2 = 1$  and  $D_3 = 0$ ; for reference material 3,  $D_2 = 0$  and  $D_3 = 1$ . These dummy variables allow us to obtain one slope for each isotope delta from multiple datasets accounting for differences in intercept, with each reference material weighted by its spread in the x-axis range. Thus, slopes  $\hat{\beta}_{31}, \hat{\beta}_{45}$ , and  $\hat{\beta}_{46}$  were calculated for  ${}^{31}\delta + 1, {}^{45}\delta + 1$ , and  ${}^{46}\delta + 1$ , respectively, each using eqn. (16). To normalize measured values of  $\delta$ +1 to a common peak area, we first calculated the  $(\delta+1)_0$  that would be measured at m/z 44 peak area  $A_0$ : 

$$(\delta + 1)_0 = \hat{\beta}(A_0) + \gamma_1 + \gamma_2 D_2 + \gamma_3 D_3 \tag{17}$$

Note that  $(\delta+1)_0$  is still a function of  $\hat{\beta}$ , the intercepts  $\gamma_1, \gamma_2, \gamma_3$ , and the dummy variables  $D_2$  and  $D_3$ . To obtain the difference  $\delta_0 - \delta$  from the measured m/z 44 peak area A, we subtract eqn. (17) from eqn. (16), to obtain: 

$$(\delta + 1)_0 - (\delta + 1) = \hat{\beta}(A_0 - A)$$

In this case, the size-corrected molecular isotope ratio,  $\delta_0$ , for each sample with measured  $\delta$  and peak area A is given by: 

$$(\delta + 1)_0 = \hat{\beta}(A_0 - A) + (\delta + 1)$$
(18)

Eqn. (18) is simply a function of the slope  $\hat{\beta}$ , the measured (A) and target (A<sub>0</sub>) m/z 44 peak areas, and the measured  $\delta$ . Thus, eqn. (18) can be applied across a range of peak areas and  $\delta$  values to normalize these  $\delta$  values to a common peak area. Using this method, we normalized the measured  ${}^{31}\delta+1$ ,  ${}^{45}\delta+1$ , and  ${}^{46}\delta+1$  of each sample to a peak area (A<sub>0</sub>) of 20 Vs (volt seconds), equivalent to 10 nmol N<sub>2</sub>O on the Lab 1 IRMS (Figure 1, Step 7). We note that the linearity 

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correction estimated here implicitly assumes that samples and reference materials are affected by the same relative blank size. 

3.2.2 Scale normalization and calculation of  ${}^{17}R$ 

After applying the linearity correction, a scale normalization was applied to  ${}^{45}\delta$  and  ${}^{46}\delta$ (Figure 1, Step 8). The scale normalization for  ${}^{45}\delta$  and  ${}^{46}\delta$  needs to be carried out before the scrambling correction (which is essentially a scale normalization of  ${}^{31}\delta$ ); otherwise, the wrong 

bulk  ${}^{15}N/{}^{14}N$  and  ${}^{18}O/{}^{16}O$  ratios are implied. Furthermore, while the  $\gamma$  and  $\kappa$  calculations constrain the differences between  $\delta({}^{15}N^{\alpha})$  and  $\delta({}^{15}N^{\beta})$ , their absolute values are governed by  $\delta$ <sup>(15</sup>N<sup>bulk</sup>), necessitating that the "correct", normalized value of <sup>45</sup> $\delta$  be input to the scrambling equations. This scale normalization is a replacement for any scale normalization or offset correction to the final output  $\delta$  values, such as the one-point and two-point offset corrections calculated and applied in Mohn et al. (2014). 

A scale normalization was calculated for each run included in the intercalibration exercise. Since assigned values of  ${}^{45}R$  and  ${}^{46}R$  for each reference gas were unavailable, assigned  $^{45}R$  and  $^{46}R$  were calculated from assigned  $^{15}R^{\alpha}$ ,  $^{15}R^{\beta}$ , and  $^{18}R$  and eqns. (3), (4), and (5) (Table 1), assuming  ${}^{17}R_{VSMOW} = 0.0003799^{69}$  and  ${}^{18}R_{VSMOW} = 0.0020052^{58}$ . Next, the assigned  ${}^{45}R$  and  ${}^{46}R$  for each reference gas were divided by the known  ${}^{45}R$  and  ${}^{46}R$  of the direct N<sub>2</sub>O reference injection to obtain assigned  ${}^{45}\delta$  and  ${}^{46}\delta$  for each reference material. Then, these assigned  ${}^{45}\delta$  and  ${}^{46}\delta$  values were compared to measured  ${}^{45}\delta$  and  ${}^{46}\delta$  values, and scale normalization coefficients were calculated following the logarithmic scale normalization outlined in Kaiser et al. (2007):  $\ln(1 + {}^{45}\delta^{n}) = m\ln(1 + {}^{45}\delta) + b$ 

where  ${}^{45}\delta^n$  is the normalized  ${}^{45}\delta$ , "m" is the slope of the regression of  $\ln(1+{}^{45}\delta^n)$  vs.  $\ln(1+45\delta)$ , and "b" is the intercept (and likewise for  $46\delta$ ). From this regression, the normalized  $\delta$ values can be obtained:

$$1 + {}^{45}\delta^{n} = e^{b}(1 + {}^{45}\delta)^{m}$$
(19)

For the working reference, the values of  ${}^{45}\delta$  and  ${}^{45}\delta^{n}$  are equal to zero, so the intercept b should be equal to or very close to zero. The benefit of the logarithmic normalization is that, unlike a linear scale normalization, it is scale-invariant<sup>62</sup>: essentially, the logarithmic scale normalization does not skew the data towards extremely high or low values, and instead equally weights all data points<sup>62</sup>.

Next, a measured  ${}^{18}R$  was derived from the scale-normalized  ${}^{45}R$  and  ${}^{46}R$  for each sample and reference material (Figure 1, Step 8). The size correction and scale normalization were carried out in the pyisotopomer spreadsheet template; the  ${}^{18}R$  derivation from the scale-normalized  ${}^{45}R$  and  ${}^{46}R$  was the first step accomplished by the pyisotopomer code<sup>65</sup>. Deriving  ${}^{18}R$ was accomplished by assuming a mass-dependent relationship between  ${}^{17}R$  and  ${}^{18}R$  (eqn. 5) and  ${}^{15}R^{\alpha} = {}^{15}R^{\beta} = {}^{15}R^{\text{bulk}}$ . These terms are then substituted into eqns. (3) and (4) to yield: 

$${}^{45}R = 2^{15}R^{\text{bulk}} + {}^{17}R_{\text{VSMOW}} \left(\frac{{}^{18}R}{{}^{18}R_{\text{VSMOW}}}\right)^{\beta} \left(\Delta^{17}0 + 1\right)$$
(20)

$${}^{46}R = {}^{18}R + 2{}^{15}R^{\text{bulk}} \left[ {}^{17}R_{\text{VSMOW}} \left( \frac{{}^{18}R}{{}^{18}R_{\text{VSMOW}}} \right)^{\beta} \left( \Delta^{17}O + 1 \right) \right] + \left( {}^{15}R^{\text{bulk}} \right)^{2}$$
(21)

Note that the slope  $\beta$  of the mass-dependent relationship between  ${}^{17}R$  and  ${}^{18}R$  is an adjustable parameter in the code (default: 0.516), and  $\Delta^{17}$ O for each reference material may be 

1		
2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17	323 324 325 326 327 328 329 330 331 332 333 334 335	entered in the data correction template and subsequently accounted for in this correction (default: 0 ‰). Eqns. (20) and (21) were then solved for <sup>18</sup> <i>R</i> and <sup>15</sup> <i>R</i> <sup>bulk</sup> to obtain an estimated <sup>18</sup> <i>R</i> and <sup>15</sup> <i>R</i> <sup>bulk</sup> for each sample and reference material, and <sup>17</sup> <i>R</i> was calculated from <sup>18</sup> <i>R</i> according to eqn. (5). The resulting <sup>18</sup> <i>R</i> , <sup>17</sup> <i>R</i> , and <sup>15</sup> <i>R</i> <sup>bulk</sup> were used in the scrambling calculation. They contain an error due to the assumption that <sup>15</sup> <i>R</i> <sup><math>\alpha</math></sup> = <sup>15</sup> <i>R</i> <sup><math>\beta</math></sup> = <sup>15</sup> <i>R</i> <sup>bulk</sup> , although the magnitude of this error should be small <sup>62</sup> . Later, the isotopomer calculation solves for <sup>15</sup> <i>R</i> <sup><math>\alpha</math></sup> and <sup>15</sup> <i>R</i> <sup><math>\beta</math></sup> separately and thus corrects this error. In the intercalibration exercise, values of <i>m</i> and <i>b</i> were calculated from the slopes of assigned <sup>45</sup> $\delta$ <sup>a</sup> vs. measured <sup>45</sup> $\delta$ and assigned <sup>46</sup> $\delta$ <sup>a</sup> vs. measured <sup>46</sup> $\delta$ from the reference materials in each run. These runs took place in February 2021 for Lab 1 and August 2020 and November 2020 for Lab 2. Combined, the scale normalization and size correction should account for any size- or isotope-ratio dependent effects, including those of a blank, linearity, or fractionation in the GasBench.
18 19 20 21 22 23	336 337 338 339 340	<b>3.2.3 Calculating</b> ${}^{31}R_{\rm m}$ of the direct N <sub>2</sub> O reference injection We used the same scrambling coefficients for the working reference gas as for the samples. We recommend that the user calculates the ${}^{31}R$ of the direct reference injection ( ${}^{31}R_{\rm wr}$ in eqns. 14 and 15) with the following sequence of steps: 1) calculate ${}^{31}R_{\rm wr}$ from eqn. (10) with
24 25 26	341 342	either $\gamma = \kappa = 0.1$ , which reflects commonly reported values <sup>36,50,54</sup> , or an <i>a priori</i> estimate, if available (Figure 1, Step 9); 2) use that <sup>31</sup> $R_{wr}$ to correct data from two reference materials and
27	343	from those reference materials, obtain $\gamma$ and $\kappa$ from eqns. (11) and (12) (Figure 1, Step 10); 3)
28 29	344	use these updated $\gamma$ and $\kappa$ to re-calculate ${}^{31}R_{wr}$ from eqn. (10) (Figure 1, Step 11). The input $\gamma$ and
30	345	$\kappa$ (used to calculate <sup>31</sup> $R_{wr}$ ) and output $\gamma$ and $\kappa$ (calculated from paired reference materials) should
31 32 33	346 347	converge quickly, so one iteration of this process should be sufficient. This value of ${}^{31}R_{\rm wr}$ can then be used to convert ${}^{31}\delta$ to ${}^{31}R_{\rm s}$ . The user should also note that there are likely to be multiple
34 35	348	pairings of input and output $\gamma$ and $\kappa$ that will consistently yield indistinguishable delta values.
36 37	349 350	3.2.4 IRMS scrambling calibration and isotopomer calculation
38 39	351	The "Scrambling" function of pyisotopomer was used to calculate $\gamma$ and $\kappa$ algebraically
40	352	from all possible pairings of reference materials CA08214 and 53504 measured on a given IRMS
41 42	353	(Lab 1 or Lab 2; Figure 1, Step 13). The reference materials CA08214 and 53504 were chosen
42	354 355	because of their 113 ‰ $\delta(^{15}N^{sp})$ difference (see Results and Discussion for a description of how to choose reference material pairings), as well as the range of $\delta(^{15}N^{\alpha})$ , $\delta(^{15}N^{\beta})$ , $\delta(^{15}N^{bulk})$ , and
44	356	$\delta(^{18}\text{O})$ spanned by the two reference materials, which represent values found typically in
45 46	357	culture <sup>52,70</sup> and nature <sup>26,31</sup> . One-week running averages of $\gamma$ and $\kappa$ were calculated to smooth their
40 47	358	variation and used to obtain position-dependent $\delta$ values for unknowns and reference materials
48	359	run as unknowns for quality control (CA06261, S2, B6, and atmosphere-equilibrated seawater),
49 50	360	using the "Isotopomers" function of pyisotopomer (Figure 1, Step 14).
51	361	For comparison, this exercise was repeated, calculating $\gamma$ and $\kappa$ iteratively with the least
52 53	362	squares optimization (Figure 1, Step 12). The mean algebraic $\gamma$ and $\kappa$ from the paired reference
54 55 56	363 364	materials CA08214 and 53504 was used as the initial guess for the least squares solver. In this case, reference materials CA08214 and CA06261 were used to calculate the least squares $\gamma$ and
57 58		

K because these reference materials are close in their calibrated isotopomer values to natural abundance unknowns. As above,  $\gamma$  and  $\kappa$  were combined into a one-week running average; these running averages of  $\gamma$  and  $\kappa$  for each system were used to obtain position-dependent  $\delta$  values for reference materials and unknowns in the intercalibration exercise (Figure 1, Step 14). The analytical precisions of  $\delta(^{15}N^{\alpha})$ ,  $\delta(^{15}N^{\beta})$ ,  $\delta(^{15}N^{sp})$ ,  $\delta(^{15}N^{bulk})$ , and  $\delta(^{18}O)$  produced by each method are presented in the Results and Discussion. N<sub>2</sub>O amounts were obtained from the m/z 44 peak area and instrument N<sub>2</sub>O sensitivity<sup>67</sup>. To obtain the conversion factor between peak area and amount of  $N_2O$ , the peak areas for reference material amounts from 1 to 40 nmol N<sub>2</sub>O were recorded. Standard deviations for inferred N<sub>2</sub>O amounts of replicate unknown samples were 0.07 nmol for Lab 1, and 0.19 nmol for Lab 2. All data corrections are described in the README documents associated with pyisotopomer on the Python Package Index<sup>65</sup>. 3.3 Lake water unknowns To validate the scrambling calibration, samples of unknown isotopic composition were collected from Lake Lugano, Switzerland in July 2020 and analyzed separately by both Lab 1 and Lab 2. The samples were collected at depths of 10 and 90 meters, including six replicate

bottles at each depth. Samples were collected into 160-mL glass serum bottles (Wheaton),

overflowing each bottle twice, closing bubble-free, and removing liquid to form a 10-mL

headspace comprised of air. Based on the northern hemisphere monthly mean tropospheric N<sub>2</sub>O

mole fraction when the samples were collected in July, 2020<sup>71</sup>, an atmospheric headspace of this

volume would have contained 0.13 nmol N<sub>2</sub>O. For Lab 2, where the full amount of N<sub>2</sub>O in the

overestimation of the amount of N<sub>2</sub>O in the sample<sup>71</sup>. For Lab 1, where 2 mL sample liquid is

left behind post-analysis, equilibration the 10-mL headspace during sample storage results in

depending on its concentration. In both cases, these errors are similar to the analytical precision

of the N<sub>2</sub>O amount measurement. Each sample was capped with a gray butyl septum (National

µL saturated mercuric chloride solution and stored at lab temperature (20-22°C). The isotope

fractionation associated with N<sub>2</sub>O partitioning, defined as the isotope ratio of the gas phase

divided by the isotope ratio of the liquid phase,  $({}^{15}\varepsilon = -0.7 \text{ }_{\infty}, {}^{18}\varepsilon = -1.1 \text{ }_{\infty}, 298.2 \text{ K})$  falls

groups of three replicate bottles to be measured by Lab 1 and Lab 2, respectively.

within the analytical uncertainty<sup>72</sup>. The six replicate bottles at each depth were split into two

Scientific) and sealed with an aluminum crimp seal. Samples were promptly preserved with 100

either an underestimate (0.12 nmol) or overestimate (0.10 nmol) of N<sub>2</sub>O in the sample,

sample is measured, incorporation of the headspace into the measurement results in a 0.13 nmol

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## **4.1 Linearity relation**

Linearity relations were calculated using the dummy variable method described in
Section 3.2.1 and applied to the intercalibration data as follows. A linearity relation was
determined for Lab 1 in February 2021 (Figure 2a-c) and applied to lake water samples run in
Lab 1 and reference materials prepared and run in Lab 1. Reference materials prepared in Lab 2
but run in Lab 1 exhibited statistically distinct linearity slopes from those both prepared and run
in Lab 1; thus, a separate linearity relation was applied to these reference materials (but not to the
lake water samples) (Figure 2d-f). A linearity relation was determined for Lab 2 in May 2020

2		
3 4 5 6 7 8	410 411 412 413 414	(Figure 2g-i) and applied to lake water samples and reference materials run in Lab 2. As previously observed <sup>73</sup> , for each linearity relation, the slopes of the fits for individual reference materials were identical within error. The linearity correction reduced the spread of measured molecular isotope ratios across size series of each given reference material (Figure S2).
9	415	4.2 IRMS scrambling calibration
10 11	416	For both labs, the "algebraic" solution produced reasonable values of $\gamma$ and $\kappa$ (i.e.,
12	417	between 0 and 1) for reference material pairings involving the reference material 53504 ( $\delta$ ( <sup>15</sup> N <sup>sp</sup> )
13 14	418	= -93 ‰). The mean $\gamma$ and $\kappa$ calculated for Lab 1 from reference materials 53504 and CA08214
15 16	419	were 0.174±0.022 and 0.083±0.022, respectively (Table S2). In August 2020, the mean $\gamma$ and $\kappa$
16 17	420	calculated for Lab 2 from the same two reference materials were 0.095±0.011 and 0.091±0.010,
18 19	421	respectively (Table S2). In November 2020, $\gamma$ and $\kappa$ for Lab 2 were slightly different but within
20	422	$1\sigma$ of the values measured in August 2020 (0.091±0.013 and 0.086±0.013, respectively; Table
21 22	423	S2). Other reference materials paired with 53504 produced similar values of $\gamma$ and $\kappa$ . The
22 23 24 25 26 27	424 425 426 427	difference $\gamma - \kappa$ was also consistent for reference material pairings with 53504: for Lab 1, $\gamma - \kappa$ was 0.090-0.091, and for Lab 2, it was 0.003-0.005 (Table S2). For pairings with 53504, the $\delta(^{15}N^{sp})$ difference between both reference materials was greater than 100 ‰. Pairs of reference materials with smaller $\delta(^{15}N^{sp})$ differences produced more
28 29 30	428 429	variable $\gamma$ and $\kappa$ values with the algebraic solution, which sometimes fell outside the physically plausible range between 0 and 1. For example, in Lab 1, the pairing of CA06261 and CA08214
31 32 33 34	430 431 432	produced $\gamma$ and $\kappa$ values of 0.01±0.23 and -0.08±0.23, respectively. In this case, the measurement uncertainty was too large — and the $\delta(^{15}N^{sp})$ values too close — for the scrambling coefficients to be adequately determined. What matters, however, is that the difference between $\gamma$
35 36	433 434	and $\kappa$ is accurate; as the results show, the absolute values are less important (and can even be negative, greater than 1, or otherwise "unphysical").
37 38	435	To understand the uncertainty in $\gamma$ and $\kappa$ calculated from equations 11 and 12, we define a
39 40 41 42	436 437	variable <i>d</i> , which allows us to express the analytical solution for $\gamma$ and $\kappa$ (eqns. 13a and 13b) in terms of $\delta(^{15}N^{sp})$ , $\delta(^{15}N^{sp})$ , and $\delta(^{15}N^{sp})$ : $d = \frac{\binom{1^5R^{\beta} + {}^{31}R - {}^{45}R}{\binom{1}{5}R^{\alpha} + {}^{15}R^{\beta} + {}^{31}R - {}^{45}R}}$ (22)
43 44		$d = \frac{1}{15R_{\text{atm}}\left(1 + {}^{15}R^{\alpha} + {}^{15}R^{\beta} + {}^{31}R - {}^{45}R\right)} $ (22)
45 46 47 48 49 50 51	438 439 440 441	The value of <i>d</i> is similar for all samples and reference gases run on a given IRMS and depends primarily on the difference ${}^{31}R - {}^{45}R$ . Using $\delta$ notation, i.e., $\delta({}^{15}N) = {}^{15}R/{}^{15}R_{atm} - 1$ , and dropping the label " ${}^{15}N$ " for brevity, eqns. (13a) and (13b) can be written as follows: $\frac{d_2}{1 + \delta_2^{\alpha}} - \frac{d_1}{1 + \delta_1^{\alpha}} - \frac{d_2}{1 + \delta_2^{\alpha}} - \frac{d_1}{1 + \delta_1^{\alpha}}$
52 53		$\kappa = \frac{\overline{1+\delta_2^{\alpha}} - \overline{1+\delta_1^{\alpha}}}{1+\delta_2^{\beta}} = \frac{\overline{1+\delta_2^{\alpha}} - \overline{1+\delta_1^{\alpha}}}{\delta_1^{\text{sp}} - \overline{\delta_2^{\text{sp}}}} $ (23a)
54 55 56 57		$\frac{1+\delta_2}{1+\delta_2^{\alpha}} - \frac{1+\delta_1}{1+\delta_1^{\alpha}} - \frac{\delta_1}{1+\delta_1^{\alpha}} - \frac{\delta_2}{1+\delta_2^{\alpha}}$
58		
59 60		pyisotopomer 1: http://mc.manuscriptcentral.com/rcm

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459 
$$\sigma_{\kappa}^2 \approx 2 \left(\frac{\sigma^{31}_R}{^{15}_R \alpha}\right)^2 \left(\frac{1}{\delta_1^{\text{sp}} - \delta_2^{\text{sp}}}\right)^2$$
460 or

$$\sigma_{\kappa} \approx \sqrt{2} \frac{\sigma(^{31}R) \quad 1}{^{15}R^{\alpha} \left|\delta_{1}^{\rm sp} - \delta_{2}^{\rm sp}\right|} \tag{24a}$$

461 Similarly, for  $\gamma$ :

$$\sigma_{\gamma} \approx \sqrt{2} \frac{\sigma^{(31R)}}{^{15}R^{\beta}} \frac{1}{\left|\delta_{1}^{\text{sp}} - \delta_{2}^{\text{sp}}\right|}$$
(24b)

where  $\sigma({}^{31}R)/{}^{15}R$  can be approximated by the measurement uncertainty in  ${}^{31}\delta$  and  $|\delta^{\text{sp}} - \delta^{\text{sp}}|$  is the absolute value of the difference in assigned site preferences between the two reference materials. This means that for a measurement uncertainty in <sup>31</sup> $\delta$  of 1 ‰ and a  $\delta$ (<sup>15</sup>N<sup>sp</sup>) difference of 10 % between the two reference materials,  $\gamma$  and  $\kappa$  would have absolute uncertainties of 0.14. This uncertainty translates into a relative uncertainty of about 30 % for the  $\delta$ <sup>(15</sup>N<sup>sp</sup>) value of an unknown sample – far too high for practical applications (Supplementary text S4). A  $\delta$ (<sup>15</sup>N<sup>sp</sup>) difference of 100 ‰ would give a more useful absolute uncertainty of 0.014 for  $\gamma$  and  $\kappa$ . These theoretical uncertainties are reflected in the experimental data. For Lab 1, the reference materials 53504 ( $\delta$ (<sup>15</sup>N<sup>sp</sup>) = -92.73 ‰) and CA08214 ( $\delta$ (<sup>15</sup>N<sup>sp</sup>) = 20.54 ‰) yielded  $\gamma$  =  $0.174 \pm 0.022$  and  $\kappa = 0.083 \pm 0.022$ . The standard deviation of <sup>31</sup> $\delta$  was 1.89 ‰ (*n* = 12). This produces an estimated uncertainty in y and  $\kappa$  of  $\sqrt{2(1.89 \%)}/(113.27 \%) = 0.024$ , which agrees well with the experimental data. Similarly, reference materials 53504 and CA06261 ( $\delta$ (<sup>15</sup>N<sup>sp</sup>) = 27.07 ‰) vielded  $\gamma = 0.163 \pm 0.018$  and  $\kappa = 0.073 \pm 0.018$ . The standard deviation of <sup>31</sup> $\delta$  was 1.58 ‰ (n = 10), and the  $\delta(^{15}N^{sp})$  difference was 119.80 ‰. This produced an estimated uncertainty in  $\gamma$  and  $\kappa$  of  $\sqrt{2(1.58 \%)}/(119.80 \%) = 0.019$ , also in line with the uncertainties in  $\gamma$  and  $\kappa$ . Rearranging eqns. (24a) and (24b), we obtain expressions for the required  $|\delta^{\text{sp}} - \delta^{\text{sp}}|$  to obtain a target level of uncertainty ( $\sigma$ ) in  $\gamma$  and  $\kappa$ , given the measurement uncertainty in <sup>31</sup>*R*:  $\left|\delta_{1}^{\mathrm{sp}}-\delta_{2}^{\mathrm{sp}}\right|=\sqrt{2}\frac{\sigma(^{31}R)\,1}{^{15}R^{\alpha}\sigma_{\kappa}}$ (25a) $\left|\delta_1^{\rm sp} - \delta_2^{\rm sp}\right| = \sqrt{2} \frac{\sigma(^{31}R) \, 1}{^{15} n^{\beta} \sigma_{\gamma}}$ (25b) Assuming  $\sigma({}^{31}R)/{}^{15}R^{\alpha} \approx \sigma({}^{31}R)/{}^{15}R^{\beta} \approx \sigma({}^{31}\delta)$ , we obtain:  $\left|\delta_1^{\rm sp} - \delta_2^{\rm sp}\right| = \sqrt{2}\sigma(^{31}\delta)\frac{1}{\sigma_{\rm vr}}$ (26)

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3 4	483	where $\sigma(^{31}\delta)$ is the $^{31}\delta$ measurement uncertainty in per mil, and $\sigma_{\gamma\kappa}$ is the target absolute
5 6	484 485	uncertainty in $\gamma$ and $\kappa$ . For example, with a measurement uncertainty of 1 ‰ in <sup>31</sup> $\delta$ , the $\delta$ ( <sup>15</sup> N <sup>sp</sup> ) values of the two reference materials must differ by at least 141 ‰ to achieve an absolute
7 8	486	-
9	480 487	uncertainty in $\gamma$ and $\kappa$ of 0.01. Based on these results, we recommend calculating $\gamma$ and $\kappa$ from reference materials with a large $\delta$ ( <sup>15</sup> N <sup>sp</sup> ) difference, as estimated from eqn. (26).
10	488	As an alternative to the algebraic solution, a least squares optimization can be used to
11 12	489	find a solution for $\gamma$ and , although that solution may find a local optimum rather than a global
13	490	optimum. The user can select a least squares optimization instead of the algebraic solution with
14 15	491	the "method" keyword argument to pyisotopomer's Scrambling function. The least squares
16	492	optimization smooths measurement uncertainty, making it useful for for fitting repeat
17	493	measurements of reference materials to a single pair of "best" values for $\gamma$ and $\kappa$ . Its disadvantage
18 19	494	is that, unlike the algebraic solution, the least squares optimization depends on the initial guess
20	495	for $\gamma$ and $\kappa$ . Using data from reference materials CA06261 and CA08214, a range of initial
21 22	496	guesses from $\gamma = \kappa = 0.000$ to $\gamma = \kappa = 0.200$ produced a range of least squares solutions, from $\gamma =$
23	497	0.090 and $\kappa = 0.000$ to $\gamma = 0.269$ and $\kappa = 0.183$ (Figure S3). Despite this range of $\gamma$ and $\kappa$ ,
24 25	498	however, the least squares optimization produced a consistent $\gamma - \kappa$ of 0.09. As shown in Section
26 27	499	4.4, $\gamma - \kappa$ governs the accuracy of $\delta(^{15}N^{sp})$ far more than the individual values of $\gamma$ and $\kappa$ .
28	500	Given an accurate initial guess, the least squares optimization will find a minimum at or
29	501	close to this initial guess, even for reference material pairings close in their $\delta(^{15}N^{sp})$ . For
30 31	502	example, when we used the algebraic $\gamma$ and $\kappa$ from reference materials CA08214 and 53504 as an
32	503	initial guess, the least squares optimization produced similar $\gamma$ and $\kappa$ for a variety of reference
33 34	504	material pairings (Table S2). Furthermore, for the same initial guess, the least squares
35	505 506	optimization finds different solutions for the Lab 1 and Lab 2 instruments, even for reference material pairings close in their $\delta(^{15}N^{sp})$ (Table S3). This demonstrates that, depending on the
36	507	measurement precision at the time, the least squares optimization searches an appropriately wide
37 38	508	solution space to resolve large differences in instrument behavior.
39 40	509	If the first-time user wishes to obtain accurate individual values of $\gamma$ and $\kappa$ , we
41	510	recommend obtaining reference materials different enough in their $\delta(^{15}N^{sp})$ to calculate $\gamma$ and $\kappa$
42 43	511	with the algebraic solution. If the user wishes to take advantage of the smoothing of the least
43 44	512	squares optimization, this algebraic $\gamma$ and $\kappa$ can then be used as the initial guess for the least
45	513	squares solver.
46 47	514	We also recommend that the user test the accuracy of the least squares $\gamma$ and $\kappa$ by
48 49	515	plugging $\gamma$ and $\kappa$ back into eqn. (10) and comparing the result to the measured <sup>31</sup> <i>R</i> for each
50	516	reference material. The two ${}^{31}R$ values should match. pyisotopomer ${}^{65}$ performs this calculation
51	517	automatically and outputs the difference as a $\delta$ value:
52 53		${}^{31}\delta^{\text{error}} = \frac{{}^{31}R_{\text{calculated}}}{-1} $ (27)
54		${}^{31}\delta^{\text{error}} = \frac{{}^{31}R_{\text{calculated}}}{{}^{31}R_{\text{measured}}} - 1 $ (27)
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3 4 5	519 520	where ${}^{31}R_{\text{calculated}}$ is calculated by plugging the least squares $\gamma$ and $\kappa$ into eqn. (10), and ${}^{31}R_{\text{measured}}$ represents the measured ${}^{31}R$ for each reference material. In the intercalibration
6 7 8 9 10 11 12 13	521 522 523 524 525 526 526 527	exercise, the mean of the absolute values of ${}^{31}\delta^{\text{error}}$ from least squares $\gamma$ and $\kappa$ solutions ranged from 0.27 ‰ to 0.86 ‰ (Table S2), similar in magnitude to the ${}^{31}\delta$ analytical uncertainty for Labs 1 and 2 (Table S5). This indicates that the amount of error introduced by using the least squares optimization is similar to the measurement error in ${}^{31}\delta$ (Table S2, Table S5). In comparison, the ${}^{3}\delta^{\text{error}}$ introduced by the algebraic solution corresponded to values of ( ${}^{31}R_{\text{calculated}}$ $-{}^{31}R_{\text{measured}}$ ) within machine precision (Table S2).
14 15	528	4.3 Variability in fragmentation behavior
16	529	As shown above, $\gamma - \kappa$ , as opposed to the individual values of $\gamma$ and $\kappa$ , is the best
17 18	530	constrained parameter in the scrambling calculation. We show below that $\gamma - \kappa$ also has the
19 20 21 22	531 532 533	greatest impact on $\delta({}^{15}N^{\alpha})$ , $\delta({}^{15}N^{\beta})$ , and $\delta({}^{15}N^{sp})$ . $\gamma - \kappa$ is proportional to ${}^{31}\delta - {}^{45}\delta$ , and thus is a metric of an instrument's scrambling behavior. To examine the change in the fragmentation behavior of a single IRMS over time, we
23	534	compiled values of $\gamma - \kappa$ for Lab 1 from June 2018 – March 2021 (Figure 3). To equally weigh
24 25	535	each day of running the instrument, first, we calculated a daily mean $\gamma - \kappa$ , then calculated a five-
26		
27 28	536	day running average of $\gamma - \kappa$ from these daily means. The value of $\gamma - \kappa$ varied throughout the
29	537 538	time series, with a mean of 0.092±0.002. High volatility in $\gamma - \kappa$ in February-April 2019 corresponded with a period when the lab temperature was poorly controlled, with strong day-
30 31	538 539	night variation (Figure 3). During periods when the lab temperature was stable, $\gamma - \kappa$ tended to
32 33	539 540	increase as the instrument box and trap currents diverged with filament age, although no linear
33 34	541	relationship emerged.
35	542	There are several reasons why the scrambling behavior of the ion source might change
36 37	543 544	over time, as well as differing between instruments. The NO <sup>+</sup> fragment ion can be produced by one of several routes from $N_2O^+$ <sup>75,76</sup> . The pathways and associated isotope effects for the
38	545	formation of fragment ions are affected by collision frequency, the distribution of excited states,
39 40	546	and the time spent in the ion source, which suggests that ion source conditions such as vapor
41	547	pressure, ionizing energy, and accelerating voltage may all influence the fragmentation behavior
42 43	548 549	of an IRMS system <sup>54,75–78</sup> . Future work could track the effect of variation in these parameters on the fragmentation behavior of the instrument, as in Westley et al. <sup>54</sup> , which may allow for
44	550	optimization of fragmentation and scrambling in the ion source.
45 46	551	For these reasons, performing the scrambling calibration only once is insufficient to
47	552	obtain high-quality N <sub>2</sub> O isotopocule data. Instead, it is important to recalibrate an IRMS system
48 49	553 554	for scrambling on a regular basis since ion source conditions may change with time and can shift abruptly with events such as filament changes. We recommend using a running average of $\gamma$ and
49 50	555	<i>k</i> over a window corresponding to 10 pairings of reference materials, corresponding to a five-day
51 52		
52 53 54 55 56 57	556 557 558 559	window if two pairs of reference materials are run per day. If there is high volatility in $\gamma$ and $\kappa$ , as seen above in March-April 2019, it may be necessary to shorten this window, to apply scrambling corrections most appropriate to instrument conditions.
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3 1	560	4.4 Sensitivity of position-dependent $\delta$ values to uncertainty in scrambling coefficients
4 5	561	The uncertainty in $\delta({}^{15}N^{\alpha})$ , $\delta({}^{15}N^{\beta})$ , and $\delta({}^{15}N^{sp})$ associated with the uncertainty in each
6	562	scrambling coefficient is less straightforward to assess than the uncertainty in ${}^{31}R$ given by eqns.
7 8	563	(23) and (24), due to the nonlinear relationship between $\delta(^{15}N^{\alpha})$ , $\delta(^{15}N^{\beta})$ , $\gamma$ , and $\kappa$ . (see eqn. (53)
9	564 565	of Kaiser and Röckmann, 2008). A first order approximation of $\delta(^{15}N^{sp})$ is given by (supplementary text S4):
10 11 12	505	(supprementary text 54): $\delta(^{15}N^{\rm sp}) \approx \frac{2(1-\gamma+\kappa)}{1-\gamma-\kappa} (^{31}\delta - {}^{45}\delta) $ (28)
13	566	From this equation, it is apparent that $\delta(^{15}N^{sp})$ is modulated primarily by the difference $\gamma$
14 15 16	567 568	- $\kappa$ , rather than the individual values of $\gamma$ and $\kappa$ . It is also apparent that $\gamma - \kappa$ is proportional to ${}^{31}\delta$ $-{}^{45}\delta$ .
17	569	A Monte Carlo simulation can be a useful way of visualizing how $\gamma$ , $\kappa$ , and, $\gamma - \kappa$ impact
18 19	570	$\delta(^{15}N^{\alpha})$ , $\delta(^{15}N^{\beta})$ , and $\delta(^{15}N^{sp})$ . We performed two sensitivity experiments with data from Lab 1:
20	571	1) sensitivity of $\delta({}^{15}N^{\alpha})$ , $\delta({}^{15}N^{\beta})$ , and $\delta({}^{15}N^{sp})$ to $\gamma - \kappa$ ,
21 22	572	2) sensitivity of $\delta({}^{15}N^{\alpha})$ , $\delta({}^{15}N^{\beta})$ , and $\delta({}^{15}N^{sp})$ to the individual values of $\gamma$ and $\kappa$ , holding
23	573	their difference constant.
24 25	574	For the first sensitivity experiment, a Monte Carlo simulation was used to introduce
26	575	random uncertainty in the $\gamma$ and $\kappa$ values used to calculate $\delta$ values of three reference materials.
27 28	576	Based on Table S2, we chose $\gamma = 0.174$ and $\kappa = 0.083$ as central values and varied $\gamma - \kappa$ such that
29	577	the standard deviation of $\gamma - \kappa$ was equal to 10 % of the mean (0.091). For the second sensitivity
30 31	578	experiment, we modeled $\gamma$ and $\kappa$ in tandem as random numbers centered around $\gamma = 0.174$ and $\kappa =$
32 33	579	0.083, with uncertainties equal to 10 % of the mean y, and held $\gamma - \kappa$ constant at 0.091. For both
34	580	experiments, we sampled 1000 pairs of $\gamma$ and $\kappa$ , and then calculated the 1000 simulated values of
35 36	581	$\delta^{(15}N^{\alpha})$ , $\delta^{(15}N^{\beta})$ , and $\delta^{(15}N^{sp})$ for the three reference materials (CA06261, 53504, CA08214).
37	582	This analysis showed that a 10 % relative uncertainty in $\gamma - \kappa$ can lead to large variations
38 39	583	in $\delta({}^{15}N^{\alpha})$ , $\delta({}^{15}N^{\beta})$ , and $\delta({}^{15}N^{sp})$ , e.g., pooled standard deviations of 17.1-18.5 ‰ for $\delta({}^{15}N^{sp})$
40	584	(Figure 4a-c). In contrast, a 10 % relative error in $\gamma$ , keeping $\gamma - \kappa$ constant, led to pooled
41 42	585	standard deviations of 1.0-4.3 ‰ in $\delta(^{15}N^{sp})$ (Figure 4d-f). In both experiments, varying $\gamma$ and $\kappa$
43	586	produced the most variability for reference material 53504, whose $\delta(^{15}N^{sp})$ was greatest in
44 45	587	magnitude.
46	588	These results reflect the earlier conclusion that $\gamma - \kappa$ is the best constrained parameter in
47 48	589	the scrambling calculation, and, conversely, that this difference has the greatest effect on
40	590	$\delta(^{15}N^{sp})$ . Thus, we recommend regular scrambling calibrations, as assuming the wrong $\gamma - \kappa$
50	591	difference may have a significant impact on site preferences calculated from these coefficients.
51 52	592 593	4.5 Comparison of results between two IRMS laboratories
53	594	The application of pyisotopomer was tested through an intercalibration including four
54 55	595	reference materials and two Lake Lugano samples measured by two IRMS laboratories, plus two
56	596	additional reference materials run in Lab 1. Using an average $\gamma$ and $\kappa$ produced by the algebraic
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- method from the pairing of reference materials 53504 and CA08214, isotopomers were calculated for lake water unknowns, four reference materials run as unknowns for quality control, and the two reference materials used in the calibration and (Table 2). This exercise was repeated, calculating y and  $\kappa$  instead with least squares method and the pairing of reference materials CA06261 and CA08214 (Table S4). The root mean square deviation (RMSD) for each reference material was calculated by comparison to the calibrated values provided by a previous intercalibration effort<sup>56</sup> (for atmosphere-equilibrated seawater), an internal standard (B6), and four gases sourced from J. Mohn (S2, CA06261, 53504, and CA08214). Almost all isotopomer values produced by the least squares optimization (Table S4) were within error of those produced by the algebraic solution (Table 2); the latter is discussed below. The  $\delta$ <sup>(15</sup>N<sup>bulk</sup>) measured by the two labs displayed good agreement for each of the four reference materials, as well as the lake water samples. The  $\delta$ (<sup>15</sup>N<sup>bulk</sup>) RMSDs ranged from 0.2 to 0.6 ‰ (Table 2), all of which were smaller than the 0.8 ‰ presented for IRMS labs by Mohn et al., 2014). The RMSD for atmospheric  $N_2O$  was highest, at 0.6 %. For both lake water samples, the  $\delta(^{15}N^{\text{bulk}})$  values measured by Lab 1 and Lab 2 were statistically indistinguishable (Table 2; Figure S4). Likewise, the  $\delta(^{18}O)$  measured by the two labs displayed good agreement for each of the four reference materials measured by both labs, as well as the lake water samples. The  $\delta$ <sup>(18</sup>O) RMSDs were slightly greater than the 1.00 % presented for IRMS labs by Mohn et al. (2014), ranging from 0.5 ‰–1.7 ‰, with the greatest RMSD for reference material 53504 (Table 2). For the lake water unknowns, the  $\delta$ <sup>(18</sup>O) values measured by the two labs were within error of each other (Table 2; Figure S4). The  $\delta({}^{15}N^{\alpha})$  measured by the two labs also showed good agreement for reference materials CA06261, CA08214, and atmosphere-equilibrated seawater: in each case, the combined RMSD was less than 2.4 ‰ (Table 2). This is similar to the data presented in Mohn et al. (2014), who find an RMSD for  $\delta(^{15}N^{\alpha})$  for IRMS laboratories of 2.47 ‰. The  $\delta(^{15}N^{\alpha})$ measured by Lab 1 for reference material 53504  $(0.0\pm1.0 \text{ }\%)$  was lower than both the calibrated value (1.71 ‰) and the value measured by Lab 2 (1.7±1.0 ‰). The values of  $\delta$ <sup>(15</sup>N<sup> $\alpha$ </sup>) measured by the two labs for the two lake water samples, however, were within error of each other. For  $\delta({}^{15}N^{\beta})$ , the RMSDs for each reference material were of a similar order of magnitude to  $\delta({}^{15}N^{\alpha})$ . ranging from 0.2 ‰-2.1 ‰, similar to the value 2.12 ‰ reported by Mohn et al. (2014). The  $\delta(^{15}N^{\beta})$  measured by Lab 1 for the lake water unknowns was within error of that measured by Lab 2 (Table 2; Figure S4). Of note, the  $\delta(^{15}N^{\beta})$  for the lake water unknown taken at 90 m depth was -32.8 ‰ (average of measurements by Lab 1 and Lab 2), which is far more negative than
  - 42 630 most values observed previously $^{26,31}$ .

The  $\delta(^{15}N^{sp})$  values measured by the two laboratories showed larger standard deviations than the  $\delta({}^{15}N^{\alpha})$  and  $\delta({}^{15}N^{\beta})$  individually, which is to be expected, since  $\delta({}^{15}N^{sp})$  is a measure of difference between the latter two parameters. The  $\delta(^{15}N^{sp})$  RMSD values, however, were all less than 3 % for atmosphere-equilibrated seawater, 53504, and CA08214 (Table 2). This represents an improvement on Mohn et al. (2014), who find an RMSD of 4.29 % for  $\delta$ (<sup>15</sup>N<sup>sp</sup>) measured by IRMS laboratories. The  $\delta(^{15}N^{sp})$  RMSD for reference material CA06261 was greater, at 4.4 ‰, which may result from this reference material having a more negative  $\delta(^{15}N^{\alpha})$  than either of the two reference materials used in the scrambling calibration. The lake water samples showed larger offsets in  $\delta$ <sup>(15</sup>N<sup>sp</sup>) than the reference materials (Figure S4). The lake water sample from 10 m depth showed an especially large difference in  $\delta(^{15}N^{sp})$  between Lab 1 and Lab 2: Lab 1 measured a mean  $\delta(^{15}N^{sp})$  of (18.8±1.6) ‰ at this depth, while Lab 2 measured a mean  $\delta(^{15}N^{sp})$ 

642 of (21.4±2.5) ‰ (Table 2). At 90 m depth, Lab 1 measured a mean  $\delta$ (<sup>15</sup>N<sup>sp</sup>) of 52.3±1.2 ‰, and 643 Lab 2 measured a mean  $\delta$ (<sup>15</sup>N<sup>sp</sup>) of (50.9±0.5) ‰.

After size correction and scale normalization, the only consistent difference between measurements made by the two labs were differences in peak area, which may reflect differences in the setup of the purge and trap system and/or differences in instrument sensitivity. The N<sub>2</sub>O amounts (in nmol) measured in the lake water samples were also similar between the two labs involved in the intercalibration exercise, indicating that this difference in sensitivity was adequately compensated for by the peak area to amount conversion factor. In the sample taken at 10 m depth, Lab 1 found (2.97±0.04) nmol; Lab 2 found (2.31±0.09) nmol. At 90 m depth, Lab 1 found (20.46±0.37) nmol; Lab 2 found (19.82±0.01) nmol N<sub>2</sub>O. The intercalibration is expressed in terms of N<sub>2</sub>O amounts instead of concentrations to eliminate uncertainties in sample volume; all bottle volumes were the same. Thus, we conclude that differences in sample pretreatment procedure were corrected for by the size correction and scale normalization steps, leaving no residual effect on the final  $\delta$  values or N<sub>2</sub>O amounts.

## **4.6 Additional considerations**

The pyisotopomer package produces good results if each of the data preprocessing steps properly account for size- and delta-dependent effects on the measured isotope ratios  ${}^{31}\delta$ ,  ${}^{45}\delta$ , and  $^{46}\delta$ . However, it will produce spurious results under the following circumstances. Firstly, varying blanks may introduce errors due to the size correction not being applicable to samples and reference materials alike. Second, if the  ${}^{45}\delta$  and  ${}^{46}\delta$  scale normalization slope and intercept differ substantially from one and zero (such as a negative slope), there likely exists an issue with the scale normalization (such as the reference materials not spanning a wide enough range in  ${}^{45}\delta$  and <sup>46</sup> $\delta$ ). A spurious scale normalization will likewise produce errors in the final isotopocule values. Thirdly, if reference materials that are too close in their site preferences are used to determine  $\gamma$ 

and  $\kappa$  with the algebraic solution, the resulting coefficients may represent "unphysical" values (i.e., not between 0 and 1); these, however, would be inconsequential if the unknown samples have  $\delta(^{15}N^{sp})$  values close to these reference materials. Finally,  $\delta(^{17}O)$  is calculated from a mass dependent relationship with  $\delta(^{18}O)$  (the parameters of which can be adjusted with keyword arguments to the Scrambling and Isotopomers functions) unless  $\Delta(^{17}O)$  is determined separately<sup>60,62,63</sup> and entered in the data corrections template.

## 5. Conclusion: How to obtain high-quality N<sub>2</sub>O isotopocule data using pyisotopomer

Using pyisotopomer and three reference materials, one can characterize the scrambling
behavior for a given IRMS and apply those scrambling coefficients to calculate the isotopocule
values of unknown samples. To ensure high-quality results from these calculations, we provide
the following recommendations. Firstly, if reference materials with suitably distinct site

 $\begin{array}{r}
 for the form the second calculating the scrambling coefficients <math>\gamma$  and  $\kappa$  from the scrambling coefficients  $\gamma$  and  $\kappa$  from the scrambling coefficients  $\gamma$  and  $\kappa$  from the scrambling function of pyisotopomer. We offer the least squares approach as an alternative, with the following the scramble scramble

<sup>51</sup> 682 caveats: 1) The least squares solver finds a minimum close to the initial guess for  $\gamma$  and  $\kappa$ . As <sup>52</sup> 683 such, if the solver is fed an initial guess other than the absolute minimum calculated from the <sup>53</sup> 684 algebraic solution, it will find the "wrong" absolute value of  $\gamma$  and  $\kappa$ . It will, however, find the <sup>55</sup> 685 correct value of  $\gamma - \kappa$ , which has a much larger impact on calculated isotopocules. 2) Using the

1				
2 3 4 5 6 7 8 9 10 11	686 687 688 689 690 691 692	$\delta({}^{15}N^{\alpha}), \delta({}^{15}N^{\beta}), \text{ and } \delta({}^{15}N^{\text{sp}})$ to those of the as the unknowns diverge in their isotopomer guess is available, such as through a calibrat as the initial guess for the least squares solve scrambling calculation twice, using the solution	nly a small effect if the unknowns are close in their reference materials but will have a deleterious effect values from the reference materials. 3) If an initial ion with the algebraic solution, this should be used or. Otherwise, we recommend iterating through the ion from the first iteration as the initial guess for n paired reference materials daily to obtain accurat	ct
12 13 14 15 16 17 18 19 20 21 22 23	<ul> <li>693</li> <li>694</li> <li>695</li> <li>696</li> <li>697</li> <li>698</li> <li>699</li> <li>700</li> <li>701</li> <li>702</li> <li>703</li> </ul>	running average and use that average to calc Using pyisotopomer in an intercalibr recommendations, we find good agreement if different IRMS labs for both reference mate while the intercalibration results demonstrat intercalibration of $\delta(^{15}N^{sp})$ using a uniform s represents an improvement upon previous N In this paper, we demonstrate the new reference gases to the community. At present	ation exercise and implementing the above between the calibrated $\delta$ values measured by two rials and natural lake samples. We conclude that e potential for further improvement in precision, the crambling calculation (pyisotopomer) presented he 20 intercalibrations. d to support efforts to generate and distribute	ere
24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39	704 705 706 707 708 709 710 711 712 713 714 715 716	to obtain precise values of $\gamma$ and $\kappa$ with the a extremely small measurement uncertainties more calibrated N <sub>2</sub> O reference gases <sup>79</sup> , but t fully funded program is needed to produce r combined with reference materials such as U precise and accurate N <sub>2</sub> O isotopocule calibra <b>Data availability statement</b> The manuscript is prepared to comply with t pyisotopomer is available for installation via (pypi.org/project/pyisotopomer). The second Zenodo (doi.org/10.5281/zenodo.7552724).	lgebraic solution unless the user is able to achieve n <sup>31</sup> R. There have been other efforts to produce hese gases are not yet commercially available. A efference materials such as 53504, which — USGS 51 and USGS 52 — should provide users with thom. The RCMS data policy. The latest version of the Python Package index release of pyisotopomer is also available via This research was supported by U.SNSF grant	
<ul> <li>39</li> <li>40</li> <li>41</li> <li>42</li> <li>43</li> <li>44</li> <li>45</li> <li>46</li> <li>47</li> <li>48</li> <li>49</li> <li>50</li> <li>51</li> <li>52</li> <li>53</li> <li>54</li> <li>55</li> <li>56</li> <li>57</li> </ul>	717	-		
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1 2				
3	718	References		
4 5	719 720	1.	Yung YL, Wang WC, Lacis AA. Greenhouse effect due to atmospheric nitrous oxide.	
6 7	720 721	1.	<i>Geophys Res Lett.</i> 1976;3(10):619-621. doi:10.1029/GL003i010p00619	
8 9	722	2.	Smith C, Nicholls ZRJ, Armour K, et al. The Earth's Energy Budget, Climate Feedbacks,	
10	723		and Climate Sensitivity Supplementary Material. In: Masson-Delmotte V, Zhai P, Pirani A,	
11	724		et al., eds. Climate Change 2021: The Physical Science Basis. Contribution of Working	
12 13	725		Group I to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change.	
13	726		Cambridge University Press; 2021. Accessed October 4, 2021.	
15	727		https://www.ipcc.ch/report/ar6/wg1/downloads/report/IPCC_AR6_WGI_Chapter_07_Supp	
16	728		lementary_Material.pdf	
17 18	729	3.	Crutzen PJ. The influence of nitrogen oxides on the atmospheric ozone content. $QJR$	
19	730	5.	Meteorol Soc. 1970;96(408):320-325. doi:10.1002/qj.49709640815	
20	, 2 0			
21 22	731	4.	Ravishankara AR, Daniel JS, Portmann RW. Nitrous Oxide (N2O): The Dominant Ozone-	
22	732		Depleting Substance Emitted in the 21st Century. Science. 2009;326(5949):123-125.	
24	733		doi:10.1126/science.1176985	
25	724	5	Wuchhles DI Nitrous Ovide: No Loughing Matter Science 2000:226(5040):56 57	
26	734 735	5.	Wuebbles DJ. Nitrous Oxide: No Laughing Matter. <i>Science</i> . 2009;326(5949):56-57. doi:10.1126/science.1179571	
27 28	155			
29	736	6.	Müller R. The impact of the rise in atmospheric nitrous oxide on stratospheric ozone.	
30	737		Ambio. 2021;50(1):35-39. doi:10.1007/s13280-020-01428-3	
31 32	720	-		
33	738	7.	Kim KR, Craig H. Nitrogen-15 and Oxygen-18 Characteristics of Nitrous Oxide: A Global	
34	739		Perspective. Science. 1993;262(5141):1855-1857. doi:10.1126/science.262.5141.1855	
35 36	740	8.	Pérez T, Trumbore SE, Tyler SC, Davidson EA, Keller M, Camargo PB de. Isotopic	
37	741		variability of N2O emissions from tropical forest soils. <i>Glob Biogeochem Cycles</i> .	
38	742		2000;14(2):525-535. doi:10.1029/1999GB001181	
39	740	0		
40 41	743	9.	Kim KR, Craig H. Two-isotope characterization of N20 in the Pacific Ocean and	
42	744 745		constraints on its origin in deep water. <i>Nature</i> . 1990;347(6288):58-61. doi:10.1038/347058a0	
43	743		do1.10.1038/347038a0	
44 45	746	10.	Dore JE, Popp BN, Karl DM, Sansone FJ. A large source of atmospheric nitrous oxide from	
46	747		subtropical North Pacific surface waters. Nature. 1998;396(6706):63-66.	
47	748		doi:10.1038/23921	
48	740	1.1		
49 50	749 750	11.	Naqvi SWA, Naik H, Jayakumar A, et al. Seasonal Anoxia Over the Western Indian	
50	750 751		Continental Shelf. In: Wiggert JD, Hood RR, Naqvi SWA, Brink KH, Smith SL, eds.	
52	751		<i>Geophysical Monograph Series</i> . Vol 185. American Geophysical Union; 2009:333-345. doi:10.1029/2008GM000745	
53	132		doi.10.1029/2008010000/45	
54 55	753	12.	Yoshida N, Hattori A, Saino T, Matsuo S, Wada E. 15N/14N ratio of dissolved N2O in the	
55 56	754		eastern tropical Pacific Ocean. Nature. Published online 1984. doi:10.1038/307442A0	
57				
58				
59 60		pyis	ttp://mc.manuscriptcentral.com/rcm 2	
55				

1 2			
3 4 5	755 756	13.	Rahn T, Wahlen M. Stable Isotope Enrichment in Stratospheric Nitrous Oxide. <i>Science</i> . 1997;278(5344):1776-1778. doi:10.1126/science.278.5344.1776
6 7 8	757 758	14.	Rahn T, Wahlen M. A reassessment of the global isotopic budget of atmospheric nitrous oxide. <i>Glob Biogeochem Cycles</i> . 2000;14(2):537-543. doi:10.1029/1999GB900070
9 10 11 12	759 760	15.	Yoshida N. 15N-depleted N2O as a product of nitrification. <i>Nature</i> . 1988;335(6190):528-529. doi:10.1038/335528a0
13 14 15 16	761 762 763	16.	Barford CC, Montoya JP, Altabet MA, Mitchell R. Steady-State Nitrogen Isotope Effects of N2 and N2O Production in Paracoccus denitrificans. <i>Appl Environ Microbiol</i> . 1999;65(3):989-994. doi:10.1128/AEM.65.3.989-994.1999
17 18 19 20 21	764 765 766	17.	Pérez T, Trumbore SE, Tyler SC, et al. Identifying the agricultural imprint on the global N2O budget using stable isotopes. <i>J Geophys Res Atmospheres</i> . 2001;106(D9):9869-9878. doi:10.1029/2000JD900809
21 22 23 24 25	767 768 769	18.	Yamulki S, Toyoda S, Yoshida N, Veldkamp E, Grant B, Bol R. Diurnal fluxes and the isotopomer ratios of N2O in a temperate grassland following urine amendment. <i>Rapid Commun Mass Spectrom</i> . 2001;15(15):1263-1269. doi:10.1002/rcm.352
26 27 28 29 30 31	770 771 772 773	19.	Lewicka-Szczebak D, Augustin J, Giesemann A, Well R. Quantifying N2O reduction to N2 based on N2O isotopocules – validation with independent methods (helium incubation and 15N gas flux method). <i>Biogeosciences</i> . 2017;14(3):711-732. doi:https://doi.org/10.5194/bg-14-711-2017
32 33 34 35 36	774 775 776 777	20.	Verhoeven E, Barthel M, Yu L, et al. Early season N <sub>2</sub> O emissions under variable water management in rice systems: source-partitioning emissions using isotope ratios along a depth profile. <i>Biogeosciences</i> . 2019;16(2):383-408. doi:https://doi.org/10.5194/bg-16-383-2019
37 38 39	778 779	21.	Yoshida N, Toyoda S. Constraining the atmospheric N2O budget from intramolecular site preference in N2O isotopomers. <i>Nature</i> . 2000;405(6784):330-334. doi:10.1038/35012558
40 41 42 43 44 45	780 781 782 783	22.	Prokopiou M, Martinerie P, Link to external site this link will open in a new window, et al. Constraining N2O emissions since 1940 using firn air isotope measurements in both hemispheres. <i>Atmospheric Chem Phys.</i> 2017;17(7):4539-4564. doi:10.5194/acp-17-4539-2017
46 47 48 49	784 785 786	23.	Yu L, Harris E, Henne S, et al. The isotopic composition of atmospheric nitrous oxide observed at the high-altitude research station Jungfraujoch, Switzerland. <i>Atmospheric Chem Phys.</i> 2020;20(11):6495-6519. doi:10.5194/acp-20-6495-2020
50 51 52 53 54 55 55	787 788 789	24.	Toyoda S, Yoshida N, Miwa T, et al. Production mechanism and global budget of N2O inferred from its isotopomers in the western North Pacific. <i>Geophys Res Lett.</i> 2002;29(3):7-1-7-4. doi:10.1029/2001GL014311
57 58			
59 60		pyıs	sotopomer http://mc.manuscriptcentral.com/rcm 2.

1 2			
3 4 5 6	790 791 792	25.	Popp BN, Westley MB, Toyoda S, et al. Nitrogen and oxygen isotopomeric constraints on the origins and sea-to-air flux of N2O in the oligotrophic subtropical North Pacific gyre. <i>Glob Biogeochem Cycles</i> . 2002;16(4):12-1-12-10. doi:10.1029/2001GB001806
7 8 9 10 11	793 794 795	26.	Yamagishi H, Westley MB, Popp BN, et al. Role of nitrification and denitrification on the nitrous oxide cycle in the eastern tropical North Pacific and Gulf of California. <i>J Geophys Res Biogeosciences</i> . 2007;112(G2). doi:10.1029/2006JG000227
12 13 14 15	796 797 798	27.	Yamagishi H, Yoshida N, Toyoda S, Popp BN, Westley MB, Watanabe S. Contributions of denitrification and mixing on the distribution of nitrous oxide in the North Pacific. <i>Geophys Res Lett</i> . 2005;32(4). doi:10.1029/2004GL021458
16 17 18 19 20	799 800 801	28.	Westley MB, Yamagishi H, Popp BN, Yoshida N. Nitrous oxide cycling in the Black Sea inferred from stable isotope and isotopomer distributions. <i>Deep Sea Res Part II Top Stud Oceanogr</i> . 2006;53(17-19):1802-1816. doi:10.1016/j.dsr2.2006.03.012
20 21 22 23 24	802 803 804	29.	Farías L, Castro-González M, Cornejo M, et al. Denitrification and nitrous oxide cycling within the upper oxycline of the eastern tropical South Pacific oxygen minimum zone. <i>Limnol Oceanogr</i> . 2009;54(1):132-144. doi:10.4319/lo.2009.54.1.0132
25 26 27 28 29 30	805 806 807 808	30.	Casciotti KL, Forbes M, Vedamati J, Peters BD, Martin TS, Mordy CW. Nitrous oxide cycling in the Eastern Tropical South Pacific as inferred from isotopic and isotopomeric data. <i>Deep Sea Res Part II Top Stud Oceanogr</i> . 2018;156:155-167. doi:10.1016/j.dsr2.2018.07.014
31 32 33 34	809 810 811	31.	Bourbonnais A, Letscher RT, Bange HW, et al. N2O production and consumption from stable isotopic and concentration data in the Peruvian coastal upwelling system. <i>Glob Biogeochem Cycles</i> . 2017;31(4):678-698. doi:10.1002/2016GB005567
35 36 37 38 39	812 813 814	32.	Toyoda S, Yoshida O, Yamagishi H, Fujii A, Yoshida N, Watanabe S. Identifying the origin of nitrous oxide dissolved in deep ocean by concentration and isotopocule analyses. <i>Sci Rep.</i> 2019;9(1):1-9. doi:10.1038/s41598-019-44224-0
40 41 42 43	815 816 817	33.	Kelly CL, Travis NM, Baya PA, Casciotti KL. Quantifying Nitrous Oxide Cycling Regimes in the Eastern Tropical North Pacific Ocean With Isotopomer Analysis. <i>Glob Biogeochem</i> <i>Cycles</i> . 2021;35(2):e2020GB006637. doi:10.1029/2020GB006637
44 45 46 47	818 819 820	34.	Toyoda S, Kakimoto T, Kudo K, et al. Distribution and Production Mechanisms of N2O in the Western Arctic Ocean. <i>Glob Biogeochem Cycles</i> . 2021;35(4):e2020GB006881. doi:https://doi.org/10.1029/2020GB006881
48 49 50 51	821 822	35.	Friedman L, Bigeleisen J. Oxygen and Nitrogen Isotope Effects in the Decomposition of Ammonium Nitrate. <i>J Chem Phys.</i> 1950;18(10):1325-1331. doi:10.1063/1.1747471
52 53 54 55 56 57	823 824 825	36.	Toyoda S, Yoshida N. Determination of nitrogen isotopomers of nitrous oxide on a modified isotope ratio mass spectrometer. <i>Anal Chem.</i> 1999;71(20):4711-4718. doi:10.1021/ac9904563
57 58 59 60		pyis	otopomer http://mc.manuscriptcentral.com/rcm 2

1 2				
2 3 4 5 6 7 8	826 827 828 829	37.	Brenninkmeijer CAM, Röckmann T. Mass spectrometry of the intramolecular nitrogen isotope distribution of environmental nitrous oxide using fragment-ion analysis. <i>Rapid Commun Mass Spectrom</i> . 1999;13(20):2028-2033. doi:10.1002/(SICI)1097-0231(19991030)13:20<2028::AID-RCM751>3.0.CO;2-J	
8 9 10 11 12 13	830 831 832 833	38.	Kaiser J, Brenninkmeijer CAM, Röckmann T. Intramolecular 15N and 18O fractionation the reaction of N2O with O(1D) and its implications for the stratospheric N2O isotope signature. <i>J Geophys Res Atmospheres</i> . 2002;107(D14):ACH 16-1-ACH 16-14. doi:10.1029/2001JD001506	in
14 15 16	834 835	39.	Kaiser J. Stable Isotope Investigations of Atmospheric Nitrous Oxide. Johannes Gutenberg University of Mainz; 2003. https://doi.org/10.25358/openscience-3976	g
17 18 19 20 21	836 837 838	40.	Röckmann T, Levin I. High-precision determination of the changing isotopic composition of atmospheric N2O from 1990 to 2002. <i>J Geophys Res Atmospheres</i> . 2005;110(D21). doi:10.1029/2005JD006066	1
22 23 24	839 840	41.	Yung YL, Miller CE. Isotopic Fractionation of Stratospheric Nitrous Oxide. <i>Science</i> . 1997;278(5344):1778-1780. doi:10.1126/science.278.5344.1778	
25 26 27 28	841 842 843	42.	Röckmann T, Kaiser J, Brenninkmeijer CAM, et al. Isotopic enrichment of nitrous oxide (15N14NO, 14N15NO, 14N14N18O) in the stratosphere and in the laboratory. <i>J Geophys Res Atmospheres</i> . 2001;106(D10):10403-10410. doi:10.1029/2000JD900822	5
29 30 31 32	844 845	43.	Toyoda S, Yoshida N, Urabe T, et al. Temporal and latitudinal distributions of stratosphere N2O isotopomers. <i>J Geophys Res Atmospheres</i> . 2004;109(D8). doi:10.1029/2003JD0043	
33 34 35 36	846 847 848	44.	Kaiser J, Engel A, Borchers R, Rockmann T. Probing stratospheric transport and chemistr with new balloon and aircraft observations of the meridional and vertical N2O isotope distribution. <i>Atmos Chem Phys.</i> Published online 2006:22.	ry
37 38 39 40 41	849 850 851	45.	Park S, Atlas EL, Boering KA. Measurements of N2O isotopologues in the stratosphere: Influence of transport on the apparent enrichment factors and the isotopologue fluxes to the troposphere. <i>J Geophys Res Atmospheres</i> . 2004;109(D1). doi:10.1029/2003JD003731	he
42 43 44 45	852 853 854	46.	Sutka RL, Ostrom NE, Ostrom PH, Gandhi H, Breznak JA. Nitrogen isotopomer site preference of N2O produced by Nitrosomonas europaea and Methylococcus capsulatus Bath. <i>Rapid Commun Mass Spectrom RCM</i> . 2003;17(7):738-745. doi:10.1002/rcm.968	
46 47 48 49 50	855 856 857	47.	Sutka RL, Ostrom NE, Ostrom PH, et al. Distinguishing Nitrous Oxide Production from Nitrification and Denitrification on the Basis of Isotopomer Abundances. <i>Appl Environ Microbiol</i> . 2006;72(1):638-644. doi:10.1128/AEM.72.1.638-644.2006	
51 52 53 54 55 56	858 859 860	48.	Sutka RL, Ostrom NE, Ostrom PH, Gandhi H, Breznak JA. Nitrogen isotopomer site preference of N2O produced by Nitrosomonas europaea and Methylococcus capsulatus Bath. <i>Rapid Commun Mass Spectrom</i> . 2004;18(12):1411-1412. doi:10.1002/rcm.1482	
57 58 59 60		pyis	sotopomer http://mc.manuscriptcentral.com/rcm	2.

2				
3 4 5 6	861 862 863	49.	Toyoda S, Mutobe H, Yamagishi H, Yoshida N, Tanji Y. Fractionation of N2O isotopome during production by denitrifier. <i>Soil Biol Biochem</i> . 2005;37(8):1535-1545. doi:10.1016/j.soilbio.2005.01.009	ers
7 8 9 10 11	864 865 866	50.	Frame CH, Casciotti KL. Biogeochemical controls and isotopic signatures of nitrous oxid production by a marine ammonia-oxidizing bacterium. <i>Biogeosciences</i> . 2010;7(9):2695-2709. doi:10.5194/bg-7-2695-2010	e
12 13 14 15	867 868 869	51.	Lazo-Murphy BM, Larson S, Staines S, et al. Nitrous oxide production and isotopomer composition by fungi isolated from salt marsh sediments. <i>Front Mar Sci</i> . 2022;9. Accessed January 3, 2023. https://www.frontiersin.org/articles/10.3389/fmars.2022.1098508	d
16 17 18 19 20	870 871 872	52.	Ostrom NE, Pitt A, Sutka R, et al. Isotopologue effects during N2O reduction in soils and pure cultures of denitrifiers. <i>J Geophys Res Biogeosciences</i> . 2007;112(G2). doi:10.1029/2006JG000287	in
21 22 23 24 25	873 874 875 876	53.	Kaiser J, Park S, Boering KA, Brenninkmeijer CAM, Hilkert A, Röckmann T. Mass spectrometric method for the absolute calibration of the intramolecular nitrogen isotope distribution in nitrous oxide. <i>Anal Bioanal Chem.</i> 2004;378(2):256-269. doi:10.1007/s00216-003-2233-2	
26 27 28 29 30	877 878 879	54.	Westley MB, Popp BN, Rust TM. The calibration of the intramolecular nitrogen isotope distribution in nitrous oxide measured by isotope ratio mass spectrometry. <i>Rapid Commun Mass Spectrom</i> . 2007;21(3):391-405. doi:10.1002/rcm.2828	1
31 32 33 34 35	880 881 882 883	55.	Ostrom NE, Gandhi H, Coplen TB, et al. Preliminary assessment of stable nitrogen and oxygen isotopic composition of USGS51 and USGS52 nitrous oxide reference gases and perspectives on calibration needs. <i>Rapid Commun Mass Spectrom</i> . 2018;32(15):1207-121 doi:10.1002/rcm.8157	4.
36 37 38 39 40 41	884 885 886 887	56.	Mohn J, Wolf B, Toyoda S, et al. Interlaboratory assessment of nitrous oxide isotopomer analysis by isotope ratio mass spectrometry and laser spectroscopy: current status and perspectives. <i>Rapid Commun Mass Spectrom</i> . 2014;28(18):1995-2007. doi:10.1002/rcm.6982	
42 43 44	888 889	57.	Kelly CL. pyisotopomer: Nitrous oxide isotopocule data corrections in Python. Published online January 5, 2023. Accessed March 13, 2023. https://pypi.org/project/pyisotopomer/	
45 46 47 48	890 891	58.	Baertschi P. Absolute18O content of standard mean ocean water. <i>Earth Planet Sci Lett</i> . 1976;31(3):341-344. doi:10.1016/0012-821X(76)90115-1	
49 50 51	892 893	59.	Jabeen I, Kusakabe M. Determination of $\delta$ 17O values of reference water samples VSMO and SLAP. <i>Chem Geol.</i> 1997;143:115-119. doi:10.1016/S0009-2541(97)00109-5	W
52 53 54 55 56 57	894 895 896	60.	Kaiser J, Röckmann T, Brenninkmeijer CAM. Complete and accurate mass spectrometric isotope analysis of tropospheric nitrous oxide. <i>J Geophys Res Atmospheres</i> . 2003;108(D15). doi:10.1029/2003JD003613	
57 58 59 60		pyis	otopomer http://mc.manuscriptcentral.com/rcm	2

1 2				
3 4 5 6	897 898 899	61.	Kaiser J, Röckmann T. Correction of mass spectrometric isotope ratio measurements for isobaric isotopologues of O2, CO, CO2, N2O and SO2. <i>Rapid Commun Mass Spectrom</i> . 2008;22(24):3997-4008. doi:10.1002/rcm.3821	
7 8 9 10 11	900 901 902	62.	Kaiser J, Hastings MG, Houlton BZ, Röckmann T, Sigman DM. Triple Oxygen Isotope Analysis of Nitrate Using the Denitrifier Method and Thermal Decomposition of N2O. <i>Anal Chem.</i> 2007;79(2):599-607. doi:10.1021/ac061022s	
12 13 14 15	903 904 905	63.	Wankel SD, Ziebis W, Buchwald C, et al. Evidence for fungal and chemodenitrification based N 2 O flux from nitrogen impacted coastal sediments. <i>Nat Commun.</i> 2017;8(1):1-11 doi:10.1038/ncomms15595	Ι.
16 17 18 19	906 907 908	64.	Magyar PM, Orphan VJ, Eiler JM. Measurement of rare isotopologues of nitrous oxide by high-resolution multi-collector mass spectrometry. <i>Rapid Commun Mass Spectrom</i> . 2016;30(17):1923-1940. doi:10.1002/rcm.7671	1
20 21 22 23	909 910	65.	Kelly CL. ckelly314/pyisotopomer: v1.0.4. Published online January 19, 2023. doi:10.5281/zenodo.7552724	
24 25 26	911 912	66.	McIlvin MR, Casciotti KL. Technical updates to the bacterial method for nitrate isotopic analyses. <i>Anal Chem.</i> 2011;83(5):1850-1856. doi:10.1021/ac1028984	
27 28 29 30 31	913 914 915	67.	McIlvin MR, Casciotti KL. Fully automated system for stable isotopic analyses of dissolv nitrous oxide at natural abundance levels. <i>Limnol Oceanogr Methods</i> . 2010;8(2):54-66. doi:10.4319/lom.2010.8.54	ed
32 33 34 35 36	916 917 918 919	68.	Scott KM, Lu X, Cavanaugh CM, Liu JS. Optimal methods for estimating kinetic isotope effects from different forms of the Rayleigh distillation equation 1 1Associate editor: J. Horita. <i>Geochim Cosmochim Acta</i> . 2004;68(3):433-442. doi:10.1016/S0016-7037(03)00459-9	
37 38 39	920 921	69.	LI W. Measurement of the absolute abundance of oxygen-17 in V-SMOW. <i>Chin Sci Bull</i> . 1988;33:1610-1613. Accessed June 10, 2021. https://ci.nii.ac.jp/naid/80004607415/	
40 41 42 43 44	922 923 924	70.	Santoro AE, Buchwald C, McIlvin MR, Casciotti KL. Isotopic Signature of N2O Produce by Marine Ammonia-Oxidizing Archaea. <i>Science</i> . 2011;333(6047):1282-1285. doi:10.1126/science.1208239	d
45 46 47 48	925 926 927	71.	Dutton GS, Elkins JW, Hall BD. Nitrous Oxide data from the NOAA/ESRL halocarbons is situ program. Published online 2021. Accessed November 19, 2021. https://data.nodc.noaa.gov/cgi-bin/iso?id=gov.noaa.ncdc:C01556	in
49 50 51 52 53	928 929 930	72.	Inoue HY, Mook WG. Equilibrium and kinetic nitrogen and oxygen isotope fractionations between dissolved and gaseous N2O. <i>Chem Geol.</i> 1994;113(1):135-148. doi:10.1016/00092541(94)90009-4	
54 55 56 57	931 932	73.	Röckmann T, Kaiser J, Brenninkmeijer CAM, Brand WA. Gas chromatography/isotope- ratio mass spectrometry method for high-precision position-dependent 15N and 18O	
58 59 60		pyis	http://mc.manuscriptcentral.com/rcm	2

1 2			
2 3 4 5	933 934		measurements of atmospheric nitrous oxide. <i>Rapid Commun Mass Spectrom</i> . 2003;17(16):1897-1908. doi:10.1002/rcm.1132
6 7 8	935 936	74.	Glover DM, Jenkins WJ, Doney SC. <i>Modeling Methods for Marine Science</i> . Cambridge University Press; 2011. doi:10.1017/CBO9780511975721
9 10 11 12 13	937 938 939	75.	Lorquet JC, Cadet C. Excited states of gaseous ions: I. Selection rules in photoelectron spectroscopy and photoionization. The case of N2O+. <i>Int J Mass Spectrom Ion Phys</i> . 1971;7(3):245-254. doi:10.1016/0020-7381(71)80020-7
14 15 16 17	940 941 942	76.	Märk E, Märk TD, Kim YB, Stephan K. Absolute electron impact ionization cross section from threshold up to 180 eV for N2O+e $\rightarrow$ N2O++2e and the metastable and collision induced dissociation of N2O+. <i>J Chem Phys.</i> 1981;75(9):4446-4453. doi:10.1063/1.442611
18 19 20	943 944	77.	Bigeleisen J. Chemistry of Isotopes. <i>Science</i> . 1965;147(3657):463-471. doi:10.1126/science.147.3657.463
21 22 23 24	945 946	78.	Begun GM, Landau L. Metastable Transitions in N2O+. <i>J Chem Phys.</i> 1962;36(4):1083-1084. doi:10.1063/1.1732641
25 26 27 28	947 948 949	79.	Mohn J, Biasi C, Bodé S, et al. Isotopically characterised N2O reference materials for use as community standards. <i>Rapid Commun Mass Spectrom</i> . 2022;36(13):e9296. doi:10.1002/rcm.9296
29 30 31 32 33 34	950 951		doi:10.1002/rcm.9296
35 36 37 38 39 40			
41 42 43 44 45 46			
47 48 49 50 51			
52 53 54 55			
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Table 1. Reference materials for N<sub>2</sub>O isotopic analysis and intercalibration. Except for one internal standard (B6),

calibrated values were provided via independent measurement by S. Toyoda, Tokyo Tech., J. Mohn, EMPA; or, in 

the case of tropospheric N<sub>2</sub>O, the 2018 annual average measured at Jungfraujoch, Switzerland, reported by Yu et al. (2020). The laboratories participating in the intercalibration exercise were at Stanford University ("Lab 1") and the

University of Basel ("Lab 2"). <sup>31</sup>R values represent the inherent, unscrambled <sup>31</sup>R of each reference material, 

calculated from eqn. (6).

Reference material	Matrix	Mole fraction	$\delta(15N^{\alpha})$	$\delta(15N^{\beta})$	$\delta(^{15}\mathrm{N^{sp}})$	$\delta(^{15}\mathrm{N}^{\mathrm{bulk}})$	$\delta(^{18}\text{O})$	$^{31}R$ ( $^{15}R^{\alpha+17}R$ )	<sup>45</sup> <i>R</i>	<sup>46</sup> <i>R</i>	Calibration
		µmol mol <sup>-1</sup>		(‰, v.	s. air N <sub>2</sub> )		(‰, vs. VSMOW)				
S2 reference gas	Synthetic air	90	5.55	-12.87	18.42	-3.66	32.73	0.004083	0.007712	0.002087	Toyoda & M
B6 reference gas	Не	900	-0.40	-0.15	-0.26	-0.28	41.95	0.004063	0.007739	0.002106	Lab 1 inter standard
Tropospheric N <sub>2</sub> O (2018 annual average)	Air	~0.33	15.6	-2.3	17.9	6.6	44.4	0.004123	0.007787	0.002111	Yu et al. (20
CA06261	Synthetic air	90	-22.21	-49.28	27.07	-35.75	26.94	0.003980	0.007475	0.002075	Toyoda & M
53504	Synthetic air	90	1.71	94.44	-92.73	48.08	36.01	0.004070	0.008093	0.002095	Toyoda & N
CA08214	Synthetic air	90	17.11	-3.43	20.54	6.84	35.39	0.004126	0.007790	0.002093	Toyoda & M
90454	Synthetic air	90	25.73	25.44	0.29	25.59	35.88	0.004158	0.007928	0.002094	Toyoda & N
94321	Synthetic air	90	50.52	2.21	48.31	26.37	35.54	0.004249	0.007934	0.002094	Toyoda & N
Lab 1 pure N <sub>2</sub> O direct injection ("A01")	Pure N2O	N/A	0.24	0.12	0.13	0.18	39.85	0.003734	0.007742	0.002101	Toyoda
Lab 2 pure N <sub>2</sub> O direct injection	Pure N2O	N/A	-4.07	3.59	-7.66	-0.24	39.25	0.004044	0.007739	0.002100	Mohn

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**Table 2.** N<sub>2</sub>O isotopic composition of reference materials and two unknowns analyzed by two IRMS laboratories,961calculated using  $\gamma$  and  $\kappa$  values determined from reference materials 53504 and CA08214 with the algebraic962solution.  $\delta(^{15}N^{\alpha})$ ,  $\delta(^{15}N^{\beta})$ ,  $a(^{15}N^{sp})$  and  $\delta(^{15}N^{bulk})$  are reported in % vs. Air N<sub>2</sub>, and  $\delta^{18}O$  is reported in % vs.963VSMOW. Uncertainties are standard deviations of replicate bottles and do not include calibration uncertainties. The964root-mean square deviation (RMSD) was calculated with respect to calibrated values.

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Reference material		n	$\delta(15N^{\alpha})$	σ	$\delta(15N^{\beta})$	σ	$\delta(^{15}\mathrm{N^{sp}})$	σ	$\delta(^{15}N^{bulk})$	σ	δ( <sup>18</sup> O)	σ
					(‰, vs. a	ir N2)					(‰, v VSMC	vs. DW)
CA06261	Calibrated value		-22.2		-49.3		27.1		-35.7		26.9	
	Lab 1	4	-20.6	1.3	-50.5	1.3	29.9	2.7	-35.6	0.2	28.4	0.8
	Lab 2	16	-20.5	1.4	-50.9	2.6	30.4	3.8	-35.7	1.0	27.6	1.8
	RMSD		2.3		2.1		4.4		0.2		1.5	
53504	Calibrated value		1.7		94.4		-92.7		48.1		36.0	
	Lab 1	4	0.0	1.0	95.7	2.1	-95.7	2.5	47.9	1.1	37.6	0.8
	Lab 2	15	1.7	1.0	94.5	1.9	-92.8	2.9	48.1	0.6	36.4	1.6
	RMSD		1.7		1.3		3.0		0.2		1.7	
CA08214	Calibrated value		17.1		-3.4		20.5		6.8		35.3	
	Lab 1	6	17.0	2.0	-2.4	0.9	19.4	2.9	7.3	0.7	36.3	1.4
	Lab 2	16	17.0	1.1	-3.2	0.7	20.2	1.3	6.9	0.6	36.0	3.6
	RMSD		0.1		1.1		1.2		0.5		1.3	
Tropospheric N <sub>2</sub> O	Calibrated value		15.6		-2.3		17.9		6.6		44.4	
	Lab 1	7	15.1	0.8	-2.5	2.3	17.5	2.8	6.3	1.0	43.1	2.1
	Lab 2	2	15.8	1.1	-3.7	0.0	19.5	1.0	6.1	0.5	44.7	1.0
	RMSD		0.6		1.4		1.7		0.6		1.3	
B6	Calibrated value		-0.4		-0.1		-0.3		-0.3		41.9	
	Lab 1	7	-2.2	0.7	1.3	1.0	-3.4	1.2	-0.4	0.7	41.5	1.6
	RMSD		1.8		1.4		3.2		0.2		0.5	
S2	Calibrated value		5.6		-12.9		18.4		-3.7		32.7	
	Lab1	6	5.0	0.5	-13.1	1.6	18.1	1.3	-4.0	1.0	31.5	1.8
	RMSD		0.5		0.2		0.3		0.4		1.2	
Lake Lugano, 10m	Lab 1	3	13.2	0.3	-5.6	1.2	18.8	1.5	3.8	0.4	44.6	1.2
	Lab 2	5	14.8	1.5	-6.6	1.3	21.4	2.5	4.1	0.5	45.5	0.6
Lake Lugano, 90m	Lab 1	3	19.2	0.5	-33.1	0.7	52.3	1.2	-6.9	0.1	56.8	0.1
	Lab 2	2	18.5	0.8	-32.4	0.3	50.9	0.5	-6.9	0.5	55.4	1.9