- 1 Regional variations in antimicrobial susceptibility of community-acquired uropathogenic
- 2 Escherichia coli in India: findings of a multicentric study highlighting the importance of local
- 3 antibiograms
- 4 Authors: Meher Rizvi*, Shalini Malhotra, Jyotsna Agarwal, Areena H. Siddiqui, Sheela
- 5 Devi, Aruna Poojary, Bhaskar Thakuria, Isabella Princess, Hiba Sami, Aarti Gupta[#], Asfia
- 6 Sultan[#], Ashish Jitendranath[#], Balvinder Mohan[#], Banashankari G. S. [#], Fatima Khan[#], Juri
- 7 Bharat Kalita[#], Mannu Jain[#], N.P. Singh[#], Renu Gur[#], Sarita Mohapatra[#], Shaika Farooq[#],
- 8 Shashank Purwar[#], Mohmed Soeb Jankhwala[#], V. R. Yamuna Devi[#], Ken Masters[#], Nisha
- 9 Goyal[#], Manodeep Sen[#], Razan Al Zadjali[#], Sanjay Jaju[#], Rugma R[#], Suneeta Meena[#], Sudip
- 10 Dutta#, Bradley Langford, Kevin A. Brown, Kaitlyn M. Dougherty, Reba Kanungo, Zaaima
- 11 Al Jabri, Sanjeev Singh, Sarman Singh, Neelam Taneja, Keith H. St John, Raman Sardana,
- 12 Pawan Kapoor, Amina Al Jardani, Rajeev Soman, Abdullah Balkhair, David M Livermore
- *Corresponding Author
- [#]These Authors have contributed equally
- 15 Meher Rizvi, Associate Professor,
- **Affiliation:** Dept of Microbiology and Immunology, College of Medicine and Health Sciences,
- 17 Sultan Qaboos University, Oman.
- 18 Email: rizvimeher@squ.edu.om
- 20 **Shalini Malhotra**, Professor
- 21 Affiliation: Dept of Microbiology, ABVIMS and Dr RML Hospital, New Delhi, India.
- **Email:** drshalinimalhotra@yahoo.com
- 24 Jyotsna Agarwal,

19

23

- 25 **Affiliation:** Professor and Head, Dept of Microbiology, Dr RMLIMS, Lucknow, India.
- 26 **Email:**jyotsnaagarwal.micro@gmail.com

29 30	Areena H. Siddiqui, Professor and Head, Affiliation: Dept of Microbiology, IIMSR, IU, Lucknow, India.
31	Email:draeenahoda@rediffmail.com
32	Charle Dark
33	Sheela Devi
34	Affiliation: Professor of Microbiology
35	Pondicherry Institute of Medical Sciences, Pondicherry, India.
36	Email: sheeladevic@yahoo.com
37	Amuna Dagiamy, Laboratomy Director
38 39	Aruna Poojary, Laboratory Director Affiliation: Dept. of Pathology & Microbiology,
	Breach Candy Hospital Trust, Mumbai, India.
40 41	Email: arunapoojary@gmail.com
42	Eman. arunapoojary(w,gman.com
43	Bhaskar Thakuria
44	Affiliation: Professor and Head
45	Microbiology AIIMS Patna, India.
46	Email: drbhaskart@aiimspatna.org
47	Linuit. drondskart@armispacia.org
48	Isabella Princess, Associate Consultant
49	Affiliation: Apollo Speciality Hospitals,
50	Vanagaram, Chennai - 600095, India.
51	Email: drisabella p@apollohospitals.com
52	<u></u>
53	Hiba Sami, Assistant Professor
54	Affiliation: Jawaharlal Nehru Medical College and Hospital,
55	AMU, Aligarh, 202002, UP, India
56	Email: hibasamizafar@gmail.com
57	
58	Aarti Gupta
59	Affiliation: Zonal Head, Lab Operations,
60	Agilus Diagnostics Limited, Fortis Memorial Research Institute,
61	Gurugram, India
62	Email: thisisaarti@hotmail.com
63	
64	Asfia Sultan, Assistant Professor
65	Affiliation: Jawaharlal Nehru Medical College and Hospital,
66	AMU, Aligarh, 202002, UP, India
67	Email:drasfia@gmail.com
68	
69	Ashish Jitendranath
70	Affiliation : Professor, Department of Microbiology, Sree Gokulam Medical College and
71	Research Foundation, Thiruvananthapuram
72	Email id: ashishjit11@gmail.com

Balvinder Mohan

Affiliation: Professor, 75 Department of Medical Microbiology, 76 77 PGIMER, Chandigarh, India. 78 Email: balvindermohan2002@yahoo.com 79 80 Banashankari G. S. **Affiliation:** Professor & Head 81 Department of Microbiology 82 M S Ramaiah Medical College 83 Bengaluru, India. 84 Email: banashankarigs@gmail.com 85 86 87 Fatima Khan, Associate Professor **Affiliation:** Jawaharlal Nehru Medical College and Hospital, 88 AMU, Aligarh, 202002, UP, India. 89 Email: fatimasalmanshah@gmail.com 90 91 Dr Juri Bharat Kalita, 92 Affiliation: Director, Laboratory Services and Consultant Microbiologist 93 Affiliation: Ayursundra Superspeciality Hospital, Guwahati, Assam, India. 94 Email: jbkalita@gmail.com 95 96 Mannu Jain Affiliation: Professor and head, Microbiology Department, 97 98 Surat Municipal Institute of Medical Education and Research (SMIMER), Surat, Gujarat, India. 99 Email: jainmannu01@gmail.com 100 101 102 Dr N.P. Singh 103 **Affiliation:** Director Professor & Head, Department of Microbiology, University College of Medical Sciences & GTB Hospital, Delhi, India. 104 Email: singhnanjna@yahoo.co.in 105 106 Renu Gur, Consultant Microbiology 107 108 **Affiliation:** Department of Microbiology, Dr. Baba Saheb Ambedkar Medical College & Hospital, Delhi, India. 109 110 Email: renugur@hotmail.com 111 Sarita Mohapatra 112 113 **Affiliation:** Dept. of Microbiology, All India Institute of Medical Sciences, New Delhi, India. 114 115 Email: drsarita2005@gmail.com 116 Shaika Farooq 117 Affiliation: Associate Professor, Dept of Microbiology 118

GMC Srinagar, India.

120	Email: drshaika1farroq@gmail.com
121	Charles December Des Constitution
122	Shashank Purwar, Professor.
123	Affiliation: Department of Microbiology.
124	All India Institute of Medical Sciences Bhopal, India.
125	Email: shashank.microbiology@aiimsbhopal.edu.in
126	Maland Carlo Tarilla ala
127	Mohmed Soeb Jankhwala
128	Affiliation: Associate Professor, Department of Microbiology,
129	Nootan Medical College and Research Centre,
130	Sankalchand Patel University, Visnagar, Gujarat, India.
131	Email: shoaibdoc4@gmail.com
132	V. D. Varrance Devi
133	V. R. Yamuna Devi Affiliation: Consultant Infection control,
134	
135	HOD CSSD & Operation theatre coordinator,
136	Apollo hospitals, Chennai, India.
137 138	Email: dryamunadevi_r@apollohospitals.com
139	Ken Masters, Associate Professor
140	Affiliation: Medical Education and Informatics Department, College of Medicine and Health
141	Sciences, Sultan Qaboos University, Oman.
141	Email: itmeded@gmail.com
143	Eman. micacula ginan.com
144	Dr Nisha Goyal
145	Affiliation: Assistant Professor, Department of Microbiology, University College of Medical
146	Sciences & GTB Hospital, Delhi, India.
147	Email: drnishagoyalucms@gmail.com
148	Eman. umsnago yaruems@gman.com
149	Manodeep Sen
150	Affiliation: Professor, Department of Microbiology,
151	Dr Ram Manohar Lohia Institute of Medical Sciences Lucknow, India.
152	Email: sen manodeep6@yahoo.com
153	Eman: sen_manoucepo(a, yanoo.com
154	Razan Zadjali
155	Affiliation: Department of biochemistry,
156	College of Medicine and Health Sciences
157	Sultan Qaboos University, Oman
158	Email: razanzadjalii@gmail.com
159	Than idealeadan(a) Shahoon
160	Sanjay Jaju
161	Affiliation: Assistant Professor
162	Family Medicine & Public Health
163	College of Medicine and Health Sciences
164	Sultan Qaboos University, Oman

Email: sanjay@squ.edu.om

166	
167	Rugma R
168	Affiliation: Assistant Professor, Department of Microbiology,
169	Sree Gokulam Medical College and Research Foundation, Kerala, India.
170	Email: dr.rugma.r@gmail.com
171	
172	Dr. Suneeta Meena
173	Affiliation: Dept of Laboratory Medicine, AIIMS, New Delhi, India.
174	Email: suneetameena@gmail.com
175	
176	Dr. Sudip Dutta
177	Affiliation: Dept of Laboratory Medicine, AIIMS, New Delhi, India.
178	Email: dr.sudipdatta@gmail.com
179	
180	Dr Bradley Langford
181	Affiliation: University of Toronto
182	Toronto, Canada.
183	Brad.langford@utoronto.ca
184	
185	Kevin Antoine Brown
186	Affiliation: Public Health Ontario, Toronto, ON
187	Email: Kevin.Brown@oahpp.ca
188	Kaitlyn Marie Dougherty
189	Affiliation: Data Analyst
190	Chicago Infection Control, Inc.
191	Chicago, USA.
192	Email: <u>katemarie18@hotmail.com</u>
193	
194	Dr. Reba Kanungo
195	Affiliation: Former Prof and Head of Dept of Microbiology and Dean Research,
196	Pondicherry Institute of Medical Sciences, Pondicherry, India
197	reba.kanungo@gmail.com
198	
199	Dr. Zaaima Al Jabri
200	Affiliation: Dept of Microbiology and Immunology, College of Medicine and Health Sciences
201	Sultan Qaboos University, Oman.
202	Email: zaeema@squ.edu.om
203	
204	Dr. Sanjeev Singh
205	Affiliation: Medical Director
206	Dept of Medicine- Infection Diseases and Epidemiology
207	Amrita Institute of Medical Sciences Faridabad
208	Amrita Vishwavidyapeetham
209	Email: sanjeevksingh@fbd.amrita.edu

212	Affiliation: Former Director, All India Institute of Medical Sciences, Bhopal
213	Email: sarman_singh@yahoo.com, sarman.singh@gmail.com
214	
215	Dr. Neelam Taneja
216	Affiliation: Professor and HOD
217	Department of Medical Microbiology
218	Postgraduate Institute of Medical Education and Research
219 220	Chandigarh Email: drneelampgi@yahoo.com
220	Eman. ameerampgita yanoo.com
221	Mr Keith H. St John,
222	Affiliation: President,
223	North Star IPC Consulting Services, LLC, USA
224	Email: kstjohn@ecri.org
225	
226	Dr. Raman Sardana
227	Affiliation: Professor (Adjunct), Clinical Microbiology and Infection Control
228	Indraprastha Apollo Hospitals New Delhi
229	Director, Board of Trustees, The IFIC, UK
230	Hon. Secretary, Hospital Infection Society-India, New Delhi, India.
231	Email: ramansardana@hotmail.com
232	Air Marshal Dr Pawan Kapoor (Retd)
233	Affiliation: Chairman Steering Committee National Accreditation Board for Hospitals and
234	Healthcare Providers,
235	New Delhi, India.
236	Email: pawankapoor56@yahoo.co.in
237	<u>purround</u>
238	Dr. Amina Al-Jardani
239	Affiliation: Central Public Health Laboratories, Directorate General for Disease Surveillance
240	and Control, Ministry of Health, Oman.
241	Email: aksaljardani@gmail.com
242	
243	Dr Abdullah Balkhair
244	Affiliation: Infectious Diseases Unit, Department of Medicine, Sultan Qaboos University
245	Hospital, Sultan Qaboos University, Oman.
246	Email: balkhair@squ.edu.om
247	
248	Dr. Rajeev Soman
249	Affiliation: Senior ID Physician
250	Jupiter Hospital Pune, India.
251	Email: rajeev.soman@yahoo.com
252	

Dr. Sarman Singh

Dr. David M Livermore

Affiliation: Norwich Medical School,

253

254

255 256	University of East Anglia, Norwich, UK.
257	Email: d.livermore@uea.ac.uk
258	Word Count Abstract: 248
259	Word Count Main Paper: 4964
260	
261	Running heads: E. coli from community UTIs in India
262	Keywords: Escherichia coli, community-acquired UTIs, India, Antimicrobial Resistance
263	Corresponding author contact details: Meher Rizvi, rizvimeher@squ.edu.om

264 Abstract

Background: Evidence-based prescribing is essential to optimise patient outcomes in cystitis.
This requires knowledge of local antibiotic resistance rates. DASH to Protect Antibiotics
(https://dashuti.com/) is a multicentric mentorship programme guiding centres in preparing,
analysing and disseminating local antibiograms to promote antimicrobial stewardship in
community UTI. Here we map the susceptibility profile of <i>Escherichia coli</i> from 22 Indian centres.
Methods: These centres spanned 10 Indian States and three Union Territories. Antibiograms for
urinary E. coli from the outpatient departments were collated. Standardisation was achieved by
regional online training; anomalies were resolved via consultation with study experts. Data were
collated and analysed. Findings: Nationally, fosfomycin, with 94% susceptibility (inter-centre
range 83-97%), and nitrofurantoin with 85% susceptibility (61-97%) retained widest
activity. Susceptibility rates were lower for co-trimoxazole (49%), fluoroquinolones (31%) and
oral cephalosporins (26%). Rates for third- and fourth- generation cephalosporins were 46% and
52%, respectively, with 54% (33-58%) ESBL prevalence. Piperacillin-tazobactam (81%) amikacin
(88%), meropenem (88%) retained better activity, but one centre in Delhi recorded only 42%
meropenem susceptibility. Susceptibility rates were mostly higher in South, West and Northeast
India; centres in the heavily-populated Gangetic plains, across North and Northwest India, had
greater resistance. These findings highlight the importance of local antibiograms in guiding
appropriate antimicrobial choices.
Interpretation: Fosfomycin and nitrofurantoin are the preferred oral empirical choices for
interpretation. Tostomyem and introducation are the preferred of an empirical enoices for
uncomplicated E. coli cystitis in India, though elevated resistance in some areas is concerning.
Empiric use of fluoroquinolones and third generation cephalosporins is discouraged whereas
piperacillin/tazobactam and aminoglycosides remain carbapenem-sparing parenteral agents.

INTRODUCTION

Urinary tract infections (UTIs) are among the most frequent infections worldwide. About 60% of women and 20% of men will experience at least one UTI during their lifetime, prompting antibiotic treatment, usually prescribed empirically [1,2]. *Escherichia coli* remains the predominant pathogen globally in both community- and hospital-acquired settings [3]. Increasing resistance complicates treatment, making outcomes uncertain, even in simple cystitis [4].

Minimising resistance needs multi-disciplinary stewardship approaches [4]. These include evidence-based prescribing, which requires knowledge of local community and hospital antibiotic resistance rates. In India, much prescribing is market-driven rather than evidence-based. This situation prompted us to develop *DASH to Protect Antibiotics* (https://dashuti.com/).

DASH is a multicentric mentorship-based study aiming to assemble disseminate antibiogram data and to promote greater interaction between microbiologists and clinical practitioners and thereby to improve antimicrobial prescribing. The present investigation involved 22 centres across India and sought to collect, review and optimise antibiogram data for community-acquired UTI due to *E. coli*. DASH's further approaches include vignette-based questionnaires and focused education.

METHODS

Centre recruitment

This ongoing study was open to all interested centres across India, including public and private medical colleges, tertiary healthcare facilities and standalone laboratories. Invitations to participate were sent by email, WhatsApp and through LinkedIn. Forty-one centres were approached, of which 29 (27 tertiary-care public and private hospitals and two private laboratories) agreed to join.

Five hospitals and both private laboratories subsequently withdrew, citing lack of time or internal support, leaving 22 sites: 11 were in North (N) India, one in Jammu & Kashmir (extreme North), four in Delhi, one in the neighbouring National Capital Region (NCR) Gurugram, one each in Aligarh and Chandigarh and three in Lucknow, five in South (S) India (two in Chennai and one each in Pondicherry, Karnataka and Kerala), three in West (W) India (two in Gujarat and one in Mumbai), along with single centres in the East (E) (Patna), Northeast (NE) (Guwahati) and Central India (Bhopal). (Figure 1). Due to proximity, Chandigarh (a Union Territory west of Delhi) and Gurugram (in Haryana but part of the National Capital Region) were analysed together with the Delhi sites (Supplementary Table S1). The 'Delhi' region sites (except Chandigarh) are located in the Gangetic plains, along with Aligarh, Lucknow (with 3 sites) and Patna. Seventeen centres were academic whereas five were non-academic. Ten states and three union territories participated. The duration of this study was one year, from 1st January 2022 to 31st December 2022.

Ethical approval for the study was obtained by the centres. Details of the centres' infrastructure and routine practices were collated via a questionnaire.

Initial actions to achieve standardisation of methods

Prior to preparing the Outpatient Department (OPD)-based antibiograms, a workshop on implementation of the WHONET and BACLINK susceptibility data analysis software (https://whonet.org) was conducted by three centres [2]. This was filmed and made available to all sites: links are:- https://youtu.be/ijSFIIy5DZ4, https://youtu.be/wh7XlsxKmJg. Centres remained free to prepare their antibiograms using other tools if preferred.

Sample processing at study sites

Microscopy for bacteria and leucocytes was the most common initial screen, used at 14 sites; five sites used the dipstick method and three screened by visual examination of urine turbidity. Twelve centres used automated bacterial identification for putatively-infected urines; 10 used classical manual methods [5]. Antimicrobial Susceptibility testing (AST) was performed according to CLSI guidelines M100-Ed33) 2022 [6]. Ten sites largely used disc diffusion testing whereas 12 used automated systems, six used a mixture of both approaches. Quality control was practiced by all laboratories. ESBLs were detected using cephalosporin/clavulanic acid synergy tests by eight centres.

CLSI urine breakpoints were used for interpretation of cefazolin and cefuroxime results. Isolates with susceptibility reaching the dose-dependent breakpoints, e.g. to cefepime, were counted as susceptible.

Data collection, handling, review and validation

Only clinical isolates from patients presenting with a symptomatic UTI at an out-patient or Emergency Department were included.

Data from such patients were collated into site antibiograms if 30 or more non-duplicate isolates were tested at the site. Only data for routinely-tested antimicrobial agents were included. CLSI guideline M39A4E CLSI 2022 was used to prepare the antibiograms [3,7]. Once the data were collected, exhaustive region-wide online sessions were conducted, involving Prof Livermore, to analyse them and to resolve anomalies (e.g.: lower percent susceptibilities for: (i) amikacin compared with gentamicin; (ii) ceftriaxone, cefotaxime and ceftazidime compared with

cefuroxime; (iii) cefuroxime compared with cefazolin; (iv) piperacillin/tazobactam compared with amoxicillin/clavulanic acid; (v) meropenem compared with ertapenem and/or piperacillin/tazobactam, and (vi) ciprofloxacin compared with levofloxacin.

Statistics

351

352

353

354

355

356

357

358

359

360

361

362

363

364

365

366

367

368

369

370

371

Antimicrobial susceptibilities of the E. coli isolates were compared across six broad geographic regions comprising N., S., E., NE, W. and Central India. Overall susceptibility was calculated, and the proportions of susceptible isolates were compared between regions (z test for proportions). Representative drugs from different antimicrobial drugs (fosfomycin, nitrofurantoin, trimethoprim-sulfamethoxazole, cefotaxime, ceftriaxone, gentamicin, meropenem, ciprofloxacin, piperacillin/tazobactam and cefepime) were subjected to detailed statistical analysis. To obtain a measure of the degree of inter-regional variability, the intra-cluster correlation (ICC) was calculated based on a random intercept logistic regression model using SPSS version 23 IBM and R version 4.0 and Excel. Medians were calculated. Arithmetic and harmonic means were calculated to average percentage susceptibility rates reported by different sites. Since percent susceptibilities are ratios, harmonic means were preferred; however, results were similar regardless of which type of average was used (see Table 1). 'Resistance to third-generation cephalosporins' is the harmonic mean of individual sites' resistance rates to ceftazidime, cefotaxime, ceftriaxone, and cefixime; that for 'β-lactam/β-lactamase inhibitors' is for piperacillin/tazobactam and cefoperazone/sulbactam (analysed vs. piperacillin/tazobactam breakpoints); for that 'carbapenems' is the average of imipenem and meropenem.

Funding

The study was unfunded and relied entirely on the existing infrastructure, manpower, motivation and goodwill.

RESULTS

372

373

374

375

376

377

378

379

380

381

382

383

384

385

386

387

388

389

390

391

Antimicrobial susceptibility profile of E. coli across India

Antimicrobial susceptibility profiles of 7790 isolates of community-acquired E. coli were analysed from a total of 51,703 samples received at the OPDs surveyed. Overall susceptibility rates across all sites are shown in Table 1, with site-by-site detail in supplementary Table S1 and regional rates, with confidence intervals for major antibiotic groups, in Table 2. Regional rates for major oral antibiotics are illustrated by site in Figure 2, with those for i.v. antimicrobial agents in Figure 3, with further detail in Supplementary Table S2. Antimicrobial susceptibilities at two centres (one in Delhi, another in Gujarat) were considered to be outliers and their data were not included in the national and regional means (Table 1); The centre in Central India (Bhopal) provided a combined antibiogram for urinary E. coli from both in- and out- patients, and their data likewise were excluded when calculating national susceptibility. Significant inter-regional variability in resistance rates was observed for all drugs, as shown in Table 2. The ICC was highest (0.92) for fosfomycin, indicating least variation, and lowest (0.26), indicating most variation, for ciprofloxacin. We review the salient features below, by antibiotic or antibiotic class. Fosfomycin: Across all the six regions, fosfomycin was the most reliably active antimicrobial, with 94% (92 to 97%) national susceptibility.

- Nitrofurantoin: The national susceptibility to nitrofurantoin was 85%. In general, W. India had
- 393 high susceptibility (88% to 97%), as did S. India (87% to 95%) whereas wide variation was
- observed for sites across N. and Central India (61% to 96%).
- 395 **Co-trimoxazole:** Antimicrobial susceptibility to co-trimoxazole was low, ranging from 36% to
- 396 68%, with a national rate of 49%. Two individual centres in S. India, (Bangalore and
- 397 Thiruvananthapuram) reported 68% susceptibility the highest in the country.
- 398 **First- and second- generation cephalosporins:** These drugs performed poorly, with only around
- 399 26% susceptibility nationally.
- 400 Third- and fourth-generation cephalosporins: Susceptibility rates ranged between 40 and 50%,
- averaging 46.3%. (Table 1, with details in Supplementary Table S1 and S2). Guwahati in the NE
- had the highest susceptibility rate, at 67%, and Patna in E. India the lowest, at 29% (Figure 2). The
- ational susceptibility rate for cefepime was 52%, with local rates ranging from 93% in Surat to
- 404 36% at the sole centre in Delhi where it was tested.
- 405 **Estimation of ESBL prevalence**: The national prevalence rate for ESBLs was thereby estimated
- at 54%, ranging from 33% in NE to 58% in N. India.
- 407 β-Lactam/β-lactamase inhibitors: Overall, susceptibility rates were 81% for
- 408 piperacillin/tazobactam and 47% for amoxicillin/clavulanic acid; cefoperazone/sulbactam lacks at
- 409 CLSI breakpoint but, if the piperacillin/tazobactam breakpoint was applied, susceptibility was
- 410 estimated at 79%. The susceptibility range among sites was extremely wide for
- amoxicillin/clavulanate, from 6% in one centre in Delhi to 83% in Guwahati (Assam). By contrast,
- 412 rates for cefoperazone/sulbactam and piperacillin/tazobactam were more narrowly spread, from

413	72 (Chandigarh) to 92% Thiruvananthapuram, Kerala) for cefoperazone/sulbactam and 81 to 94%
414	in W., NE and S. India to 82% in E. India for piperacillin/tazobactam. Lower rates were observed
415	from N. India (64%) and Delhi (79%).
416	Carbapenems: National susceptibility rates were 88% for both imipenem and meropenem (Table
417	1 and Figure 3). Significantly higher susceptibility rates to meropenem were observed in S. (90 to
418	98%) and W. India (92 to 95%) compared with other regions (p<0.05).
419	There were several outliers: one site in Lucknow had a meropenem susceptibility rate of 68%, one
420	in Bhopal (Central India) had a rate of 64%. An extreme outlier in Delhi recorded 42% meropenem
421	susceptibility; this was not included in the calculation of averages.
422	Fluoroquinolones: The national susceptibility rate for ciprofloxacin was 29% with only three
423	centres reporting susceptibility rates exceeding 50% (Table 1); fewer centres tested levofloxacin,
424	with only a slightly higher (35%) susceptibility rate recorded.
425	Aminoglycosides: High rates susceptibility rates were observed to gentamicin (75 to 84%) and
426	amikacin (88 to 96%) in S. India and also in W. India (gentamicin: 74 to 85% and amikacin: 97 to
427	98%. Rates by region are given in Figure 3. Two outliers, one in Delhi and another in Gujarat,
428	reported less than 50% susceptibility to amikacin; the Delhi site was the same one that had
429	unusually low susceptibility to meropenem. Given the frequent genetic linkage of metallo (NDM)-
430	carbapenemases and aminoglycoside-compromising ArmA and Rmt ribosomal
431	methyltransferases, this parallel pattern lends confidence in both the outlying results [8].

DISCUSSION

The rapid emergence and proliferation of multi-drug resistant uropathogens – often harbouring ESBLs, AmpC enzymes and carbapenemases – makes the treatment of even simple UTIs more challenging, often rendering empirically-used antimicrobials inactive [9]. Providing relevant antibiograms to clinicians is vital to addressing this issue; it is vital also to stratify by whether UTI isolates are from in- or out- patients [10]. Treatment of UTIs in India follows national and international guidelines, but the large regional variations observed in our study suggest that management should be tailored to reflect local resistance rates [11,12]. E. coli is considerably the commonest uropathogen worldwide [13]. Here we tracked antimicrobial susceptibility among isolates of the species recovered from patients with UTI attending outpatient departments in 22 centres across India. High resistance rates were seen, especially in N. India, where many centres (i.e., those in Delhi, Lucknow, Aligarh, Patna) are located across the 'Gangetic Plains'. Two of the outliers, with particularly high resistance rates, lie in this region. As illustrated e.g., by https://vividmaps.com/india-maps/ this region has a burgeoning population, many of whom lack safe water and sanitation, and who quite possibly experience extensive inappropriate antimicrobial prescribing. Fosfomycin, with 94% overall susceptibility, emerged as the most-reliably active antimicrobial in vitro, though with significantly greater susceptibility in S. compared with N. India, p<0.05. These findings are consistent with other studies in India, including recently published data from the Odisha State, where susceptibility rates of 99% and 91.3% were recorded for E. coli and K. pneumoniae, respectively [14]. Fosfomycin, prescribed as a single oral dose of 3 grams, maintains good in-vitro activity regardless of the presence of other resistances [15]; however clinical

outcomes in cystitis were reportedly poorer than with a five-day high-dose (100 mg q8h) course

of nitrofurantoin [16]. A complicator is that the standard regimen for nitrofurantoin is 100 mg

433

434

435

436

437

438

439

440

441

442

443

444

445

446

447

448

449

450

451

452

453

454

q12h, not q8h; moreover, it is plausible that two- or three- dose fosfomycin regimens may be more effective than the licensed single-dose therapy [16]. Advocating mainstream use of fosfomycin does raise concerns about emergence of resistance, especially as it is a useful salvage drug for infections involving extremely- and pan- drug resistant bacteria [17].

Surprising rates of resistance were seen to nitrofurantoin, which shows near 100% activity in surveys of urinary *E. coli* collected in Europe [18]. The overall susceptibility rate was 85%, but with rates as low as 61 to 74% in Aligarh, Patna and Lucknow, which are widely-separated cities across northern India. Susceptibility in W. India (93.1%) was significantly greater (p<0.05) than in N., S., or E. India or in the Delhi-NCR region, while susceptibility in NE India was significantly greater than in E. India Perhaps of note, the sites with the lowest susceptibility rates were higher tertiary centres, receiving more referrals. Other studies have reported susceptibility rates of 90.3% for *E. coli* from N. India, 91% for Rajasthan, 94.2% for S. India, 93.9% for E. India and 93.4% for W. India [13,19]. Mohapatra *et al.* [13] reported 94.2% susceptibility for *E. coli* from community-acquired UTIs across four centres in different regions of India; however, recent data from Guntur in Andhra Pradesh suggests only 60% susceptibility of *E. coli* to nitrofurantoin in outpatient settings [20]. In the UK resistance to nitrofurantoin in *E. coli*, though uncommon, is associated with chromosomal mutations [21]. Work is urgently needed to explore whether these or other modes of resistance have evolved and are accumulating in India.

Resistance rates to other orally-administrable antibiotics were very high, suggesting that their empirical use will be associated with frequent failure. Co-trimoxazole, retained activity against only 49% of isolates. Bhargava *et al.* in 2022 [22] reported even lower susceptibility, at 39.8%, and Vijayganapathy *et al.* in 2021 [23] reported 24% susceptibility; their datasets for *E. coli* were from N. and S. India respectively. In pairwise comparisons, isolates from S. and W.

India independently demonstrated greater susceptibility compared with those from N., E. or Central India, or from the Delhi-NCR (p<0.05).

In the case of fluoroquinolones, data were most complete for ciprofloxacin, with a national susceptibility rate of only 29%. Similar rates were seen for norfloxacin and levofloxacin. Rates for ciprofloxacin ranged from 11 to 55% in N. India, 24 to 52% in W. India, 11 to 40% in S. India and 11 to 36% in Delhi, indicating little clear regional difference despite considerable site-to-site differences within regions, reflected in the low ICC. These results are in keeping with the findings of others: Bharara *et al.* [24] reported 50% and 33% susceptibility to levofloxacin and ciprofloxacin, respectively, for *E. coli* in Delhi in 2018 whilst, in S. India, Vijayganapathy *et al.* [23] reported 38% and 26% susceptibility, respectively. All these fluoroquinolone rates were lower than for co-trimoxazole and amoxicillin/clavulanic acid. Losada *et al.* [25] in Spain likewise reported greater susceptibility to co-trimoxazole and amoxicillin/clavulanic acid (70% and 77%, respectively) than to fluoroquinolones (67%) for *E. coli*. Given additional concerns regarding fluoroquinolone safety [26] and their propensity to cause collateral damage to the gut flora, there seems no good reason to still advocate these agents for empirical use in UTIs in India.

Turning to intravenous agents, likely to be used for an ascending UTI, the national susceptibility rate to third-generation cephalosporins was 46.3%, whilst that to cefepime was 52%. W. India exhibited significantly greater susceptibility to cefotaxime (85.1%) compared with other regions, where it varied between 27% and 53% (p<0.05). Similar patterns were seen for ceftazidime, ceftriaxone and cefepime, with the highest susceptibility observed in W. India. Cefepime susceptibility was notably higher in W. India, at 78 to 91%. For comparison, Jangid *et al.* 2021 [9], in a multicentric study spanning many Indian centres, reported 33.6% susceptibility for *E. coli* to cefixime, while Bhargava *at al.*, 2022 [22] reported less than 10% susceptibility for cefepime in

N. India. At least one centre in each region tested prevalence of ESBLs directly. Whilst this is limited coverage, these ESBL data were entirely consistent with cephalosporin resistance data, which were extensive. Such cross-referencing of two data sets adds confidence. Moreover, the similarly high rates of resistance to third-generation cephalosporins and cefepime suggest that most cephalosporin resistance is attributable to ESBLs rather than to AmpC enzymes, though W. India, with its higher cefepime susceptibility, may be an exception. An exceptionally high ESBL prevalence (72%) was reported by the site in Patna, Bihar, perhaps reflecting the hospital being a major referral centre. Paul et al. 2021 [27] previously reported 26.2% ESBL prevalence in Assam (NE. India) whilst Behera et al. 2022 [28] reported 43% combined prevalence in E. coli and Klebsiella pneumoniae from community UTIs from E. India and, in 2021, Kumar et al [29] reported 46.6% ESBL prevalence in E. coli from Uttarakhand in N. India. In 2022, Mohapatra et al. [13] reported an ESBL prevalence of more than 50% across four centres in E. coli. Our observation of higher apparent susceptibility rates to ceftazidime than to cefotaxime (Table 1) suggested that much 'ESBL-mediated resistance' there was due to CTX-M type ESBLs, though this requires molecular confirmation. Piperacillin/tazobactam susceptibility was recorded as 81% overall, almost matched by cefoperazone/sulbactam at 79%, whereas amoxicillin/clavulanic acid was active only against 47% of the isolates. Overall, NE India followed by S., W. and E. India exhibited significantly higher susceptibility to piperacillin/tazobactam compared to N. India (p < 0.05). Mohapatra et. al. 2022 reported similar (75.1%) susceptibility data for piperacillin/tazobactam but much higher susceptibility (74.7%) for amoxicillin/clavulanic acid among Gram-negative uropathogens [13]. Based upon testing at only a few sites, S. India reported higher susceptibility (89%) to cefoperazone/sulbactam than to piperacillin/tazobactam (81%), reversing the national pattern,

502

503

504

505

506

507

508

509

510

511

512

513

514

515

516

517

518

519

520

521

522

523

though caution is needed owing to the lack of international breakpoints for the sulbactam combination. Vijayaganapathy *et al.* 2018 reported 80% susceptibility to piperacillin/tazobactam and 78% to cefoperazone/sulbactam for urinary *E. coli* from out-patients in S India, also suggesting the near equal activity of these combinations [23].

Nationwide, susceptibility to aminoglycosides was around 80% (gentamicin, 76%; amikacin, 87%). In S. and W. India, however, amikacin susceptibility rates were as high as 88 to 96% and 97 to 98%, respectively, whereas at two centres in N. India – in Lucknow and Aligarh – susceptibility was only *c.* 60%. The S. (78.0%); E. (78.6%) and W. Regions (80.0%) recorded significantly higher proportion of susceptibility to gentamicin (p<0.05) than in N. India (70%) and Delhi NCR (71.0%). Previously, Bhargava *et al.* 2022 reported 77% susceptibility for amikacin among *E. coli* from N. India [22].

Despite concerns about the community spread of NDM carbapenemases in India, susceptibility to carbapenems remained at 88% nationally, with high rates reported from S. (90 to 98%) and W. India (92 to 95%) (30). Similarly, in a four-centre study, Mohapatra *et al*, 2022 reported 90.4% carbapenem susceptibility for *E. coli* [13] whilst Vijayganapathy *et al*. 2018 reported 99% susceptibility in S. India and Nair *et al*. reported 87.8% susceptibility in W. India [23,31]. Disturbingly, much lower susceptibility rates were seen at the outlier centre in Delhi (42%), and at single centres in Lucknow, N. India (68%), and Bhopal (64%). Bhargava *et al*. likewise reported low susceptibility for 37.2% for meropenem and 57.4% for imipenem from Allahabad, N. India, testing *E. coli* from both in- and out- patients [22].

On the basis of our results, we recommend nitrofurantoin and fosfomycin as first-line antibacterial agents for uncomplicated community-acquired UTIs in India. Both these agents have the further

benefit of causing little collateral damage to the gut flora [32]. Caveats and cautions are: (i) whereas the susceptibility data favour fosfomycin, trial data indicate nitrofurantoin may be a more effective agent;[16] (ii) several centres reported significant (>20%) rates of resistance to nitrofurantoin and one had only 85.3% susceptibility to fosfomycin, and (iii) neither agent is reliably effective in complicated or ascending infection. For such infections, warranting intravenous therapy, both aminoglycosides and the more potent β-lactam/β-lactam inhibitor combinations (i.e., piperacillin-tazobactam and cefoperazone-sulbactam) remain widely active, as do carbapenems – though we advocate reserving these where possible. Geographic variability underscores the need to generate and utilise local antibiograms to support appropriate empirical prescribing, exactly as DASH seeks to support [20]. The higher resistance in N. India may be linked to several factors: greater over the counter sale of antibiotics, indiscriminate prescription of antibiotics, large population with low per capita income, higher burden of disease and substandard drugs [33,34,35]. It also underscores the likely weakness of any global surveillance that only includes three or four centres to 'represent' a country as large and diverse as India.

Limitations

This study used hospitals' routine data, allowing us to assemble a large amount of geographically representative information without additional testing. The approach does, however, leave the study vulnerable to site-to-site variations in methodology. We sought to control and correct these as much as possible but cannot be certain that they were completely eliminated. As with almost all studies of community UTIs, the study is likely to be subject to the problem that microbiological sampling is skewed towards complicated, unresponsive and recurrent cases, who are more likely to have resistant pathogens [36]. Moreover, because most primary and secondary care hospitals do little or no culture and susceptibility testing from urines, we were obliged to largely use tertiary

centres and, even at their outpatient departments, these may serve a more complex patient population, more likely to harbour resistant pathogens.

Conclusions

570

571

572

573

574

575

576

577

578

579

580

581

582

583

584

585

586

587

588

589

590

591

As antibiotic susceptibility rates vary strikingly across a large country like India, local antibiograms should guide empirical treatment for simple UTIs. India is a large, diverse country with large variations in population, per capita income, literacy. The variations extend to healthcare infra-structure, adoption of best practices and also antimicrobial resistance. W. and S. India are more prosperous and are less densely populated than N. India, with better healthcare infra-structure and wider scale adoption of best practices including judicious use of antimicrobials. Maybe these important indicators are being reflected in the significant variations in resistance observed in different regions of India. This study confirms that fosfomycin and nitrofurantoin remain excellent oral empirical choices for uncomplicated community UTIs due to E. coli in India, including when these are due to strains resistant to other agents. Both nitrofurantoin and fosfomycin have the further benefit of causing little collateral damage to the gut flora. Nonetheless, notably raised rates of resistance to nitrofurantoin were recorded at several sites and, for fosfomycin, at one site. Such data need to be considered alongside the trial showing better outcomes for nitrofurantoin [16]. Our findings strongly discourage the empirical use of fluoroquinolones and third-generation cephalosporins in simple cystitis. β-Lactam/β-lactamase inhibitor combinations aminoglycosides likely remain the best carbapenem-sparing agents where ascending infection demands i.v. therapy.

Contributors: MR contributed to the conceptualization of the study, analysis, drafting and editing of the manuscript; AG,AS,AJ, BM, BGS, FK, JBK, MJ, NPS, RG, SM, SF, SP, MSJ, VRYD, NG,

MS, participated in the study and shared the data and MR, AHS, HS, ID, VRYD, KM, SM, SD,

drafted and edited the manuscript; MR, RAZ, DL, AHS, AP, IP, RK, HS, analysed the data and SJ,

RR, SM, SD, BL, KAB, KMD, RK, ZAJ, SS, SS, NT, KHSJ, RS, PK, AAR, RS, ABK, DML

supervision and intellectual input; KAB, KM, SJ did statistical analysis

Data sharing statement

 Data supporting the findings of this study is available.

Conflict of Interest: None to declare

Acknowledgement

We are grateful to Dr. Arjun Srinivasan US (CAPT), Centers for Disease Control and Prevention, Atlanta, Georgia, USA., for his constant support and guidance throughout the study. We acknowledge the help extended by Prof. Anil Kumar, Department of Microbiology, Amrita Institute of Medical Sciences, Kochi, India, Dr Lilian Abbo, MD, FIDSA, Chief Infection Prevention and Antimicrobial Stewardship, Jackson Health System, Professor of Infectious Diseases, University of Miami Miller School of Medicine for their inputs in preparing the Microbiology Questionnaire. We thank Sdr Simranjit Singh and Mr Adil Jamal, India for the infographics. We appreciate the data analysis carried out by Yousuf Al Mamari and Sara Al Obeidani, MD students, College of Medicine & Health Sciences, Sultan Qaboos University, Oman

References

[1] Medina M, Castillo-Pino E. An introduction to the epidemiology and burden of urinary tract infections. Ther Adv Urol. 2019 Jan;11:175628721983217.

[2] Farrell K, Tandan M, Hernandez Santiago V, Gagyor I, Braend AM, Skow M, et al.
Treatment of uncomplicated UTI in males: a systematic review of the literature. BJGP
Open. 2021 Apr;5(2):bjgpopen20X101140.

- [3] Flores-Mireles AL, Walker JN, Caparon M, Hultgren SJ. Urinary tract infections: epidemiology, mechanisms of infection and treatment options. Nat Rev Microbiol. 2015 May;13(5):269–84.
 - [4] Calvo-Villamañán A, San Millán Á, Carrilero L. Tackling AMR from a multidisciplinary perspective: a primer from education and psychology. Int Microbiol. 2022 Oct 13;26(1):1–9.
- [5] Tille PM. Bailey & Scott's diagnostic microbiology. Fifteenth edition. St. Louis, Missouri: Elsevier; 2022.
 - [6] M100 PERFORMANCE STANDARDS FOR ANTIMICROBIAL SUSCEPTIBILITY TESTING, 33RD EDITION,M100ED33. S.l.: CLSI; 2023.
 - [7] Hindler JA. Analysis and presentation of cumulative antimicrobial susceptibility test data. 5th edition. Pennsylvania: Clinical and Laboratory Standards Institute; 2022.
 - [8] Hidalgo L, Hopkins KL, Gutierrez B, Ovejero CM, Shukla S, Douthwaite S, et al. Association of the novel aminoglycoside resistance determinant RmtF with NDM carbapenemase in Enterobacteriaceae isolated in India and the UK. J Antimicrob Chemother. 2013 Jul 1;68(7):1543–50.
 - [9] R J. Antimicrobial Sensitivity Patterns of Uropathogens in India: A Nationwide, Multicentric, Big-Data, Retrospective Study. Open Access J Urol Nephrol. 2021 Aug 6;6(2):1–12.
 - [10] Wesolek JL, Wu JY, Smalley CM, Wang L, Campbell MJ. Risk Factors for Trimethoprim and Sulfamethoxazole-Resistant Escherichia Coli in ED Patients with Urinary Tract Infections. Am J Emerg Med. 2022 Jun;56:178–82.
 - [11] Treatment guidelines for antimicrobial use in common syndromes. Indian Council of Medical Research, department of health research,; Delhi, India.: https://www.icmr.nic.in/sites/default/files/guidelines/treatment_guidelines_for_antimicrobial.pdf/ last accessed on 27th January 2024.
 - [12] Home-Sanford Guide-Antimicrobial Stewardship. Available online: https://www.sanfordguide.com/ (accessed on 27 January 2024).
 - [13] Mohapatra S, Panigrahy R, Tak V, J. V. S, K. C. S, Chaudhuri S, et al. Prevalence and resistance pattern of uropathogens from community settings of different regions: an experience from India. Access Microbiol [Internet]. 2022 Feb 9 [cited 2023 Aug 28];4(2). Available from: https://www.microbiologyresearch.org/content/journal/acmi/10.1099/acmi.0.000321
- Mohanty S, Behera B, Sahu S, Praharaj A. In vitro activity of fosfomycin against multidrug-resistant urinary and nonurinary gram-negative isolates. Indian J Crit Care Med. 2018 Jul;22(7):533–6.

Sultan A, Rizvi M, Khan F, Sami H, Shukla I, Khan H. Increasing antimicrobial resistance among uropathogens: Is fosfomycin the answer? Urol Ann. 2015;7(1):26.

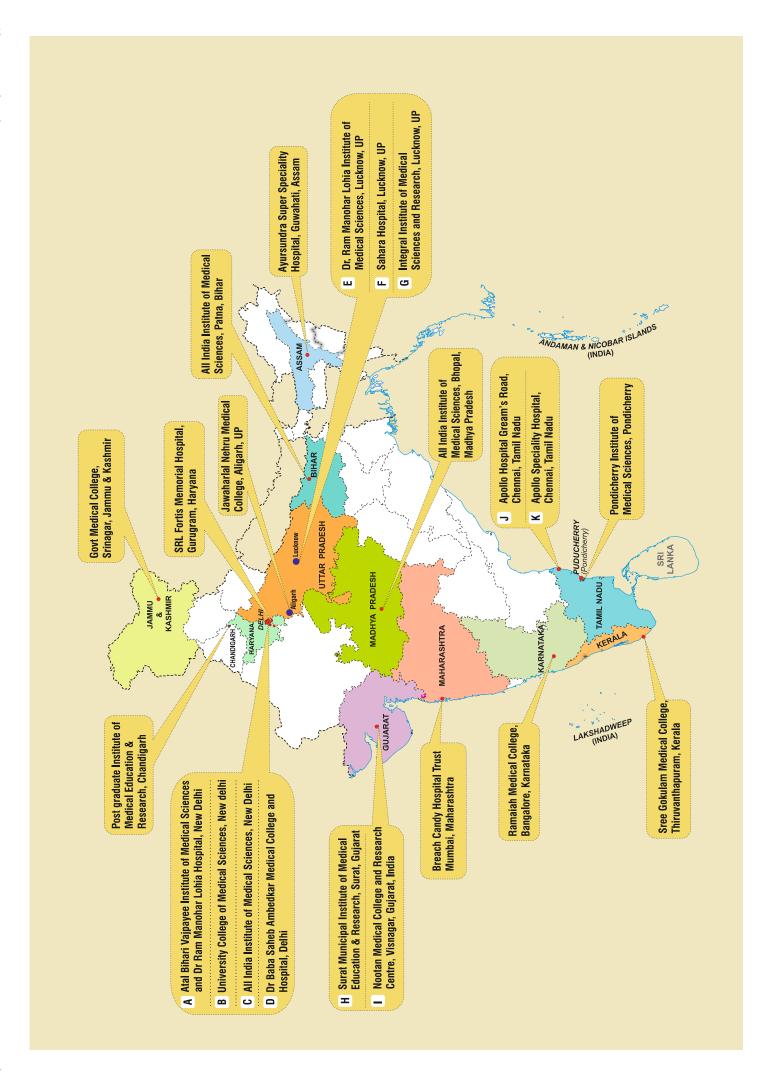
- [16] Huttner A, Kowalczyk A, Turjeman A, Babich T, Brossier C, Eliakim-Raz N, et al. Effect of 5-Day Nitrofurantoin vs Single-Dose Fosfomycin on Clinical Resolution of Uncomplicated Lower Urinary Tract Infection in Women: A Randomized Clinical Trial. JAMA. 2018 May 1;319(17):1781.
- [17] Abdelraheem WM, Mahdi WKM, Abuelela IS, Hassuna NA. High incidence of fosfomycin-resistant uropathogenic E. coli among children. BMC Infect Dis. 2023 Jul 17;23(1):475.
- [18] Tutone M, Bjerklund Johansen TE, Cai T, Mushtaq S, Livermore DM. SUsceptibility and Resistance to Fosfomycin and other antimicrobial agents among pathogens causing lower urinary tract infections: findings of the SURF study. Int J Antimicrob Agents. 2022 May;59(5):106574.
- [19] Sharma R, Jain M, Vyas L, Singhal A. Hospital and community isolates of uropathogens and their antibiotic sensitivity pattern from a tertiary care hospital in North West India. Ann Med Health Sci Res. 2014;4(1):51.
- [20] Ranakishor P, Paditham M, Harika P, Chandrika Reddy A, Bhuvaneswari K, Srujana M, et al. Antibiotics susceptibility pattern and prevalence of isolated uropathogens in inpatient and out patients with lower urinary tract infections. J Appl Pharm Sci [Internet]. 2022 Jan 5 [cited 2023 Aug 28]; Available from: https://japsonline.com/abstract.php?article_id=3547&sts=2
- [21] Wan Y, Mills E, Leung RCY, Vieira A, Zhi X, Croucher NJ, et al. Alterations in chromosomal genes nfsA, nfsB, and ribE are associated with nitrofurantoin resistance in Escherichia coli from the United Kingdom. Microb Genomics [Internet]. 2021 Dec 3 [cited 2023 Sep 6];7(12). Available from: https://www.microbiologyresearch.org/content/journal/mgen/10.1099/mgen.0.000702
- [22] Bhargava K, Nath G, Bhargava A, Kumari R, Aseri GK, Jain N. Bacterial profile and antibiotic susceptibility pattern of uropathogens causing urinary tract infection in the eastern part of Northern India. Front Microbiol. 2022 Aug 9;13:965053.
- [23] Vijayganapathy S, Karthikeyan V, Mallya A, Mythri K, Viswanatha R, Keshavamurthy R. Antimicrobial resistance patterns in a tertiary care nephro-urology center in South India. J Integr Nephrol Androl. 2018;5(3):93.
- [24] Bharara T, Sharma A, Gur R, Duggal S, Jena P, Kumar A. Comparative analysis of extended-spectrum beta-lactamases producing uropathogens in outpatient and inpatient departments. Int J Health Allied Sci. 2018;7(1):45.
- [25] Losada I, Barbeito G, García-Garrote F, Fernández-Pérez B, Malvar A, Hervada X, et al. Estudio de sensibilidad de Escherichia coli productores de infecciones del tracto urinario comunitarias en Galicia. Período: 2016-2017. Aten Primaria. 2020 Aug;52(7):462–8.

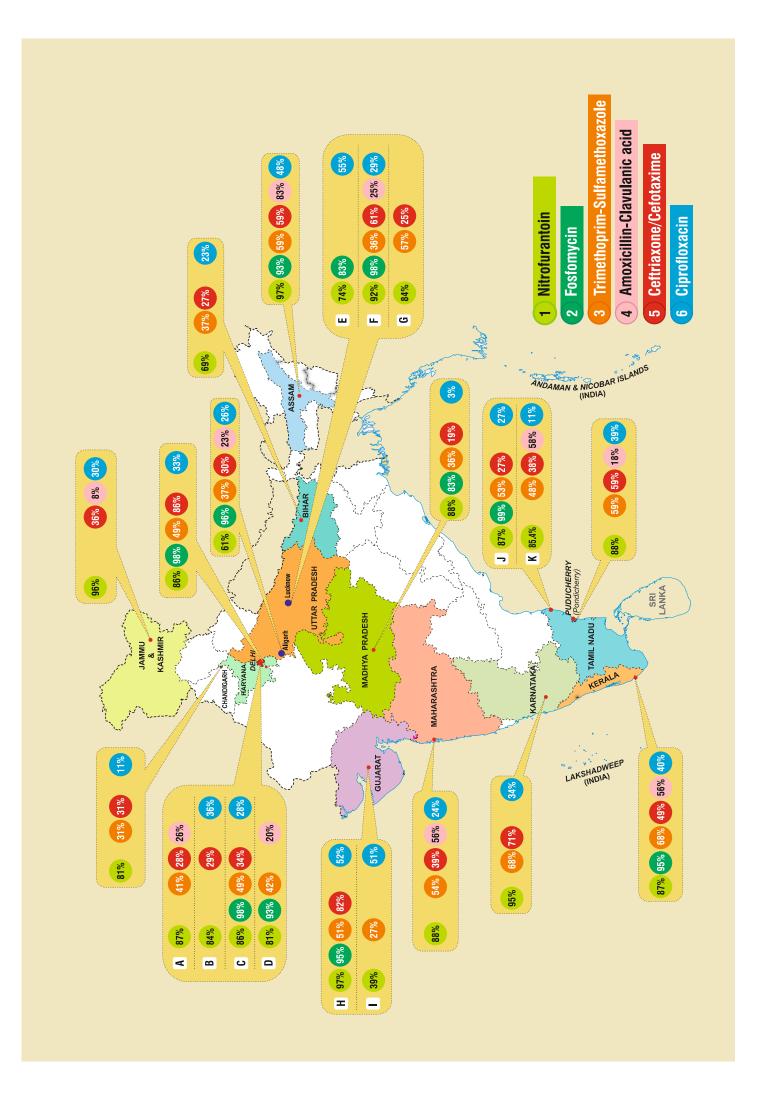
Wagenlehner F, Nicolle L, Bartoletti R, Gales AC, Grigoryan L, Huang H, et al. A global perspective on improving patient care in uncomplicated urinary tract infection: expert consensus and practical guidance. J Glob Antimicrob Resist. 2022 Mar;28:18–29.

- [27] Paul D, Anto N, Bhardwaj M, Prendiville A, Elangovan R, Bachmann TT, et al. Antimicrobial resistance in patients with suspected urinary tract infections in primary care in Assam, India. JAC-Antimicrob Resist. 2021 Sep 30;3(4):dlab164.
- [28] Behera B, Debbarma M, Rout B, Baral P, Das S, Jena L, et al. Prevalence of ESBL Producing Bacteria in Community-Acquired UTI from Eastern Part of India. J Pure Appl Microbiol. 2022 Sep 1;16(3):1682–8.
- [29] Kumar N, Chatterjee K, Deka S, Shankar R, Kalita D. Increased Isolation of Extended-Spectrum Beta-Lactamase-Producing Escherichia coli From Community-Onset Urinary Tract Infection Cases in Uttarakhand, India. Cureus [Internet]. 2021 Mar 11 [cited 2023 Aug 28]; Available from: https://www.cureus.com/articles/50456-increased-isolation-of-extended-spectrum-beta-lactamase-producing-escherichia-coli-from-community-onset-urinary-tract-infection-cases-in-uttarakhand-india
- [30] Gajamer VR, Bhattacharjee A, Paul D, Deshamukhya C, Singh AK, Pradhan N, Tiwari HK. Escherichia coli encoding blaNDM-5 associated with community-acquired urinary tract infections with unusual MIC creep-like phenomenon against imipenem. J Glob Antimicrob Resist. 2018 Sep;14:228-232. doi: 10.1016/j.jgar.2018.05.004. Epub 2018 Aug 9. PMID: 29775789
- [31] Nair PK. Prevalence of carbapenem resistant Enterobacteriaceae from a tertiary care hospital in Mumbai, India. J Microbiol Infect Dis. 2013 Dec 1;03(04):207–10.
- [32] Gardiner BJ, Stewardson AJ, Abbott IJ, Peleg AY. Nitrofurantoin and fosfomycin for resistant urinary tract infections: old drugs for emerging problems. Aust Prescr. 2019 Feb;42(1):14–9.
- [33] Kotwani A, Joshi J, Lamkang AS. Over-the-Counter Sale of Antibiotics in India: A Qualitative Study of Providers' Perspectives across Two States. Antibiotics (Basel). 2021 Sep 17;10(9):1123. doi: 10.3390/antibiotics10091123. PMID: 34572705; PMCID: PMC8472180.
- [34] Mukherjee A, Surial R, Sahay S, Thakral Y, Gondara A. Social and cultural determinants of antibiotics prescriptions: analysis from a public community health centre in North India. Front Pharmacol. 2024 Jan 25;15:1277628. doi: 10.3389/fphar.2024.1277628. PMID: 38333004; PMCID: PMC10850286.
- [35] Gupta, P., Patel, S.A., Sharma, H. et al. Burden, patterns, and impact of multimorbidity in North India: findings from a rural population-based study. BMC Public Health 22, 1101 (2022). https://doi.org/10.1186/s12889-022-13495-0
- [36] McNulty CAM, Richards J, Livermore DM, Little P, Charlett A, Freeman E, et al. Clinical relevance of laboratory-reported antibiotic resistance in acute uncomplicated urinary tract infection in primary care. J Antimicrob Chemother. 2006 Sep 6;58(5):1000–8.

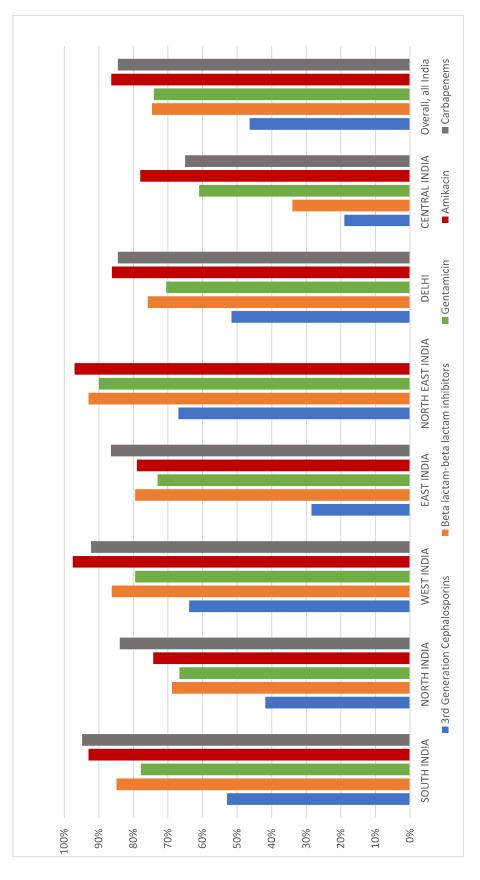
Legends

- Figure 1: Participating States and Centres
- **Figure 2:** Antimicrobial Susceptibility profile of Escherichia coli to the major antibiotic groups. Number of strains tested for each group were as follows: nitrofurantoin (7790), fosfomycin (4165), trimethoprim-sulphamethoxazole (6639), amoxicillin-clavulanic acid (4307), ceftriaxone/cefotaxime (6014), ciprofloxacin (6712)
- Figure 3: Average susceptibility of *Escherichia coli* to five major antimicrobial groups.
- Figure 4: Estimated regional prevalence of ESBLs and carbapenem resistance









Carbapenems: average of imipenem and meropenem. Number of strains tested were as follows: Third-generation cephalosporins: ceftazidime (3871), cefotaxime (3369), ceftriaxone (2645), and cefixime (530). Beta-Lactam-beta-lactamase inhibitors: piperacillin-3-Lactam- β -lactamase inhibitors: average of piperacillin-tazobactam and cefoperazone/sulbactam Third-generation cephalosporins: average of ceftazidime, cefotaxime, ceftriaxone, and cefixime

tazobactam (5242) and cefoperazone/sulbactam (4094). Aminoglycosides: gentamicin (6834), amikacin (6945) Carbapenems: imipenem (6203) and meropenem (7064)

Table 1: Antibiotics tested, and nationwide antimicrobial susceptibility profile of Escherichia coli isolated from outpatients

					Resistan	Resistance rates across different centres	s different ce	ntres
Total no. centres = 22	No. of centres testing the drug	% of centres testing the drug	Total no. of isolates tested	% of all isolates tested with drug	Nationwide average % susceptibility	Arithmetic mean %	Harmonic mean %	Median
Nitrofurantoin	22	100	7790	100%	%98	%98	85%	%98
Fosfomycin	10	45	4165	23%	%56	%56	94%	826
Trimethoprim- Sulfamethoxazole	19	98	6639	85%	49%	49%	48%	48%
Ampicillin	11	50	3871	50%	21%	21%	20%	21%
Cefazolin	4	18	3369	43%	76%	26%	26%	26%
Cefuroxime	6	41	3166	41%	76%	79%	728	26%
Cefoxitin	2	6	538	2%	41%	41%	41%	41%
Cefepime	11	20	4999	64%	%85	23%	25%	51%
Ceftazidime	11	20	3871	20%	%55	25%	54%	54%
Cefotaxime	6	41	3369	43%	47%	47%	%17	47%
Ceftriaxone	11	20	2645	34%	48%	48%	46%	48%
Cefixime	3	14	230	%2	41%	41%	41%	41%
Cefoperazone	1	5	222	3%	18%	18%	18%	18%

Ampicillin-Sulbactam	4	18	683	%6	31%	31%	78%	31%
Amoxicillin-Clavulanic acid	11	20	4307	25%	47%	47%	47%	48%
Piperacillin- Tazobactam	17	77	5242	%29	81%	81%	81%	81%
Cefoperazone/sulbact am	7	32	4094	53%	79%	%62	79%	79%
Imipenem	16	73	6203	80%	88%	88%	88%	89%
Meropenem	19	98	7064	91%	89%	89%	88%	91%
Gentamicin	19	98	6834	%88	76%	%9/	76%	77%
Amikacin	20	91	6945	%68	88%	%88	87%	89%
Norfloxacin	6	41	3425	44%	29%	29%	28%	29%
Levofloxacin	8	36	3313	43%	41%	41%	35%	41%
Ciprofloxacin	19	98	6712	%98	33%	33%	79%	34%

Key:	
Fewer than	Fewer than 33% of isolates tested
Adequate pr	Adequate proportion of isolates tested: % susceptibility
%06<	
81-90%	
71-80%	
61-70%	
40-60%	
<40%	

Drug	North	South	West	East	North-	Delhi-	Overall	ICC
	\mathbf{A}	В	\mathbf{C}	D	East	NCR	Susceptibility	
					E	F		
Fosfomycin	92.0%	97.0%	95.4%		93.1%	95.0%	93.6%	0.92
(N=3187)	(90.5; 93.5)	(92.3; 97.7)	(91.0; 99.7)		(83.9; 102.3)	(93.7; 96.3)	(88.6; 96.6)	
		*A						
Nitrofurantoin	81.0%	88.0%	93.1%	69.0%	96.6%	86.7%	86.6%	0.85
(N=6570)	(79.3; 82.7)	(86.8; 89.2)	(91.4; 94.8)	(64.9; 73.0)	(90.0; 103.2)	(85.4; 88.0)	(79.8; 92.0)	
	*D	*A *D	*A*B*D*F		*D	*A*D		
Trimethoprim-	43.0%	59.0%	52.0%	36.8%	58.6%	41.0%	45.6%	0.46
Sulfamethoxazole (N=5472)	(39.2; 46.8)	(57.2; 60.8)	(46.7; 55.3)	(32.6; 40.9)	(40.7; 76.5)	(39.0; 42.9)	(38.4; 53.6)	
		*A*C*D*F	*A*D*F					
Cefotaxime	52.8%	39.9%	85.1%	27.0%		29.0%	47.1%	0.36
(N=3336)	(45.7; 59.9)	(38.0; 41.7)	(77.7; 92.4)	(23.3; 30.7)		(26.8; 31.2)	(24.0; 72.2)	
	*B*D*F	*D*F	*A*B*D*F					
Ceftriaxone	38.0%	46.9%	61.1%		58.6%	36%	59.7%	0.50
(N=2645)	(35.7; 40.3)	(43.5; 50.3)	(58.2; 64.1)		(40.8; 76.4)	(81.5; 91.1)	(38.6; 78.0)	
		*A	*A*B					
Gentamicin	66.9%	78.1%	80.0%	78.6%	89.7%	71.0%	74.3%	0.71
(N=5674)	(63.7; 70.1)	(76.6; 79.6)	(77.3; 82.7)	(75.0; 82.2)	(78.7; 100.7)	(69.2; 72.8)	(67.8; 80.8)	
		*A*F	*A*F	*A*F				
Meropenem	81.2%	95.0%	94.0%	86.1%		85.9%	86.9%	0.85
(N=5989)	(79.5; 82.9)	(94.2; 95.8)	(92.4; 95.6)	(83.1; 89.1)		(84.2; 87.6)	(75.8; 93.4)	
		*A*D*F	*A*D*F			*A		
Ciprofloxacin	33.0%	30.0%	37.9%	22.9%	48.3%	27.0%	24.5%	0.26
(N=5702)	(30.6; 35.4)	(28.4; 31.7)	(34.7; 41.1)	(19.3; 26.6)	(30.2; 66.4)	(24.9; 29.1)	(12.0; 43.7)	
		*D	*B*D*F		*D			
Piperacillin-	65.0%	81.0%	86.9%	82.0%	93.1%	80.0%	76.7%	0.71
Tazobactam (N=4970)	(63.2; 66.8)	(79.6; 82.4)	(84.7; 89.1)	(78.7; 85.3)	(83.9; 102.3)	(77.8; 82.2)	(59.6; 88.6)	
` '		*A	*A*B*F	*A	*A	*A		
		1 (0 00/	1 04 00/	43.9%		36.0%	48.1%	0.48
Cefepime (N=4021)	40.9% (37.7; 44.1)	60.0% (57.9; 62.1)	84.0% (81.6; 86.4)	(39.7; 48.1)		(32.7; 39.3)	(27.8; 69.1)	0.10

^{*} Results are based on two-sided z-tests with a significance level p <0.05. For pair-wise comparison of susceptibility profile between regions, the region with lower susceptibility (labelled by the bold capital alphabet) is placed within the region which has significantly higher susceptibility compared to it. (i.e. E.coli showed a statistically significantly higher susceptibility to fosfomycin in the South region than in the North region). Tests are adjusted for all pairwise comparisons within a row of each innermost subtable using the Bonferroni correction. 95% confidence interval is provided in parenthesis.

Supplementary Table S1: Detailed analysis of antimicrobial susceptibility profile of Escherichia coli across India

Cefoperazone	ı	1	ı								
nitziloO				%66		%66	%66	%66	1153		
Meropenem	95%	%86	%06	93%	97%	82%	82%	82%	1946	%89	%68
mənəqiml	%26	%86	%06	93%	%26	82%	%56	82%	1946	74%	%06
\enoseracofe) metaedlus	1	92%	ı	%98	ı	%68	%68	%68	1293	ı	1
-nillioeragiq metoedoseT	74%	92%	%92	81%	81%	81%	81%	%08	1946	61%	57%
-nillioixomA bios oinsluvslO	1	26%	1	1	58%	21%	21%	21%	246	1	25%
-nillioiqmA metoedlu2	1	1	1	1	1					26%	1
Ciprofloxacin	39%	40%	34%	27%	11%	30%	78%	24%	1946	55%	16%
Levofloxacin	20%	1	1	ı	13%	32%	25%	21%	480	53%	1
niosacin	28%	1	42%	27%	ı	42%	40%	38%	1700	1	ı
Mikacin	1	%88	93%	%96	95%	%86	%86	83%	1572	%99	87%
nioimetnee	75%	84%	75%	%62	76%	%82	%8/	%8/	1946	53%	70%
əmiqə l əϽ	-	78%		25%	51%	61%	%09	%69	1399	34%	54%
әтіхітәӘ	1	%89	ı	1	ı	%89	%89	%89	140	1	1
Seftriaxone	53%	49%		1	38%	47%	46%	46%	620	1	61%
əmixstofəƏ	41%	1	1	38%	ı	40%	39%	39%	1527	1	53%
əmibizafləƏ	1	1	71%	ı	44%	28%	%95	54%	279	37%	61%
ntiixofəO	1	1	1	1	ı					49%	
Sefuroxime	20%	48%	38%	1	25%	33%	31%	762	793	1	19%
nilozetəƏ			30%	ı	,	30%	30%	30%	173	14%	
nilliziqmA	23%	30%	17%	1	26%	24%	23%	23%	793	,	18%
elozexodiemeilus	29%	%89	%89	53%	48%	%65	29%	28%	1946		36%
Trimethoprim-	25	89 %56	39	83 %66	48	97% 59	92% 26	85 %/6	1293 19	- 83%	38% 36
Fosfomycin	1		,		, ,0						
niofnerufortiN	%88 t	87%	3 95%	83 87%	85%	%88	%88	%88	1946	74%	3 92%
Total		140	173	1153	106	_				451	178
E.coli	Pondicherry Institute of Medical Sciences, Pondicherry	Sree Gokulam Medical College, Thiruvanthapuram, Kerala	Ramaiah Medical College, Bangalore, Karnataka	Apollo Hospital Gream's Road, Chennai, Tamil Nadu	Apollo Speciality Hospital, Chennai, Tamil Nadu	Average	Geometric mean	Harmonic mean	Total isolates tested	Dr. Ram Manohar Lohia Institute of Medical Sciences, Lucknow, UP	SAHARA Hospital, Lucknow, UP
			South India:	Pondicherry, Tamil Nadu, Karnataka, Kerala						North India: Uttar Pradesh and	Kashmir

Govt Medical College, Srinagar, Jammu & 471 96% Kashmir Jawaharlal Nehru Medical College, Aligarh Average Geometric mean Harmonic mean Total isolates tested Total solates		37% 43% 42% 41% 593	2%																			
Jawaharlal Nehru 303 61% Medical College, Aligarh 81% Average 81% Geometric mean 79% Harmonic mean 79% Total isolates tested 1515 Breach Candy Trust 663 88% Maharashtra 663 88% Surat Municipal Institute of Medical Education & Research, Surat, Gujarat 87 97% Average Average 93% Geometric mean 92%		42% 42% 41% 593		1		1	j	36%	8				26%	19%	30%		%8		73%		%08	
Average 81% 80% Harmonic mean 79% 79% 79% 70tal isolates tested 15.15 15.15		43% 42% 41% 593						30%	33%	30		25%	24%				23%				92%	
Geometric mean Harmonic mean Total isolates tested Breach Candy Trust Hospital, Mumbai, Maharashtra Surat Municipal Institute of Medical Education & 87 Research, Surat, Gujarat Average Geometric mean 92%		42%	10%	20% 1	19% 4	49% 43	43% 53	23% 38%	33%	41%	%19	74%	25%	36%	33%	39%	19%	. %59	73%	84%	84%	
Harmonic mean Total isolates tested Breach Candy Trust Hospital, Mumbai, Maharashtra Surat Municipal Institute of Medical Education & 87 Research, Surat, Gujarat Average Geometric mean 92%		593	%9	19% 1	19% 4	49% 42	42% 53	23% 36%	33%	40%	%99	73%	25%	32%	30%	%98	17%	. 64%	73%	84%	83%	
Total isolates tested 1515		593	4%	18% 1	19% 4	49% 40	40% 53	53% 34%	% 33%	% 39%	%59	71%	25%	78%	27%	34%	14%	. 64%	73%	84%	83%	
Breach Candy Trust Hospital, Mumbai, Maharashtra Surat Municipal Institute of Medical Education & 87 Research, Surat, Gujarat Average Geometric mean 92%	%56 82%		649	563	178 4	451 74	741 17	178 1064	64 303	3 741	741	1044	774	922	1212	563	952	741	471	741	1515	
Breach Candy Trust Hospital, Mumbai, Maharashtra Surat Municipal Institute of Medical Education & 87 Research, Surat, Gujarat Average Geometric mean 92%	%56 828	Z 40/																				
Surat Municipal Institute of Medical Education & 87 97% Research, Surat, Gujarat Average Geometric mean 92%	95%	34%	21%		29%			39%	<u>%</u>	78%	85%	826			24%		26%	81%	85%	91%	62% 5	%66
93%		51%	18%	29%	32%	33% 87%		85% 82%	% 53%	91%	74%	%86		61%	52%	23%		94%		-	%56	
85%	%56	23%	70%	29%	31% 3	33% 87%		85% 61%	% 23%	%58 %	%08	%86		61%	38%	23%	%95	%88	%58	91%	94% 5	%66
_	%56	25%	19%	29%	30%	33% 87	87% 85	85% 57%	% 23%	84%	%62	%26		61%	35%	23%	%95	87%	%58	91%	63%	%66
Harmonic mean 92% 95	%56	25%	19%	36%	30%	33% 87	87% 85	85% 53%	% 23%	84%	%62	%26		61%	33%	23%	%95	81%	85%	91%	63%	%66
Nootan Medical College (Outlier) and Research Centre, 33 39% Visnagar, Gujarat, India		27%					88	_%			36%	36%	%06		51%	% 6		3%				
Total isolates tested 783 87	87	783	750	87 7	750 8	87 87		120 750	0 87	750	783	783	33	87	783	120	663	783	663	. 699	750 (663
-																						-
All India Institute of Medical Sciences, Patna, 467 69% East India: Bihar Bihar		37%	11%		21%		30% 27	27%		44%	73%	79%	28%		23%			85%	77%	87%	%98	
Average 69%		37%	11%		21%	30	30% 27	27%		44%	73%	%62	28%		23%			82%	%//	87%	%98	
Geometric mean 69%		37%	11%		21%	30%		27%		44%	73%	%62	78%		23%			. %78	%//	81%	%98	

		1										18%	18%	18%	18%		222
										%66		1%	20%	10%	7%		404
%98	467							72%	93%	95%		%88	%98	%98	82%	42%	1408
87%	467							74%	74%	93%		%06	83%	82%	82%	35%	1408
%//	467											72%	72%	72%	72%		222
85%	467		93%	88%	886	93%	29	%89	%88	83%			%08	%62	%62		1037
			83%	83%	83%	83%	29	25%			20%		23%	22%	22%	% 9	1439
23%	467		48%	48%	48%	48%	29		%98	33%	28%	11%	27%	25%	22%		1297
										40%	27%		34%	33%	32%		846
28%	467								25%			16%	21%	20%	20%		451
%62	467		%26	%26	%26	%26	29	74%	91%	94%	%88	84%	%98	%98	%98	42%	2072
73%	467		%06	%06	%06	%06	29	%29	81%		74%	%09	71%	%02	%02	29%	1890
44%	467										%9E		%98	36%	36%		664
			29%	29%	29%	29%	29			36%			%98	%98	%98		182
27%	467							28%	29%			31%	78%	79%	78%		1077
30%	467		75%	75%	75%	75%	29	45%			34%		40%	39%	39%		1290
21%	467																
11%	467		45%	45%	45%	45%	29				16%		16%	16%	16%		664
37%			29%	29%	29%	29%		41%		49%	42%	31%	41%	40%	40%	31%	1843
<u></u>	467		93% 26	93% 26	93% 26	93% 26	9 29	4.		98% 46	93% 47	Ж	96% 4:	95% 4(95% 4(Ж	846 18
	_						29	%	%			%				%	
%69	467		%26	826	826	%26	29	.6 87%	.9 84%	.2 86%	.4 84%	2 81%	84%	84%	84%	%92 6	2072
			29					626 al,	, 229	182	, 664	2 22				149	
ic mean	Total isolates tested		Ayursundra Super Speciality Hospital, Guwahati, Assam		Geometric mean	ic mean	Total isolates tested	Atal Bihari Vajpayee Institute of Medical Sciences and Dr Ram Manohar Lohia Hospital, New Delhi	University College of Medical Sciences, New delhi	SRL Fortis Memorial Hospital, Gurugram, Haryana	All India Institute of Medical Sciences, New Delhi	Post graduate Institute of Medical Education & Research, Chandigarh		Geometric mean	ic mean	Dr Baba Saheb Ambedkar Medical College and Hospital, Delhi	Total isolates tested
Harmonic mean	Total iso		Ayursun Specialit Guwaha	Average	Geomet	Harmonic mean	Total iso	Atal Bihari Institute of Sciences ar Manohar L New Delhi	Universi Medical delhi	SRL Fortis Hospital, Haryana	All India Medical Delhi	Post gra of Medio Research	Average	Geometi	Harmonic mean	Dr Baba Saheb Ambedkar Mec College and Ho Delhi	Total iso
			_		India: Assam					<u> </u>	Haryana, Chandigarh					(Outlier)	1

Central India: Madhya	All India Institute of Medical Sciences, Bhopal, Madhya Pradesh	%88	83%	36%	13	13%	19%		21%	61%	78%	9	8% 3	3%	18	18% 32%	36%	%99 %	64%	~	
Pradesh	Average	%88 83%		%98	13	13%	19%		21%	61%	%82	9	8 89	3%	18	18% 32%	%98 %	%99 %	% 64%	%	
	Total isolates tested	978	876 876 876	826	978	<u></u>	826		8/6	826	826	J,	6 8/6	826	826	8.48	8. 978	8 978	8 978	~	

Key:	
%06<	
81-90%	
71-80%	
61-70%	
40-60%	
<40%	

Supplementary Table S2. Antimicrobial susceptibility rates for urinary Escherichia coli, by region

Colistin	66		100	1			66	100	1	ı	1	1	50	0	100
Meropenem	95	06	- 86	84	89	- 6	94	92 - 95 1	98	98	1	-	98	2 (93 1
mənəqiml	95	06	- 86	84		- 06		91 9	87 8	87 8	1	-	83 8	74 7	6
metoedlus/ənozerəqofəD	68	98	- 92	73		73	85	85 (77	77	1	-	72		72
Piperacillin - tazobactam	81	74	- 36	65	22	- 92	88	81 - 94	82	40	93	93	80	89	- 88
bioA oinsluvslO - nillioixomA	57	56	- 82	19	∞	25	56	99	,	ı	83	83	23	(- 25
metoedlu2 - nillioiqmA			1	39	56	- 51	23	23	,	ı	-	-	,		-
Ciprofloxacin	30	11	- 04	33	16	- 55	38	24 - 52	23	23	48	48	27	11	- 36
Levofloxacin	32	13	- 20	36	19	53	61	61	,	,	-	-	34		- 40
Norfloxacin	42	27	- 28	25	24	- 26			28	28	1	-	21		- 25 4
Amikacin	93	88	- 96	74	25	- 68	86	97 - 98	79	19	97	16	98	_	94
nicimetnad	78	75	- 84	29	53	- 12	80	74 -	73	73	90	06	71	09	- 81
əmiqə 1 əO	61	51	- 8/	41	34	- 54	85	78 - 91	44	44	,	-	36		36
əmixifəƏ	89		89	33		33	23	23	1	,	1	-	1		1
Seffriaxone	47	38	- 53	38	25	- 61	61	39 - 82	1	ı	59	59	98		98
emixetofeO	40	38	- 41	53		53	85	85	27	27	,		29	28	31
əmibizettəD	58	44	71	43	32	- 61	87	87	30	30	75	75	40	34	- 45
nitixofəO			1	49		49	33	33		ı		-	1		1
əmixorufəƏ	33	20	- 84	19		19	31	29 - 31	21	21	1	-	1		-
nilosefa	30		30	20	14	- 25	29	29	,	ı	1	-	1		1
nillisiqmA	24	17	30	10	2	- 18	20	18 - 21	11	11	45	45	16		16
elosexominto	59	48	- 89	43	36	- 57	53	51 - 54	37	37	59	59	41	31	- 49
Fosfomycin	97	92	- 66	92	83	- 86	95	95	1		93	93	96	93	- 98
Nitrofurantoin	88	87	- 95	81	61	- 96	93	88 - 97	69	69	97	97	84	81	- 87
Harmonic Mean and sange	Mean		range	Mean		range	Mean	range	Mean	range	Mean	range	Mean		range
	HIIIOS	INDIA			NORTH	E C	TOOM	INDIA	EAST	INDIA	NORTH	EASI		DELHI	

		-
	64	64
	99	99
	36	36
	32	32
	18	18
	-	
	3	3
	9	9
	-	-
	78	78
	61	61
	21	21
	-	ı
	-	1
	-	
	19	19
		-
	13	13
	-	ı
	_	-
	36	36
	83	83
	88	88
	Mean	range
CENTR	AL	INDIA