

SUceptibility and Resistance to Fosfomycin and other antimicrobial agents among pathogens causing lower urinary tract infections: findings of the SURF study

Authors: Michaela Tutone,^a Truls E. Bjerklund Johansen,^b Tommaso Cai,^c Shazad Mushtaq^d, David M Livermore^e

Affiliations:

^a Global Medical Affairs, Zambon SpA, Bresso (MI), Italy

Postal address: Zambon SpA, Via Lillo del Duca, 10, 20091 Bresso (MI), Italy.

Michaela.Tutone@gmail.com

^b Department of Urology, Oslo University Hospital and Institute of Clinical Medicine, University of Oslo, Norway and Institute of Clinical Medicine, University of Aarhus, Denmark

Postal address: Institutt for klinisk medisin, Universitetet i Oslo, Postboks 1171 Blindern 0318, Oslo. tebj@medisin.uio.no

^c Department of Urology, Santa Chiara Regional Hospital, Trento, Italy and Institute of Clinical Medicine, University of Oslo, Norway

Postal address: Institutt for klinisk medisin, Universitetet i Oslo, Postboks 1171 Blindern 0318, Oslo. ktommy@libero.it, tommaso.cai@apss.tn.it

^d Antimicrobial Resistance and Healthcare-Associated Infections Reference Unit, UK Health Security Agency, Colindale Avenue, London, UK, NW9 5EQ. SHAZAD.MUSHTAQ@PHE.gov.uk

^e Norwich Medical School, University of East Anglia, Norwich, UK, NR4 7TJ

Postal address: Norwich Medical School, Bob Champion Research and Educational Building,

Rosalind Franklin Road, University of East Anglia, Norwich Research Park, Norwich, UK, NR4
7UQ. d.livermore@uea.ac.uk

Corresponding author:

Michaela Tutone
Zambon SpA,
Via Lillo del Duca, 10
20091 Bresso (MI)
Italy
Michaela.Tutone@gmail.com

Abbreviations

AMRHAI, antimicrobial resistance and healthcare associated infections

API, Analytical Profile Index

CLED, cysteine lactose electrolyte deficient

CLSI, Clinical and Laboratory Standards Institute

EMA, European Medicines Agency

ESBLs, extended-spectrum β -lactamases

EUCAST, European Committee on Antimicrobial Susceptibility Testing

MALDI-ToF, matrix-assisted laser desorption/ionization time-of-flight

MIC, minimum inhibitory concentration

NA, not applicable

REWIND, REal World INternational Database

SURF, Surveillance sUsceptibility and Resistance to Fosfomycin in comparison with other antimicrobial agents study

UTI, urinary tract infection

uUTI, uncomplicated urinary tract infection

1 **Abstract**

2 **Background:** Urinary tract infections (UTIs) are prevalent world-wide, particularly
3 among women. Their incidence increases with age, and treatment is increasingly
4 challenging owing to antibiotic resistance and the lack of new agents. We investigated
5 the susceptibility of current urinary isolates to fosfomycin and other antibiotics across
6 Europe.

7 **Methods:** This cross-sectional study collected consecutive urinary isolates from non-
8 hospitalised women at 20 centres in Belgium, UK, Italy, Spain and Russia. Bacteria were
9 tested by disk diffusion with relevant antibiotics. As a quality control, a central
10 laboratory re-tested, by agar dilution: (i) isolates found resistant to fosfomycin, and (ii)
11 every tenth isolate; all non-Russian sites were included.

12 **Results:** A total of 2848 isolates were analysed, principally *Escherichia coli* (2064,
13 72.5%), *Klebsiella* spp. (275, 9.7%) and 103 *Proteus* spp. (103, 3.6%). For *E. coli*, agents
14 active against >90% of isolates were nitrofurantoin (98.5%), fosfomycin (96.4%), and
15 mecillinam (91.8%). Fosfomycin and nitrofurantoin remained active against >90% of
16 cephalosporin-resistant *E. coli*. Among 143 *E. coli* recorded as susceptible locally by
17 disk tests, 138 (96.5%) were confirmed susceptible by MIC tests, however resistance
18 was only confirmed in 29/58 (50%) of those reported resistant by local disk tests.

19 **Conclusion:** *E. coli* was found to be the most common uropathogen isolated and was

20 highly susceptible to fosfomicin, nitrofurantoin and mecillinam, all used effectively for
21 more than 30 years. Guidelines advocating fosfomicin for uUTIs in women remain
22 microbiologically valid.

23

24 **Keywords:** urinary tract infection, uropathogen, fosfomicin trometamol, antibiotic
25 susceptibility, cystitis, *Escherichia coli*

26 **1 Introduction**

27 Urinary tract infections (UTIs) – principally uncomplicated cystitis in women – are
28 among the most common bacterial diseases in humans [1]. They are a significant cause
29 of morbidity at all ages [2] but there is an increased prevalence in women aged 15–24
30 years and aged ≥ 45 years [3, 4]. The most prevalent pathogen is *Escherichia coli*,
31 accounting for 80% of cases, but other Enterobacterales are also frequent, notably
32 *Klebsiella pneumoniae* [5, 6].

33 If untreated, or if the treatment fails, cystitis can precipitate ascending infections,
34 including pyelonephritis and sepsis, with renal damage [2]. The main reason for failure
35 is resistance to the antibiotics used, which standardly include β -lactams, trimethoprim,
36 and co-trimoxazole [7, 8]; fluoroquinolones also are still widely used, as revealed in the
37 multi-national REWIND (REal World INternational Database) study [9], although they
38 are no longer recommended in European or international guidelines. Resistance to
39 these standardly used antibiotics is increasing, although its prevalence varies among
40 countries [10-13]. Most resistance, except to fluoroquinolones, is determined via
41 acquired plasmids, including those encoding extended-spectrum β -lactamases (ESBLs),
42 which inactivate cephalosporins [8, 11, 14].

43 The challenge of resistance, along with the paucity of novel antibiotics, highlight the
44 need to re-evaluate older alternatives [15]. These include fosfomycin, an agent known
45 for over 40 years, which is available as an oral trometamol salt as well as in parenteral

46 formulations. A systematic review and meta-analysis has shown that a single dose of
47 fosfomycin trometamol was as effective as longer courses of alternative agents for
48 uncomplicated UTIs (uUTIs) in women [16].

49 Fosfomycin has an inherently broad spectrum of activity; however, the European
50 Committee on Antimicrobial Susceptibility Testing (EUCAST) now only has breakpoints
51 (S <8, R >8 mg/L) for *E. coli* in respect of the trometamol formulation used for uUTI
52 [17].

53 The aim of the European-wide ‘Surveillance sUsceptibility and Resistance to
54 Fosfomycin in comparison with other antimicrobial agents study’ (SURF) was to
55 provide a current snapshot of the prevalence of resistance to fosfomycin compared
56 with that to other oral antibiotics frequently prescribed to treat uUTIs in women.

57 **2 Materials and methods**

58 **2.1 Study design and isolates**

59 SURF was a cross-sectional epidemiological study on bacteria isolated from urine
60 samples collected from women between April 2019 and November 2019. Twenty
61 laboratories located across five countries participated, comprising three in Belgium,
62 two in UK, five in Italy, four in Spain and six in Russia (see Acknowledgements). To
63 avoid selection bias, the study protocol required collection of urine samples from all
64 consecutively sampled non-hospitalised women who (according to clinical referral or

65 the International Classification of Diseases coding system) were believed to have a
66 lower UTI. No clinical data were collected and since the 'study subjects' were the
67 uropathogens, not the patients, only limited institutional review was required and
68 obtained.

69 **2.2 Sample processing and analysis**

70 Isolates were sub-cultured and streaked for single colonies on MacConkey or Cysteine
71 Lactose Electrolyte Deficient (CLED) agar. Local laboratories used the following
72 methods to identify the organisms: matrix-assisted laser desorption/ionization time-of-
73 flight (MALDI-ToF) mass spectroscopy (14 sites), Vitek or other automated systems (5
74 sites) or Analytical Profile Index (API) strips (1 site). Diffusion susceptibility tests were
75 performed for each isolate using disks containing fosfomycin (plus glucose-6-
76 phosphate) 200 µg, amoxicillin/clavulanate 20/10 µg, ampicillin 10 µg, cefpodoxime
77 10 µg, cefalexin 30 µg, ciprofloxacin 5 µg, trimethoprim 5 µg, mecillinam 10 µg and
78 nitrofurantoin 100 µg. These disks were obtained centrally from Thermofisher (Life
79 Technologies Italia Fil. Life Technologies Europe BV Via G.B. Tiepolo, 18 I-20900 Monza
80 MB, Italy) and distributed to the sites to ensure consistent quality. Sites used Mueller-
81 Hinton agar and confluent growth, following the test method shared by both EUCAST
82 and the CLSI. Disk diffusion zone diameters were interpreted according to current
83 EUCAST breakpoints (EUCAST 2021), following the amendments to guidance on
84 fosfomycin susceptibility testing [18].

85 Given that *E. coli* is the most commonly identified uropathogen and that EUCAST 2021
86 only provides zone diameter breakpoints for fosfomicin applied to *E. coli* [18], our
87 analysis focused on this species.

88 **2.3 Quality control**

89 As a representative quality control sample, laboratories in all participating countries
90 except Russia sent a subculture of every tenth bacterial isolate to a central laboratory
91 (Antimicrobial Resistance and Healthcare Associated Infections [AMRHAI] Reference
92 Unit, Public Health England, now UK Health Security Agency) for re-testing. They also
93 sent all isolates, irrespective of species, with a zone diameter <24 mm, corresponding
94 to the then (and current) EUCAST disk breakpoint.

95 Isolates received by AMRHAI were re-identified by MALDI-ToF mass spectroscopy
96 (Biotyper, Bruker, Bremen Germany), then minimum inhibitory concentrations (MICs)
97 were determined using the CLSI agar dilution method [19]. Fosfomicin, glucose-6-
98 phosphate, nitrofurantoin, ampicillin, clavulanate, ciprofloxacin, trimethoprim and
99 cephalexin were purchased from Merck Life Sciences (Gillingham, UK); amoxicillin,
100 cefpodoxime and mecillinam were purchased from Alpha Aesar (Heysham, UK).

101 **2.4 Statistical analysis**

102 All statistical analyses were conducted using SAS[®] release 9.4 (SAS Institute, Inc., Cary,
103 NC, USA). No formal hypothesis was formulated; rather the analyses were exploratory

104 and contained analytical and descriptive aspects. No formal sample size estimation
105 was computed. Categorical data are presented as absolute and relative frequencies (n
106 and %). Chi-squared tests were used to compare proportions of resistant and
107 susceptible isolates between countries; data validation/sensitivity analyses considered
108 all isolates with paired (local and central laboratory) results.

109 Results were assigned to a not applicable (NA) category when there was no EUCAST
110 breakpoint defined for a particular organism/antibiotic combination, or when there
111 were missing values.

112 **3 Results**

113 **3.1 Isolates collected and tested**

114 A total of 2848 isolates were collected and tested: 473 (16.6%) in Belgium, 581 (20.4%)
115 in Italy, 565 (19.8%) in Spain, 393 (13.8%) in the UK and 836 (29.4%) in Russia (Figure
116 1). Among the 2012 non-Russian isolates, 543 (19.1%) were sent to the central
117 laboratory for re-testing as part of the quality control, and 542 (19.0%) were re-tested
118 there, with MICs determined. Ultimately, a total of 534 isolates (18.8%) had matching
119 paired local and central laboratory susceptibility results, comprising 116 (21.7%) from
120 Belgium, 150 (28.1%) from Italy, 150 (28.1%) from Spain and 118 (22.1%) from the UK
121 (Figure 1). Of the 534 paired isolate samples, 333 had been sent for central analysis
122 because they had an inhibition zone diameter measurement of <24 mm for fosfomycin
123 and 201 as part of the representative 10% sampling (26 fulfilled both criteria and, to

124 avoid double inclusion, are counted only within the 10% sample).

125 The most prevalent uropathogen was *E. coli* (n= 2064, 72.5%), followed, among
126 Enterobacterales, by *Klebsiella* spp. (n=275, 9.7%) and Proteeae (n=103, 3.6%);
127 substantial groups among the remaining 406 (14.3%) included *Enterococcus* spp.
128 (n=134), *Streptococcus* spp. (n=45), *Staphylococcus* spp. (n=28) and *Pseudomonas*
129 *aeruginosa* (n=21).

130 **3.2 Susceptibility and resistance among eligible *E. coli* isolates**

131 Based on the disk diffusion tests by local laboratories, only three agents –
132 nitrofurantoin, fosfomycin and mecillinam – were active against >90% of *E. coli*
133 isolates, with resistance rates of 0.5%, 3.6% and 8.2%, respectively. Of these, only
134 nitrofurantoin and fosfomycin retained activity against over 90% of the 348 (16.9%) *E.*
135 *coli* isolates that were resistant to cefpodoxime and which accordingly were inferred to
136 be ESBL producers or AmpC hyperproducers. Mecillinam appeared active against
137 84.2% of these cefpodoxime-resistant isolates, though its clinical efficacy against ESBL
138 producers remains contentious [20, 21] even when these appear susceptible *in vitro*.
139 Cephalosporins, amoxicillin/clavulanate and ciprofloxacin were active against around
140 80% (79.2–83.1%) of all *E. coli* isolates, whereas trimethoprim and ampicillin inhibited
141 fewer than 70%. Under 60% of the cefpodoxime-resistant *E. coli* isolates were
142 susceptible to agents besides fosfomycin, nitrofurantoin and mecillinam.

143 Comparisons covering all species are challenging due to the lack of fosfomicin and
144 nitrofurantoin breakpoints, except for *E. coli*. Among agents with breakpoints for all
145 uropathogens, only mecillinam remained active against >90% of isolates;
146 cephalosporins, amoxicillin/clavulanate and ciprofloxacin were active against 80–
147 83.9%, trimethoprim against 71%, and ampicillin against 48.2% (Table 1).

148 Resistance rates differed significantly ($p < 0.01$) between countries for all the antibiotics
149 studied except trimethoprim. Fosfomicin and nitrofurantoin nonetheless retained
150 activity against >90% of *E. coli* in all the countries (Table 2), while mecillinam only
151 narrowly failed to do so, with resistance rates of 10.0–10.8% in Italy, Belgium and the
152 UK. Overall, the highest resistance rates were seen in Italy (highest for five of nine
153 antibiotics tested) and the UK (highest for four antibiotics); Russia had the lowest
154 resistance rates for five of the nine agents included. Resistance prevalence rates for
155 cefpodoxime and cephalexin closely tracked each other across countries, being highest
156 (27.5% and 26.3%, respectively) in the UK and lowest (6.4% and 8.0%, respectively) in
157 Spain (Table 2).

158 **3.3 Data validation**

159 Two hundred and one *E. coli* isolates were sent to the central laboratory for MIC
160 testing (Table 3). Among the 143 isolates submitted as fosfomicin susceptible, based
161 on zone diameters ≥ 24 mm, 138 were confirmed susceptible by MIC tests, with MICs
162 ≤ 8 mg/L, indicating a false susceptible rate from local laboratories of 3.5%. Four of the

163 five isolates sent as susceptible but found resistant by the central laboratory were
164 inhibited by fosfomycin at 16–32 mg/L, with only one isolate found substantially more
165 resistant (MIC 64 mg/L). The rate of false susceptible results was also low ($\leq 5\%$) for
166 cefpodoxime, cephalexin, ciprofloxacin, trimethoprim and nitrofurantoin, but was 9.3%
167 for mecillinam (17/183), 15.5% (14/90) for ampicillin and 21.6% (34/157) for
168 amoxicillin/clavulanate.

169 False resistance rates were higher. Only half (29/58, 50%) of the *E. coli* submitted as
170 ‘fosfomycin resistant’, based on zones < 24 mm were confirmed as resistant by dilution
171 testing (MIC > 8 mg/L), whereas the other 29 isolates were found susceptible, 22 of
172 them with MICs of ≤ 1 mg/L; for nitrofurantoin, 4/5 isolates submitted as resistant were
173 found susceptible, indicating a false resistance rate of 80%, albeit based on a tiny
174 group. Mecillinam (7/18, 39%), cefpodoxime (10/46, 22%), trimethoprim (10/71, 14%)
175 and cephalexin (5/44, 11%) also had false resistance rates $> 10\%$, whereas rates were
176 under 10% for amoxicillin/clavulanate, ampicillin and ciprofloxacin.

177 Three hundred and thirty-three non-*E. coli* isolates were re-tested for susceptibility to
178 fosfomycin at the central laboratory (Table 4), 36 with zones equal to or larger than
179 the *E. coli* breakpoint of 24 mm and 297 with smaller zones. Among the former 36,
180 ‘susceptible’ MICs ≤ 8 mg/L were confirmed for 14, including 3/4 *Klebsiella* spp. and 6/7
181 Proteae, though only for 5/25 isolates of other species. MICs > 8 mg/L were seen for
182 248/297 non-*E. coli* reported to give zones < 24 mm, including 20/23 Proteae and

183 116/123 'others'; 'discordances' of an MIC <8 mg/L but a zone <24 mm were
184 predominantly seen for *Klebsiella* spp. (39 cases among 151 *Klebsiella* tested with
185 zones <24 mm).

186 **4 Discussion**

187 This study aimed to identify the pathogens responsible for community uUTIs in women
188 and to assess their current antimicrobial resistance profiles across Europe, including
189 Russia. *In vitro* susceptibility is a strong predictor of the likely success of antimicrobial
190 treatment in uUTI [22]. Although most resistance-contingent treatment failures are not
191 seriously consequential, a minority do lead to more severe disease, mostly in the
192 elderly; in particular, *E. coli* bacteraemias are strongly associated with failure of
193 therapy of prior UTIs [23].

194 A total of 2848 isolates were analysed and, as expected, considerably the most
195 common uropathogen was *E. coli* (72.5%), followed by *Klebsiella* spp. (9.7%) and
196 *Proteus* spp. (3.6%). The dominance of *E. coli* is consistent with prior studies [12, 24,
197 25], especially in Europe and the USA [12].

198 Although the Infectious Diseases Society of America (IDSA) no longer advocates a
199 single threshold for the rate of resistance, as it previously did specifically for co-
200 trimoxazole [25], it is widely agreed that an antibiotic ceases to be appropriate as
201 empirical therapy for uUTI when the resistance rate reaches 15–20% [25-27]. A cut-off

202 of approximately 20% is also supported by cost-effectiveness studies [28, 29]. On this
203 criterion, the only agents considered here that retained acceptable activity were
204 nitrofurantoin, fosfomycin and mecillinam, with cefpodoxime, cephalexin,
205 ciprofloxacin and amoxicillin/clavulanate 20/10 µg marginal, having resistance rates
206 around 15–20% and trimethoprim and ampicillin unacceptable, owing to much higher
207 resistance rates. Ciprofloxacin is now also discouraged by the European Medicines
208 Agency owing to toxicity concerns [30].

209 The resistance of *E. coli* to antimicrobials has increased in both developed and
210 developing countries [31], partly owing to spread of the sequence type (ST)131
211 lineage, which is often fluoroquinolone resistant and carries cephalosporin-hydrolysing
212 ESBLs along with the inhibitor-resistant OXA-1 penicillinase [32, 33]. Among the
213 present 2848 isolates, 14% were cefpodoxime resistant, suggesting likely ESBL
214 production and, among these, over 90% remained susceptible to fosfomycin and
215 nitrofurantoin.

216 Resistance rates among *E. coli* isolates differed between countries, being highest in
217 Italy and the UK and lowest in Russia and Belgium. These patterns are somewhat
218 counterintuitive: resistance rates for bloodstream isolates are generally highest in
219 Mediterranean Europe rather than, for example, the UK [34]. This dissonance may
220 reflect either differences in antimicrobial use between countries, or differences in the
221 extent of testing biases: treatment is often empirical, with culture reserved for first-

222 regimen failures. Consequently, laboratory testing may be biased towards recurrent
223 and recalcitrant infections, more likely to harbour resistant bacteria [35]. We do not
224 know the extent of this confounder in different countries, nor how much it varied
225 between them. Nevertheless, despite its effect, over 90% of *E. coli* isolates were
226 susceptible to fosfomycin and nitrofurantoin in *all* countries, supporting their broad
227 utility, and consistent with the results of other recent surveys [36].

228 Site-to-site reproducibility is a challenge for all decentralised surveys of antimicrobial
229 susceptibility; moreover, fosfomycin is a challenging drug to test in the laboratory,
230 requiring addition of glucose-6-phosphate to the disks and the discounting of isolated
231 colonies within inhibition zones [37]. We addressed these issues by central re-testing,
232 using the reference agar dilution method, for every tenth isolate and for all those that
233 gave zones <24 mm. For *E. coli* the results were reassuring: only 5 of the 143 re-tested
234 isolates that had given zones \geq 24 mm proved resistant at 8 mg/L and only one of these
235 5 were resistant at the pre-2021 EUCAST fosfomycin trometamol breakpoint of 32
236 mg/L. 'False susceptibility' was a greater issue for amoxicillin/clavulanate and
237 mecillinam, with incidence rates of 34/157 and 17/183 respectively. The more
238 frequent error in the case of fosfomycin was that resistance was over-estimated by
239 disk diffusion, being confirmed by MIC testing in only half (29/58) *E. coli* isolates where
240 it was claimed from disk testing. No other agent had such a degree of resistance over-
241 estimation, except nitrofurantoin, where resistance was extremely rare.

242 When this project was initiated, EUCAST had an S \leq 32/R >32 breakpoint for fosfomycin
243 trometamol with *all* Enterobacterales; while the SURF study was in progress, this
244 breakpoint was lowered to S \leq 8/R >8 and narrowed to *E. coli*. The present data support
245 the view that disk testing has little reliability beyond *E. coli*, with poor MIC/zone
246 concordance found. Moreover, MIC distribution collated by EUCAST indicate that
247 modal values for fosfomycin for other relevant pathogens besides *E. coli* either equal
248 (*Klebsiella* spp.) or exceed (*Enterococcus* spp., and *Staphylococcus* spp.) an 8 mg/L
249 breakpoint [38]. The one notable and pertinent exception to these generalisations,
250 from the present data, is that the *E. coli* susceptibility criteria potentially might be
251 extended to Proteeae, where 6/7 collected isolates with zones >24 mm were
252 confirmed inhibited at 8 mg/L whereas 20/23 with zones <24 mm were confirmed
253 resistant at 8 mg/L (Table 4). Further zone/MIC correlation studies, along with
254 outcome data, are needed to resolve this issue and the role for fosfomycin trometamol
255 in uUTIs involving Proteeae.

256 Due to the dominance of *E. coli* and low rates of antimicrobial resistance, fosfomycin is
257 recommended as first-line treatment in many European countries, including Belgium
258 and Italy, and also in Russia and Brazil [9]. In addition, fosfomycin is recommended as a
259 first-line treatment for uUTIs in the latest European Association of Urology and IDSA
260 guidelines on urological infections [25, 39, 40]. The present results support such
261 guidance.

262 A strength of this study is that it included a large sample, covering five countries
263 (Belgium, Italy, Russia, Spain and UK), and should prove to be a useful source of quality
264 data on antibiotic resistance to guide empirical therapy. Limitations are that it
265 recruited consecutive laboratory isolates, rather than consecutive women presenting
266 with uUTI, and that individual patient characteristics were not collected. This
267 precluded exploration of reasons for differences in resistance rates between countries
268 and, as discussed earlier, raises questions on whether the extent of routine testing –
269 and the contingent sample bias towards difficult cases – may vary between countries.
270 Lastly, only four of the five participating countries sent samples to the central
271 laboratory to be re-tested.

272 In summary, stewardship is crucial to maintaining the utility of antibiotics, and should
273 be a key consideration for physicians managing UTIs. Good stewardship has two key
274 aspects: (i) ensuring that patients who need antibiotics swiftly receive active,
275 proportionate ones, and (ii) preventing over-use and disproportionate use of
276 antibiotics. In this context, fosfomycin and nitrofurantoin represent important tools for
277 the management of uUTIs owing to their low prevalence of resistance.

278 **5 Conclusion**

279 *E. coli* remains the most common causative uropathogen in all countries included in
280 this study. Due to the high susceptibility rates and acceptable resistance, especially
281 against *E. coli* isolates, fosfomycin, like nitrofurantoin, seems to be a good candidate to

282 effectively address antibiotic-resistant UTIs Europe-wide.

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314

315 **Conflicts of interest**

- 316 DML: Advisory Boards or ad hoc consultancy Accelerate, Antabio, Centauri, Entasis,
317 GSK, Integra-Holdings, Meiji, Menarini, Mutabilis, Nordic, Paion, ParaPharm, Pfizer,
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320 Menarini, Nordic, Pfizer and Shionogi. Relevant shareholdings – Dechra, GSK, Merck,
321 Pfizer and Perkin Elmer amounting to less than 10% of portfolio value; Share options:
322 T.A.Z. He also has nominated holdings in Avacta, Diaceutics, Evgen, Faron, Genedrive,

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324 Schemes but has no authority to trade these shares directly.

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328 **Author contributions**

329 **MT** contributed to study design and management, preparation of the manuscript,

330 review of all drafts and approval

331 **TEBJ** contributed to study design, preparation of the manuscript, review of all drafts

332 and approval

333 **TC** contributed to study design, preparation of the manuscript, review of all drafts and

334 approval

335 **SM** led the central laboratory 'quality control' testing and interpretation of results, and

336 reviewed drafts

337 **DML** contributed extensively to study design, review of results, and writing of this

338 manuscript

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451

452 **Figure legend**

453 **Figure 1.** Disposition of samples by country and by laboratory in the analysis

454 population.

455

Tables

Table 1. Number and percentages of resistant and susceptible isolates (eligible and cefpodoxime-resistant isolates)

Antibiotic	All <i>E. coli</i>		Cefpodoxime-resistant <i>E. coli</i>		All isolates	
	N ^a	Resistant isolates, n (%)	N ^a	Resistant isolates, n (%)	N ^a	Resistant isolates, n (%)
Nitrofurantoin 100 µg	2060	31 (0.5)	348	15 (4.3)	2766	EUCAST breakpoints only for <i>E. coli</i>
Fosfomycin 200 µg	2062	74 (3.6)	348	28 (8.0)	2816	EUCAST breakpoints only for <i>E. coli</i>
Mecillinam 10 µg	2061	169 (8.2)	348	61 (17.6)	2804	203 (7.2)
Cefpodoxime 10 µg	2062	348 (16.9)	348	348 (100.0)	2813	454 (16.1)
Cephalexin 30 µg	2062	358 (17.4)	348	319 (91.7)	2811	500 (17.8)
Amoxicillin/clavulanate 20/10 µg	2062	383 (18.6)	348	165 (47.4)	2830	506 (17.9)
Ciprofloxacin 5 µg	2062	428 (20.8)	348	208 (59.8)	2841	568 (20.0)
Trimethoprim 5 µg	2062	632 (30.6)	348	172 (49.4)	2824	820 (29.0)
Ampicillin 10 µg	2062	1007 (51.2)	348	333 (95.7)	2827	1463 (51.8)

Table 2. Number and percentage of resistant *E. coli* isolates by antibiotic and country (eligible isolates)

	Fosfomyci n	Amoxicillin/ clavulanate	Ampicillin	Cefpodoxime	Cephalexin	Ciprofloxacin	Trimethoprim	Mecillinam	Nitrofurantoin
Italy (N=325) ^a	27 (8.3)	91 (28.0)	169 (52.0)	71 (21.8)	70 (21.5)	105 (32.3)	101 (31.1)	35 (10.8)	8 (2.5)
Spain (N=435)	20 (4.6)	70 (16.1)	225 (51.7)	<i>28 (6.4)</i>	<i>35 (8.0)</i>	73 (16.8)	132 (30.3)	37 (8.5)	0
United Kingdom (N=240)	7 (2.9)	66 (27.5)	153 (63.8)	66 (27.5)	63 (26.3)	36 (15.0)	87 (36.3)	24 (10.0)	5 (2.1)
Belgium (N=367) ^a	<i>7 (1.9)</i>	82 (22.3)	176 (48.0)	43 (11.7)	46 (12.5)	<i>53 (14.4)</i>	110 (30.0)	37 (10.1)	5 (1.4)
Russia (N=695)	<i>13 (1.9)</i>	<i>74 (10.6)</i>	<i>332 (47.8)</i>	140 (20.1)	144 (20.7)	161 (23.2)	<i>202 (29.1)</i>	<i>36 (5.2)</i>	13 (1.9)
Chi square	<.0001	<.0001	0.0005	<.0001	<.0001	<.0001	0.3496	0.0068	0.0067

N = total number of isolates per country; *E. coli* resistance data are presented n (%) of samples. Percentages are computed on eligible isolates tested for each antibiotic within each country.

^a Numbers of isolates tested are one or two lower than N in some cases owing to failed tests

Bold: highest rates; *italic*, lowest rates for each antibiotic

Table 3. Comparison of central and local results for *E. coli* (n=201) retested as a QC sample.

	Isolates categorised as susceptible at local laboratory			Isolates categorised as resistant at local laboratory		
	Total submitted to central lab.	Central lab found susceptible	Central lab found resistant	Total submitted to central lab.	Central lab found susceptible	Central lab found resistant
Fosfomycin 200 µg	143	138	5	58	29	29
Amoxicillin/clavulanate 20/10 µg	157	123	34	44	4	40
Ampicillin 10 µg	90	76	14	111	7	104
Cefpodoxime 10 µg	155	148	7	46	10	36
Cephalexin 30 µg	157	152	5	44	5	39
Ciprofloxacin 5 µg	147 ^a	137 + 8 found I ^b	2	50	4	46
Trimethoprim 5 µg	130	128	2	71	10	61
Mecillinam 10 µg	183	166	17	18	7	11
Nitrofurantoin 100 µg	196	196	0	5	4	1

^a Includes isolates found 'I', defined by EUCAST as high-dose susceptible.

^b I, high dose susceptible.

Concordance between local and central laboratory results for each tested uropathogen by antibiotic (paired isolates subgroup)

Table 4. Concordance between local and central laboratory results for fosfomycin only, shown for non-*E. coli* bacteria

Local Laboratory	Central Laboratory	
	MIC \leq 8 mg/L Susceptible, n	MIC >8 mg/L Resistant, n
All non-<i>E. coli</i>		
Zone \geq 24 mm, Susceptible	14	22
Zone <24 mm, Resistant	49	248
<i>Klebsiella</i> spp.		
Zone \geq 24 mm, Susceptible	3	1
Zone <24 mm, Resistant	39	112
Proteeae (i.e. <i>Proteus</i>, <i>Morganella</i> and <i>Providencia</i> spp.)		
Zone \geq 24 mm, Susceptible	6	1
Zone <24 mm, Resistant	3	20
Other		
Zone \geq 24 mm, Susceptible	5	20
Zone <24 mm, Resistant	7	116