





Genome Sequences of Plant-Associated Rhodococcus sp. **Isolates from Tunisia**

Sabrine Dhaouadi,^a Joe Win,^b Amira Hamdane Mougou,^a Adeline Harant,^b DSophien Kamoun,^b Ali Rhouma^c

a Laboratory of Bio Aggressors and Integrated Pest Management, Department of Plant Health and Environment, National Institute of Agronomy, University of Carthage, Tunis, Tunisia

^bThe Sainsbury Laboratory, University of East Anglia, Norwich, United Kingdom

^cPartnership for Research and Innovation in the Mediterranean Area (PRIMA), Barcelona, Spain

Sabrine Dhaouadi and Joe Win contributed equally to this work. Author order reflects the first author's overall leadership of the project.

ABSTRACT The draft genome sequences of plant-associated *Rhodococcus* spp. from Tunisia are reported here. Two Rhodococcus fascians strains were obtained from almond rootstocks, and one Rhodococcus kroppenstedtii strain was obtained from a pistachio tree. The fourth Rhodococcus sp. strain was isolated from an ornamental plant.

lant-pathogenic *Rhodococcus* spp. are known to cause disease on herbaceous and woody species (1-3). On ornamental plants, major disease symptoms caused by Rhodococcus fascians were described as leafy galls and stem fasciation (4), while on woody trees, the symptoms included stunted growth and proliferation of misshapen shoots (2, 3). Four distinct Rhodococcus species isolates from pistachio and almond rootstocks and ornamental plants in Tunisia were used in this study, one of which was shown to cause disease on ornamental plants (5). It is well known that the population structure of R. fascians tends to be diverse from one host to another and from one region to another (6). Here, we provide insight into the genetic diversity of plantassociated Rhodococcus sp. isolates in Tunisia through their genome sequencing and assemblies.

Rhodococcus strains (Table 1) were isolated from plant tissues following the protocol used by Dhaouadi et al. (5) and grown at 27°C on agar plates of D2 medium (7). Genomic DNA extraction and sequencing were outsourced to MicrobesNG (Birmingham, UK). Briefly, three beads were washed with DNA extraction buffer containing lysozyme and RNase A and incubated for 25 min at 37°C. Proteinase K and RNase A were added and incubated for 5 min at 65°C. Genomic DNA was purified using an equal volume of solid-phase reversible immobilization (SPRI) beads (ABM, Richmond, Canada) and resuspended in EB buffer (10 mM Tris-Cl, pH 8.5). DNA was quantified in triplicate with the Quant-IT double-stranded DNA (dsDNA) high-sensitivity (HS) assay in an Eppendorff AF2200 plate reader. Genomic DNA libraries were prepared using a Nextera XT library prep kit (Illumina, San Diego, CA, USA) following the manufacturer's protocol. DNA quantification and library preparation were carried out on a Hamilton Microlab STAR automated liquid-handling system. Pooled libraries were quantified using the Kapa Biosystems library quantification kit for Illumina on a Roche light cycler 96 quantitative PCR (qPCR) machine. Libraries were sequenced on the Illumina HiSeq instrument using a 250-bp paired-end protocol. The reads were trimmed using Trimmomatic version 0.39 (8) with a sliding window quality cutoff of Q15. Sequence reads were assembled into contigs using SPAdes version 3.7 (9). The assembly metrics in Table 1 were calculated using QUAST version 5.0.2 (10). The genomes were annotated with Prokka version 1.14.3 (https://github.com/tseemann/prokka). Protein-coding fea-

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Address correspondence to Sabrine Dhaouadi. sabrinedhaouadi@outlook.com.

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TABLE 1 Summary statistics for Rhodococcus genomes assembled from Illumina readsa

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		Host common	RioSample	No. of	Largest	Largest Total GC Mean No. of contin length content covera	GC	Mean	No. of	×	No. of	No. of	No. of	GenBank No. of No. of accession no.	SRA
Organism	Host		ċ	contigs	(dq)	(pp)	(%)	(×)	(%) (x) reads (bp)	(pp)	CDS	tRNAs	tmRNAs	CDS tRNAs tmRNAs (assembly)	accession no.
Rhodococcus fascians GS6	Prunus dulcis	Prunus dulcis Bitter almond	SAMN13734959 122 318,389 5,441,276 64.56 40.6029 552,329 120,644 5,112 51 1	122	318,389	5,441,276	64.56	40.6029	552,329	120,644	5,112	51	l	JAAFYX0000000000	SRR11109608
Rhodococcus fascians SB10	Prunus persica × Prunus amygdalus	runus persica Garnem rootstock SAMN13734960 199 \times Prunus amygdalus	SAMN13734960	199	352,664	352,664 5,548,817 64.39		54.8541	703,905 133,430 5,158 50	133,430	5,158	20	-	JAAFYW000000000 SRR11109607	SRR11109607
Rhodococcus sp. strain B10	<i>Iresine herbstii</i> Hook	Iresine herbstii Herbst's bloodleaf SAMN13734961 156 Hook	SAMN13734961	156	338,193	338,193 5,495,644 65.11		109.753	1,580,360 112,702 5,154	112,702	5,154	51	-	JAAFYV0000000000	SRR11109606
Rhodococcus Pistacia v kroppenstedtii K5 L. cv.	Pistacia vera 5 L. cv.	Pistachio	SAMN13734962 157	157	168,870	168,870 4,040,887 70.26		84.8393	836,282 57,161 3,697	57,161	3,697	51	_	JAAFYU000000000 SRR11109605	SRR11109605
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^a CDS, coding DNA sequences; tmRNA, transfer-messenger RNA.

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tures and tRNA were predicted using Prodigal version 2.6 (11), and rRNA was predicted using ARAGORN version 1.2 (12). For taxonomic identification of the bacterial genomes, we used the average nucleotide identity (ANI) test (13). Default settings were used for all software unless otherwise specified.

The assembly statistics and total number of genes are shown in Table 1. The results based on the ANI test and current taxonomic nomenclature revealed an identity of over 90% of the submitted genome sequence to *Rhodococcus* species. The sequence of the isolate K5 genome is 97.9% identical to the type strain of *Rhodococcus kroppenstedtii*, the isolate B10 genome is identical to multiple *Rhodococcus* type strains, the isolate SB10 genome is 97.7% identical to *Rhodococcus fascians* NBRC 12155, and the isolate GS6 genome is 97.8% identical to *Rhodococcus fascians* NBRC 12155.

Data availability. This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under accession numbers JAAFYU000000000, JAAFYV0000000000, JAAFYV0000000000, and JAAFYX0000000000. The versions described in this paper are JAAFYU010000000, JAAFYV010000000, JAAFYW010000000, and JAAFYX010000000. The raw sequence reads have been deposited in the NCBI Sequence Read Archive under BioProject number PRJNA598862 and run numbers SRR11109605, SRR11109606, SRR11109607, and SRR11109608.

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REFERENCES

- Putnam ML, Miller ML. 2007. Rhodococcus fascians in herbaceous perennials. Plant Dis 91:1064–1076. https://doi.org/10.1094/PDIS-91-9-1064.
- Stamler RA, Heerema R, Randall JJ. 2015. First report of phytopathogenic Rhodococcus isolates on pistachio bushy top syndrome 'UCB-1' rootstock in New Mexico. Plant Dis 99:1854–1855. https://doi.org/10.1094/PDIS-04 -15-0471-PDN.
- 3. Stamler RA, Kilcrease J, Kallsen C, Fichtner EJ, Cooke P, Heerema RJ, Randall JJ. 2015. First report of *Rhodococcus* isolates causing pistachio bushy top syndrome on 'UCB-1' rootstock in California and Arizona. Plant Dis 99:1468–1476. https://doi.org/10.1094/PDIS-12-14-1340-RE.
- Stes E, Francis I, Pertry I, Dolzblasz A, Depuydt S, Vereecke D. 2013. The leafy gall syndrome induced by *Rhodococcus fascians*. FEMS Microbiol Lett 342:187–194. https://doi.org/10.1111/1574-6968.12119.
- Dhaouadi S, Hamdane AM, Bahri BA, Rhouma A, Fichtner EJ. 2019. First report of *Rhodococcus* spp. isolates causing stunting and lateral stem proliferation of *Iresine herbstii* 'Aureo-Reticulata' in Tunisia. Phytopathol Mediterr 58:391–394. https://doi.org/10.14601/Phytopathol_Mediter -10626.
- Creason AL, Vandeputte OM, Savory EA, Davis EW, II, Putnam ML, Hu E, Swader-Hines D, Mol A, Baucher M, Prinsen E, Zdanowska M, Givan SA, El Jaziri M, Loper JE, Mahmud T, Chang JH. 2014. Analysis of genome sequences from plant pathogenic *Rhodococcus* reveals genetic novelties in virulence loci. PLoS One 9:e101996. https://doi.org/10.1371/journal pone.0101996.
- 7. Kado Cl, Heskett MG. 1970. Selective media for Agrobacterium, Coryne-

- bacterium, Erwinia, Pseudomonas, and Xanthomonas. Phytopathology 60:969–976. https://doi.org/10.1094/phyto-60-969.
- 8. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10.1093/bioinformatics/btu170.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. Bioinformatics 29:1072–1075. https://doi.org/10.1093/bioinformatics/btt086.
- Hyatt D, Chen GL, LoCascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. BMC Bioinformatics 11:119. https://doi.org/10.1186/1471 -2105-11-119.
- Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. Nucleic Acids Res 32:11–16. https://doi.org/10.1093/nar/gkh152.
- Ciufo S, Kannan S, Sharma S, Badretdin A, Clark K, Turner S, Brover S, Schoch CL, Kimchi A, DiCuccio M. 2018. Using average nucleotide identity to improve taxonomic assignments in prokaryotic genomes at the NCBI. Int J Syst Evol Microbiol 68:2386–2392. https://doi.org/10.1099/ ijsem.0.002809.