## Vitamin D and the hepatitis B vaccine response: A prospective cohort study and a

# randomized, placebo-controlled oral vitamin D<sub>3</sub> and simulated sunlight supplementation

## trial in healthy adults

Daniel S Kashi<sup>1,2</sup>, Samuel J Oliver<sup>1</sup>, Laurel M Wentz<sup>3</sup>, Ross Roberts<sup>1</sup>, Alexander T Carswell<sup>1</sup>, Jonathan C Y Tang<sup>4</sup>, Sarah Jackson<sup>5</sup>, Rachel M Izard<sup>6</sup>, Donald Allan<sup>7</sup>, Lesley E Rhodes<sup>8</sup>, William D Fraser<sup>4</sup>, Julie P Greeves<sup>4,5</sup> and Neil P Walsh<sup>2</sup>

<sup>1</sup>College of Human Sciences, Bangor University, Bangor, UK.

<sup>2</sup> Faculty of Science, Liverpool John Moores University, Liverpool, UK.

<sup>3</sup>Beaver College of Health Sciences, Appalachian State University, Boone, USA.

- <sup>4</sup> Norwich Medical School, University of East Anglia, Norwich, UK.
- <sup>5</sup> Department of Army Health and Physical Performance Research, Army HQ, Andover, UK.

<sup>6</sup>Occupational Medicine, HQ Army Recruiting and Initial Training Command, Upavon, UK.

<sup>7</sup> Medical Physics Department, Salford Royal NHS Foundation Trust, and University of

Manchester, Manchester Academic Health Science Centre, Manchester, UK.

<sup>8</sup> Faculty of Biology, Medicine and Health, University of Manchester, and Dermatology Centre, Salford Royal NHS Foundation Trust, Manchester Academic Health Science Centre, Manchester, UK.

**Source(s) of support:** This work was funded by The Ministry of Defence (MOD)

**Conflict of interest and funding disclosure**: None of the authors report a conflict of interest related to the study.

Corresponding author:	Dr Samuel Oliver
	College of Human Sciences,
	Bangor University,
	Bangor,
	LL57 2PZ, United Kingdom
	Tel: + 44 1248 383965
	E-mail: <u>s.j.oliver@bangor.ac.uk</u>
	ORCID: 0000-0002-9971-9546

Running head: Vitamin D and the hepatitis B vaccine response

Clinical trial registry number: Study 1 NCT02416895; <u>https://clinicaltrials.gov/ct2/show/study/NCT02416895</u>; Study 2 NCT03132103; <u>https://clinicaltrials.gov/ct2/show/NCT03132103</u>

# 1 Abstract

2	Purpose: To determine serum 25(OH)D and 1,25(OH) <sub>2</sub> D relationship with hepatitis B vaccination
3	(study-1). Then, to investigate the effects on hepatitis B vaccination of achieving vitamin D
4	sufficiency (serum 25(OH)D $\geq$ 50 nmol/L) by a unique comparison of simulated-sunlight and oral
5	vitamin D <sub>3</sub> supplementation in wintertime (study-2). Methods: Study-1 involved 447 adults. In
6	study-2, 3 days after the initial hepatitis B vaccination, 119 men received either placebo,
7	simulated-sunlight (1.3x standard-erythema dose, 3x/week for 4-weeks and then 1x/week for 8-
8	weeks) or oral vitamin $D_3$ (1,000 IU/day for 4-weeks and 400 IU/day for 8-weeks). We measured
9	hepatitis B vaccination efficacy as percentage of responders with anti-hepatitis B surface antigen
10	immunoglobulin G $\geq$ 10 mIU/mL. <u>Results:</u> In study-1, vaccine response was poorer in persons
11	with low vitamin D status (25(OH)D $\leq$ 40 vs 41–71 nmol/L mean difference[95% confidence
12	interval] -15%[-26, -3%]; 1,25(OH) <sub>2</sub> D ≤120 <i>vs</i> ≥157 pmol/L -12%[-24%, -1%]). Vaccine
13	response was also poorer in winter than summer (-18%[-31%, -3%]), when serum 25(OH)D and
14	$1,25(OH)_2D$ were at seasonal nadirs, and 81% of persons had serum $25(OH)D < 50$ nmol/L. In
15	study-2, vitamin D supplementation strategies were similarly effective in achieving vitamin D
16	sufficiency from the winter vitamin D nadir in almost all (~95%); however, the supplementation
17	beginning 3 days after the initial vaccination did not effect the vaccine response (vitamin D vs
18	placebo 4%[-21%, 14%]). Conclusion: Low vitamin D status at initial vaccination was
19	associated with poorer hepatitis B vaccine response (study-1); however, vitamin D
20	supplementation commencing 3 days after vaccination (study-2) did not influence the vaccination
21	response.

22

**Keywords:** Cholecalciferol, Vitamin D, 25-hydroxyvitamin D, Hepatitis B, vaccination, UVB.

#### 24 Introduction

Discovery of the vitamin D receptor in almost all immune cells, and the many roles vitamin D 25 has in innate and adaptive arms of immunity [1-3], highlight the importance of vitamin D in the 26 regulation of immune responses [4]. As such, avoiding low serum 25-hydroxyvitamin D 27 (25(OH)D) and achieving vitamin D sufficiency  $(25(OH)D \ge 50 \text{ nmol/L})$  may be important for 28 29 the development of vaccine responses and consequently public health [5]. Cell and animal studies indicate that vitamin D may modulate vaccine responses through 1,25-dihydroxyvitamin D 30 31  $(1,25(OH)_2D)$  interaction with antigen presentation [6], dendritic cell migration, and the subsequent activation of T and B cell antibody responses [7-9]. Indeed, vitamin D 32 supplementation that corrected wintertime vitamin D status to achieve sufficiency before a 33 tetanus toxoid booster vaccination resulted in higher IgG antibody concentration compared to a 34 placebo [10]. 35

36

The influence of vitamin D on the development of the hepatitis B vaccination response in humans 37 remains unclear; previous investigations have only studied chronic kidney patients and report 38 conflicting findings [11,12]. Moreover, the relationship between the biologically active form of 39 vitamin D, 1,25(OH)<sub>2</sub>D, and hepatitis B vaccine is yet to be examined. Hepatitis B vaccination 40 has previously been shown to be influenced by genetics and lifestyle factors [13-15] with 10-41 15% of adults responding inadequately by producing too few antibodies, as dictated by an anti-42 hepatitis B surface antigen immunoglobulin G (IgG) concentration of less than 10 mIU/mL [16]. 43 Conversely, those responding to the vaccination with IgG concentration of 10 mIU/mL or more 44 are generally accepted to be protected against infection clinically [16,17]. Whether vitamin D 45 influences the development of hepatitis B vaccination in healthy adults is unknown, but important 46

to understand given that more than 50% fail to achieve vitamin D sufficiency during winter 47 months [18-20]; and many adults remain unvaccinated because childhood vaccine coverage is 48 ~90% or less and routine infant hepatitis B vaccination began only recently in some countries 49 (e.g. UK [21-23]). The hepatitis B vaccination course presents a suitable model to study the 50 influence of vitamin D on the secondary immune response because there is widespread inter-51 52 individual variability in the magnitude of the antibody response after the second vaccination, and it is more possible to control prior exposure than with other commonly experienced vaccines (e.g. 53 54 influenza) [24].

55

Here we present results from two studies examining the influence of vitamin D on hepatitis B 56 57 vaccine response. In these studies we measured 1,25(OH)<sub>2</sub>D, vitamin D's biologically active form, and 25(OH)D, which with their respective 4-6 h and 2-3 week half lives can be considered 58 acute and chronic vitamin D status markers, respectively [25]. In study 1, a prospective cohort 59 study of 447 healthy young men and women conducted during all seasons, we examined for the 60 first time serum 1,25(OH)<sub>2</sub>D and 25(OH)D relationship with hepatitis B vaccination in healthy 61 adults. We hypothesized that low serum 1,25(OH)<sub>2</sub>D and 25(OH)D at the time of initial 62 vaccination would be associated with poorer secondary antibody response to hepatitis B 63 vaccination. In study 2, a randomized placebo-controlled trial, we determined the effect of 12-64 weeks wintertime vitamin D supplementation on the hepatitis B vaccination response. The 65 supplementation was a unique comparison of simulated sunlight in accordance with 66 recommendations on safe (non-sunburning), low-level sunlight exposure [26], and oral vitamin 67 D<sub>3</sub> to achieve vitamin D sufficiency (serum  $25(OH)D \ge 50 \text{ nmol/L}$ ). Vitamin D sufficiency was 68 targeted as maintaining serum 25(OH)D concentration  $\geq 50$  nmol/L has been recommended for 69

70	multiple health outcomes [27] by the Institute of Medicine (IOM) and European Food Safety
71	Authority (EFSA) and is achieviable using safe doses [19,20]. The comparison was also made as
72	vitamin D can be obtained from dietary sources but is predominately synthesized by skin
73	exposure to solar ultraviolet (UV) B radiation; UV radiation has a range of vitamin D-dependent
74	and -independent effects on immunity [28,29]. We hypothesized that vitamin D supplementation
75	that achieves vitamin D sufficiency during winter when vitamin D status is usually low would
76	lead to superior secondary antibody response to hepatitis B vaccination compared to placebo
77	supplementation.

78

## 79 Methods

The Ministry of Defence (UK) Research Ethics Committee approved these studies, and protocols were conducted in accordance with the Declaration of Helsinki (2013). All participants provided written informed consent.

83

85 Participant recruitment, inclusion and exclusion criteria

86 Between June 2014 and November 2015, 1268 men and women who entered the British Army

were assessed for eligibility for this prospective cohort study. Eligible participants were  $\geq 18$ 

years of age. One thousand one hundred and three recruits volunteered (men from the Infantry

- 89 Training Centre, Catterick, UK; latitude 54°N, and women from the Army Training Centre,
- 90 Pirbright, UK; latitude 51°N). Participants were excluded from the final analysis if they failed the
- 91 initial medical assessment, followed an atypical hepatitis B vaccination schedule (the first two

<sup>84</sup> Study 1

vaccine doses were not administered within 4 weeks of each other), or did not provide a blood 92 sample to assess the secondary hepatitis B vaccine response. Participants were also excluded 93 from statistical analysis if their medical records documented previous exposure to hepatitis B 94 vaccination; or, if this was later confirmed by measurable antibody titers against hepatitis B 95 surface antigen detected in baseline samples (anti-HBs titers >0 mIU/mL). The baseline 96 97 demographics, anthropometrics, and lifestyle behaviors for the 447 participants included in the final analysis are summarized in **Table 1** (Supplemental Table 1 includes details of the larger 98 99 recruited sample).

100

#### 101 *Procedures*

Before participants commenced Basic Military training they completed an initial medical 102 assessment. During the initial medical assessment, participants received their first 20-µg dose of 103 recombinant hepatitis B vaccine into the deltoid muscle (Engerix-B, Smithkline Beecham 104 Pharmaceuticals, Uxbridge, UK) and a venous blood sample was collected for the determination 105 of hepatitis B antibody titer, serum 25(OH)D and 1,25(OH)<sub>2</sub>D concentrations (Figure 1). At the 106 initial medical assessment, we also collected baseline measures of participant demographics (e.g. 107 ethnicity) and anthropometrics; height and body mass were assessed in light clothing with shoes 108 removed by stadiometer and digital platform scale, respectively (SECA 703, Birmingham, UK). 109 Lifestyle factors previously shown to influence the vaccination response were also assessed by 110 questionnaire; including alcohol and smoking use, sleep and mood [13,15,14]. To assess sleep 111 duration and quality the night before vaccination participants completed a questionnaire based on 112 the procedures of Prather et al [15]. Sleep duration was calculated as the number of hours and 113 114 minutes elapsed between the time they reported going to sleep and the time they reported waking.

7

Sleep quality was reported on a scale from 1 = very poor to 4 = very good. Before receiving their 115 initial hepatitis B vaccination participants also completed a Brunel mood scale (BRUMS) [30], 116 which measures 6 moods (vigor, anger, tension, confusion, depression, fatigue). Each mood is 117 assessed by 4 items scored from 0 = not at all to 4 = extremely and therefore the maximum score 118 per mood is 20, with greater scores indicating a greater feeling of the mood. In line with the 119 typical hepatitis B vaccination schedule, participants received a second 20-µg hepatitis B vaccine 120 dose one month after the first. A second venous blood sample was collected 8 weeks after the 121 122 second hepatitis B vaccine dose (3-months after the first hepatitis B vaccine dose) to determine secondary serum hepatitis B antibody titers, the primary outcome measure. The serum hepatitis B 123 antibody titer (anti-HBs) was assessed as this is the routine serological test to determine if a 124 person has been successfully vaccinated against hepatitis B [16]. We focused on the antibody 125 response to the second vaccination because there is widespread inter-individual variability in the 126 magnitude of antibody response following the second vaccination of the typical three-dose series 127 [24]. This variability is in distinct contrast with the antibody response to the first vaccination, 128 when <10% of individuals have detectable levels of antibody, or the third, when the majority of 129 individuals have mounted maximal antibody responses, respectively [15]. 'All-cause illness' 130 consisting of physician diagnosed cases of upper and lower respiratory tract infection and 131 gastrointestinal infection were also retrieved from medical records for the period of basic 132 133 training.

134

135 *Study 2* 

136 Participant recruitment and exclusion criteria

137	Healthy men were recruited in a double-blind randomized, placebo-controlled trial upon entering
138	the British Army Combat Infantryman's Course, Catterick, UK during January and February of
139	2016 and 2017, when ambient UVB is negligible at UK latitudes (50–60°N), and serum 25(OH)D
140	is at a seasonal low. Eligible participants were $\geq 17$ years of age and had passed the initial medical
141	assessment; had no history of skin cancer, photosensitivity, or lupus erythematosus; and had sun-
142	reactive skin type I-IV [31]. Participants were excluded for the same reasons as in study 1, plus
143	current consumption of vitamin D in dietary supplements; use of a sunbed or travel to a sunny
144	climate 3-months before the study.

145

#### 146 *Experimental procedures*

Participants had the same baseline assessments and hepatitis B vaccination schedule as study 1 147 (Figure 1). Following this, we block randomized participants within their platoons to one of four 148 intervention groups: 1) solar simulated radiation (SSR); 2) solar simulated radiation placebo 149 (SSR-P); 3) oral vitamin D<sub>3</sub> (ORAL); or 4) oral placebo (ORAL-P). Block randomization by 150 151 randomizer.org resulted in an equal distribution of intervention groups within each platoon, and ensured any differences in training conditions between platoons did not influence the study 152 outcomes. An independent researcher completed the randomization and investigators were blind 153 to the randomization until statistical analyses were completed. The interventions began 3 days 154 after the initial hepatitis B vaccine dose. The intervention strategy for the SSR and ORAL groups 155 was to restore and then maintain vitamin D sufficiency (serum  $25(OH)D \ge 50 \text{ nmol/L}$ ) as 156 recommended by Institute of Medicine (IOM) and the European Food Safety Authority (EFSA) 157 [19,20]. Participants completed a 4-week restoration phase, necessary because serum 25(OH)D 158 was at its winter nadir, followed by an 8-week maintenance phase (Figure 1). Blood samples 159

160	were obtained at baseline, and after 5 and 12 weeks for the determination of serum 25(OH)D and
161	1,25(OH) <sub>2</sub> D (Figure 1). Vitamin D from solar UV radiation exposure was estimated in weeks 4
162	and 11 using polysulphone badges and from the diet in week 12 using a food frequency
163	questionnaire [32]. On completion of the study, participants completed an 'exit survey,' which
164	required them to guess the intervention they thought they had been receiving.

165

## 166 Simulated sunlight intervention

In accordance with guidelines on safe, low-level sunlight exposure for vitamin D synthesis [26], 167 and as described previously to achieve vitamin D sufficiency (serum  $25(OH)D \ge 50 \text{ nmol/L}$ ) in 168 the majority of white skinned persons [33], those assigned to the SSR intervention were exposed 169 three-times-a-week, during the restoration phase to an investigator controlled constant UV 170 radiation dose using a whole body irradiation cabinet (Hapro Jade, Kapelle, The Netherlands) 171 fitted with Arimed B fluorescent tubes (Cosmedico, Stuttgart, Germany). The fluorescent tubes 172 emitted a UV radiation spectrum similar to sunlight ( $\lambda$ : 290–400 nm; 95% UVA: 320–400 nm, 173 5% UVB: 290–320 nm) that was characterized by a spectroradiometer (USB2000+, Ocean Optics 174 BV, Duiven, The Netherlands) radiometrically calibrated with traceability to UK national 175 standards. During each exposure participants received a 1.3 standard erythemal dose (SED), and 176 wore shorts and a T-shirt to expose  $\sim 40\%$  of skin surface area. This dose is equivalent to  $\sim 15$ 177 minutes midday summer sun exposure in northern England (latitude 53.5°N) [33] and taking 178 account of pre-vitamin D irradiance at different latitudes, can be related to exposure times at 179 other world locations [34]. For example, the equivalent exposure time in Philadelphia, 180 Pennsylvania, USA (40°N) would be ~12 minutes; and that for Oslo, Norway (60°N) would be 181

182	~18 minutes. During the maintenance phase, we exposed SSR participants to the same $1.3x$ SED
183	dose only once-a-week: pilot investigations confirmed the required dose to maintain sufficiency
184	(serum 25(OH)D $\geq$ 50 nmol/L). A constant SSR dose was maintained during the study by
185	monitoring irradiance using a spectroradiometer (USB2000+, Ocean Optics BV) and adjusting
186	for any decrease in measured irradiance emitted by increasing exposure time (mean duration of
187	SSR exposures was 229 $\pm$ 17 s). We controlled the exposure time by using an electronic timer.
188	Participants undergoing SSR-P treatment received the same number of intervention exposures
189	each week and the exposure duration as SSR except the irradiation cabinet fluorescent tubes were
190	covered with transparent UV radiation blocking film (DermaGard UV film, SunGard, Woburn,
191	Massachusetts, USA) [35] in a manner invisible to the participants and experimenters.
192	Spectroradiometry confirmed the UV radiation blocking film was effective at preventing
193	transmission of 99.9% of UV radiation.

194

### 195 *Oral vitamin* $D_3$ *intervention*

196 Participants receiving the ORAL intervention consumed a daily vitamin D<sub>3</sub> supplement

197 containing 1000 IU and 400 IU vitamin D<sub>3</sub> during the restoration phase and maintenance phases,

respectively (Pure Encapsulations, Sudbury, Massachusetts, USA) [35]. The restoration dose

199 (1000 IU/day) was based on previous predictive modelling to achieve serum  $25(OH)D \ge 50$ 

- nmol/L [36], and pilot investigations that showed it achieved similar serum 25(OH)D
- 201 concentrations to SSR; and was less than the tolerable upper intake recommended by IOM and
- EFSA [19,20]. The ORAL maintenance dose (400 IU/day) was in accordance with
- recommendations [19]. For 12-weeks, ORAL-P participants consumed a daily oral cellulose

placebo capsule, identical in size, shape and color to the vitamin D<sub>3</sub> capsules (Almac Group,
County Armagh, UK). Independent analysis found the vitamin D<sub>3</sub> content of the 1000 and 400 IU
capsules to be 1090 and 460 IU, respectively and confirmed the placebo did not contain vitamin
D (NSF International Laboratories, Ann Arbor, Michigan, USA).

208

209 Biochemical analyses (Study 1 and 2)

210 Whole blood samples were collected by venepuncture from an antecubital vein into plain

vacutainer tubes (Becton Dickinson, Oxford, UK) and left to clot for one hour. Subsequently,

samples were centrifuged at 1500 g for 10 min at 4°C and the serum aliquoted into eppendorf

tubes before being immediately frozen at -80°C for later analysis. Baseline and secondary serum

antibody titers were determined using a hepatitis B antibody enzyme-linked immunoassay kit

215 (DiaSorin, Saluggia, Italy). The intra-assay coefficient of variation was 4.9% (study 1) and 5.9%

(study 2). Total serum 25(OH)D was measured with high-pressure liquid chromatography tandem

mass spectrometry [37]; and serum 1,25(OH)<sub>2</sub>D using the DiaSorin LIAISON XL 1,25(OH)<sub>2</sub>D

chemiluminescent immunoassay (Stillwater, Minnesota, USA) method. Analyses were performed

in a Vitamin D External Quality Assurance Scheme certified laboratory (Bioanalytical Facility,

220 University of East Anglia, Norwich, UK).

221

222 Statistical analysis

Secondary antibody titers have a non-normal distribution and therefore, in line with previous research [17], we categorized the development of secondary antibody response to the hepatitis B vaccine as the percentage of participants with serum antibody titer response to hepatitis  $B \ge 10$ 

mIU/mL. Those participants with anti-HBs titers  $\geq 10$  mIU/mL were categorized as vaccine 226 'responders' whilst those with antibody titers <10 mIU/mL were categorized as vaccine 'non-227 responders' [17]. Further, those responding to the vaccination with anti-HBs titers of 10 mIU/mL 228 or more are generally accepted to be protected against infection clinically [17,16]. The sample 229 size estimation for study 1 and 2 was calculated as a minimum of 152, using the anticipated 230 difference in hepatitis B vaccine responder rate of 20% (Cohen's h = 0.4; small-medium effect 231 size) between individuals displaying low and high vitamin D status [11], with a type 1 error (one 232 233 tailed) of 5%, and a power of 80%. For study 1, we used chi-square analysis to compare the percentage of hepatitis B vaccine responders in those with IOM defined vitamin D sufficient 234 status (serum 25(OH)D  $\geq$ 50 nmol/L) compared to those with serum <50 nmol/L. However, as 235 there is no consensus to the optimal vitamin D threshold for immune function [18,38], we 236 conducted Kruskal Wallis tests to compare the percentage of hepatitis B vaccine responders 237 across 25(OH)D, 1,25(OH)<sub>2</sub>D and 24,25(OH)<sub>2</sub>D terciles. One-way ANOVA and Kruskal-Wallis 238 tests were used, where appropriate, to compare serum vitamin D (25(OH)D and 1,25(OH)<sub>2</sub>D), 239 percentage of participants displaying serum  $25(OH)D \ge 50$  nmol/L and the percentage of hepatitis 240 B vaccine responders across seasons. Independent t-test, chi-square, One-way ANOVA and 241 Kruskal-Wallis tests, were used, where appropriate, to compare demographic, anthropometric, 242 alcohol and smoking use, sleep, mood, contraception use in women, 'all-cause illnes 'data across 243 seasons and between participants with serum  $25(OH)D \ge 50$  nmol/L and <50 nmol/L. For study 2, 244 Kruskal-Wallis was used to compare the percentage of secondary hepatitis B vaccine responders 245 after SSR, ORAL, SSR-P and ORAL-P. In addition, the percentage of secondary hepatitis B 246 vaccine responders was compared between vitamin D supplementation (SSR and ORAL 247 combined) and placebo groups (SSR-P and ORAL-P combined) using chi-square analysis. Serum 248

249	25(OH)D and 1,25(OH) <sub>2</sub> D were compared between vitamin D and placebo groups using mixed
250	model ANOVA (4 group (SSR, ORAL, SSR-P and ORAL-P) x 3 time points (baseline, week 5
251	and 12) and 2 group (SSR and ORAL combined, SSR-P and ORAL-P) x 3 time points. Post hoc
252	comparisons were conducted using Bonferroni corrected <i>t</i> -tests. Chi-square tests were conducted
253	to compare the percentage of participants displaying total serum $25(OH)D \ge 50 \text{ nmol/L}$ at
254	baseline, week 5 and week 12 between vitamin D and placebo groups. Independent samples t-
255	test, Mann-Whitney $U$ and chi-square tests were used to compare demographic, anthropometric,
256	alcohol and smoking use, sleep, and mood data between vitamin D and placebo supplement
257	groups. All statistical analyses were completed using SPSS Statistics 22 (IBM, Armonk, New
258	York, USA).

259

#### 260 **Results**

261 Study 1

262 *Participant flow* 

A total of 1103 men and women were recruited from June 2014 to November 2015. Participants

began the study throughout the year: 20% in winter (December–February), 14% in spring

265 (March–May), 26% in summer (June–August), and 40% in autumn (September–November).

266 Participant flow, drop out and exclusion before biochemical and statistical analysis are

summarized in Figure 2. There was no significant difference in demographics, anthropometrics,

lifestyle behaviors, sleep, mood, contraception use, or all cause illness between participants

included and excluded in the final analysis (Supplemental Table 2).

270

271 Vitamin D and secondary hepatitis B vaccine response

272	At the time of the initial vaccination 43% of participants had serum 25(OH)D <50 nmol/L, 26%
273	were vitamin D insufficient (serum 25(OH)D 30–50 nmol/L), and 17% were vitamin D deficient
274	(serum 25(OH)D <30 nmol/L). Only 1 participant presented with severe vitamin D deficiency
275	(serum 25(OH)D <12.5 nmol/L). Fewer participants tended to respond to the hepatitis B
276	vaccination who had 25(OH)D <50 nmol/L than those who were vitamin D sufficient at the time
277	of initial vaccination (50% vs 58%, mean difference [95% confidence interval], -8% [-17%, 1%],
278	P = 0.09, h = 0.16, <b>Figure 3A</b> ). Moreover, hepatitis B vaccine response was poorer in those with
279	serum 25(OH)D $\leq$ 40 nmol/L (mean 30 $\pm$ 7 nmol/L) compared to participants with 25(OH)D
280	between 41–71 nmol/L (mean 56 $\pm$ 9 nmol/L) at the time of initial vaccination (mean difference
281	[95% confidence interval -15% [-26%, -3%], $P = 0.01$ , <b>Figure 3B</b> ). Fewer participants were also
282	hepatitis B vaccine responders when they presented with low serum 1,25(OH) <sub>2</sub> D compared to
283	participants who presented with high serum $1,25(OH)_2D$ at the time of initial vaccination (50% vs
284	62%, mean difference [95% confidence interval] -12% [-24%, -1%,], <i>P</i> < 0.05, h = 0.24, <b>Figure</b>
285	<b>3C</b> ). Furthermore, fewer participants were hepatitis B vaccine responders when they presented
286	with combined low $1,25(OH)_2D$ and $25(OH)D$ compared to combined medium-high $25(OH)D$
287	and 1,25(OH) <sub>2</sub> D (43% vs 65%, mean difference [95% confidence interval], -22% [-39%, -5%], P
288	= 0.01). No differences were observed between those who presented with low serum
289	24,25(OH)D compared to participants who presented with high serum 24,25(OH)D at the time of
290	initial vaccination (52% vs 60%, mean difference [95% confidence interval] -8% [20%, 3%], P =
291	0.14).
292	

There were no differences between participants with  $25(OH)D \ge 50 \text{ nmol/L}$  and <50 nmol/L in demographics, anthropometrics, lifestyle behaviors, sleep, mood, contraception use, or all cause

illness before the initial hepatitis B vaccination (**Table 2**). Anthropometrics, lifestyle behaviors, sleep, mood and all cause illness also did not predict vaccine response (P > 0.05). Additionally, contraception use did not influence the vaccine response (P > 0.05, e.g. none *vs* oral

- contraception, 68% vs 62% mean difference [95% confidence interval] 6% [-9, 21%]). Further
- regression analysis controlling for BMI, smoking, alcohol, sleep and mood indicated that vitamin
- D sufficient men, but not women, were 1.8 times more likely to be vaccine responders than those
- with serum 25(OH)D <50 nmol/L (OR [95% confidence interval], men 1.8 [1.0, 3.2] and women
- 302 0.8 [0.4, 1.7]). Serum 25(OH)D, 1,25(OH)<sub>2</sub>D, 24,25(OH)<sub>2</sub>D, vitamin D sufficiency and hepatitis
- B response was lower in men than women (P < 0.05, men vs women: 25(OH)D, 56 ± 30 vs 69 ±
- 304 32 nmol/L; 1,25(OH)<sub>2</sub>D, 126  $\pm$  32 *vs* 165  $\pm$  43 pmol/L; 24,25(OH)<sub>2</sub>D, 4.4  $\pm$  2.8 *vs* 6.5  $\pm$  3.7
- nmol/L; vitamin D sufficiency, 49% vs 69%; hepatitis B response, 49% vs 65%).
- 306

295

296

- 307 Seasonal variation in vitamin D and hepatitis B vaccine response
- Serum 25(OH)D, 1,25(OH)<sub>2</sub>D and vitamin D sufficiency (25(OH)D  $\geq$ 50 nmol/L) was lower in
- winter than spring, summer and autumn (P < 0.05, Figure 4A, 4B & 4C). In winter, 81%
- participants had 25(OH)D <50 nmol/L (Figure 4B) with 32% of participants vitamin D deficient
- 311 (serum 25(OH)D <30 nmol/L). The percentage of hepatitis B vaccine responders was also lower
- in winter than summer (44% vs 62%, mean difference [95% confidence interval] -18% [-31%, -
- 313 3%], P < 0.05, h = 0.36, Figure 4D). With the exception of all cause illness, participants
- recruited in the different seasons were similar as indicated by no differences in demographic,
- anthropometrics, lifestyle behaviors, sleep, mood or use of contraception in women before the
- initial hepatitis B vaccination (Table 1). Similar seasonal variations in serum 24,25(OH)<sub>2</sub>D were
- also observed with winter serum  $24,25(OH)_2D$  contrations lower than summer and autumn (P <

318 0.05, winter 2.9  $\pm$  2.2 nmol/L, spring 4.2  $\pm$  2.8 nmol/L, summer 6.5  $\pm$  3.2 nmol/L, autumn 5.9  $\pm$ 

- 319 3.4 nmol/L)
- 320

321 *Study 2* 

322 *Participant flow and blinding* 

323 Two hundred and thirty-one men were assigned to the interventions in January and February of

2016 and 2017. The study ended after reaching its scheduled date of closure. Participant flow,

drop out and exclusion before biochemical and statistical analysis is summarized in Figure 5.

326 There was no significant difference in demographics, anthropometrics, lifestyle behaviors, sleep

or mood between participants included and excluded in the final analysis (Supplemental Table 3).

328 There were no adverse events reported relating to vitamin D or placebo supplementation.

Participants were sufficiently blinded to the intervention since only 35% correctly guessed their

allocated group, 30% were incorrect, and 35% said they did not know whether they had received

an active (SSR and ORAL) or placebo (SSR-P and ORAL-P) intervention.

332

333 The influence of low-level simulated sunlight and oral vitamin  $D_3$  on vitamin D status

At baseline, 75% of the volunteers had 25(OH)D <50 nmol/L, 45% were vitamin D insufficient

(serum 25(OH)D 30–50 nmol/L), and 30% were vitamin D deficient (serum 25(OH)D <30

nmol/L). Only 1 participant presented with severe vitamin D deficiency (serum 25(OH)D <12.5

nmol/L). There was no difference between vitamin D and placebo supplementation groups'

demographics, anthropometrics, lifestyle behaviors, sleep, mood (Table 3), or vitamin D status

(Figure 6, P > 0.05). There were also no differences in these variables between combined

vitamin D and placebo supplemented groups (Supplemental Table 4 & Figure 6). During the 12-

17

341	week intervention, daily sunlight exposure was low, as expected considering the latitude and time
342	of year [39], with similar sunlight exposure (0.22 $\pm$ 0.33 SED/day; $P > 0.05$ ) and dietary vitamin
343	D intake (112 $\pm$ 84 IU/day, $P > 0.05$ ) in vitamin D and placebo supplement groups.

344

345	The vitamin D supplementation was successful in achieving vitamin D sufficiency and
346	maintaining serum 25(OH)D concentrations so that at week 5 and 12 serum 25(OH)D
347	concentrations in the vitamin D supplementation groups were higher than the placebo groups ( $P$
348	< 0.05, Figure 6A & D). By week 5, 95% of participants in the vitamin D supplementation
349	groups were vitamin D sufficient (25(OH)D $\geq$ 50 nmol/L, <b>Figure 6E</b> ). There was no difference in
350	serum 25(OH)D or percentage of participants achieving vitamin sufficiency between vitamin D
351	supplementation groups ( $P > 0.05$ ). Serum 1,25(OH) <sub>2</sub> D was similar in all groups at baseline ( $P >$
352	0.05) and increased with supplementation ( $P < 0.05$ , Figure 6C & F), with greater responses in
353	the vitamin D supplementation groups compared to the placebo groups at week 5 ( $P < 0.05$ ).
354	There was no difference between groups at week 12 ( $P > 0.05$ ) because 1,25(OH) <sub>2</sub> D increased
355	from week 5 to 12 in placebo groups ( $P < 0.05$ ). Serum 24,25(OH) <sub>2</sub> D responded similarly to
356	supplementation as serum $25(OH)D$ so that at week 5 and 12 serum $24,25(OH)_2D$ concentrations
357	in the vitamin D supplementation groups were higher than the placebo groups ( $P < 0.05$ ,
358	Supplemental Table 5).

359

360 The influence of simulated sunlight and oral vitamin D<sub>3</sub> on secondary hepatitis B vaccine
361 response

362	Vitamin D supplementation beginning 3 days after the initial vaccination did not influence the
363	secondary antibody response as the percentage of secondary hepatitis B vaccine responders was
364	similar among the vitamin D and placebo groups (SSR 60%, SSR-P 57%, ORAL 56%, ORAL-P
365	52%, $P > 0.05$ , Figure 7A). Analyses comparing combined vitamin D to placebo also revealed
366	no effect of vitamin D supplementation on secondary hepatitis B vaccine response (SSR and
367	ORAL vs SSR-P and ORAL-P, 58% vs 54%, mean difference [95% confidence interval], 4% [-
368	21%, 14%], $P > 0.05$ , h = 0.08, <b>Figure 7B</b> ). Furthermore, a secondary analysis including only
369	men who had 25(OH)D <50 nmol/L at baseline also revealed no effect of vitamin D
370	supplementation on secondary hepatitis B vaccine response ( $P > 0.05$ ).
371	

## 372 **Discussion**

We determined the influence of vitamin D on the development of the hepatitis B vaccination in 373 healthy adults. In study 1, vitamin D status (25(OH)D and 1,25(OH)<sub>2</sub>D) at the time of initial 374 vaccination influenced the subsequent secondary hepatitis B vaccine response: low vitamin D 375 status was associated with poorer hepatitis B vaccine response (Figure 3). Analysis controlling 376 for demographic, anthropometric, and lifestyle factors, revealed that vitamin D sufficient men, 377 but not women, were nearly 2 times more likely to be responders to the hepatitis B vaccine than 378 those with serum 25(OH)D of <50 nmol/L. These differences may be explained by lower serum 379 25(OH)D and 1,25(OH)<sub>2</sub>D in men and a lower proportion of men achieving vitamin D 380 sufficiency compared to women. Indeed, the hepatitis B vaccine response was poorer in men than 381 women. Furthermore, hepatitis B vaccine response was associated with seasonal alterations in 382 serum 25(OH)D and 1,25(OH)<sub>2</sub>D, with poorer hepatitis B vaccine responses in winter than 383 summer (Figure 4D). The findings of study 1 indicated a possible immunomodulatory role of 384

vitamin D in the development of hepatitis B vaccine response. Given these findings, and 385 the high prevalence of serum 25(OH)D < 50 nmol/L during winter (81% of persons had serum 386 25(OH)D < 50 nmol/L in study 1, in study 2 we examined the effect of winter vitamin D 387 supplementation on hepatitis B vaccine response. Study 2, a randomized, placebo-controlled 388 trial, involved a unique comparison of safe, simulated, casual skin sunlight exposure and oral 389 vitamin D<sub>3</sub> supplementation specifically designed to achieve vitamin D sufficiency. Contrary to 390 our hypothesis, and despite achieving and maintaining IOM and EFSA defined vitamin D 391 392 sufficiency in 95% of participants (Figure 6), vitamin D supplementation beginning 3 days after the initial hepatitits B vaccination did not influence the hepatitis B vaccine response (Figure 7). 393

394

The divergent findings of study 1 and 2 are contrary to our hypothesis; however, they are 395 consistent with animal and human studies that have identified it is the early (within 24 h), rather 396 than later, stages of orchestrating the development of immunity that are most sensitive to 397 intervention [40,41]. Indeed, vitamin D, and specifically 1,25(OH)<sub>2</sub>D, may influence the hepatitis 398 B vaccine response by stimulating antigen presenting cells, which are pivotal for the initial 399 capturing, processing and presenting of the antigen at the site of vaccination [42,43]. In animal 400 models, it has been observed that locally produced  $1.25(OH)_2D$  induced migration of dendritic 401 cells from the site of vaccination to non-draining lymphoid organs, where they can stimulate 402 antigen specific T and B-cells to mount a strong and persistent antibody response to diphtheria 403 vaccination [7,8]. Co-administration of 1,25(OH)<sub>2</sub>D with trivalent influenza vaccine in mice was 404 shown to enhance both mucosal and systemic specific antibody response [44,45], and highlights 405 vitamin D as a potential vaccine adjuvant. In addition, previous research in humans has shown 406 higher IgG antibodies in response to tetanus toxoid vaccination after 9 weeks of vitamin D 407

20

supplementation compared to a placebo group [10], which lends further support to the notion ofvitamin D as a potential adjuvant for vaccines more generally.

410

In both studies, we were unable to collect an additional blood sample after the third, and final, 411 hepatitis B vaccine dose; therefore, it remains to be determined whether vitamin D influences the 412 final development of the hepatitis B vaccine response. As non-responders to initial vaccine dose 413 tend to be poorer responders to subsequent doses [15], it is reasonable to hypothesise that persons 414 415 low in vitamin D at the initial hepatitis B vaccination are more likely to be vaccine nonresponders after the full hepatitis B vaccine course (Figure 3). Future studies should however 416 confirm the influence of vitamin D status at the time of initial vaccination on final antibody status 417 after the full hepatitis B vaccine course. Study 1 was a prospective cohort study, and it is 418 therefore possible factors other than vitamin D may explain the associations observed between 419 vitamin D, season and the hepatitis B vaccine response. Previously, body mass index, mood, 420 sleep and lifestyle (alcohol and smoking use) have been shown to influence the hepatitis B 421 vaccination response [13-15]. Further, seasonal alterations in infectious disease and compromised 422 host immunity might influence seasonal alterations in hepatitis B vaccination independent of 423 vitamin D status [46]. A strength of our studies is that we took account of these factors and 424 showed they were similar across the seasons (Table 1), and between persons who were vitamin D 425 sufficient and not (Table 2) and supplementation groups (Table 3). Furthermore, all cause illness, 426 a marker of host immunity (Tables 1 & 2), and living conditions were also similar. These 427 similarities strengthen the argument that vitamin D, rather than other factor(s), is responsible for 428 observed association with hepatitis B vaccination in study 1. Nonetheless, future randomized 429 control studies using similar supplementation methods as study 2 that improve vitamin D status 430

21

before the initial vaccination would verify whether vitamin D status at the time of initialvaccination is important in the development of the hepatitis B response.

433

434	The objective of these original studies was to explore the influence of vitamin D status on the
435	hepatitis B vaccination response, with the interventions designed to achieve vitamin D
436	sufficiency including a 4-week period of low-level SSR (12 exposures) followed by 8-weeks of
437	maintenance SSR (8 exposures). While vitamin D synthesis is the major established health
438	benefit of UVR, the latter has immunomodulatory (both suppressive and augmenting) effects,
439	which may be mediated through vitamin D-dependent and -independent pathways [28,29]. Thus a
440	previous human study of contrasting design examined for a possible effect of prior acute higher-
441	level UVR exposure (UVB therapy lamps; daily exposures given at the individual's sunburn
442	threshold for 5 days) on the first hepatitis B vaccination response [47]. The investigators did not
443	relate their findings to vitamin D status. They found no overall impact of UVR on cellular
444	(lymphocyte stimulation test) or humoral (antibody titre) response to hepatitis B surface antigen,
445	despite the UVR regime being adequate to reduce other immune responses, i.e. contact
446	hypersensitivity and natural killer cell activity. Further analysis found individual difference in
447	susceptibility, with a reduced vaccination response observed in those individuals with a minor
448	variant of IL-1beta polymorphism; prevalence of the variant is low and further studies are
449	suggested [48].

450

In combination with findings in elderly chronic kidney disease patients [11], our findings in
healthy adults highlight the potential importance of preventing low vitamin D status at the time of

the initial vaccination for the adequate development of the hepatitis B vaccination. Future 453 research is merited to confirm the influence of vitamin D on the hepatitis B vaccination response 454 in infants and the elderly, who are at greater risk of poor vitamin D status than healthy young 455 adults [49], and because the hepatitis B vaccination is mandatory during infancy in several 456 countries [21,22]. This does not reduce the impact of the current studies findings as many adults 457 remain unvaccinated because childhood vaccine coverage is ~90% or less and routine infant 458 hepatitis B vaccination began only recently in some countries (e.g. UK [21-23]). Adult 459 460 vaccination is recommended for persons at increased risk of exposure to bodily fluids such as health care professionals, patients, and those travelling to areas of the world where hepatitis B is 461 widespread e.g. sub-Saharan Africa, east and southeast Asia and the Pacific Islands [16]. The 462  $1,25(OH)_2D$  findings from study 1 also provide a mechanism by which maintaining vitamin D 463 sufficiency and high 1,25(OH)<sub>2</sub>D may be important for vaccine immunogenicity beyond hepatitis 464 B. As more than 50% fail to achieve vitamin D sufficiency during winter months [24-26] future 465 research to further understand the role of vitamin D on vaccination more broadly is warranted. 466 The 8% difference in hepatitis B vaccination response between people who were vitamin D 467 sufficient and 25(OH)D <50 nmol/L, and the 18% difference between winter and summer 468 (Figures 3A & 4D) are comparable with the effects on the hepatitis B vaccine response shown for 469 other lifestyle factors e.g. smoking, obesity and poor sleep [13,15]. Of particular clinical interest, 470 the winter vaccine response (44% anti-HBs titers  $\geq 10$  mIU/mL) was poorer than typically 471 expected after two hepatitis B vaccine doses (50–90%: Figure 4) [50]. Therefore, rather than 472 restoring vitamin D sufficiency from its winter nadir, as in study 2, we suggest maintaining year-473 round vitamin D sufficiency, and where necessary preventing the decline in the end of summer 474 serum 25(OH)D by commencing vitamin D supplementation in late summer or early autumn and 475

476	continuing until spring (~6 months). To maintain end of summer serum 25(OH)D individuals
477	should aim to achieve current IOM and EFSA vitamin D dietary intake recommendations
478	[19,20]. We achieved this in study 2 with a daily 400 IU oral vitamin $D_3$ dose (Figure 6). Oral
479	vitamin D <sub>3</sub> supplementation is recommended in the autumn and winter because unlike simulated
480	sunlight there is no time burden for an individual; no requirement for bulky irradiation cabinets;
481	and oral vitamin $D_3$ supplementation is effective regardless of sun reactive skin type [51].
482	Further, even very low sub-sunburn UVR doses were recently shown to cause skin cell DNA
483	damage in easy-burning skin types [52]. Low-level sunlight exposure, as used in study 2, may
484	however provide benefits to human health additional to vitamin D synthesis, and this is an active
485	area of research [29].

486

#### 487 *Conclusions*

In a prospective cohort study of 447 healthy adults (study 1), vitamin D sufficiency was rare 488 during the UK winter, and fewer people responded to the hepatitis B vaccination than during the 489 summer. In study 1, poorer vitamin D status (serum  $1,25(OH)_2D \le 120 \text{ pmol/L}$  and  $25(OH)D \le 40$ 490 nmol/L) at the time of initial vaccination was associated with fewer healthy adults responding to 491 the hepatitis B vaccine. In a subsequent randomized control trial (study 2), vitamin 492 D supplementation (oral or via simulated sunlight exposure) that began 3 days after the initial 493 vaccination, and achieved vitamin D sufficiency within 5 weeks, did not influence the hepatitis B 494 vaccination response. Randomized control trials that manipulate vitamin D status before the 495 initial vaccination are warranted to confirm the influence of vitamin D status at the time of initial 496 vaccination on the hepatitis B vaccine response. In accordance with the findings of the 497 prospective cohort study (study 1), avoiding low vitamin D status at the time of the initial 498

499	hepatitis B vaccination, by maintaining year-round vitamin D sufficiency, might
500	be recommended to optimise the response to hepatitis B vaccination. This is particularly
501	important for persons that rely on effective vaccination prophylaxis such as health care
502	professionals and patients regularly exposed to bodily fluids.
503	
504	Acknowledgements
505	This work was funded by the Ministry of Defence, UK. LER is supported by the National
506	Institute for Health Research (NIHR) Manchester Biomedical Research Centre. We would like to
507	thank Xin Hui Aw Yong, Mark Ward, Claire Potter, Anna Ferrusola-Pastrana, Dr Gabriella
508	Rossetti, Jason Edwards, Sophie Harrison, and Dr Thomas O'Leary for their assistance with data
509	collection. We also thank Dr Michael Zurawlew for his assistance with intervention
510	randomization and Prof Ann Webb and Dr Richard Kift for providing and analyzing the
511	polysulphone badges.
512	
513	The authors' responsibilities were as follows: NPW, JPG and SJ conceived the project and had
514	primary responsibility for the final content; SJO, RMI, DA, LER, WDF, JPG, and NPW
515	developed the overall research plan; SJO, RMI, SJ, JPG, and NPW had study oversight; DSK,
516	SJO, LMW, RR, ATC, JCYT, SJ, RMI, DA, LER, WDF and NPW conducted the research and
517	analyzed the samples; DSK, SJO, and RR performed the statistical analysis; DSK, SJO, and

518 NPW wrote the manuscript with LMW, RR, ATC, JCYT, SJ, RMI, DA, LER, WDF and JPG. All

519 authors read and approved the final manuscript.

References 521 522 1. Chang SH, Chung Y, Dong C (2010) Vitamin D suppresses Th17 cytokine production by 523 inducing C/EBP homologous protein (CHOP) expression. J Biol Chem 285 (50):38751-38755. 524 doi:10.1074/jbc.C110.185777 525 2. He CS, Handzlik M, Fraser WD, Muhamad A, Preston H, Richardson A, Gleeson M (2013) 526 Influence of vitamin D status on respiratory infection incidence and immune function during 4 527 months of winter training in endurance sport athletes. Exerc Immunol Rev 19:86-101 528 3. Wang TT, Nestel FP, Bourdeau V, Nagai Y, Wang Q, Liao J, Tavera-Mendoza L, Lin R, 529 Hanrahan JW, Mader S, White JH (2004) Cutting edge: 1,25-dihydroxyvitamin D3 is a direct 530 inducer of antimicrobial peptide gene expression. J Immunol 173 (5):2909-2912 531 4. Baeke F, Takiishi T, Korf H, Gysemans C, Mathieu C (2010) Vitamin D: modulator of the 532 immune system. Curr Opin Pharmacol 10 (4):482-496. doi:10.1016/j.coph.2010.04.001 533 5. Lang PO, Aspinall R (2015) Can we translate vitamin D immunomodulating effect on innate 534 and adaptive immunity to vaccine response? Nutrients 7 (3):2044-2060. doi:10.3390/nu7032044 535 6. Lemire JM (1995) Immunomodulatory actions of 1,25-dihydroxyvitamin D3. J Steroid 536 Biochem Mol Biol 53 (1-6):599-602 537

7. Enioutina EY, Bareyan D, Daynes RA (2008) TLR ligands that stimulate the metabolism of
vitamin D3 in activated murine dendritic cells can function as effective mucosal adjuvants to

subcutaneously administered vaccines. Vaccine 26 (5):601-613.

541 doi:10.1016/j.vaccine.2007.11.084

- 542 8. Enioutina EY, Bareyan D, Daynes RA (2009) TLR-induced local metabolism of vitamin D3
- plays an important role in the diversification of adaptive immune responses. J Immunol 182
- 544 (7):4296-4305. doi:10.4049/jimmunol.0804344
- 545 9. von Essen MR, Kongsbak M, Schjerling P, Olgaard K, Odum N, Geisler C (2010) Vitamin D
- controls T cell antigen receptor signaling and activation of human T cells. Nat Immunol 11
- 547 (4):344-349. doi:10.1038/ni.1851
- 10. Heine G, Drozdenko G, Lahl A, Unterwalder N, Mei H, Volk HD, Dorner T, Radbruch A,

549 Worm M (2011) Efficient tetanus toxoid immunization on vitamin D supplementation. Eur J Clin

550 Nutr 65 (3):329-334. doi:10.1038/ejcn.2010.276

11. Zitt E, Sprenger-Mahr H, Knoll F, Neyer U, Lhotta K (2012) Vitamin D deficiency is

associated with poor response to active hepatitis B immunisation in patients with chronic kidney

553 disease. Vaccine 30 (5):931-935. doi:10.1016/j.vaccine.2011.11.086

12. Jhorawat R, Jain S, Pal A, Nijhawan S, Beniwal P, Agarwal D, Malhotra V (2016) Effect of

vitamin D level on the immunogenicity to hepatitis B vaccination in dialysis patients. Indian J

556 Gastroenterol 35 (1):67-71. doi:10.1007/s12664-016-0621-8

- 13. Averhoff F, Mahoney F, Coleman P, Schatz G, Hurwitz E, Margolis H (1998)
- 558 Immunogenicity of hepatitis B Vaccines. Implications for persons at occupational risk of
- hepatitis B virus infection. Am J Prev Med 15 (1):1-8
- 14. Glaser R, Kiecolt-Glaser JK, Bonneau RH, Malarkey W, Kennedy S, Hughes J (1992) Stress-
- induced modulation of the immune response to recombinant hepatitis B vaccine. Psychosom Med

562 54 (1):22-29

- 15. Prather AA, Hall M, Fury JM, Ross DC, Muldoon MF, Cohen S, Marsland AL (2012) Sleep
- and antibody response to hepatitis B vaccination. Sleep 35 (8):1063-1069.
- 565 doi:10.5665/sleep.1990
- 16. Public Health England (2017) Hepatitis B: the green book, Chapter 18. vol 7. Retrieved from
- 567 https://www.gov.uk/government/publications/hepatitis-b-the-green-book-chapter-18#history
- 17. Huzly D, Schenk T, Jilg W, Neumann-Haefelin D (2008) Comparison of nine commercially
- available assays for quantification of antibody response to hepatitis B virus surface antigen. J
- 570 Clin Microbiol 46 (4):1298-1306. doi:10.1128/JCM.02430-07
- 18. He CS, Aw Yong XH, Walsh NP, Gleeson M (2016) Is there an optimal vitamin D status for
- immunity in athletes and military personnel? Exerc Immunol Rev 22:42-64
- 19. Institute of Medicine (2011) Dietary reference intakes for calcium and vitamin D. National
- Academies Press, Washington, DC. doi:<u>https://doi.org/10.17226/13050</u>

- 20. European Food Safety Authority (2016) Scientific opinion on dietary reference values for
  vitamin D. EFSA J 14 (10):1-145
- 577 21. Schillie S, Vellozzi C, Reingold A, Harris A, Haber P, Ward JW, Nelson NP (2018)
- 578 Prevention of Hepatitis B Virus Infection in the United States: Recommendations of the Advisory
- 579 Committee on Immunization Practices. MMWR Recomm Rep 67 (1):1-31.
- 580 doi:10.15585/mmwr.rr6701a1
- 581 22. Bozzola E, Spina G, Russo R, Bozzola M, Corsello G, Villani A (2018) Mandatory
- vaccinations in European countries, undocumented information, false news and the impact on
- vaccination uptake: the position of the Italian pediatric society. Ital J Pediatr 44 (1):67-67.

584 doi:10.1186/s13052-018-0504-y

- 585 23. Public Health England (2019) Historical vaccine development and introduction of vaccines in
- the UK. Vaccination timeline. Public Health England, Retrieved from
- 587 <u>https://www.gov.uk/government/publications/vaccination-timeline</u>
- 588 24. Szmuness W, Stevens CE, Harley EJ, Zang EA, Oleszko WR, William DC, Sadovsky R,
- 589 Morrison JM, Kellner A (1980) Hepatitis B vaccine: demonstration of efficacy in a controlled
- clinical trial in a high-risk population in the United States. N Engl J Med 303 (15):833-841.
- 591 doi:10.1056/NEJM198010093031501
- 592 25. Holick MF (2009) Vitamin D status: measurement, interpretation, and clinical application.
- 593 Ann Epidemiol 19 (2):73-78. doi:10.1016/j.annepidem.2007.12.001

- 594 26. Advisory Group on Non-ionising Radiation (2017) Ultraviolet radiation, vitamin D and
- 595 health. Public Health England, London

596 27. Bischoff-Ferrari HA (2014) Optimal serum 25-hydroxyvitamin D levels for multiple health

- 597 outcomes. Adv Exp Med Biol 810:500-525
- 598 28. Hart PH, Gorman S, Finlay-Jones JJ (2011) Modulation of the immune system by UV
- radiation: more than just the effects of vitamin D? Nat Rev Immunol 11 (9):584-596.

600 doi:10.1038/nri3045

29. Hart PH, Norval M, Byrne SN, Rhodes LE (2019) Exposure to Ultraviolet Radiation in the

Modulation of Human Diseases. Annu Rev Pathol 14:55-81. doi:10.1146/annurev-pathmechdis012418-012809

- 30. Terry PC, Lane AM, Lane HJ, Keohane L (1999) Development and validation of a mood
  measure for adolescents. J Sports Sci 17 (11):861-872. doi:10.1080/026404199365425
- 31. Fitzpatrick TB (1988) The validity and practicality of sun-reactive skin types I through VI.
  Arch Dermatol 124 (6):869-871

32. Webb AR, Kift R, Durkin MT, O'Brien SJ, Vail A, Berry JL, Rhodes LE (2010) The role of
sunlight exposure in determining the vitamin D status of the U.K. white adult population. Br J
Dermatol 163 (5):1050-1055. doi:10.1111/j.1365-2133.2010.09975.x

- 33. Rhodes LE, Webb AR, Fraser HI, Kift R, Durkin MT, Allan D, O'Brien SJ, Vail A, Berry JL
- (2010) Recommended summer sunlight exposure levels can produce sufficient (> or =20 ng ml(-
- 613 1)) but not the proposed optimal (> or =32 ng ml(-1)) 25(OH)D levels at UK latitudes. J Invest
- 614 Dermatol 130 (5):1411-1418. doi:10.1038/jid.2009.417
- 615 34. Webb AR, Kift R, Berry JL, Rhodes LE (2011) The vitamin D debate: translating controlled
- experiments into reality for human sun exposure times. Photochem Photobiol 87 (3):741-745.
- 617 doi:10.1111/j.1751-1097.2011.00898.x
- 618 35. Carswell AT, Oliver SJ, Wentz LM, Kashi DS, Roberts R, Tang JCY, Izard RM, Jackson S,
- Allan D, Rhodes LE, Fraser WD, Greeves JP, Walsh NP (2018) Influence of vitamin D
- supplementation by sunlight or oral D3 on exercise performance. Med Sci Sports Exerc 50

621 (12):2555-2564. doi:10.1249/MSS.000000000001721

- 622 36. Cashman KD, Hill TR, Lucey AJ, Taylor N, Seamans KM, Muldowney S, Fitzgerald AP,
- Flynn A, Barnes MS, Horigan G, Bonham MP, Duffy EM, Strain JJ, Wallace JM, Kiely M

624 (2008) Estimation of the dietary requirement for vitamin D in healthy adults. Am J Clin Nutr 88

- 625 (6):1535-1542. doi:10.3945/ajcn.2008.26594
- 626 37. Tang JCY, Nicholls H, Piec I, Washbourne CJ, Dutton JJ, Jackson S, Greeves J, Fraser WD
- 627 (2017) Reference intervals for serum 24,25-dihydroxyvitamin D and the ratio with 25-
- hydroxyvitamin D established using a newly developed LC-MS/MS method. J Nutr Biochem
- 629 46:21-29. doi:10.1016/j.jnutbio.2017.04.005

- 630 38. Ross AC, Manson JE, Abrams SA, Aloia JF, Brannon PM, Clinton SK, Durazo-Arvizu RA,
- Gallagher JC, Gallo RL, Jones G, Kovacs CS, Mayne ST, Rosen CJ, Shapses SA (2011) The
- 632 2011 report on dietary reference intakes for calcium and vitamin D from the Institute of
- Medicine: what clinicians need to know. J Clin Endocrinol Metab 96 (1):53-58.
- 634 doi:10.1210/jc.2010-2704
- 39. Macdonald HM (2013) Contributions of sunlight and diet to vitamin D status. Calcif Tissue
  Int 92 (2):163-176. doi:10.1007/s00223-012-9634-1
- 40. Fleshner M, Watkins LR, Lockwood LL, Bellgrau D, Laudenslager ML, Maier SF (1992)
- 638 Specific changes in lymphocyte subpopulations: a potential mechanism for stress-induced
- 639 immunomodulation. J Neuroimmunol 41 (2):131-142
- 41. Harper Smith AD, Coakley SL, Ward MD, Macfarlane AW, Friedmann PS, Walsh NP (2011)
- 641 Exercise-induced stress inhibits both the induction and elicitation phases of in vivo T-cell-
- mediated immune responses in humans. Brain Behav Immun 25 (6):1136-1142.
- 643 doi:10.1016/j.bbi.2011.02.014
- 42. Penna G, Adorini L (2000) 1 Alpha,25-dihydroxyvitamin D3 inhibits differentiation,
- maturation, activation, and survival of dendritic cells leading to impaired alloreactive T cell
  activation. J Immunol 164 (5):2405-2411
- 43. D'Ambrosio D, Cippitelli M, Cocciolo MG, Mazzeo D, Di Lucia P, Lang R, Sinigaglia F,
  Panina-Bordignon P (1998) Inhibition of IL-12 production by 1,25-dihydroxyvitamin D3.

Involvement of NF-kappaB downregulation in transcriptional repression of the p40 gene. J Clin
Invest 101 (1):252-262. doi:10.1172/JCI1050

- 44. Daynes RA, Araneo BA, Hennebold J, Enioutina E, Mu HH (1995) Steroids as regulators of
- the mammalian immune response. J Invest Dermatol 105 (1 Suppl):14S-19S
- 45. Daynes RA, Araneo BA (1994) The development of effective vaccine adjuvants employing
- natural regulators of T-cell lymphokine production in vivo. Ann N Y Acad Sci 730:144-161
- 46. Fisman DN (2007) Seasonality of infectious diseases. Annu Rev Public Health 28:127-143.
- 656 doi:10.1146/annurev.publhealth.28.021406.144128
- 47. Sleijffers A, Garssen J, de Gruijl FR, Boland GJ, van Hattum J, van Vloten WA, van Loveren
- 658 H (2001) Influence of ultraviolet B exposure on immune responses following hepatitis B
- vaccination in human volunteers. J Invest Dermatol 117 (5):1144-1150. doi:10.1046/j.0022-
- 660 202x.2001.01542.x
- 48. Sleijffers A, Yucesoy B, Kashon M, Garssen J, De Gruijl FR, Boland GJ, van Hattum J,
- Luster MI, van Loveren H (2003) Cytokine polymorphisms play a role in susceptibility to
- 663 ultraviolet B-induced modulation of immune responses after hepatitis B vaccination. J Immunol
- 664 170 (6):3423-3428. doi:10.4049/jimmunol.170.6.3423
- 49. Roth DE, Abrams SA, Aloia J, Bergeron G, Bourassa MW, Brown KH, Calvo MS, Cashman
- 666 KD, Combs G, De-Regil LM, Jefferds ME, Jones KS, Kapner H, Martineau AR, Neufeld LM,

- Schleicher RL, Thacher TD, Whiting SJ (2018) Global prevalence and disease burden of vitamin
  D deficiency: a roadmap for action in low- and middle-income countries. Ann N Y Acad Sci
  1430 (1):44-79. doi:10.1111/nyas.13968
- 50. Joines RW, Blatter M, Abraham B, Xie F, De Clercq N, Baine Y, Reisinger KS, Kuhnen A,
- Parenti DL (2001) A prospective, randomized, comparative US trial of a combination hepatitis A
- and B vaccine (Twinrix) with corresponding monovalent vaccines (Havrix and Engerix-B) in
- adults. Vaccine 19 (32):4710-4719
- 51. Farrar MD, Kift R, Felton SJ, Berry JL, Durkin MT, Allan D, Vail A, Webb AR, Rhodes LE
- 675 (2011) Recommended summer sunlight exposure amounts fail to produce sufficient vitamin D
- status in UK adults of South Asian origin. Am J Clin Nutr 94 (5):1219-1224.
- 677 doi:10.3945/ajcn.111.019976
- 52. Shih BB, Farrar MD, Cooke MS, Osman J, Langton AK, Kift R, Webb AR, Berry JL, Watson
- 679 REB, Vail A, de Gruijl FR, Rhodes LE (2018) Fractional Sunburn Threshold UVR Doses
- 680 Generate Equivalent Vitamin D and DNA Damage in Skin Types I-VI but with Epidermal DNA
- Damage Gradient Correlated to Skin Darkness. J Invest Dermatol 138 (10):2244-2252.
- 682 doi:10.1016/j.jid.2018.04.015

This article was accepted in its current	; form to on 24th April 2020. This	s is a post-peer-review, pre-copy	edit version of an articl	e published in European 34
<b>Table 1.</b> Study 1 baseline participant	demographics, anthropometrics,	lifestyle behaviors, sleep, mood	and all cause illness in g	cohorts recruited across seasons
Journal of Nutrition. The final authent	icated version is available online	<del>at: http://dx.doj.org/Der: 10.10</del>	<del>07/S00394-020-02261-</del>	N

		Winter	Spring	Summer	Autumn
	All	n = 88	n = 63	n = 115	n = 181
	n = 447				
Demographics					
$\Delta qe (years)$	$21.7 \pm 3.0$	$21.5 \pm 3.0$	221 + 32	$21.9 \pm 3.0$	$21.5 \pm 3.1$
Ethnicity Couposion $[n_{0}(0)]$	424(07)	$21.5 \pm 5.0$	$22.1 \pm 3.2$	$21.9 \pm 3.0$	$21.3 \pm 3.1$ 179 (09)
Eminerty, Caucasian $[n(\%)]$	434 (97)	83 (97)	02 (98)	109 (90)	178 (98)
Anthropometrics					
Height (m)	$1.73\pm0.08$	$1.73\pm0.09$	$1.71\pm0.09$	$1.75\pm0.08$	$1.71\pm0.08$
Body mass (kg)	$71.8 \pm 10.8$	$72.1 \pm 11.3$	$70.8 \pm 10.8$	$74.0\pm10.7$	$70.5 \pm 10.5$
BMI $(kg/m^2)$	$24.0 \pm 2.7$	$23.9 \pm 2.8$	$24.2 \pm 2.7$	$24.1 \pm 2.6$	$23.9 \pm 2.7$
2 ( <b>g</b> )		2017 - 210		2	2007 - 201
Lifestyle behaviors					
Alcohol user, $[n(\%)]$	376 (88)	76 (93)	50 (82)	99 (87)	151 (88)
Smoker, $[n(\%)]$	259 (58)	53 (61)	38 (60)	71 (62)	97 (54)
Sleep night before initial vaccination					
Duration (h)	$6.4 \pm 0.8$	$6.3 \pm 0.7$	$6.4 \pm 0.5$	$6.3 \pm 0.9$	$6.6 \pm 0.9$
Ouality (very poor $= 1$ to very good $= 4$ )	$1.7 \pm 0.8$	$1.7 \pm 0.8$	$1.6 \pm 0.7$	$1.8 \pm 0.8$	$1.6 \pm 0.8$
Contraception $(n = 138)^{l}$					
None	36 (26)	7 (19)	4(11)	5 (14)	20 (56)
COCP	50 (36)	9 (18)	15 (30)	5 (10)	21 (42)
POP	9 (7)	2(22)	2(22)	1(11)	4 (45)
Injection	8 (6)	2 (25)	ົ້	1(12)	5 (63)
Implant	35 (25)	9 (26)	6 (17)	5(14)	15(43)
Implant	55 (25)	) (20)	0(17)	5 (14)	15 (45)
Mood before initial vaccination <sup>2</sup>					
Vigor	$8.4 \pm 3.0$	$8.5\pm3.0$	$7.3 \pm 3.1$	$8.6 \pm 2.8$	$8.7 \pm 3.1$
Anger	$0.9 \pm 1.6$	$0.6 \pm 1.1$	$0.7 \pm 1.5$	$1.0 \pm 1.6$	$0.9 \pm 1.7$
Tension	$4.8 \pm 3.4$	$4.1 \pm 3.1$	$4.7 \pm 3.7$	$4.3 \pm 3.1$	$5.3 \pm 3.5$
Confusion	$2.3 \pm 2.4$	$2.4 \pm 2.8$	$1.8 \pm 1.9$	$2.5 \pm 2.4$	$2.3 \pm 2.5$
Depression	$0.7 \pm 1.6$	$0.6 \pm 1.1$	$0.6 \pm 2.0$	$0.7 \pm 1.3$	$0.8 \pm 1.7$
Fatigue	42 + 30	36 + 29	43 + 33	42 + 28	44 + 30
i uuguo	$1.2 \pm 5.0$	$5.0 \pm 2.7$	1.5 ± 5.5	$1.2 \pm 2.0$	1.1 ± 5.0
All cause illness $[n (\%)]^3$	71 (16)	$8(9)^{*}$	10 (16)	10 (9)*	43 (24)

Values presented as mean  $\pm$  SD, unless otherwise stated. COCP, combined oral contraceptive pill, POP, progesterone-only pill. \* P < 0.05 lower than autumn. Notes: <sup>1</sup>Female contraception data collected from a female specific questionnaire (n = 37 excluded from final data analysis). <sup>2</sup>Greater scores indicate a greater feeling of the mood (maximum per mood = 20). <sup>3</sup>Physician diagnosed cases of respiratory and gastrointestinal tract infection.

35

**Table 2.** Study 1 baseline participant demographics, anthropometrics, lifestyle behaviors, sleep,mood and all cause illness in those with serum 25(OH)D <50 nmol/L and  $\geq$ 50 nmol/L

	Serum 25(OH)D						
	<50 nmol/L	≥50 nmol/L					
	n = 194	n = 253					
Demographics							
Men [n (%)]	139 (72)	133 (53)					
Women $[n(\%)]$	55 (28)	120 (47)					
Age (vears)	$21.3 \pm 2.9$	$22.0 \pm 3.2$					
Ethnicity, Caucasian $[n (\%)]$	186 (96)	248 (98)					
Anthropometrics							
Height (m)	$1.74 \pm 0.08$	$1.71 \pm 0.09$					
Body mass (kg)	$73.4 \pm 10.8$	$70.1 \pm 10.7$					
BMI (kg/m <sup>2</sup> )	$24.2 \pm 2.8$	$23.9 \pm 2.6$					
Lifestyle behaviors							
Alcohol user. $[n(\%)]$	167 (86)	209 (83)					
Smoker, [ <i>n</i> (%)]	122 (63)	137 (54)					
Sleep night before initial vaccination							
Duration (h)	$6.6 \pm 0.7$	$6.3 \pm 0.9$					
Quality (very poor = 1 to very $good = 4$ )	$1.7 \pm 0.7$	$1.7 \pm 0.8$					
Contraception $(n = 138)^{1}$							
None	14 (33)	22 (23)					
COCP	10 (23)	40 (43)					
POP	2 (5)	7 (7)					
Injection	4 (9)	4 (4)					
Implant	13 (30)	22 (23)					
Mood before initial vaccination <sup>2</sup>							
Vigor	$8.4 \pm 3.1$	$8.4 \pm 3.0$					
Anger	$0.8 \pm 1.4$	$0.9 \pm 1.6$					
Tension	$4.7 \pm 3.5$	$4.8 \pm 3.3$					
Confusion	$2.5 \pm 2.6$	$2.2 \pm 2.3$					
Depression	$0.8 \pm 1.8$	$0.7 \pm 1.4$					
Fatigue	$4.2 \pm 3.0$	$4.3 \pm 3.0$					
All cause illness $[n (\%)]^3$	29 (15)	42 (17)					

Values presented as mean  $\pm$  SD unless otherwise stated. COCP, combined oral contraceptive pill, POP, progesterone-only pill. There were no significant differences between vitamin D sufficient and insufficient participants in demographic, anthropometrics, lifestyle behaviors, sleep, mood or all cause illness before the initial hepatitis B vaccination at baseline. Notes: <sup>1</sup>Female contraception data collected from a female specific questionnaire (n = 37 excluded from final data analysis). <sup>2</sup>Greater scores indicate a greater feeling of the mood (maximum per mood = 20). <sup>3</sup>Physician diagnosed cases of respiratory and gastrointestinal tract infection.

	SSR	SSR-P	ORAL	ORAL-P
	n = 30	n = 28	n = 32	n = 29
Demographics				
Age (years)	$21.5\pm3.1$	$21.7 \pm 3.4$	$20.9\pm2.7$	$21.4\pm3.0$
Ethnicity (Caucasian) [n (%)]	29 (97)	28 (100)	32 (100)	29 (100)
Skin type (I, II, III, IV) $[n (\%)]^1$	3 (10), 8 (27), 14 (46), 5 (17)	2 (7), 10 (36), 13 (46), 3 (11)	3 (9), 11 (34), 13 (41), 5 (16)	2 (7), 9 (31), 15 (52), 3 (10)
Anthropometrics				
Height (m)	$1.78\pm0.05$	$1.77\pm0.05$	$1.78\pm0.07$	$1.78\pm0.06$
Body mass (kg)	$76.7 \pm 11.6$	$76.8\pm9.7$	$75.7 \pm 12.3$	$77.5\pm10.8$
BMI (kg/m <sup>2</sup> )	$24.3\pm3.3$	$24.4\pm2.8$	$24.9\pm2.8$	$24.9\pm2.8$
Lifestyle behaviors				
Alcohol user $[n(\%)]$	23 (77)	22 (79)	26 (81)	23 (77)
Smoker [ <i>n</i> (%)]	17 (57)	16 (57)	17 (53)	11 (38)
Sleep night before initial vaccination				
Duration (h)	$6.2 \pm 0.8$	$5.9 \pm 1.4$	$5.8 \pm 1.5$	$5.8 \pm 1.8$
Quality (very poor = 1 to very good = 4)	$2.9\pm0.7$	$2.8\pm0.7$	$2.8\pm0.7$	$2.8\pm0.7$
Mood before initial vaccination <sup>2</sup>				
Vigor	$8.0 \pm 3.4$	$9.0\pm2.9$	$7.1\pm2.9$	$8.2\pm3.2$
Anger	$1.0 \pm 1.8$	$1.5 \pm 2.5$	$1.2 \pm 2.0$	$0.7 \pm 1.6$
Tension	$3.0 \pm 2.2$	$3.6 \pm 3.4$	$3.2 \pm 3.3$	$2.6 \pm 2.1$
Confusion	$2.6 \pm 3.2$	$2.5 \pm 2.9$	$1.7 \pm 2.1$	$1.5 \pm 1.9$
Depression	$0.7 \pm 1.8$	$1.4 \pm 2.7$	$0.6 \pm 1.6$	$0.3 \pm 0.6$
Fatigue	$3.6 \pm 2.7$	$4.9 \pm 3.2$	$4.1 \pm 3.5$	$4.1 \pm 3.1$

**Table 3.** Study 2 baseline participant demographics, anthropometrics, lifestyle behaviors, sleep and mood in solar simulated radiation (SSR), SSR placebo (SSR-P) oral vitamin  $D_3$  (ORAL) and oral placebo (ORAL-P) supplemented groups

Values presented as mean  $\pm$  SD unless otherwise stated. There were no significant differences between supplemented groups in demographics, anthropometrics, lifestyle behaviors, sleep or mood before the initial hepatitis B vaccination at baseline (P > 0.05). Notes: <sup>1</sup>Skin types are based on Fitzpatrick scale [31]. <sup>2</sup>Greater scores indicate a greater feeling of the mood (maximum per mood = 20).

#### **Figure legends**

- FIGURE 1. Schematic of Study 1 and 2 procedures. Study 1 investigated the influence of vitamin D status at the time of the initial hepatitis B vaccination on the secondary antibody response to hepatitis B vaccination. Study 2 investigated the effect of vitamin D supplementation by solar simulated radiation (SSR), oral vitamin D<sub>3</sub> (ORAL), or placebo (SSR-P or ORAL-P) after the initial hepatitis B vaccination on secondary hepatitis B vaccine response. Needle and bottle icon represents hepatitis B vaccination doses. Blood tube icon represents when blood samples were obtained for serum 25(OH)D, 1,25(OH)<sub>2</sub>D and hepatitis B antibody titer measurements.
- FIGURE 2. Flow diagram indicating the numbers of participants assessed for eligibility, recruited, available at follow-up, and analyzed as part of Study 1. Anti-HBs; antibodies against hepatitis B antigen.
- FIGURE 3. Secondary hepatitis B vaccine response in those with serum 25(OH)D <50 nmol/L (n = 194) and serum 25(OH)D  $\geq$ 50 nmol/L (n = 253 adults, panel A), and low, medium and high serum 25(OH)D (panel B, n = 447) and low, medium and high 1,25(OH)<sub>2</sub>D terciles (panel C, n = 444). † *P* < 0.1, lower percentage of secondary hepatitis B vaccination responders (anti-HBs  $\geq$ 10 mIU/mL) in participants with 25(OH)D <50 nmol/L than vitamin D sufficient participants. ‡ *P* < 0.05, lower percentage of secondary hepatitis B vaccination responders (anti-HBs  $\geq$ 10 mIU/mL) in low 25(OH)D and 1,25(OH)<sub>2</sub>D terciles compared to medium 25(OH)D and high serum 1,25(OH)<sub>2</sub>D terciles.

- FIGURE 4. Seasonal variation in serum 25(OH)D (panel A), percentage of participants categorized as vitamin D sufficient (25(OH)D  $\geq$ 50 nmol/L; panel B), serum 1, 25(OH)<sub>2</sub>D (panel C), and percentage of secondary hepatitis B vaccination responders (anti-HBs  $\geq$ 10 mIU/mL; panel D) in 447 healthy, young men (n = 272) and women (n = 175) residing in the UK. Panels A and C data are mean  $\pm$  SD. Panels B and D are percentages represented by vertical bars. a, lower than summer (P <0.05). b, lower than autumn (P < 0.05). c, lower than spring (P < 0.05).
- FIGURE 5. CONSORT flow diagram indicating the numbers of participants assessed, recruited, randomly assigned, and analyzed as part of Study 2. Anti-HBs; antibodies against hepatitis B antigen. Vitamin D = SSR; solar simulated radiation, ORAL; oral vitamin D<sub>3</sub>. Placebo = SSR-P; solar simulated radiation placebo, ORAL-P; oral placebo.
- FIGURE 6. Serum 25(OH)D (panels A & D), percentage of participants categorized as vitamin D sufficient (serum 25(OH)D ≥50 nmol/L, panels B & E), serum 1,25(OH)<sub>2</sub>D (panels C & F) in response to 12-weeks of vitamin D supplementation by solar simulated radiation (SSR) and oral vitamin D<sub>3</sub> (ORAL). Panels A, B & C show comparisons of individual vitamin D and placebo supplementation groups (SSR, SSR-P, ORAL & ORAL-P). Panels D, E & F show combined vitamin D supplementation (SSR & ORAL) vs combined placebo (SSR-P & ORAL-P) groups. † P < 0.05, greater than baseline. ‡ P < 0.05, greater than week 5. \* P < 0.05, greater than SSR-P. § P < 0.05, greater than ORAL-P & SSR-P. # P < 0.05, greater than</li>

combined SSR-P & ORAL-P. Data are mean  $\pm$  SD (panels A, C, D & F) and vertical bars represent percentages (panels B & E).

Figure 7 Percentage of participants categorized as secondary hepatitis B vaccine responders (anti-HBs  $\geq 10$  mIU/mL, panels A & B) after 12-weeks of vitamin D supplementation by solar simulated radiation (SSR) and oral vitamin D<sub>3</sub> (ORAL). Panel A compares individual vitamin D and placebo supplementation groups (SSR, SSR-P, ORAL & ORAL-P). Panel B shows combined vitamin D supplementation (SSR & ORAL) *vs* combined placebo (SSR-P & ORAL-P) groups. There was no difference in vaccine response between individual vitamin D and placebo supplementation groups (panel A, SSR 60%, SSR-P 57%, ORAL 56%, ORAL-P 52%, P > 0.05) or between combined **vitamin D** and placebo groups (panel B, SSR and ORAL 58% *vs* SSR-P and ORAL-P 54%, P > 0.05).

40

# 685 Figure 1

Weeks															
Baseline				1	2	3	4	5	6	7	8	9	10	11	12
Study 1		I I I I I I I I I I I I I I I I I I I					I B B B								
				Restoration phase (4-weeks)		Maintenance phase (8-weeks)									
				S G	SR or 3-times	placel -a-we	bo ek	SSR or placebo once-a-week							
Study 2	Study 2		ORAL/ ORAL-P	1,	000 IU vitamii plac	/day c n D₃ o xebo	oral r	400 IU/day oral vitamin D <sub>3</sub> or placeb					00		
Study 2	Sludy Z	π					Ŧ								
		Ü					1 B	ŧ							

### 687 Figure 2







692 Figure 4



#### 695 Figure 5



696

698 Figure 6







