#### **1** No evidence for a reduction of genetic diversity despite a strong population

### 2 decline in the Corncrake *Crex crex*

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### 11 Abstract

The preservation of genetic diversity is an important aspect of conservation biology, since small populations frequently suffer from inbreeding and loss of genetic diversity that can increase their risk of extinction. Here, we report changes in various measures of genetic diversity over 12 years in a declining population of corncrake *Crex crex*, a grassland bird species of high conservation concern throughout Europe. Despite a twofold demographic decline during the same period, we found no evidence for a reduction of genetic diversity. The maintenance of genetic diversity is an opportunity that may help the implementation of effective conservation actions.

#### 19 Introduction

20 The preservation of genetic diversity is recognised as an important aspect of conservation biology 21 (Haig et al. 2016). Small populations frequently suffer from inbreeding and loss of genetic diversity. 22 Inbreeding reduces survival and fecundity as a result of an increase in the frequency of deleterious alleles or a reduction of heterozygote advantage, thus directly increasing the extinction risk of small 23 24 populations (Reed & Frankham 2003). On the long-term, maintaining sufficiently high genetic 25 variation within species and populations is key to make them able to adapt to new environmental conditions (Crandall et al. 2000). In an era of rapid environmental changes driven by human activity, 26 27 the observed decline of many populations is expected to reduce their adaptive potential and 28 jeopardises their long-term persistence. Hence, monitoring temporal changes in the genetic diversity 29 of wild populations can serve to prioritize management actions and as an indicator of the strength of 30 human impact.

31 The corncrake *Crex crex* is a grassland bird species of high conservation concern throughout 32 Europe (Schäffer & Koffijberg 2004). Its main threat is the intensification of agricultural practices, 33 and especially early mowing, that strongly reduces the survival of fledglings (Tyler et al. 1998). It has 34 resulted in a severe population decline in several western European countries (Koffijberg et al. 2016), such as France that experienced a 90% reduction of corncrake numbers in the last 30 years (Hennique 35 36 et al. 2013). So far, we ignore whether this demographic decline of corncrake populations has affected 37 their genetic diversity. Reduced population sizes and demographic bottlenecks are usually associated 38 with a loss of allelic richness and a reduction of heterozygosity (Reed & Frankham 2003), but 39 incoming gene flow from larger, not genetically depleted, populations may contribute to maintaining 40 genetic diversity. In the corncrake, a recent population genetic study revealed high genetic diversity 41 within, and large levels of gene flow among, European populations (Fourcade et al. 2016). However, 42 temporal changes in genetic diversity have not been investigated. It is therefore unknown whether the 43 most threatened corncrake populations gradually lose genetic diversity as a result of their 44 demographic decline.

In this study, we examined the changes in genetic diversity that occurred over a 12-years period in a corncrake population in western France that experienced a 2-fold decline during the same period. Individuals were sampled and genotyped in 2000, 2011 and 2012, and we estimated allelic richness, heterozygosity and effective population size in each year to test for a decline in genetic diversity over this period. We also analysed temporal population structure to assess whether this population became gradually genetically different over time.

#### 51 Methods

52 The corncrake is a migratory bird that breeds in floodplain meadows and extensive grasslands across 53 the Palearctic. The mechanisation of mowing and the abandonment of traditional haymaking practices 54 has led to a decline of corncrake populations throughout western Europe during the last century 55 (Schäffer & Koffijberg 2004). In France, a national monitoring scheme of singing males implemented 56 from the 1980s revealed that, at the national level, the number of individuals declined from ca. 2000 57 calling males in 1983 to ca. 200 in 2016. At the same time, the national distribution of corncrakes 58 became highly fragmented and contracted in the floodplain meadows around the city of Angers (-59 0.1154°W, 47.4216°N, see Figure 1) in western France, which now host the majority of breeding 60 corncrakes in France. In this region, the number of singing males has severely declined in the past 61 decades, from ca. 500 in 1983 to only ca. 60 in 2016 (Figure 1).

62 We used 55 blood samples of male corncrakes collected respectively from 25 and 30 63 individuals in 2011 and 2012 around Angers, and previously used to assess the genetic structure of 64 populations at the European scale (Fourcade et al. 2016). We also analysed 24 new blood samples 65 collected in 2000. Birds were captured using playbacks between May and July as described in Fourcade et al. (2016). All birds were ringed which ensured that no individual was sampled twice in 66 67 this study. We extracted genomic DNA from blood samples using a salt extraction protocol and genotyped all individuals at 15 microsatellite markers, including eight corncrake-specific markers 68 69 (Gautschi et al. 2002) and seven markers conserved across many bird species (Dawson et al. 2010;

Dawson et al. 2013). The full genotyping procedure followed the protocol described in Fourcade et al.(2016).

72 We computed the observed and expected heterozygosity, as well as the rarefied allelic 73 richness, of each locus in each sampling year, using the "hierfstat" R package (Goudet 2005). 74 Effective population size was estimated using the linkage disequilibrium approach implemented in 75 NEESTIMATOR 2.1 (Do et al. 2014), excluding rare alleles with frequency < 0.05. Additionally, we 76 tested for a temporal differentiation of the sampled population. First we computed pairwise indices of 77 population differentiation between sampling years: G''s<sub>T</sub>, an unbiased and standardized analogue of 78  $F_{ST}$  (Meirmans & Hedrick 2011), and D (Jost 2008), a measure of population differentiation based on 79 the number of alleles instead of on heterozygosity. Confidence intervals around these indices were 80 calculated based on a bootstrap approach with 10000 permutations as implemented in the "diveRsity" 81 R package (Keenan et al. 2013). Finally, we implemented the clustering algorithm of the program 82 STRUCTURE (Pritchard et al. 2000), which uses a Bayesian approach to assign to each individual a 83 membership probability to an *a priori* number of genetic clusters. We ran 10 STRUCTURE replicates 84 for K = 1, K = 2 and K = 3 clusters with the following settings: admixture model with correlated allele 85 frequencies (Falush et al. 2003), and 100 000 burn-in steps followed by 500 000 iterations. We also 86 turned the LOCPRIOR option on, which makes use of prior information to facilitate the detection of 87 weak genetic structures, usually the sampling location (Hubisz et al. 2009). Here, instead, we used the 88 years of sampling as prior genetic clusters.

#### 89 Results

90 There was no evidence for a decline in genetic diversity over time (Figure 2). Observed

91 heterozygosity was 0.70 ( $\pm$  0.07 s.e.m) in the oldest sample (2000), then 0.62 ( $\pm$  0.05 s.e.m) in 2011

and 0.68 ( $\pm$  0.06 s.e.m) in 2012. Similarly, there was almost no difference between sampling years in

terms of expected heterozygosity, with a mean of 0.70 ( $\pm$  0.07 s.e.m) in 2000, 0.76 ( $\pm$  0.04 s.e.m) in

94 2011 and 0.75 ( $\pm$  0.05 s.e.m) in 2012. There was a weak decrease in allelic richness over time, from

95 9.50 ( $\pm$  1.44 s.e.m) alleles per locus in 2000 on average, to 9.15 ( $\pm$  0.10 s.e.m) in 2011 and 8.87 ( $\pm$ 

1.08 s.e.m) in 2012. Finally, although the mean estimate of effective population size was highly
variable between years (183.3 in 2000, 60.5 in 2011 and 643.9 in 2012), the upper confidence interval
was always infinity, showing that there was no signal of linkage disequilibrium in our data that can
distinguish it from being indeterminably large.

100 Pairwise measures of genetic differentiation between years were not significant, as bootstrap 101 confidence intervals always included zero (Table 1). However, we note that  $G''_{ST}$  and D calculated 102 between the samples collected in 2000 and those collected in 2011 or 2012 were considerably larger 103 that between 2011 and 2012 (Table 1). STRUCTURE provided no evidence for a temporal genetic 104 structure neither, as shown by a larger likelihood for one genetic cluster and an estimated membership 105 of individuals to two or three genetic clusters that did not match the temporal structure of the data 106 (Supplementary material, Figure S1). Nevertheless, we observed that samples collected in 2011 and 107 2012 were more closely related with each other than with the 2000 sampling. For instance, assuming 108 K = 2, samples from 2000 were assigned to the red cluster of Figure S1 by 80 % on average, while the 109 mean membership for this cluster was 62 % both for the samples collected in 2011 and in 2012.

#### 110 Discussion

111 We demonstrated that, despite a strong and continuous demographic decline, the corncrake population 112 of Western France did not simultaneously face a reduction of its genetic diversity. Not only did 113 genetic diversity remained stable, it was also remarkably high for a population estimated around 180 114 males (in 2012). There is generally a strong correlation between microsatellite heterozygosity and 115 population size in birds (Evans & Sheldon 2008). At first, it is therefore surprising to observe that 116 genetic diversity did not reflect the population drop recorded in the field. Several hypotheses can 117 explain this result, which have profound implications for the management of this endangered 118 population.

In light of the available data, the most likely scenario is that genetic diversity was maintainedby gene flow from distant populations that have not suffered from the same demographic decline.

121 Although the available evidence remain scarce, there are records of occasional, within-season, long-122 distance movements (up to 1,500 km) revealed by ring recoveries (Schäffer & Koffijberg 2004; 123 Koffijberg et al. 2016) and song analyses (Mikkelsen et al. 2013). We also previously described large 124 levels of gene flow among European populations of corncrakes that we attributed to the dispersal of 125 individuals from the most productive sites of eastern Europe to the declining western European populations (Fourcade et al. 2016). (Koffijberg et al., 2016)(Koffijberg et al., 2016)In this regard, the 126 127 estimates of effective population size estimates could not be distinguished from infinity, suggesting 128 that the samples originated from a larger population than the few hundreds individuals recorded in the 129 study area. We also observed what might be a sign of a slow temporal differentiation. It may reveal 130 the gradual change in the genetic characteristics of the original population, in agreement with the 131 hypothesis that the maintenance of genetic diversity was achieved by regular immigration events.

132 The fact that the extreme decline of corncrake numbers was not followed by a similar 133 decrease of their genetic diversity is good news in a conservation perspective. For example, it is 134 known that low levels of heterozygosity, when they reflect a reduced genetic diversity at immune-135 related loci, can result in a higher susceptibility to pathogens (Hawley et al. 2005). In this regard, we 136 already observed that western European corncrake populations, including France, did not suffer from 137 higher malaria prevalence than the larger populations of eastern Europe (Fourcade et al. 2014). It 138 suggests that, as the population is not genetically depleted, it may be able to recover successfully if 139 effective management actions are implemented.

140 Longer genetic monitoring may however be needed to rule out the possibility that a slow 141 decline of genetic diversity is occurring. Indeed, it is possible that the population decline was still too 142 recent to result in a reduction of heterozygosity and that in a clear genetic signal will be detectable 143 later. In the absence of annual monitoring between 1998 and 2006, it is also unknown whether population decline started before or after 2000 (Figure 1). Samples from the 1980s or earlier, when 144 145 corncrake numbers where high both at the local and national levels (Hennique et al. 2013), would help 146 identifying the association between demography and genetic diversity in corncrake populations. 147 Interestingly, here, allelic richness showed a slight trend towards decline, which might be an early

148 warning of a loss of genetic diversity, since allelic richness is usually more sensitive to a decrease of 149 population size than heterozygosity (Allendorf 1986).(Reif & Vermouzek 2018) Accordingly, 150 previous analyses using approximate Bayesian computation suggested that contemporary genetic data 151 were compatible with a scenario of ongoing population decline (Fourcade et al. 2016). So far, the 152 agri-environmental schemes implemented to protect the corncrake in the region have failed to halt its 153 decline. Future environmental policies should now take advantage of the fact that genetic diversity has 154 been maintained to implement effective conservation strategies that may reverse the unfavourable demographic trend. 155

#### 156 Acknowledgements

157 The authors acknowledge funding from Plan Loire Grandeur Nature, European Regional

158 Development Fund, Région des Pays de la Loire, Angers Loire Métropole, Direction Régionale de

159 l'Environnement, de l'Aménagement et du Logement (DREAL) and Département Maine-et-Loire,

and thank people who helped collecting corncrake samples in the field: Gilles Mourgaud, Édouard

161 Beslot, Emmanuel Séchet.

#### 162 References

- Allendorf FW (1986) Genetic Drift and the Loss of Alleles Versus Heterozygosity. Zoo Biol, 5, 181164 190.
- 165 Crandall KA, Bininda-Emonds ORP, Mace GM, Wayne RK (2000) Considering evolutionary

166 processes in conservation biology. Trends Ecol Evol, 15, 290-295.

- 167 Dawson DA, Ball AD, Spurgin LG, Martín-Gálvez D, Stewart IRK, Horsburgh GJ, Potter J, Molina-
- 168 Morales M, Bicknell AWJ, Preston SAJ, Ekblom R, Slate J, Burke T (2013) High-utility
- 169 conserved avian microsatellite markers enable parentage and population studies across a wide
- 170 range of species. BMC Genomics, 14, 176-176.

171	Dawson DA, Horsburgh GJ, Küpper C, Stewart IRK, Ball AD, Durrant KL, Hansson B, Bacon I, Bird
172	S, Klein Á, Krupa AP, Lee J-W, Martín-Gálvez D, Simeoni M, Smith G, Spurgin LG, Burke T
173	(2010) New methods to identify conserved microsatellite loci and develop primer sets of high
174	cross-species utility - as demonstrated for birds. Molecular Ecology Resources, 10, 475-494.
175	Do C, Waples RS, Peel D, Macbeth GM, Tillett BJ, Ovenden JR (2014) NEESTIMATOR v2: re-
176	implementation of software for the estimation of contemporary effective population size (N-e)
177	from genetic data. Molecular Ecology Resources, 14, 209-214.
178	Falush D, Stephens M, Pritchard JK (2003) Inference of population structure using multilocus
179	genotype data: linked loci and correlated allele frequencies. Genetics, 164, 1567-1587.
180	Fourcade Y, Keiss O, Richardson DS, Secondi J (2014) Continental-scale patterns of pathogen
181	prevalence: a case study on the corncrake. Evolutionary Applications, 7, 1043-1055.
182	Fourcade Y, Richardson DS, Keišs O, Budka M, Green RE, Fokin S, Secondi J (2016) Corncrake
183	conservation genetics at a European scale: the impact of biogeographical and anthropological
184	processes. Biol Conserv, 198, 210-219.
185	Gautschi B, Klug Arter M, Husi R, Wettstein W, Schmid B (2002) Isolation and characterization of
186	microsatellite loci in the globally endangered Corncrake, Crex crex Linné. Conserv Genet, 3,
187	451-453.
188	Goudet J (2005) HIERFSTAT, a package for R to compute and test hierarchical F -statistics. Mol
189	Ecol, 2, 184-186.
190	Haig SM, Miller MP, Bellinger R, Draheim HM, Mercer DM, Mullins TD (2016) The conservation
191	genetics juggling act: integrating genetics and ecology, science and policy. Evolutionary
192	Applications, 9, 181-195.
193	Hawley DM, Sydenstricker KV, Kollias GV, Dhondt Aa (2005) Genetic diversity predicts pathogen
194	resistance and cell-mediated immunocompetence in house finches. Biol Lett, 1, 326-329.

- 195 Hennique S, Mourgaud G, Deceuninck B, Chanson C (2013) Deuxième plan national d'actions en
- faveur du Râle des genêts (Crex crex) 2013-2018. LPO, LPO Anjou, Ministère de l'Écologie, du
  Développement Durable et de l'Énergie, DREAL des Pays de la Loire.
- 198 Hubisz MJ, Falush D, Stephens M, Pritchard JK (2009) Inferring weak population structure with the
- assistance of sample group information. Molecular Ecology Resources, 9, 1322-1332.
- 200 Jost L (2008) GST and its relatives do not measure differentiation. Mol Ecol, 17, 4015-4026.
- 201 Keenan K, McGinnity P, Cross TF, Crozier WW, Prodöhl PA (2013) diveRsity: An R package for the
- 202 estimation and exploration of population genetics parameters and their associated errors.
- 203 Methods Ecol Evol, 4, 782-788.
- Koffijberg K, Hallman C, Keišs O, Schäffer N (2016) Recent population status and trends of
   Corncrakes Crex crex in Europe. Vogelwelt, 136, 75 87.
- 206 Meirmans PG, Hedrick PW (2011) Assessing population structure : Fst and related measures. Mol
  207 Ecol, 11, 5-18.
- Mikkelsen G, Dale S, Holtskog T (2013) Can individually characteristic calls be used to identify long distance movements of Corncrakes Crex crex? J Ornithol, 154, 751-760.
- 210 Pritchard JK, Stephens M, Donnelly P (2000) Inference of population structure using multilocus
- 211 genotype data. Genetics, 155, 945-959.
- Reed DH, Frankham R (2003) Correlation between Fitness and Genetic Diversity. Conserv Biol, 17,
  230-237.
- Reif J, Vermouzek Z (2018) Collapse of farmland bird populations in an Eastern European country
   following its EU accession. Conservation Letters.
- 216 Schäffer N, Koffijberg K (2004) Crex crex Corncrake. BWP Update, 6, 57-78.
- 217 Tyler GA, Green RE, Casey C (1998) Survival and behaviour of Corncrake Crex crex chicks during
- the mowing of agricultural grassland. Bird Study, 45, 35-50.

## 220 Tables

- 221 **Table 1:** Pairwise genetic differentiation between sampling years, expressed as *G*''<sub>ST</sub> below diagonal
- and *D* above diagonal. Numbers in brackets show the 95% confidence intervals obtained through
- 223 10000 permutations.

	2000	2011	2012
2000		0.0145 [-0.0216 - 0.0632]	0.0054 [-0.0270 - 0.0469]
2011	0.0474 [-0.0105 - 0.1132]		0.0010 [-0.0285 - 0.0440]
2012	0.0297 [-0.0148 - 0.0761]	0.0115 [-0.0345 - 0.0735]	

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### 226 Figures

Figure 1: Location of sampling site in France shown as a black rectangle, with the estimated European distribution of the Corncrake represented in light blue (left); population trend in the study region based on the number of singing males recorded during annual surveys (right). For visualisation purpose, the blue line shows the estimated trend  $\pm$  confidence interval according to a Poisson generalized additive model with k = 4 for the smooth term. The years of sampling (2000, 2011 and 2012) are represented as arrows.



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# 239 Supplementary material

- 240
- 241 **Figure S1:** Results of the STRUCTURE analysis. A: Likelihood of the data to belong to one, two or

three genetic clusters (mean and standard deviation across 10 replicates). B: Estimated membership of

243 each individual (represented as vertical bars) to two (top) or three (bottom) genetic clusters.

