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Executive Summary

The Working Party make more than 100 tabulated recommendations in antimicrobial prescribing for the treatment of infections caused by MDR GNB and suggest additional further research, and algorithms for hospital and community antimicrobial usage in urinary infection. The international definition of multi-drug resistance is complex, unsatisfactory and hinders the setting and monitoring of improvement programmes. We give a new definition of multi-resistance. The background information on the mechanisms, global spread, and the UK prevalence of antibiotic prescribing and resistance has been systematically reviewed. The treatment options available in hospitals using intravenous antibiotics and in primary care using oral agents have been reviewed, ending with a consideration of antibiotic stewardship and recommendations given. The guidance has been derived from current peer-reviewed publications and expert opinion with open consultation. Methods for systematic review were NICE compliant and in accordance with the SIGN 50 Handbook; critical appraisal was applied using AGREE II. Published guidelines were used as part of the evidence base and to support expert consensus.
The guidance includes recommendations for stakeholders, including prescribers, and antibiotic-specific recommendations. The clinical efficacy of different agents is critically reviewed. We found there are very few good quality comparative randomized clinical trials to support treatment regimens, particularly for licensed older agents.

Susceptibility testing of MDR GNB causing infection to guide treatment needs critical enhancements. Meropenem- or imipenem resistant Enterobacteriaceae should have their carbapenem MICs tested urgently, and any carbapenemase class determined: mandatory reporting of these isolates from all anatomical sites and specimens would improve risk assessments. Broth microdilution methods should be adopted for colistin susceptibility testing.

Antimicrobial stewardship programmes should be instituted in all care settings, based on resistance rates and audit of compliance with guidelines, but should be augmented by improved surveillance of outcome in Gram-negative bacteraemia, and feedback to prescribers. Local and national surveillance of antibiotic use, resistance and outcome should be supported and antibiotic prescribing guidelines should be informed by these data.
The diagnosis and treatment of both presumptive and confirmed cases of infection by GNB should be improved. This guidance, with infection control to arrest increases in MDR, should be used to improve outcome of infections with such strains. Anticipated users include medical, scientific, nursing, antimicrobial pharmacy and paramedical staff where they can be adapted for local use.

Lay Summary

Multi-drug resistant (MDR) Gram-negative bacteria (GNB) are bacteria (or germs) that remain susceptible to only one or two antibiotics. Gram-negative bacteria usually live in the gut (or in the environment), where they do no harm, but can appear and cause infection at other body sites that normally lack any bacteria, for example in the bladder or blood. This especially occurs in patients who are made vulnerable by underlying disease, injury or hospitalization. MDR GNB may be acquired from other patients who have received antibiotics. Infections caused by MDR GNB are difficult to treat and so may cause more prolonged symptoms in the site of infection and can cause additional complications such as pneumonia or infection in the blood. This can prolong the length of stay in hospital, and in some cases, can cause death. Some types of MDR GNB e.g. Acinetobacter spp. can be carried on the skin rather than the gut, again with no obvious signs or symptoms. ‘Colonization’ describes carriage of bacteria on body surfaces or in the gut without infection. When patients develop infection and require antibiotic treatment, selecting the correct antibiotic can be difficult. This report provides advice on the best choice of antibiotics currently available.

1 Introduction

This guidance has been prepared by a joint Working Party of the British Society for Antimicrobial Chemotherapy (BSAC), the Healthcare Infection Society (HIS) and the British Infection Association (BIA) to advise on the treatment of infections caused by MDR GNB. It also describes best practice in antimicrobial prescribing. There is an accompanying guideline describing appropriate infection prevention and control.
precautions, including hand hygiene, equipment and environmental cleaning and guidance on screening for MDR GNB. The infection control and prevention guideline should be used in conjunction with the present document. There is a glossary for technical terms (See Appendix 1).

The Working Party comprised a group of medical microbiologists and scientists, infectious disease physicians, infection control practitioners, epidemiologists, and patient representatives nominated by the Societies. The patient representatives were lay members and had direct experience of the treatment of healthcare-associated infections through personal experience, membership of SURF (Healthcare-acquired Infection Service Users Research Forum), patient charities or through involvement in the development of NICE guidelines. The representatives were:

Susan Bennett, Member of Health Care Acquired Infections, Service Users Research Forum, Leicester, UK
Jennifer Bostock, Member of Health Care Acquired Infections, Service Users Research Forum, Leicester, UK
Maria Cann, Trustee, MRSA Action, Kirkham, UK

They were involved in the preparation of the remit of the Working Party remit (Appendix 3), were invited to all meetings, invited to comment on the final draft prepared by the authors and endorsed the final version.

2 Guideline Development Team

2.1 Guideline Advisory Group

Phil Wiffen, Cochrane Pain, Palliative and Supportive Care Group Pain Research, Churchill Hospital Oxford, Nuffield Dept. of Clinical Neurosciences, Oxford.

Karla Soares-Wieser, Enhance Reviews, Ltd, Wantage.
2.2 Responsibility for Guidelines

The views expressed in this publication are those of the authors and have been endorsed by the three sponsoring societies following consultation. Patient representatives confirmed the guidelines addressed the questions raised in setting the Working Party’s remit.

3 The Working Party report

Date of publication: TBC 2017 (Published online TBC)

3.1 What is The Working Party Report?

This Report is a set of recommendations covering the treatment of infections caused by MDR GNB (i.e. herein defined as susceptible to only to one or two different antibiotics). Strains internationally defined as MDR GNB by possession of resistance to three or more classes of antibiotics can nevertheless be treated with a wide range of antibiotics so we argue the case for a re-definition below (See Section 6.2.).

The Working Party recommendations have been developed systematically through a multi-professional group based on published evidence. They should be used to develop local protocols for acute and long-term healthcare settings.

3.2 Why do we need a Working Party Report for these infections?

MDR GNB have become more prevalent internationally, including in the UK and Europe. The increased use of broad-spectrum agents encourages their proliferation. The spread of these bacteria causes infections that can increase the length of hospital stay and adversely affect the quality of life of patients. Public awareness has been increasing, and the relative lack of new antimicrobial agents to treat infections due to Gram-negative bacteria has resulted in the formulation of the five-year Antimicrobial Resistance Strategy by the UK Department of Health. Outbreaks are associated with
considerable, physical, psychological and financial costs. Evidence-based treatment regimens are effective in improving the outcome of infections due to these bacteria.

3.3 What is the purpose of the Report’s recommendations?

The Report describes appropriate antimicrobial chemotherapy for infections due to MDR Gram-negative bacteria.

3.4 What is the scope of these guidelines?

We examine the background information on the mechanisms, global spread, and the UK prevalence of resistance, prescribing, and then discuss treatment i) in hospitals using antibiotics intravenously and ii) in primary care using agents given orally, ending with a consideration of antibiotic stewardship. Data (and doses, where given) usually refer to adults as there are few data for children and neonates. Extrapolation from adult data for β-lactams seems reasonably secure but this is not necessarily the case for other agents. Another set of guidelines considers appropriate infection control principles, best practice hand hygiene, screening and environmental cleaning\(^3\). For the detailed scope for this guideline see Appendix 2.5 and for the review questions see Appendix 3.7.

3.5 What is the evidence for these guidelines?

In the preparation of these recommendations, systematic reviews were performed of peer-reviewed research using the searches show in Appendix 4. Expert opinion was also derived from published guidelines subjected to validated appraisal\(^2\). Evidence was assessed for methodological quality and clinical applicability according to protocols of the Scottish Intercollegiate Guidelines Network (SIGN) initially using SIGN 2011\(^1\) guidelines and then updating this as the working party continued to comply with the SIGN 2014 guidance\(^6\).
3.6 Who developed these guidelines?
A group of medical microbiologists, scientists, infectious disease physicians, infection control practitioners, epidemiologists, and patient representatives.

3.7 Who are these guidelines for?
Any hospital or general practitioner can use these guidelines and adapt them for local use. Expected users include clinical medical, nursing, antimicrobial pharmacy and paramedical staff. Paediatric licenses and formulation may limit the suitability of some of the discussed agents for children and neonates. Where there are specific issues relating to dosage, outcome or toxicity that are outside current license information, these are discussed. The guidelines should be used to improve the treatment of both presumptive and confirmed cases of infection by MDR GNB.

3.8 How are the guidelines structured?
Most areas (defined by questions) comprise an introduction, a summary of the evidence base with levels and a recommendation graded according to the available evidence. The guidelines are not organised by clinical indication.

3.9 How frequently are the guidelines reviewed and updated?
The guidelines will be reviewed and updated every four years if warranted by sufficient changes in the evidence or by the availability of new agents or formulations.

3.10 Aim
The primary aim of the review was to assess the current evidence for antimicrobial prescribing in the treatment of MDR Gram-negative infections. The secondary aims were: (a) to evaluate the efficacy of antibiotics to treat community, and hospital infections caused by MDR GNB (b) to evaluate the impact of educating and providing support to professionals and patients to reduce unnecessary use of antibiotics leading
to a reduction in the selective pressure for resistance, thereby assisting antibiotic stewardship.

4 Summary of Guidelines

The guidance has been derived from current best peer-reviewed publications and expert opinion. Each recommendation is graded according to standard grades and is associated with a class of supporting evidence, or it is presented as a Good Practice Point. General recommendations for stakeholders, including prescribers are made in Table 1. Specific antibiotic recommendations are made in Table 2.

4.1 How can the guidelines be used to improve clinical effectiveness?

The Guidelines can be used to direct and formulate antibiotic policies and to aid the prescribing practice of infection specialists and other clinicians. They provide a framework for clinical audit tools for quality improvement.

4.2 How much will implementation of the guidelines cost?

The majority of antimicrobial agents that are described in these guidelines are generic and are currently widely used. Newer β-lactam/β-lactamase inhibitors (BL/BLI) are more expensive than older BL/BLIs and most alternatives to carbapenems against MDR GNB are also more expensive. Extra financial support will be required for the surveillance of outcomes of bacteraemia. Implementation of these guidelines should enable better-focused therapy, with no increase in drug utilization and possibly a modest decrease.

4.3 Summary of suggested audit measures

Patients with infections with MDR GNB, should receive empirical (best guess) or definitive (i.e. after results of laboratory tests) appropriate antibiotic treatment (alone or in combination) and the former should be active in at least 80% of cases. It is
important to note that the basis on which resistance was defined was changed by EUCAST from predicting failed clinical response to deviation from the normal susceptibility of the species. In an era of multiple resistance, continuing to select for such resistant strains even when the patient has clinically responded to antibiotics to which the organism is resistant is undesirable. Control groups with infections at the same site and caused by the same species, but not MDR, or infections without known aetiology should not receive definitive treatment reserved for patients with MDR GNB. This audit should be conducted first for bacteraemias.

To reduce total antibiotic consumption, measured as defined daily doses. Quarterly use of carbapenems and piperacillin/tazobactam should be reduced if either is in the top quintile/1000 patient days as assessed in each quarter. Specialist and tertiary care units may have special needs and should be excluded from the quintile assessment. Reductions of use in such units should be undertaken but should be tailored by consideration of their speciality case mix.

Trimethoprim use should be reduced and nitrofurantoin use increased in primary care. Risk assessment tools for colonization and infection with MDR GNB in patients should be developed for the UK and put in place in all settings. Only infected patients known to be, or at risk of being (by these assessments), colonized with these bacteria should receive empirical treatment with drugs reserved for MDR GNB.

No antibiotic prescriptions for treating the elderly with asymptomatic bacteriuria (ASB), or urinary tract infection (UTI) in the presence of a urinary catheter unless bacteraemia or renal infection suspected.

No antibiotic prophylaxis for urinary catheter insertion or change unless previous history of symptomatic urinary infection (UTI) associated with a change of catheter, or
if there is trauma during catheter insertion, or if a urinary continence device has been inserted.

Gram-negative bacteraemia incidence should be decreased and outcome should be improved both in cases which developed in primary care, wider healthcare settings, and secondary and tertiary units.

Enhancements to surveillance should be planned and supported by information technology (IT) that allows record linkage and simplification of surveillance from the laboratory to national level.

4.4 E-learning tools

Continuing Professional Development questions and model answers are listed for self-assessment in Appendix 5.

5 Methodology

5.1 Evidence appraisal

Methods were in accordance with SIGN 50 and Cochrane Collaboration criteria \(^1\)\(^-\)\(^7\) and critical appraisal was applied using AGREEII.\(^2\) Accepted guidelines were used as part of the evidence base and to support expert consensus. Questions for review (See Appendix 3.7.) were derived from the Working Party Group which included patient representatives in accordance with Patient Intervention Comparison Outcome (PICO)\(^6\).

K Soares-Wiesner of Enhance Reviews Ltd. and Dr P Wiffen of Pain Research and Nuffield Department of Clinical Neurosciences, Oxford University used a systematic review process. Guidelines and research studies were identified for each search question. Systematic reviews, randomized controlled trials (RCT) and observational studies were included. The latter comprised cohort non-RCT, controlled before- and after-studies, and interrupted time series. All languages were searched. Search
strategies for each area are given in the sections below and in Appendix 4. MeSH
headings and free text terms were used in the Cochrane Library (Issue 11 2012),
Medline (1946-2012), Embase (1980-2012) and Cumulated Index of Nursing and Allied
Health Literature (CINAHL) (1984-2012). On 23rd May 2014, an update search was
conducted on Medline alone using the same strategy for references after 1st January
2013. Reference lists of included studies were searched. Additional references were
added in October 2016 and June 2017 to cover specific issues. Two review authors
independently screened all citations and abstracts identified, and screened full reports
of potentially eligible studies (those that addressed the review questions in primary or
systematic secondary research, or a clinical, in vitro, or in use study). Disagreements
were resolved by discussion, and rationales for exclusion of studies were documented.
Pre-tested data extraction forms were used, and study characteristics and results
collected. Data were extracted from observational studies for multiple effect estimates:
these included the number of cases analyzed, adjusted and unadjusted effect estimates,
with standard error or 95% confidence interval (CI), confounding variables and
methods used to adjust the analysis. If available, data were extracted from contingency
tables. Risk of bias was assessed using SIGN critical appraisal checklists. Interrupted
time series were assessed using the Cochrane Effective Practice and Organisation of
Care (EPOC) Group. Quality was judged by report of details of protection against
secular changes (intervention independent of other changes) and detection bias
(blinded assessment of primary outcomes and completeness of data). For outbreak
patterns associated with particular pathogens, the Working Party made additional
searches of descriptive studies to extract effective treatments for infections caused by
bacteria with specific resistance.
5.2 Data analysis and interpretation

Clinical outcomes were mortality, effectiveness of treatment, and length of hospital stay.

Microbial outcome measures were decreases in the prevalence of MDR GNB, or decreases in colonization or infection by specific GNB. Risk ratios (RR) were used for dichotomous variables, and mean differences with 95% CI were used for continuous variables. Analyses were performed in Revman 5.2. SIGN summary tables were used. Evidence tables and judgment reports were presented and discussed by the Working Party and the guidelines were prepared according to the nature and applicability of the evidence, patient preference and acceptability and likely costs. The level of evidence was as defined by SIGN (Table 3), and the strength of recommendation was based upon GRADE (Grading of Recommendations Assessment, Development and Evaluation) (Table 4). The grading relates to the strength of the supporting evidence and predictive power of the study designs, rather than the importance of the recommendation. Any disagreements between members were resolved by discussion.

For some areas and recommendations, only expert opinion is available; in such cases, a good practice recommendation has been made. A flow chart of the systematic review process is given in Figure 1.

5.3 Consultation process

These guidelines were opened to consultation with circulation to the stakeholders listed (See Appendix 6). The draft report was placed on the BSAC website for one month in June 2016 for open consultation. Views were invited on format, content, local applicability, patient acceptability and recommendations. The Working Party considered and collated comments, and agreed revisions.
6 Rationale for recommendations

6.1 Usage

It is beyond the scope of this guideline to define optimal quantitative usage of antibiotics by hospital beds or community populations and the UK is not an exceptionally high antibiotic user in international terms. Equally, measures to reduce antibiotic usage will depend on what apparent over usage is occurring in any community or hospital department. For this reason, the assessment of reduction measures whilst based on comparative epidemiology must also consider both clinical outcome measures and usage at the local level. Suggestions for reducing overall usage must therefore be largely implemented at the local level where risk to patients and benefit can be adequately assessed and lie beyond the practical scope of this guideline.

6.2 What is the definition of multi-drug-resistant Gram-negative bacteria?

Multi-drug resistant (MDR) is a vexed term. From 1980 it was used to mean, ‘resistant to multiple agents’ without the number or types of agents being specified. More recently the European Centre for Disease Prevention and Control (ECDC) has attempted to formalise the term as ‘resistant to three or more antibiotic classes’, whilst extremely drug resistant (XDR) is ‘susceptible only to one or two drug classes. These definitions, based on those for tuberculosis, are epidemiologically attractive, but can prove to be impractical. An international consensus is difficult to achieve, as not all products are available and tested by laboratories in all countries, and there is no universal testing policy for laboratories which make pragmatic decisions on what to test. Some antibiotic resistances are now very common and stable, e.g. to ampicillin and sulphonamides, so they are seldom tested, but if they are present the organism needs only one further resistance to count as MDR GNB by the “three classes of resistance” rule. There also is scope for disagreement on which antibiotics should be considered as separate classes, for example, monobactams behave similarly to oxyimino-cephalosporins in respect of
most resistance mechanisms but very differently in the case of metallo-lactamases (MBL).

Difficulties arise also if in vitro “susceptibility” is poorly defined e.g. with the absence of EUCAST breakpoints as, for example, for i) *Acinetobacter* spp. and sulbactam, and ii) for temocillin. Furthermore differences between European (EUCAST) and US (CLSI or FDA) breakpoints can affect fundamentally whether isolates are regarded as MDR or XDR and recruitment to, and results in, clinical trials. Separate breakpoints for urinary isolates although needed to take account of high urinary concentrations with some antibiotics also complicate assessments. Lack of laboratory uniformity in breakpoints can make comparisons and data aggregation meaningless. For example, EUCAST and CLSI breakpoints differ for piperacillin/tazobactam and amoxicillin/clavulanate. EUCAST defines Enterobacteriaceae isolates as piperacillin/tazobactam susceptible if they have an MIC=8mg/L (R>16mg/L) compared with <=16+4mg/L (R>=128+4 mg/L) in CLSI guidance. For amoxicillin/clavulanate susceptibility is defined by EUCAST as <=8+2mg/L (R>8mg/L (or 32+2mg/L for uncomplicated UTI) and by CLSI as <=8+4mg/L (R>=32+16mg/L). The FDA regard *Pseudomonas aeruginosa* isolates as susceptible to piperacillin/tazobactam if the MIC is <=64mg/L (the historical CLSI breakpoint for piperacillin) whereas EUCAST and CLSI now consider the breakpoint should be S=<=16+4mg/L. The EUCAST and CLSI definitions have changed with time and from previous national guidelines e.g. the pre-EUCAST BSAC breakpoint for amoxicillin/clavulanate in systemic infections was 8+4mg/L. Cefepime is a further example of an antibiotic with breakpoint changes: the old CLSI breakpoint for Enterobacteriaceae was <=8mg/L but is now <=2mg/L based on 1g.twice daily doses. Organisms with MICs of 4or 8mg/L are viewed as being “susceptible but dose-dependent” by CLSI. EUCAST categorises an MIC <=1mg/L as susceptible and >4mg/L as resistant. A failure rate of 83% in a prospective trial of cephalosporins for “susceptible”
serious infections due to ESBL-producing *Klebsiella spp.* and *E. coli* partly reflected the use of high breakpoints. Breakpoint differences and changes over time in the categorization of isolates with the same MIC as “susceptible” or “resistant” profoundly challenge conclusions in the clinical literature, including reports of regulatory trials on the response to be expected of infections due to “susceptible” or “resistant” strain or indeed which patients have been included in trials where susceptibility of the organism is a selection criterion.

For all these reasons, the international definitions have not lead to better surveillance of MDR strains and their usefulness must still be questioned. In our literature search routines, we have employed the international definitions but have had to augment these with literature on specific resistances. A useful pragmatic approach to the definition of MDR is to consider oral and parenteral drugs separately as, in the UK, these will be largely used in primary, and secondary with tertiary, care respectively, with multi-resistance constituting different challenges in each setting. Furthermore, one should base definitions on susceptibility rather than resistance as the former is more likely to be sought clinically by further testing with MDR strains. This gives a basis for alternative definitions for MDR which we would advocate. For oral drugs, multi-resistance can usefully be defined as an organism susceptible to only one or no readily available oral agent active against infections systemically or in the upper urinary tract. This definition is vulnerable to the introduction of new, or newly re-licensed, oral agents, but this is appropriate and may emphasise the importance of new agents to the licensing authorities. By this definition the following would be classed as multi-resistant isolates for the community:

1) *Escherichia coli* resistant to co-amoxiclav (amoxicillin with clavulanic acid), oral cephalosporins, quinolones, trimethoprim but susceptible to nitrofurantoin, mecillinam and fosfomycin. Although providing options in cystitis these oral agents lack evidence of...
achieving systemically active concentrations and efficacy in upper and complicated UTIs, which is particularly relevant if these are caused by ESBL- and AmpC-producing strains. ii) *P. aeruginosa* resistant to quinolones. This approach could be modified to exclude agents where the mutation frequency is sufficiently high so that resistance commonly emerges during treatment.

For parenteral antibiotics a similar approach can be considered. Susceptibility to oral agents that have no licensed, or available, parenteral form e.g. pivmecillinam and nitrofurantoin should not be taken into account. Specific agents to which impaired susceptibility might be significant include carbapenems, relevant cephalosporins (cefotaxime for Enterobacteriaceae, ceftazidime for *P. aeruginosa*), aztreonam, ceftolozane/tazobactam, ceftazidime/avibactam, temocillin, piperacillin/tazobactam, colistin, quinolones, fosfomycin, tigecycline and aminoglycosides (including amikacin).

Given this greater number of agents and the paucity of new pipeline antibiotics active against Gram-negative bacteria, it is pragmatic to consider ‘multi-resistant’ as isolates where only two, or fewer, unrelated antibiotics are active against the bacterium. By such a definition the following would be considered multi-resistant isolates in hospitals:

i) *Acinetobacter baumannii* susceptible to two or fewer of meropenem or imipenem, (third generation cephalosporins), piperacillin/tazobactam, (tigecycline), aminoglycosides, quinolones, (trimethoprim), colistin, where agents in brackets lack EUCAST breakpoints.

ii) *Klebsiella spp.*, *Enterobacter spp.*, *Serratia spp.* and *Citrobacter spp.* that are susceptible to two or fewer of carbapenems, third-generation cephalosporins, including with β-lactamase inhibitors, piperacillin/tazobactam, temocillin, tigecycline, aminoglycosides, quinolones, trimethoprim or colistin.
iii) *Proteus* spp., *Morganella* spp. and *Providencia* spp. that are resistant to third-generation cephalosporin, piperacillin/tazobactam, and aminoglycosides and susceptible only to carbapenems, and the new beta-lactam/beta-lactam inhibitors (BL/BLI) combinations (ceftolozane/tazobactam or ceftazidime/avibactam). Unlike the species considered in ii) above, these Proteaeae are inherently resistant to tigecycline and colistin.

The following would not be regarded as multi-resistant:

i) *E. coli* that is susceptible to carbapenems, ceftolozane/tazobactam, ceftazidime/avibactam, colistin and fosfomycin but resistant to unprotected third-generation cephalosporins, co-amoxiclav, piperacillin/tazobactam, quinolones, and trimethoprim.

The effect of new parenteral antibiotic introductions on the definition of MDR GNB in hospitals is illustrated by the licensing of ceftazidime/avibactam and the availability of parenteral fosfomycin. Both drugs join temocillin, tigecycline or colistin, as potentially effective agents against some Enterobacteriaceae with KPC carbapenemases. Such strains would no longer be classified as MDR GNB by our definition. Clearly acquired resistance of KPC-producing strains to colistin, ceftazidime/avibactam, fosfomycin and tigecycline may all arise so some will be MDR GNB and some will not. From a therapeutic view this is probably appropriate although all should remain major targets for infection control, given the cost of new agents and the need to conserve their usefulness, along with plasmid-mediated transmission of *bla*<sub>KPC</sub> gene, and transmission of their host strains. The use of alternative β-lactams or new BL/BLIs rather than carbapenems may be expensive but might reduce the selective pressure for carbapenem-resistant MDR GNB. These antimicrobials, with activities against different β-lactamases, may have differential effects on the prevalence of particular β-lactamases.
and other carbapenem-resistant bacteria. They may select more for MBLs which are particularly resistant to β-lactams which will limit their ultimate usefulness in a locality. The activity of different β-lactamase inhibitors against, and stability of β-lactams to, different β-lactamases is shown in Table 5.

The difficulty in international surveillance of MDR GNB need not preclude the establishment of surveillance for specific organism-antibiotic resistance combinations. This has been adopted by Public Health England for the English Surveillance Programme for Antibiotic Use and Resistance (ESPAUR) and is weighted towards resistance to third-generation cephalosporins, quinolones and carbapenems of *E. coli*, *Klebsiella* spp., and *P. aeruginosa*.

### 6.3 What is the global epidemiology of MDR GNB?

#### 6.3.1 Origins and impact of multi-resistance

Resistance to multiple agents can develop via successive mutations, through the dissemination of multi resistance plasmids/genes (e.g. transposons), or through a combination of both processes. Resistance narrows antibiotic choices for definitive therapy. More critically, it increases the likelihood that empirical therapy will prove ineffective, increasing mortality in septic patients. Plasmids are the main source of multi-drug resistance in Enterobacteriaceae and *Acinetobacter* spp., except for mutations in DNA gyrase genes *gyrA/B* conferring fluoroquinolone resistance, mutational up-regulation of *arcA/B*-mediated efflux compromising tigecycline, and for mutational derepression of AmpC β-lactamases giving resistance to third-generation cephalosporins in *Enterobacter* spp., *Citrobacter* spp., *Serratia* spp., *Morganella morgani* 13 14. By contrast, sequential accumulation of mutations is paramount in *Pseudomonas* spp.
A recent review has discussed the emergence of specific resistance lineages and the role of different plasmid groups in emerging resistance problems in *E. coli*. Some clones have spread widely for reasons that are not clear. Resistance may increase their competitiveness, but some strains are adept at acquiring multi-drug resistance. Several strands of evidence support this view. First, some 'high-risk clones', e.g. *E. coli* ST131, frequently acquire diverse resistance determinants, including different extended-spectrum β-lactamases (ESBLs), AmpC and even carbapenemases. Secondly, there is co-selection of hypermutability with resistance in *P. aeruginosa* in patients with cystic fibrosis, facilitating development of further resistance. Thirdly, it is commonplace for plasmids and resistance islands to carry multiple genes encoding resistance to an antibiotic via two or more different mechanisms not all of which can remain under effective selection pressure. Fourthly the presence of toxin-antitoxin systems in plasmids may prevent loss of plasmids even when selective pressure is removed. Fifthly, integrons, which provide efficient gene-capture and expression systems, and which are now frequent in plasmids but were not present prior to the widespread use of antibiotics, provide a mechanism whereby resistance acquisition has accelerated. Finally, the presence of MDR GNB in the environment including foodstuffs and water sources provides important pathways for amplification and the spread of some resistance genes to man.

Until recently, environmental sources of carbapenemase genes did not appear to exist but the description of high levels of NDM-producing *E. coli* in chicken in China suggests this position will not be maintained with current international practices and biosecurity of food as a source. Surprisingly, the ST131 clone of *E. coli* did not seem to have significant environmental sources in its initial spread although it has now been described occasionally in chickens.
6.3.2 Epidemiological trends among multi-drug resistant Enterobacteriaceae: cephalosporin and quinolone resistance

Countries historically varied in the prevalence of different CTX-M ESBLs conferring cephalosporin resistance and in the plasmids encoding these enzymes. The prevalence of different CTX-M enzymes has changed with time and latterly in Europe and North America CTX-M-15 has become the dominant enzyme, often associated with *E. coli* ST131. Whole genome sequencing suggests that the acquisition of CTX-M enzymes occurred a number of times in clade C of *E. coli* ST131. Frequent co-carryage of OXA-1 penicillinases impairs susceptibility to combinations of clavulanate and tazobactam with penicillins. Ceftolozane appears stable to this OXA-1 enzyme. Other factors associated with the rise of multi-drug resistant Enterobacteriaceae include the spread of plasmids encoding AmpC β-lactamase. These seem around 10-fold less frequent than plasmids encoding ESBLs in the UK although more recently, in Canada a plasmid-mediated AmpC enzyme (CMY-2 which shares a promoter gene, ISEcp1, with CTX-M-15) was almost half as common as ESBL production and one third of such strains belonged to *E. coli* ST131. Distinguishing AmpC and ESBL cephalosporin-resistant strains is important epidemiologically and in routine testing, although both EUCAST and CLSI do not recommend it for guiding treatment. However early information on AmpC/ESBL status in Enterobacteriaceae may predict respectively ceftolozane/tazobactam resistance/susceptibility. Mutations can augment multi-drug resistance: for example, porin loss can engender resistance to ertapenem (and, sometimes, other carbapenems) in ESBL- and AmpC- producing Enterobacteriaceae.

6.3.3 Carbapenem resistance

Carbapenem resistance was initially slow to emerge in Enterobacteriaceae but is now steadily increasing, and mediated more and more by acquired carbapenemases (predominantly by KPC, VIM, IMP, NDM and OXA-48-like types). Internationally
there has been a considerable spread of *K. pneumoniae* clonal complex (CC) 258 isolates with KPC carbapenemases. The rise of NDM and OXA-48 carbapenemases is more often associated with the spread of their encoding plasmids or transposons among bacterial strains. Carbapenem resistance due to ESBL or AmpC enzymes combined with Omp K35 porin loss, may lead to treatment failure but is often unstable and may impose a fitness cost on bacteria, meaning that spread of such strains among patients is rare, though not unknown. Loss of the Omp K36 porin conferred resistance to new carbapenem-β-lactamase inhibitor combinations, relebactam with imipenem/cilastatin and meropenem with vaborbactam. Resistance conferred by acquired carbapenemases is of much greater concern, and is generally associated with considerable resistance to other agents.

Data from EARS-Net suggest that the prevalence of carbapenem-resistant Enterobacteriaceae causing bacteraemia markedly increased in most parts of Europe between 2013 and 2015. European prevalence of carbapenem-resistant *K. pneumoniae* was higher than 5% in 2015 (and much higher in some of the countries) in Greece, Italy, Cyprus and Romania. In Greece, the proportion of bloodstream *K. pneumoniae* isolates resistant to carbapenems increased from 27.8% in 2005 to 62.3% in 2014. VIM enzymes dominated early in this period but were replaced by KPC types, often carried by CC258. The rise of carbapenem-resistant *K. pneumoniae* in Italy has been dramatic and recent: from 1% of bacteraemias in 2009, to 15% in 2010 to 32.3% in 2014. This increase again is mainly due to CC258 *K. pneumoniae* with KPC enzymes. This clone also spread widely earlier in the USA and then in Israel, where an aggressive, nationwide infection control intervention was successful in bringing it under control. In Romania the major problem is *K. pneumoniae* producing OXA-48 carbapenemase.
Outbreaks of carbapenemase-producing Enterobacteriaceae (CPE) have been reported in many other parts of the world, including all US states (where KPC enzymes dominate), South Asia (predominantly NDM enzymes), the Middle East (OXA-48), Brazil and Colombia (KPC). The MBL IMP-4 has spread widely in China, often together with KPC-2. IMP-4, without KPC, is the dominant carbapenemase in Australia. Further global spread is to be expected as IMP-4 has now been observed in South London (unpublished observations, Prof. D. Livermore). In the absence of comprehensive international prevalence data for infection and carriage, risk factors for CPE are difficult to derive, but seem to include travel to high prevalence areas, notably including the Indian subcontinent for NDM-producers, and exposure to healthcare and antimicrobials. Travel locations are becoming convergent with those where ESBLs are prevalent. Case-number trigger points for carbapenem-resistant isolates and regional coordination in control action has recently been modeled in the USA to show the high importance of early intervention with effective control measures for K. pneumoniae strains and other Enterobacteriaceae. Carbapenem resistance in Enterobacteriaceae has been associated with increased attributable mortality probably owing to the greater likelihood that initial empirical therapy proves inadequate.

6.3.4 Global resistance issues with oral drugs with low resistance rates in the UK

A 2008 study of clinical isolates from women aged 18–65 years with symptoms of uncomplicated lower UTI in ten countries, found susceptibility rates above 90% only for fosfomycin (98%), mecillinam (96%), and nitrofurantoin (95%). Nitrofurantoin resistance in E. coli as assessed on European and Canadian isolates made in 1999-2000 and 2007-8 was associated with a very diverse range of sequence types although many strains showed multiple resistances: mecillinam resistance was similarly diverse but not associated with multiple-resistance. A further study from Munster and Seattle
suggests nitrofurantoin resistance is particularly common in ST58. Nitrofurantoin resistance is now described in 11% of the dominant H30 sub-clone of ST131 suggesting the drug may be selective in the upper intestine although this drug does not usually eliminate Enterobacteriaceae from the faecal flora of patients receiving it. In Canada, nitrofurantoin resistance rates in ESBL-producing E. coli were 16% but in ESBL-producing Klebsiella spp. were 71% (nosocomial) and 93% (non-nosocomial).

Well-described mutations in nitrofuran reductases confer resistance and plasmid-mediated resistance due to an efflux pump (oqxAB) has recently been described from Hong Kong. This efflux pump and its encoding plasmid (with the oqxAB gene flanked by IS26 insertion sequences) was found in 26/103 nitrofurantoin resistant or intermediate human isolates (by CLSI criteria) and was commoner in ESBL-producing isolates. The combination of oqxAB with the nitroreductase genes caused high-level nitrofurantoin resistance. This two level resistance process is analogous to the hypothetical role of AAC-6'-1b-cr in aiding the emergence of quinolone resistance by chromosomal mutation. Notably oqxAB also mediates resistance to mequindox, which is used in China as a growth promoter in animal feed. In China 322/1123 veterinary isolates of E. coli carried this gene but these mainly belonged to phylogroups A and B1 that are less associated with extra intestinal pathogenicity in man.

Fosfomycin use has been complicated by the emergence of resistance in some populations. In Spain when use increased some fifty percent between 2005 and 2008, resistance rates in CTX-M-15 ESBL producing E. coli rose to 16% and among all ESBL-producing isolates increased from 4.4% in 2005 to 11.4% in 2009. The increase was particularly associated with nursing homes. Fosfomycin resistance developed in E. coli ST131 (previously present there but not typed) and was not associated with described mutational mechanisms of fosfomycin resistance. Such mutations involve inactivation of genes encoding the hexose and triose sugar phosphate transport impairing drug.
uptake. A different mechanism is present in the acquired \textit{fosA} gene, which encodes a drug-inactivating metalloglutathione transferase \textsuperscript{60}. Fosfomycin resistance was present in 2009-2010 in 7.8\% human \textit{E. coli} in mainland China and approximately half of this was due to \textit{fosA}\textsubscript{3}\textsuperscript{64}. A recent survey of food animals in Hong Kong found plasmid-mediated \textit{fosA} to be increasing in frequency and associated with CTX-M ESBL-encoding plasmids \textsuperscript{65}. A recent Chinese survey of isolates collected from 2010 to 2013 detected fosfomycin resistance in 12\% of ESBL-producing \textit{Klebsiella} and 169/278 (61\%) of KPC-producing \textit{Klebsiella pneumoniae}: 94 KPC-producing strains carried \textit{fosA} \textsubscript{3} flanked by two IS26 insertions and were clonally related\textsuperscript{66}. Similar genetic findings were made in non-clonally related \textit{E. coli} and \textit{Klebsiella sp.} in Korea\textsuperscript{67}.

Mecillinam resistance is said to remain uncommon in the clinic – at 5-7\% of ESBL-producing \textit{E coli} in Sweden\textsuperscript{68}. In a wider European study, overall susceptibility was similar with 4.8\% resistance in \textit{E coli} from uncomplicated UTI, although gradually rising \textsuperscript{69}, notably in Spain where the resistant proportion of strains rose from 1\% in 2000 to 6.5\% in 2014.

\subsection*{6.4 How do multi-drug resistant Enterobacteriaceae differ from non-fermenters in terms of their prevalence and associated resistance genes?}

Carbapenem resistance is more common in non-fermenting Gram-negative bacteria than in Enterobacteriaceae. In \textit{A. baumannii}, it was common by the year 2000, to see isolates resistant to all treatment options except carbapenems, colistin and tigecycline. Subsequently, carbapenem resistance has proliferated, reaching c. 30\% of bloodstream isolates. It is largely associated with acquired OXA-23, -40 or 58-like carbapenemases or with insertion-sequence mediated upregulation of the chromosomal OXA-51-like carbapenemase. The strain structure of \textit{A. baumannii} is extremely clonal, making it difficult, without a history of patient transfers, to distinguish place-to-place spread from repeated independent selection of lineage variants that were previously circulating at
low frequency. UK *A. baumannii* isolates producing OXA-23 carbapenemases often co-
produce *ArmA* encoded 16S ribosomal methyltransferases conferring pan-
aminoglycoside resistance. Multi-drug resistant *Acinetobacter spp.* largely cause 
outbreaks in ICU settings \(^{70-72}\), whereas carbapenem-resistant Enterobacteriaceae, 
principally *E. coli* and *Klebsiella spp.*, cause infection in a wider group of patients, and 
have far greater potential to spread rapidly when introduced into wider patient 
populations \(^{36, 44, 45, 48, 73, 74}\).

Most UK *P. aeruginosa* remain susceptible to β-lactams, including ceftazidime, 
piperacillin/tazobactam and carbapenems, aminoglycosides and fluoroquinolones, with 
resistance rates of 5-10% for these agents; and fewer than 1% for 
ceftolozane/tazobactam \(^{75}\). Nevertheless, single multi-drug resistant lineages, some 
with carbapenemases, have persisted in a few UK hospitals for up to 9 years, causing 
multiple infections widely scattered over time and possibly reflecting colonisation of the 
hospital water systems. The most frequently encountered carbapenemase is VIM, which 
may be plasmid-mediated, with multiple gene copies conferring high level meropenem 
resistance \(^{76}\) but is usually integron associated. IMP-9, another MBL is as common as 
VIM in China \(^{77}\), and has been shown to be derived (as probably are many 
carbapenemase genes) from environmental bacteria by horizontal gene transfer \(^{78}\).

Multi-drug resistance is also a major problem in *P. aeruginosa* from cystic fibrosis (CF), 
with resistance increasing over time in the individual patient’s lung microflora. Multi-
drug resistance profiles are extremely variable even within widely successful CF 
lineages, e.g. the Liverpool Epidemic Strain, which has circulated in multiple CF patients 
and units. Rates of carbapenem-resistance in *P. aeruginosa* vary greatly across Europe, 
with high rates in Eastern Europe – Lithuania, Poland, Slovakia, Hungary, Croatia, 
Romania, Bulgaria and Greece all having rates of resistance >25% and sometimes 
>50%) \(^{40}\). More generally, these rates of resistance show a gradient, rising from NW to
SE Europe, with extensive spread of carbapenemase-producing clones in Belarus, Kazakhstan and Russia, which are outside the EU surveillance area. In contrast to Enterobacteriaceae rates of resistance to carbapenem are generally higher than those to ceftazidime, piperacillin/tazobactam or aminoglycosides.

6.5 Prevalence of antibiotic resistance in Gram-negative bacilli in the UK and relevant antibiotic prescribing

There are no epidemiological reports in the UK that specifically study defined MDR GNB. In this section, we discuss information on resistance to individual antibiotics and, where available, their associated resistances. Analysis is complex. Different reports from English, Welsh, Northern Irish and Scottish devolved administrations need drawing together to give a UK summary: bacteria and antibiotic resistances do not respect national boundaries.

Reduced prescribing may be followed by reduced resistance (See 11.1) but this is not invariable at a national level. Such reduced resistance has not occurred as older antibiotics (e.g. sulphonamides and streptomycin) have been abandoned, perhaps because of resistance linkage and for reasons already discussed in (See 6.3.) Reduced prescribing may reduce the likelihood of new resistance becoming prevalent but this is only a hypothesis set within the modern issues of travel and migration, which may import and spread resistance. Overall antibiotic consumption in England has fallen by 4.5% between 2012 and 2015 to 21.8 DDD/1000 population/day. It has yet to decline in general practice to the levels seen in 2010. After 5 years of increases in prescribing, hospital antibiotic use declined by 5% in 2014 from 5190 to 4933 DDD/1000 admissions and is now at approximately 2010 and 2011 levels. This decrease is concentrated in teaching hospitals which may reflect their case-mix or different pressures in other hospitals.
In Scotland antibiotic use in primary care fell for the third consecutive year in 2015 (by 2.4%) and is now 9.5% lower than the peak rate of use in 2012. The level of prescribing was related to population deprivation scores and to residence in nursing homes where antibiotic use among those aged over 65 years was 83% than for similarly-aged patients not resident in nursing homes\textsuperscript{81}. Since 2012, antibiotic use in Scottish nursing homes has fallen by 7.8% compared with 5.1% in all patients aged >65 years. Nevertheless, hospital use rose by 3.5% and is now 9.9% higher than it was in 2012. The rate of 5880 DDDs/1000 admissions is now 19% higher than in England\textsuperscript{81}. Of course, this may reflect use of less selective combination regimens such as penicillin, metronidazole and gentamicin rather than the number of days a patient receives antibiotics which is a weakness both of using Defined Daily Doses and the number of admission to estimate the number of people exposed to an individual antibiotic. Although England has the lowest antibiotic consumption in the UK, Scottish hospitals show significantly less consumption of carbapenems and piperacillin/tazobactam.

Information on primary and secondary care prescribing for Wales for 2015\textsuperscript{82,83} is only available at the level of health board and hospital respectively, and has not been reported as aggregate totals.

An overview of current antibiotic-resistance in Gram-negative serious infections in the UK can be secured in various ways. The BSAC Bacteraemia Surveillance Programme (http://www.bsacsurv.org) provides historical and current information with a marked time lag for centrally-tested isolates from a restricted sample of 24-40 hospitals and can be examined on a national or regional basis by species. It has an archive of organisms that can be studied in retrospect, which is an important strength. Other surveillance depends on collection of local data rather than isolates. In England reporting is mandatory for all cases of \textit{E. coli} bacteraemia with an improvement in case ascertainment. However mandatory data are needed for \textit{Klebsiella}, other...
Enterobacteriaceae and Proteae, *Acinetobacter* spp. and *P. aeruginosa* if early national interventions in emerging problems are to be reliably detected. Mandatory reporting of MRSA bacteraemia in England was established in 2001 and has improved with more comprehensive data capture from 2005 onwards. Health Protection Scotland now has mandatory reporting of *E. coli* bacteraemia but other species of Gram negative bacilli are only reported across the UK on a voluntary basis. Such voluntary laboratory reporting of all bacteraemias has been in place since the Devonport incident of contaminated intravenous infusions in 1972 and is believed now to capture data for 82% of all bacteraemias. This data includes antibiotic susceptibility data which has not been present in mandatory data. The collection of voluntary and mandatory data suggests that voluntary reporting should be replaced by mandatory reporting as soon as possible to reduce the laboratory workload. Most laboratories in England and Wales examining human samples now download bacteria identified and their antibiotic susceptibilities irrespective of anatomical site to regional and national repositories where trends but not additional information e.g. demographic details of patients’ residence etc. can be analysed.

Bacteraemia due to *E. coli* has increased over the last ten years in England and Wales, and analysis of the data-set showed that receipt of antibiotics in the 4 weeks preceding bacteraemia was the most important risk factor, followed by age over 65 years, and occurrence during summer months. A study by the *E. coli* subgroup of the UKs DH Advisory Committee on Antimicrobial Prescribing, Resistance and Healthcare Associated Infection on the first 891 cases of *E coli* bacteraemia with enhanced surveillance data are available in Committee papers for 28 March 2014 on line. This showed that urinary catherisation was a factor in only 10% of cases but that in 72% of episodes from a urogenital source involved individuals aged >=65 years. A urogenital infection had been treated in 310/891 (34.8%) cases in the 4 weeks preceding
bacteraemia and this sub-population differed very significantly in its antibiotic
resistances. Resistance in this subpopulation to ciprofloxacin was 80% vs. 17% overall,
76.9% vs. 39% to trimethoprim, and 49.3% vs. 45% to co-amoxiclav. The 3rd generation
cephalosporin resistance rate in the population overall was 10% but no figure was
provided for the resistance rate in this sub-population treated. Although the rates for
ciprofloxacin seem surprising, the figures show a marked selection for multiply
resistant, if not necessarily MDR, strains because of either failed treatment that did not
cover the multi-resistant organisms or selection of resistant organisms in the gut flora
that subsequently caused a urinary infection which then progressed to bacteraemia.
Approximately half of the bacteraemias appeared to be associated only with a lower UTI
but this probably represents symptomatically silent upper UTI giving rise to
bacteraemia, either initially, or through spread to the upper tract despite treatment. The
implication of this important study is that failure to give effective antibiotics may be the
reason for 70% of E. coli bacteraemias whilst 30% of cases are associated with
antibiotic resistance and, possibly, directly with treatment failure. The former requires
detailed study which is beyond the scope of this guideline. The consistent use of an
active antibiotic regimen for those either aged over 65 years or with signs and
symptoms of an upper UTI, would make a sizeable contribution to the target of a 50%
reduction in the rate of in E. coli bacteraemias by 2020 that was announced as a target
by the then UK Prime Minister at the Japan 2016 G7 meeting\textsuperscript{86}. This enhanced
surveillance study has now been analysed and published\textsuperscript{87}. Most patients (69.6%) were
aged over 65 years. Most patients (68.3%) had a positive blood culture taken within 24
hours of admission but 46.7% of these had a healthcare exposure within the previous
month and 546 out of these 930 (58.7% of this subgroup, 31.5% overall) had received
antibiotics in the preceding month, In 281 there was a clear urinary focus for the
bacteraemia for which 145 had received antibiotics (most commonly trimethoprim or
co-amoxiclav). The largest independent risk factor for a bacteraemia’s focus being the urogenital tract was previous treatment for UTI within 4 weeks of the bacteraemia’s onset (adjusted Odds Ratio: 10.7 & (95% CI 3.6-8.1) but details of antibiotic resistance in this subpopulation for the whole study was not given. Twenty one per cent of patients had either a urinary catheter in situ or had one inserted, removed or manipulated in the previous 7 days. Since the 2014 initial report, Public Health England has changed its recommendation for first line treatment of UTI in all but those under 50 years from trimethoprim to nitrofurantoin which is a urinary antiseptic that is only effective for treating lower UTI although it can be effective for preventing pyelonephritis associated with bacteriuria of pregnancy. It is too early to tell whether this will be effective in reducing bacteraemia or whether an oral combination regimen that attains systemically active concentrations will be necessary to achieve the desired outcome. APRHAI (The UK Advisory Committee on Antimicrobial Prescribing, Resistance, and Healthcare Associated Infection) on 28th March 2014 opined that in suspected pyelonephritis or upper UTI, the patient should be admitted if a) ciprofloxacin, piperacillin/tazobactam or co-amoxiclav had been used in the previous 2 months and b) the patient’s symptoms worsened or did not improve in the 12-48 hours after prescription. In UK strains of E. coli ST131 from various sources collected in 2011-2, when O16 and non-typeable strains are excluded, there is evidence that trimethoprim resistance occurs in at least 69% of CTX-M positive strains which comprised 32% of recent UK strains studied but 39%, at most, of CTX-M-negative strains. All CTX-M producers were ciprofloxacin resistant and 71% of non-CTX-M producers were quinolone resistant. Quinolones are not therefore useful if ST131 strains are prevalent even if these strains are not ESBLs. A study reported that sequence typed E. coli isolates from the BSAC Bacteraemia Surveillance Programme showed that the significant change in E. coli bacteraemia was almost exclusively due to an increase in clonal complexes 12, 69, 73, 95 and 131. This
reflects the sequence types in these clonal complexes. The clonal complexes, which each
may contain more than one sequence type, belong to phylogroups B2 and D that have
the virulence factors associated with extraintestinal spread. Phylogroup A and B1
strains, which may be more antibiotic resistant are usually confined to the gut and lack
these virulence factors. Clonal Complex 131 unlike the other clonal complexes includes
multi-resistant isolates (of ST131) hosting CTX-M ESBLS with almost invariably now,
resistance to quinolones. In a 2010-2012 Yorkshire study of bacteraemias 129/768,
39/129 ESBL producers, were ST131 confirming the importance of ST131 strains even
in the absence of production of ESBLs. 142/768 were ST73 (3/142 ESBL producers), 81
were ST69 (1 an ESBL producer), 73 were ST95 (1 an ESBL producer), 31 were ST12
(no ESBL producer, quinolone-resistant), 27 ST127 (no ESBL producers or quinolone-
resistant strains). Phylogroup D-ST69 strains (which include the previously
designated clonal group A) were not fluoroquinolone-resistant in a recent Italian
study although they were commonly detected in Italy in a previous cystitis study.
ST69 is usually ampicillin, trimethoprim and suphamethoxazole resistant. Quinolone-
resistant D-ST69 strains were also uncommon in a Spanish survey with isolates from
2009 accounting for 3% of quinolone-resistant strains respectively, compared with 26%
for O25:H4-B2 ST131 strains. We did not consider it feasible to introduce control
measures for ST131 when preparing our earlier guidance on infection control and
indeed cephalosporin resistance has spread into many other STs.

More recent data from 2012 to 2014 on antibiotic resistance in *E. coli* bacteraemia in
England were collected on 82% (54,301/66,512) of cases recorded by mandatory
surveillance by record-linking with the national records of all bacterial isolates. 74%
were classified as community onset whereas 16% of cases occurred 7 or more days
after hospital admission. Antibiotic resistances reported were 8439 (18.4%) to
ciprofloxacin, 4256 (10.4%) to third generation cephalosporin, 4694 (10.2%) to
piperacillin/tazobactam, 4770 (9.7%) to gentamicin and 91 (0.2%) to carbapenems\textsuperscript{94}.

Non-susceptibility to quinolones and cephalosporins decreased by 10% and 11% respectively over the two years in hospital onset cases whereas third-generation cephalosporin resistance increased by 10% in community onset cases. Trends in hospital or community onset changes in antibiotic susceptibility in other species such as \textit{Klebsiella} are precluded by lack of mandatory surveillance of bacteraemia.

A 12 year single centre-study in England suggested that the increase in \textit{E. coli} bacteraemias was essentially confined to ciprofloxacin, co-amoxiclav, cefotaxime and aminoglycoside resistance and accompanied a similar change in urinary isolates\textsuperscript{95}. The major rise in cephalosporin and multi-drug resistant \textit{E. coli} in the UK occurred between 2000 and 2007 largely reflecting the spread of IncF (pEK499 or similar) plasmids, and was associated initially with the internationally-successful \textit{E. coli} ST131 lineage with chromosomal fluoroquinolone resistance. These \textit{IncF} plasmids encoding the CTX-M-15 \(\beta\)-lactamase, along with resistances to trimethoprim, sulphonamides, tetracyclines and aminoglycosides (often associated with \textit{aac(6\prime)}-Ib-cr also augmenting ciprofloxacin resistance) also spread in other \textit{E. coli} Sequence types and other Enterobacteriaceae notably \textit{K. pneumoniae}. Since approximately 2007 (the date varies with the species and resistance) the rise of cephalosporin- and fluoroquinolone-resistant Enterobacteriaceae has slowed and fluctuated (\textit{E. coli}) or reversed (\textit{Klebsiella spp.} and \textit{Enterobacter spp.}) in the UK, though not in continental Europe \textsuperscript{96}. This shift in percentage resistance may reflect the reduction in prescribing of cephalosporins and quinolones in the UK, predicated not only by the Enterobacteriaceae problem but also by concern about \textit{Clostridium difficile}. It is important to know if this reflects an absolute decrease in numbers. Some data suggests that increased quinolone use largely mirrored the selection of such strains \textsuperscript{97}. An increase in quinolone resistance in bacteraemias preceded the arrival of ESBL-producing strains. Cephalosporin use in England is now
Cephalosporin usage fell by a further 9.2% between 2012 and 2015 following larger previous declines from a peak in 2006-7 because of the national \textit{C. difficile} problems. From 2012-5, oral cephalaxin use fell by 25.7% but parenteral cefotaxime use by only 1.6%, whilst parenteral ceftriaxone use increased by 37.4% probably reflecting use of this once daily antibiotic in outpatient parenteral antibiotic therapy \cite{4}. The microbiological need for preferring this broad-spectrum agent to teicoplanin or daptomycin, which are only active against Gram-positive bacteria, should be critically reassessed.

General practice quinolone use in terms of DDDs/1000 inhabitants/day has fallen consistently since 2012 reducing by 3.6% between 2014 and 2015. However the national overall usage of ciprofloxacin has declined only slightly from approximately 0.48 DDDs/1000 inhabitants/day in 2012 to 0.43 in 2015: quinolone use in hospitals has increased despite an 18.4% incidence of ciprofloxacin resistance in \textit{E. coli} bacteremia\cite{94}. A 53.6% rise in the respiratory quinolone levofloxacin which is the L isomer of ofloxacin seems unjustifiable but reflects a recommendation for use in penicillin-allergic patients with pneumonia. A similar increase (50.3%) was seen in Scotland accompanied by a 17% increase in ofloxacin use. An English target of a 10% reduction on 2013-4 levels of cephalosporin, quinolone, and co-amoxiclav use in primary care or a reduction in use to be below the 2013-4 median value (11.3%) of Clinical Commissioning Groups (CCGs) for antibiotic prescribing of these agents, was achieved in 189/209 CCGs \cite{4}. Prescribing of these antibiotics is substantially lower in Scotland and is not the subject of targets. Scottish reductions in primary care use in 2015 were 4.9% for co-amoxiclav, 5.8% for fluoroquinolones, and 6.0% for cephalosporins, with an 8% overall reduction in use\cite{81}.

Despite these reductions, cephalosporin and quinolone resistances continues to be seen frequently in UK bloodstream and urinary \textit{E. coli} and \textit{K. pneumoniae} isolates, with
significant circulation in older patients who move between hospitals, nursing homes, and the community and who have frequent exposure to cross-infection and antibiotics. Resistance to both quinolones and third generation cephalosporins in *E. coli* bacteraemias is concentrated in those aged over 65 years and over and in England is at least twice as prevalent in those aged over 74 years compared with those aged 65 to 74 years. An Italian scoring system for carriage of ESBL-producing organisms has not been tested in the UK or modeled to see if the group of patients at risk of carrying these strains on admission to hospital is increasing. The total number of *E. coli* bacteraemias in England and therefore the absolute burden of resistance, continues to rise – by 4.6% from 35659 to 37310 between 2014 and 2015 in England. The same publication notes an increase in *Klebsiella* bacteraemias by 9% over the same period. Over the period from 2000 to 2014 the incidence of *E. coli* bacteraemia in England has risen inexorably from 20 to 50 cases/100,000 population. In England, rates of resistance to piperacillin/tazobactam are said to have increased in *E. coli* bacteraemias from 8.5% to 11.7% and in *Klebsiella* ssp. bacteraemias from 12.6% to 18.5% over the period from 2011 to 2015. Equivalent rises in resistance to co-amoxiclav from 31% to 42% in *E. coli* bacteraemias and 18.7% to 28.2% in *Klebsiella* spp. bacteraemias over the same period have occurred. Record linkage for *E. coli* bacteraemias between 2012 and 2014 showed piperacillin/tazobactam resistance increasing by 15.1% for hospital onset cases compared with 8.7% for community-onset cases. This study also revealed significant variations in resistance rates by age and sex. Similar trends were seen in Scotland with an 8.6% increase for piperacillin tazobactam resistance and 6.1% for co-amoxiclav resistance in *E. coli* bloodstream isolates and 14.8% and 28.7% respectively in *Klebsiella* sp. in 2015. Changes from CLSI to EUCAST criteria may have produced these large rises.
in resistance in Scotland (See 6.2.) but there were no changes in EUCAST criteria for
these antibiotics between 2013 and 2015\(^{81}\) and in England few laboratories use CLSI
criteria In Wales 11/18 hospitals in 2015 recorded an increase in
piperacillin/tazobactam resistance in \textit{E. coli} in 2015\(^{100}\). In England
piperacillin/tazobactam use rose linearly by 62% between 2010 and 2015 to 135
DDD/1000 admissions across all hospital types\(^4\). In Scotland, use fell by 7.9% in 2015\(^{81}\).

These changes are important. The main antibiotics used in a recent prospective study in
10 English hospitals of treatment of Gram negative bacteraemia were co-amoxiclav in
32% of patients and piperacillin/tazobactam in 34%\(^{101}\). Despite empirical therapy
being inactive against responsible organisms based on \textit{in vitro} tests in 34% of cases, all-
cause mortality was said to be low, 8% assessed at 7 days and 15% at 30 days. Given the
increasing resistance rates and use, explorations of comparative outcome in relation to
resistance and use are needed at each national level and also by source of infection (See
11.2). Mortality in \textit{E. coli} bacteraemia throughout England was measured between July
2011 and June 2012 as 18.2% at 30 days or 10.34/100,000 population in 1 year. These
data were derived by record linkage of \textit{E coli} bacteraemia cases mandatorily reported to
Public Health England; voluntary reporting of antibiotic susceptibilities on all isolates to
Public Health England, and records at the Office for National Statistics Death
Registrations and at the NHS Spine.\(^{102}\) Mortality is high as compared with Finland (8%),
and inpatient only mortality in Canada (11%), and New Zealand (9%). Analysis showed
important associated features: 30% of deaths occurred on, or on the day after, the blood
sample was taken and 76.3% within 14 days making the separate mortality analysis of
community-onset and hospital-onset bacteraemia important. Overall 19,174/26216
(73.1%) patients had their bacteraemia recorded within 1 day of admission. Mortality
was higher (34.0%) if a respiratory focus of infection was diagnosed or the focus of
infection was unknown (25.9%) than if a urogenital focus was diagnosed (13.2%). No
information was available on the antibiotics prescribed precluding any test of whether higher mortality was correlated with failure to provide adequate Gram-negative cover in suspected respiratory or unknown foci of infection; moreover, there was no audit data to show if the reported foci of infection was supported by evidence. A recent audit of coding and diagnosis of pneumonia by the British Thoracic Society did not support the diagnosis in 15.8% of cases and noted a 14.3% rate of mortality in this group. At a population level the high burden of urogenital-related infection for *E. coli* was such as to make this the largest cause of deaths, even though mortality in this group was lower. The lower rate of mortality with urogenital infection correlates with information in an earlier study which showed that the excess mortality for bacteraemia with ESBL-producing Enterobacteriaceae was confined to non-urinary infections. The study by Abernethy and colleagues identified a urogenital source for 55.3% of community-onset cases of bacteraemia and 45.1% of healthcare-onset cases. In 17.3% of cases the source was unknown. Mortality was lowest in those aged 1 to 44 years (5.4%) versus those aged 45-84 (17.9%) and >85 years (25.2%). Mortality rates varied by the susceptibility of the isolated causative bacterium; ciprofloxacin S 17.0% (95%CI 16.4%-17.5%), ciprofloxacin I or R 21.9% (95%CI 20.5%-23.2%); cephalosporin S 17.5% (95%CI 16.9%-18.1%), cephalosporin I or R 21.3% (95%CI 19.4%-23.2%). The inclusion of a factor in the adjusted model to allow for hospital and case mix related mortality eliminated any significance to the difference in mortality by cephalosporin susceptibility. Cephalosporins are unlikely to have been used in infections due to ESBL-producing organisms in England, but piperacillin/tazobactam may have been used and the absence of a difference in mortality may reflect some improved outcome in urinary infection, despite the presence of bacteraemia. Different cephalosporins are not equally associated with *C. difficile*. Oral first generation cephalosporins would be useful in early treatment. It might be appropriate, whilst keeping *C. difficile* under review, to
abandon downward pressure on the whole class of antibiotics and introduce a cephalosporin-specific approach. There were no data on mortality in relation to susceptibility to piperacillin/tazobactam, co-amoxiclav, or aminoglycosides: carbenem-resistance rates were too low for robust assessment.

Resistance to any one of quinolones, cephalosporins or carbapenems was associated with a 30% increase in mortality. The association of increased mortality in quinolone-resistant strains needs explanation and it is not clear if this relates to hospital case-mix. Furthermore, if reduced use of oral quinolones is attempted, care is needed in the controversial area of prophylaxis in neutropenia where quinolones are widely used. Studies of withdrawing quinolones for this indication show an increase in Gram negative bacteraemia with susceptible strains without any diminution at least initially in resistant strains and recent Cochrane reviews support the efficacy of quinolone prophylaxis.

Rates of carbapenemase-production by Enterobacteriaceae (<2%) remain low in the UK but reference laboratory submissions of these organisms are growing annually (Figure 2), with many of the isolates coming from clinical rather than screening samples. It is noteworthy that surveillance of carbapenem-resistant strains depends on voluntary submission to reference laboratories and that regional molecular testing necessary for rapid turnaround has not been converted into national surveillance. Given the importance of reducing carbapenem resistance, consideration should be given to introducing mandatory reporting of all isolates of carbapenem-resistant Enterobacteriaceae so the evolving picture can be properly assessed. English data suggests the proportion of carbapenem-resistant *Klebsiella sp.* rose from 0.2% to 1.1% between 2011 and 2015. There are pockets of local endemicity, especially of *K. pneumoniae* and other Enterobacteriaceae with KPC enzymes around Manchester or with VIM and OXA-48 in north Cheshire. These have persisted for 5-6 years (D.M.)
Livermore, unpublished data). Many other sites, notably London teaching hospitals, are currently being repeatedly challenged with a diversity of carbapenemase producers, many imported from overseas. Clonal complex 258 *K. pneumoniae* with KPC carbapenemase remains rare in the UK, despite repeated introduction, and the greater issue, particularly in NW England is dissemination of plasmids encoding KPC carbapenemases among different *K. pneumoniae* and Enterobacteriaceae. Carbapenem-resistant isolates submitted to reference laboratories in Scotland increased from 47 in 2014 to 63 in 2015. The dual loss of both quinolone and cephalosporin susceptibility has driven increased usage of carbapenems particularly meropenem from some 75 DDD/1000 admissions in 2010 to 104 DDD/1000 admissions in 2015 in England, a 38.6% increase, but in 2015 the increase was only 1%. In Scotland the picture is different, there was a 6.5% increase in use of carbapenems between 2014 and 2015 but this is now only 9.3% higher than in 2012.

Phenotypic information on aminoglycoside susceptibility is available. Frequent gentamicin-resistance was noted in ESBL-producing strains of *E. coli* from all sites in one region, representative of the UK, with resistance rates of 48.7% for *E. coli* ST131 and 55.1% for *E. coli* non-ST131. The record linkage data previously discussed shows that overall gentamicin-resistance rates (i.e. irrespective of ESBL production) varied by region between 5.5% and 15.4% in the years 2012 to 2014 and that the overall rate in community-onset cases was 8.6%. The region with lowest rate of resistance had a 34% higher incidence of *E. coli* bacteraemias than that with the highest rates, which suggests the possibility of dilution of the denominator by an increase in more susceptible bacteraemias (e.g. ST73 in northern England). In Wales in 2015 only 5/18 hospitals reported gentamicin resistance rates less than 8.6% in *E. coli* bacteraemia and two had rates over 20%. Rates of 8.6% to 15% would seem too high for empirical use of gentamicin alone. However, the 8.6% rate of gentamicin resistance in community
onset bacteraemia is very similar to the 8.7% resistance rate to piperacillin/tazobactam which is widely used alone\textsuperscript{94}. National data on amikacin are hard to interpret because fewer laboratories test it as well as gentamicin and the amount of testing that is second line because of resistance on first line testing remains unresolved, potentially skewing the data. Nevertheless, as expected, amikacin resistance is rarer than gentamicin resistance (2% in 2015) in England\textsuperscript{4}.

Rates of co-resistance in bacteraemia isolates for 2015 for gentamicin and third generation cephalosporins were 4.6% for \textit{E. coli} and 5.9% for \textit{Klebsiella sp.} compared with resistance rates to third-generation cephalosporins alone of 7.5% and 5.2% suggesting some useful activity for gentamicin against ESBL-producing \textit{E. coli} but less against ESBL-producing \textit{Klebsiella sp.}. Rates of co-resistance in bacteraemia isolates for 2015 to gentamicin with co-amoxiclav are 7.8% in both \textit{E. coli} and \textit{Klebsiella sp.} compared with resistance rates to co-amoxiclav alone of 35.2% and 19.3%\textsuperscript{4}. This confirms the potential utility of an aminoglycoside compared with co-amoxiclav alone for both \textit{E. coli} and \textit{Klebsiella spp.} bacteraemias. The same data source indicates a somewhat different situation with ciprofloxacin-gentamicin combinations. For \textit{E. coli} and \textit{Klebsiella spp.} rates of co-resistance were respectively 6.8% and 5.8% whereas resistance to ciprofloxacin alone occurred in 11.8% and 5.0% suggesting that addition of an aminoglycoside was seldom advantageous in \textit{Klebsiella} infection. Overall this co-resistance data\textsuperscript{4} suggests only a modest improvement on gentamicin monotherapy and the benefit compared with the harm of continuing selection of resistance by the non-aminoglycoside may not be great.

Consumption of aminoglycosides is now low in England in hospital inpatients (approximately 0.08 DDD/1000 population/day) and fell in 2015. By contrast use rose in Scotland by 5.9% becoming 16.9% more frequent than in 2012. Falls in use are likely to reflect concern about resistance in ESBL-producers and about potential toxicity; they...
may also reflect a change in clinical contacts with microbiologists as antibiotic assays are increasingly undertaken by clinical chemistry departments. A comparison with Scotland to understand the differences would be informative.

Bacteraemia represents a group of community infections selected for virulence factors sometimes but not always by antibiotics. Antibiotic resistance in Gram-negative infections in the community was thought, even a decade ago, to be quite uncommon in the UK. A historical European study of acute, community-acquired, uncomplicated, non-recurrent UTI in 2008 caused by *E. coli* involved 12 GP practices in the UK and enrolled 200 unselected women aged 18-65 years. Resistance was rare to mecillinam (1%), nitrofurantoin (0%), fosfomycin (0.5%) amoxicillin/clavulanic acid (2.0%) and ciprofloxacin (0.5%), but commoner to amoxicillin (32%), sulfamethoxazole (26%), trimethoprim (15%) and trimethoprim/sulfamethoxazole (14%) \(^{111}\). In this survey the co-amoxiclav resistance rate seems low in relation to the amoxicillin resistance rate. Reported resistance rates to co-amoxiclav in lower urinary infections have increased since the time of this study partly because of the substitution of EUCAST’s (32+2mg/L) breakpoint for the previous BSAC (16+8mg/L) value. A contemporaneous UK study with a large community sample reported 12.0% resistance to co-amoxiclav versus 54% for ampicillin \(^{112}\). Welsh data in 2014 reports the following resistance rates in “coliforms” from urine in different communities:: co-amoxiclav 12.9% (Range:5.1% to 25.4%), third-generation cephalosporin (ESBL) 6.8% (Range 3.3% to 17.9%), nitrofurantoin 10.0% (range 8.7% to 22.4%), trimethoprim 36.7% (Range:30.3 to 41.8%) and fluoroquinolone 10% (range 7.6% to 16.4%) \(^{113}\). A 2010-3 large UK study \(^{114}\) of all community urinary isolates from a UK region with a population of 5.6 million found that by 2013 resistance to third generation cephalosporins in *E. coli* had risen to 5.5% and ciprofloxacin resistance to 15.5%; for *Klebsiella spp.* the cephalosporin resistance rate was higher at 10.1%. Only 0.06% of the *E. coli* isolates were reported as resistant to one
or more carbapenems as were 0.32% of the *Klebsiella spp.* isolates. In this regional
survey, VIM enzymes were found in *Pseudomonas spp.* whereas among *E. coli* and
*Klebsiella spp.*, 16 had NDM genes, 5 KPC and 2 OXA-48. These findings support the view
that carbapenemases are rare in the community in the UK. A further study of isolates in
the same English region over the period 2007-2014 showed, after deduplication 69 with
*bla*$_{NDM}$, 26 with *bla*KPC, 16 with *bla*$_{OXA-48}$-like, and 7 with *bla*$_{VIM}$.

A historical audit of urine samples taken at presentation from primary and secondary
care in South London before the widest dissemination of ESBL positive *E. coli* ST131
occurred, found that 22.6% of isolates were resistant to trimethoprim, 43.3% to
amoxicillin, and 10.3% nitrofurantoin. Since this audit resistance to trimethoprim
has slowly risen across the UK, and in Wales is significantly commoner in isolates from
patients over 65 years. Trimethoprim resistance rates vary widely by CCG in England. In
2011 it ranged in these from 16.3% to 66.7% but by 2015 86% showed >25%
resistance with an almost uniform median of 29% in CCGs. The reason for these
variations in a minority of CCGs remains uncertain. In Wales resistance rates of 38.2%
overall are currently reported. A caveat is that high resistance rates may reflect
selective testing of previously treated patients in the community and different local
policies for submitting samples, and the true rate of resistance to trimethoprim in
patients presenting in the community with uncomplicated UTI may be lower than
current figures suggest. Trimethoprim use in England fell by 14.5% between 2014
and 2015 reversing the increase seen between 2012 and 2014. This fall should be many
times larger in 2016 if there is expeditious compliance with the Public Health England
recommendation in 2014 to substitute nitrofurantoin for trimethoprim as the first line
antimicrobial for cystitis in the older patient. A Swedish trimethoprim-sparing switch in
one region resulted in an 86% decline in trimethoprim use between 2004 and 2006.

In 2015 in England rates of trimethoprim prescribing were approximately
1.1 DDDs/1000 population/day compared with 0.8 DDDs/1000 population for nitrofurantoin\(^4\).

UK data on resistance to nitrofurantoin, fosfomycin and mecillinam is scanty. In a single centre study nitrofurantoin resistance was commoner in *Klebsiella* spp. of community origin (around 15%) than *E. coli* (3%) \(^{119}\). English national data for the 2\(^{\text{nd}}\) quarter of 2016 suggests resistance in *E. coli* in community UTIs varied with CCG between 0.3% and 12.8% with a median of 3.8% \(^4\) whilst in Scotland, 5.9% of isolates tested in 2015 showed nitrofurantoin resistance\(^8\). Nitrofurantoin resistance is also common in UK CPE isolates\(^1\). Proteaeae are inherently resistant to nitrofurantoin and data on their prevalence in UTI and resistance linkage for nitrofurantoin resistance in England is needed given the recommendation to use this antimicrobial first-line (See 9.1 for previous experience of changes in prevalent phylogroups and STs of *E. coli*). There are no recent data on fosfomycin resistance in the UK. A survey of fosfomycin resistance in Leeds found *fosA* in 2 urinary tract isolates collected months after its UK introduction in 1994 despite a lack of use in the study hospital \(^{121}\). In the same publication, a study of foods in Leeds in 1995 identified 2 Enterobacteriaceae isolates carrying *fosA* in vegetables imported from Spain. Fosfomycin resistance (MIC\(\geq\)64 mg/L was present in 32/81 strains of CPE in 2011; 27 of these were *Klebsiella* spp. \(^{120}\). In Wales, only 6.2% of cefpodoxime-resistant *E. coli* (i.e. probably ESBL- and AmpC-producing strains) were apparently resistant to mecillinam \(^{122}\) but this is discussed further later in the article (See 9.4.).

The impact of the successful clone ST131 clone of *E. coli* on multiple resistances has been assessed. In one 2011 UK study, resistance rates in ESBL-producing *E. coli* ST131 (mostly with CTX-M-15 enzyme) compared with non ST131 (producing CTX-M-15 or CTX-M-14) were respectively 99% versus 83% respectively for ciprofloxacin, and 92% vs. 86% for trimethoprim \(^9\). Fluoroquinolone resistance alleles *gyrA/B* and *parC* are
characteristic on whole genome sequencing of the Clade C of *E. coli* ST131, which is
almost exclusively the clade carrying CTX-M ESBLs \(^{29}\).

There is no reliable information on acquired colistin resistance. Usage sharply increased
by 30% between 2013 and 2015 in England, entirely in specialist and teaching
hospitals\(^4\). Given i) the growing use of colistin as a drug of last resort, ii) the prevalence
of colistin resistance in KPC-producing *Klebsiella pneumoniae*, especially in Italy, but
also in the USA. iii) the lack of mandatory surveillance of *Klebsiella sp.* and iv) the
recognition of plasmid-mediated colistin resistance due to *mcr1* and *mcr2*, there is an
urgent need for enhanced surveillance of colistin resistance at a national level \(^4\). *Mcr-1*
has been isolated from British pigs \(^{123}\) but is widespread in the European food chain
including additionally turkeys and veal calves \(^{124}\) and *mcr-2* has been found in pork and
cattle products \(^{125}\).

### 6.6 What impact have returning travelers made on UK epidemiology?

Whilst mutational resistances often emerge locally, strains with acquired resistance
genes are often clearly imported to the UK from other countries. Examples include
multi-drug resistant *K. pneumoniae* with OXA-48 carbapenemases with Libyan conflict
casualties and with patient transfers from elsewhere in the Middle East; *K. pneumoniae*
with KPC carbapenemases from Greece, and Israel and, also most significantly,
Enterobacteriaceae with the NDM MBL, from south Asia and China \(^{126}\). Colonisation of
travellers may be frequent, although precise rates are largely unknown. A systematic
review confirms travel to certain areas is a significant risk factor \(^{127}\). Most data concerns
ESBL-producing strains and there is a notable dearth of information on other important
resistances including aminoglycosides, carbapenems, colistin, and fosfomycin.

Nevertheless an Australian study suggests that travel associated aminoglycoside- and
quinolone- resistance may be even commoner than travel associated cephalosporin.
Interestingly prolonged carriage was significantly associated with the pathogenic phylogroups B2 and D rather than A and B1 but strains of ST131 were rare even with Asian travel. A Canadian study showed that bacteraemia due to CTX-M-14 ESBL-producing *E. coli* was associated with travel to Europe and Africa whilst CTX_M-15-producing strains were associated with travel to Asia \(^{129}\). Analysis of risk factors in Norway for new cases of ESBL-producing infection was undertaken in a case-control study of adults who had been resident for 1 year or more, with no previous hospital or nursing home residence >24 hours in the previous 31 days. It identified as risk factors travel to Asia, the Middle East or Africa within the past 6 weeks (OR=21 95% CI 4.5-97) or 6 weeks to 24 months (OR=2.3 95% CI 1.1-4.4), recent use of fluoroquinolones (OR=16 95%CI 3.2-80) or recent use of β-lactams other than pivmecillinam (OR=5.0 95%CI 2.1-12), diabetes (OR=3.2 95%CI 1.0-11), and freshwater swimming in the last year (OR=2.1 95% CI 1.0-4.0) were associated with UTI due to ESBL-producing *E. coli* or *Klebsiella spp.*. Factors associated with decreased risk were the number of fish meals/week (OR=0.68/fish meal 95%CI 0.51-0.90) and increasing age (OR=0.89/5 year increase 95% CI 0.82-0.97). Almost 1 in 4 (23%) ESBL-positive patients had travelled to the risk countries within the previous 6 weeks and 39% in the 6 week to 24 month period compared with 1% and 19% respectively. Travel to Europe (11% and 67% in ESBL producers and 7% and 57% non ESBL producers), America or Oceania (including Japan) was not a risk factor \(^{130}\). This emphasises that there is a longer-term effect of travel or migration that is often not considered. A placebo-controlled trial of ciprofloxacin to prevent traveller’s diarrhoea showed that the prophylaxis selected for quinolone- and other-drug resistant GNB suggesting that such practices need review \(^{131}\). Previous travel to destinations where resistance is prevalent is a risk factor for acquired multi-drug resistant bacteria and should be considered in respect of empirical therapy. However many patients with multi-drug resistant organisms lack any relevant travel.
and it is not known if their organisms represent spread from carriers, especially in the same household, who have a history of high risk travel, or who have asymptotically acquired the organism in hospital.

The most significant impact that the movement of people can have on the problem of resistance in Gram-negative bacteria is the maintenance of higher levels of resistance in commensal bacteria after return from high incidence areas. Data on faecal carriage rates may mislead when compared with correlates of clinical infection since it will include phylogroup A and B1 strains of lower pathogenicity than the B2 and D strains seen commonly in urinary and bacteraemia. Tangden in Sweden showed that 7/8 previously uncolonised travellers to South Asia and 10/32 to East Asia returned with gut carriage of ESBL E. coli. One study in Birmingham showed that 22% of individuals with names of Middle Eastern or south Asian origin had faecal carriage of CTX-M ESBL-producing E. coli compared with 8.1% in those with names of European origin. A very recent large scale survey studying 2,430 healthy individuals in four areas in England found similar carriage rates of 25% and 5.6%, respectively. In a multivariable logistic regression model the percentage contribution made to risk of colonisation was apportioned. Being born in South Asia (India, Pakistan, Bangladesh) or coming from those countries was 26.6%, travel to those countries 12.1%. In contrast being born in UK of UK origin 9.9% and travel to all other parts of the world was 17.8% (McNulty et al. (2017) submitted for publication). Hence, the choice of antibiotics for empirical treatment may need to take into account recent travel history and cultural background.

The second ESPAUR report (2016) includes details from a research study of faecal carriage rates of ESBL-producing Enterobacteriaceae in England. This showed variations in carriage from 4.9% in Shropshire to 16% in Heart of Birmingham Primary Care Trust with intermediate rates in Southampton and Newham (East London). Risk
factors in this study, which is yet to be published in full, included birth in India, Pakistan, Bangladesh, Sri Lanka, Afghanistan (which collectively accounted for 24% of all carriage) or the Middle East (including Egypt, Iraq, Saudi Arabia and other countries in the Persian Gulf) and travel in the last year to Africa, South Asia (Indian sub-continent and Afghanistan), South East Asia (Thailand, Burma, Cambodia, Laos, Malaysia, Singapore or Pacific Asia (including Vietnam, Koreas, China), South or Central America,(WHO regions). Until control measures reduce prevalence and at present only, (given the rate of change) travel to, and most particularly healthcare in, the following countries are also risk factors for either ESBL carriage or carbapenemase acquisition or both: the Eastern Mediterranean (the Balkans, Greece, Cyprus, Turkey, and Syria) and Eastern Europe and Russia, Belarus and Kazakhstan, and Italy.

There is a need for further studies with controls (non-travellers from different households of the same ethnic background) on the carriage of antibiotic-resistant *E. coli*, with strain typing and phylogroup allocation to better predict the potential for extraintestinal infection. This is further reviewed in elsewhere. Studies are needed also of *Klebsiella sp.* and on the time elapsed since travel to specified locations of high prevalence. Information on healthcare and antibiotic exposure is required as well as details of many non-ESBL antibiotic resistance mechanisms.

**Evidence:**

There is a clear indication of association of infection with ESBL-producing *E. coli* and travel. There is no information on other antibiotic resistances in association with travel and minimal information on carriage duration after travel.

Evidence level: 3

**Recommendation:**
Need to quantify risks of infection with/ carriage of, extraintestinal pathogenic *E. coli* and of *Klebsiella sp.* resistant to all antibiotics and relate to time since travel to countries with high prevalence of MDR GNB and incorporate in risk assessments for clinical infection with MDR GNB in the community and on admission to hospital to guide therapy.

**Grading:** Strong recommendation for

### 6.7 What is the clinical importance of carbapenemase- versus CTX-M- and AmpC-producing strains

ESBL-producing Enterobacteriaceae, multi-drug-resistant *P. aeruginosa* and *A. baumannii* are associated with increased mortality, length of stay and expense in most but not all studies evaluating the impact of antibiotic resistance in Gram-negative bacteria. Nevertheless, variability in the setting (mainly ICU), study design, organisms included (most notably, which Enterobacteriaceae species), resistance profile, and site of infection make the studies difficult to compare.

Fluoroquinolone resistance in *P. aeruginosa* was associated with increased hospital costs, and, if associated with imipenem resistance (MDR strains), increased mortality. Four of eight studies in one review of MDR strains of *P. aeruginosa* showed increased mortality. With *A. baumannii*, carbapenem-resistance was generally associated with increased length of stay and expense of care; mortality was generally increased, most clearly if blood-stream infection was involved. However, two studies of MDR, but carbapenem-susceptible, *A. baumannii* did not identify a significant increase in mortality, whereas studies of carbapenem-resistance in *A. baumannii* consistently identify a significant increase in mortality only partly due to use of inactive carbapenems.
More recently, studies have emerged evaluating the impact of carbapenem resistance in Enterobacteriaceae. Pooled analysis of nine studies comparing mortality in Enterobacteriaceae infections including bacteraemia found that mortality was more than two fold higher when infections were caused by CPE. Broad-spectrum antibiotics other than carbapenems can select for colonization (detectable by active surveillance) that precedes later infection with bacteria resistant to a range of other antibiotics because of linkage of with multiple resistance factors. Carbapenem resistance in Acinetobacter spp. is similarly linked with multiple resistances that can be selected for by antibiotics that are not carbapenems, and can be detected as colonization prior to development of infection and this is likely to be the case with Enterobacteriaceae.

Carbapenem resistance is an increasing problem in Enterobacter spp. in the absence of carbapenemases. In E. aerogenes ertapenem resistance is associated with loss of Omp35, a porin, and meropenem resistance with loss of Omp36 together with derepressed overproduction of AmpC.

Bacteria producing CTX-M are of international importance. In the community they are usually MDR with few and hitherto little used antibiotics offering the sole effective treatment. The spread of these strains requires widespread changes in primary care prescribing practice which can be slow to take effect. Further, systemic infection with these strains usually requires parenteral drugs involving additional hospital admissions or outpatient parenteral antibiotics. Particular successful clones such as E. coli ST131 and ST69 are frequently involved. The fundamental reason for the success of these clones remains obscure and strategies to counter their spread nationally and internationally have so far been based on antibiotic restriction alone.

AmpC-producing strains of Enterobacteriaceae were a problem when third generation cephalosporins and monobactams were widely used because stable derepression of this
enzyme occurred by mutation at the regulatory gene *ampD* in *Enterobacter* spp., *Serratia* spp., *Citrobacter freundii* and *Morganella morganii*. Selection of such mutants during cephalosporin treatment of bacteraemia with these species can cause treatment failure. Amoxicillin/clavulanate, both components of which are strong inducers of AmpC in such species is not active against such species but piperacillin although inactive against derepressed mutants seems less prone than third generation cephalosporins to select such strains from the induced population. Genes encoding AmpC enzymes have also escaped to plasmids that have spread into *E. coli*; such plasmid carrying strains are widespread in food stuffs. The main enzyme is CMY-2. In the UK it remains considerably rarer than ESBLs. Cefepime is more stable than other third-generation cephalosporins to AmpC but in *E. cloacae* high-level cefepime resistance is associated with mutation in AmpC. Carbapenems and temocillin are active against AmpC-β-lactamase whether of chromosomal or plasmid origin but ertapenem is more labile and, if OmpK35 porin loss occurs, resistance arises from this enzymes action.

### 7 Intravenous treatment options for MDR GNB: What is the efficacy of carbapenems, temocillin, fosfomycin, colistin and other antibiotics against specific MDR GNB and what are the recommended antibiotics for secondary/tertiary care? The evidence base (and grading) for all agents is generally weak, as most studies were retrospective case series, only rarely including a comparator agent. Our suggestions for intravenous treatment are summarized in the algorithm in Figure 3. Each intravenous agent is individually further considered.

### 7.1 Carbapenems Carbapenems should be regarded as the drugs of choice for serious infections with ESBL-producing Enterobacteriaceae and they are the drugs of choice for the
empirical therapy of patients with serious sepsis caused by Gram-negative bacteria, depending on local resistance rates and clinical experience.

Meropenem was found to be narrowly superior to imipenem/cilastatin (cilastatin prevents degradation of imipenem by urinary and ileal dehydropeptidase) in both clinical and bacteriological outcomes in one meta-analysis of 27 RCTs. The clinical response rates (complete remission or improvement in signs and symptoms of sepsis) for meropenem and imipenem were 91.4% and 87.2%, whereas bacteriological response rates were 85.1% and 82.8% respectively. There was no significant difference in mortality in the nine trials reporting data (7.4% for meropenem, 9.7% for imipenem).

Meropenem and imipenem (sometimes referred to as ‘Group 2’ carbapenems, based upon activity against Gram-negative non-fermentative bacteria) are typically preferred to ertapenem for the empirical treatment of bacteraemia (often arising from the urinary tract) because of the broader spectrum (see below). A switch to ertapenem may be rational with susceptible isolates if it leads to earlier discharge with OPAT but without this, is not a mechanism for reducing selection for carbapenem resistance. In Singapore, de-escalation of meropenem-regimens by ID physicians (including in a small proportion to ertapenem) was associated with no increased in clinical failure rates or hospital mortality, reduced duration of carbapenem treatment from 8 to 6 days, less diarrhoea and C. difficile infection and less carbapenem-resistant Acinetobacter baumannii acquisition.

Meropenem or imipenem select respectively for carbapenem-resistant Gram-negative organisms including pre-existing carbapenem-resistant A. baumannii, and porin oprD mutants, the commonest mechanism of imipenem resistance, arising during imipenem treatment of P. aeruginosa. Overproduction of AmpC type enzymes, and efflux pumps which are common, are implicated, in meropenem resistance in P. aeruginosa: MBLs usually of a VIM type occur but are much less common. A multi-
centre Spanish study of isolates in 2008 from *P. aeruginosa* bacteraemia showed similar resistance rates to piperacillin/tazobactam, ceftazidime and meropenem. Meropenem resistance was more commonly associated with *mexB* or *mexY* and *AmpC* overexpression whereas resistance to piperacillin/tazobactam and ceftazidime was more commonly associated with *AmpC* overexpression alone, making non-carbapenems preferable agents for avoidance of MDR strains. Nevertheless, *AmpC* overexpression was associated with quinolone resistance, which with aminoglycoside resistance is already known to be associated with efflux pumps. Whilst both imipenem and meropenem have a similar spectrum of activity, use of imipenem has declined and meropenem is now the most widely prescribed carbapenem in the UK.

Widespread usage particularly internationally, has driven the emergence of resistance and careful and considered empirical usage is essential. If the bacteria responsible for the infection are subsequently shown to produce neither ESBLs nor *AmpC* β-lactamase, carbapenem use reasonably should be stepped down to narrower spectrum agents. An Italian cohort study across 5 hospitals showed that rectal carriage of KPC-producing *Klebsiella* was predictive of bacteraemia with such strains in the subsequent 2 years; sensitivity and specificity were 93% and 42% respectively; positive and negative predictive values were 29% and 93% respectively. Bacteraemia was associated with ICU admission, invasive abdominal procedures, cancer chemotherapy or radiation therapy and the number of colonization sites. This suggest that screening may play a role in anticipating a requirement for treatment other than carbapenems active against such strains but this will not necessarily apply to other bacteria with carbapenemases.

The ominous changes and increase in meropenem resistance in Enterobacteriaceae in the UK (Evidenced in 8.4), and the clinical importance of such resistance and the need to know the resistance mechanism to use appropriate chemotherapy, mean that an accurate overall view of the emerging picture is essential so appropriate action can be
taken. We include recommendations on this epidemiological matter because of its importance. We recommend the introduction of mandatory reporting of carbapenem-resistant Enterobacteriaceae from all anatomical sites and specimens. Such isolates should be tested contemporaneously to determine the responsible carbapenemase and meropenem MIC. Isolates should be submitted to reference laboratories to determine susceptibility to a wider range of appropriate agents and for those agents, such as colistin or ceftazidime-avibactam, for which susceptibility testing is technically demanding. The determination of susceptibilities is a part of essential surveillance. Appropriate patient treatment also depends on performing these susceptibilities in an expeditious manner but the methodology required may be beyond the scope of most routine diagnostic laboratories.

Ertapenem is licensed in Europe for the treatment of intra-abdominal and gynecological infections and community-acquired pneumonia. In the rest of the world, including in the USA, it is also licensed for skin and skin structure infections and for complicated urinary tract infections (for which it is widely used 'off-label' in the UK). Ertapenem shares the broad spectrum of imipenem and meropenem against Enterobacteriaceae, some Gram-positive species .and anaerobes, but is inactive against Acinetobacter spp. and P. aeruginosa 162. It is sometimes called a Group 1 carbapenem on this basis. Its main benefit is its once-daily mode of administration.

Use of ertapenem for the treatment of infections caused by Enterobacteriaceae is less well established than for imipenem or meropenem but it has good in vitro activity. A retrospective cohort study compared outcomes of bacteraemias due to ESBL-producing E. coli and K. pneumoniae treated with ertapenem and group 2 carbapenems. Outcomes were equivalent between patients (mortality rates of 6% and 18%, respectively; \( P=0.18 \)). However, more patients treated with group 2 carbapenems had severe sepsis / septic shock / multi-organ failure - 5/49 (10.2%) for ertapenem versus 36/109 (33.3%)
for other carbapenems (odds ratio of 0.23; 95% confidence intervals 0.08–0.62; p<0.002), suggesting clinicians were more likely to treat “sicker” patients with a group 2 carbapenem than ertapenem. A retrospective study in Taiwan evaluated 251 patients with bacteraemia caused by ESBL-producing *E. coli* and *K. pneumoniae* isolates treated with a carbapenem. Two hundred and thirty patients received carbapenems appropriately – 57 ertapenem, 136 imipenem and 37 meropenem: 21 received carbapenems inappropriately, 18 received ertapenem and 3 imipenem when the MICs were respectively >0.5mg /L and >1mg/L. Among the isolates, rates of susceptibility to ertapenem (MIC ≤0.5 mg/L EUCAST) were 83.8% in *E. coli*, and 76.4% in *Klebsiella spp.*, respectively and those to meropenem were 100% and 99.3%. Sepsis-related mortality varied if the lower breakpoint CLSI breakpoint, for susceptibility (≤0.25mg/L) was used. By this criterion, mortality was 5.3% (3/57) in those patients infected with an ertapenem-susceptible strain versus 33% (6/18) for an ertapenem non-susceptible isolate if they were treated with ertapenem. If categorisation was based on the EUCAST breakpoints MIC <=0.5mg/l or >0.5mg/l, there was no significant difference in mortality. Propensity matching of patients showed that patients with isolates that were ertapenem non-susceptible by CLSI criteria had a similar raised mortality if treated with imipenem or meropenem but numbers were small. A recently published multinational retrospective cohort study of 195 patients given empirical carbapenem and 509 given targeted therapy for bacteraemia with ESBL producing *Enterobacteriaceae* found ertapenem to be equivalent to other carbapenems. The authors recognized that as in other similar studies ertapenem was more frequently used in lower risk patients and that more studies are needed in the severely ill patient populations.

Resistance (MIC=>1mg/L) and high-level resistant (taken here as MIC>16mg/L) by EUCAST breakpoints to ertapenem in *Klebsiella spp.* and *Enterobacter spp.* were well recognised before CPE began to spread and were associated with combinations of a β-
lactamase (often a CTX-M ESBL in *Klebsiella* spp. or AmpC in *Enterobacter* spp.) plus impermeability due to omp K35 porin loss. Despite the results of Lee *et al.* (2012)\(^1\) imipenem and meropenem appear to remain active against most isolates with low-level ertapenem resistance caused by these mechanisms but with raised MICs compared with normal levels for the species. An *in vitro* study showed the frequent emergence of this type of resistance in ESBL-producing *E. coli* in a pharmacokinetic model\(^ 2\) but most resistant isolates are *Klebsiella* spp. or *Enterobacter* spp. not *E. coli*. In a survey of UK isolates in 2007 only one of 95 ertapenem-resistant isolates of *Klebsiella pneumoniae* produced a defined carbapenemase, namely IMP-1 with the remainder inferred to have impermeability (porin-loss) mediated resistance.\(^3\) However, this situation has changed radically as KPC, OXA-48 and NDM are enzymes now regularly encountered in the UK\(^ 4,5\). A retrospective case-control study from the Eastern USA found that risk factors for infection caused by ertapenem-resistant Enterobacteriaceae with such impermeability-mediated resistance included exposure to any antibiotic (not just β-lactams and carbapenems) during the 30 days before a positive culture result.\(^6\) A study from Singapore found that hospitalization and fluoroquinolone treatment were predictors for the appearance of ertapenem resistant imipenem susceptible variants\(^ 7\). The use of ertapenem has no detrimental effect in terms of selecting for *P. aeruginosa*\(^ 8\). Results from ten clinical studies showed that use of ertapenem did not result in decreased susceptibility to carbapenems in *Pseudomonas*. This was confirmed in study of hospitals in Queensland\(^ 9\). A further study found that one hospital’s use of ertapenem was balanced by less use of imipenem and ciprofloxacin, and this may have contributed to a reduced prevalence of resistance of *P. aeruginosa* to imipenem\(^ 10\). In contrast to these findings a study in Singapore associated increasing consumption of ertapenem with a rising incidence density of carbapenem-resistant *P. aeruginosa*\(^ 11\). Ertapenem use had no impact on the susceptibility of *A. baumannii* to imipenem\(^ 12\).
Prolonged infusion therapy with meropenem for MDR GNB including carbapenem resistant organisms has been advocated on pharmacokinetic grounds in children for A. baumannii, P. aeruginosa and Enterobacteriaceae with meropenem MICs up to 8mg/l. 177 There is a general trend towards considering continuous infusion of beta-lactams in critically ill patients with severe Gram-negative sepsis (See 7.18) 178. Continuous infusion meropenem has been assessed in 375 obese patients for its ability to produce steady state levels above the MIC at levels from 2mg/L to >16mg/L 179. Dosing nomograms to sustain this had previously been constructed in critical care patients 180.

Meropenem combined with vaborbactam (RPX7009); a boronic acid derived β-lactamase inhibitor is progressing through Phase 111 trials and may cover Enterobacteriaceae strains with KPC producing carbapenemases but not those with MBLs or OXA-48-like enzymes. Some isolates with ompK36 porin loss (See 6.3.3 & 6.7) are resistant 38. Relebactam in combination with imipenem/cilastatin is entering Phase 3 trials with trials against imipenem-resistant bacteria compared with a combination of colistin and imipenem/cilastatin and a comparative study against piperacillin/tazobactam in ventilator-associated pneumonia. Phase 2 studies are as yet unpublished. In vitro studies show no enhanced activity against Acinetobacter spp. but activity against KPC-producing K. pneumoniae (unless it has an OmpK 36 porin loss which is responsible for meropenem resistance (See 6.3.3 & 6.7), and many but not all P. aeruginosa with enhanced AmpC production and depressed oprD 37.

Evidence

Carbapenems are drug of choice for treatment of serious infection with Enterobacteriaceae including those producing ESBLs or AmpC.

Evidence level: 1+
Imipenem use is associated with emergence of resistance in *P. aeruginosa*

Evidence level: 3

Ertapenem treatment is associated with emergence of resistance via porin loss in ESBL- and AmpC-producing Klebsiella *spp.* and *Enterobacter spp.*

Evidence level 3

**Recommendations**

- Use meropenem, imipenem or ertapenem to treat serious infections with ESBL and AmpC-producing Enterobacteriaceae.

  Grading: Strong recommendation for

- Apply antibiotic stewardship to use of all carbapenems to minimize the risk of developing resistance either by acquisition of carbapenemase-producing strains or, with ertapenem, by porin loss.

  Grading: Strong recommendation for

- Do not use imipenem to treat susceptible Pseudomonas infections

  Grading: Conditional recommendation for

- Introduce in the UK mandatory reporting of meropenem- or imipenem- resistant Enterobacteriaceae from all anatomical sites and specimens.

  Grading: Strong recommendation for

- Test immediately for the precise level of meropenem resistance and for an indication of the responsible class of carbapenemase (e.g. MBL/KPC/OXA48-like) all meropenem- or imipenem- resistant isolates of Enterobacteriaceae. Submit to
agreed reference laboratories to determine susceptibility to a wide range of
potentially active agents including, as appropriate colistin, ceftazidime/avibactam, temocillin, aminoglycosides, fosfomycin and tigecycline.

Grading: Strong recommendation for

- Prefer ertapenem for outpatient antibiotic treatment (OPAT) of susceptible infections in view of the once daily dosing regimen.

Grading: Conditional recommendation for

### 7.2 Ceftazidime

Observational studies of ceftazidime-susceptible ESBL-producing *E. coli* and *Klebsiella* spp. infections treated with ceftazidime frequently show treatment failure, mainly during bacteraemias. One study of 7 patients treated with ceftazidime in China suggested useful activity but this may reflect the type of ESBL; CTX-M-14, -27 and -9 enzymes predominate in parts of China (and Spain) and have weak activity against ceftazidime as compared with CTX-M-15 enzymes with lower ceftazidime MICs. The higher CLSI susceptible breakpoint (≤4mg/L) was found to classify 34% of CTX-M positive *E. coli* as susceptible to ceftazidime with normal inocula. Most CTX-M-14 isolates became resistant at higher inocula. The EUCAST breakpoint for susceptibility is <1mg/L reducing this problem. Early problems arose with apparent ceftazidime susceptibility by disc testing of CTX-M-15-producing *E. coli* ST131 isolates in the UK down regulated by an IS26 insertion between promoter and structural gene. Ceftazidime is active against some OXA-48-producing CPE principally those that do not co-produce ESBLs or AmpC enzymes. Ceftazidime retains activity against many isolates of *P. aeruginosa* including in the presence of mutation to imipenem or ciprofloxacin resistance. However strains with derepressed class C (AmpC) β-lactamases or
strongly upregulated efflux mechanisms are resistant, as are strains producing MBLs, other carbapenemases or ESBLs.

**Evidence**

Ceftazidime is usually ineffective in treating multi-resistant infections with Enterobacteriaceae except against some OXA-48 carbapenemase-producing strains.

Evidence level: 3

Ceftazidime remains useful for infections due to quinolone or imipenem resistant h P. aeruginosa

Evidence level: 3

**Recommendations**

- Use ceftazidime for susceptible infections with *P. aeruginosa* including quinolone- or some imipenem- resistant strains

Grading: Strong recommendation for

- Do not use ceftazidime to treat infections due to ESBL-or AmpC-producing Enterobacteriaceae or CPE (other than OXA-48 producers), even if *in vitro* tests suggest the isolate is susceptible.

Grading: Conditional recommendation against use

**7.3 Ceftazidime/avibactam**

Ceftazidime has recently been combined with the β-lactamase inhibitor avibactam. This combination has broad Gram-negative activity including Enterobacteriaceae and *P. aeruginosa*. Ceftazidime-susceptible bacteria remain susceptible to the combination, but
avibactam protects additionally against class A (TEM, SHV, CTX-M, KPC) class C (AmpC) and some class D (OXA) β-lactamases. Ceftazidime/Avibactam has no inhibitory activity against the MBLs (NDM-1, IMP and VIM) but it is the first BL/BLI combination to retain activity against KPC-2 carbapenemase-producing and most OXA-48 carbapenemase producing strains. Ceftazidime/avibactam has minimal activity against Acinetobacter spp., anaerobic or Gram-positive organisms. A recent susceptibility study that included 120 KPC-producing Enterobacteriaceae collected from US hospitals found that ceftazidime/avibactam had MIC<sub>50</sub> values of 0.5/2mg/L. The first case series of use of ceftazidime/avibactam against carbapenem-resistant Enterobacteriaceae has recently been published. Among 37 patients with severe infections due to these organisms 31 had strains with KPC carbapenemases. Resistance to ceftazidime/avibactam emerged independently in 3 cases infected by K. pneumoniae ST258 with KPC-3 enzymes. In 2 of these isolates meropenem MICs were reduced >=4-fold to the susceptible range in parallel with the rise in ceftazidime-avibactam MICs. The overall clinical success rate was 59% of patients whilst microbiological failure occurred in 10 patients, including the 3 patients where resistant mutants were selected. An earlier epidemiological study had shown that ceftazidime/avibactam median MICs of ceftazidime/avibactam are higher for KPC3-producing isolates than those with KPC-2 enzymes although it was unclear if this represents enzyme specificity or quantity. Isolates that produce KPC3 enzyme are internationally widespread including in South America and Southern Europe. Ceftazidime/avibactam resistant isolates with similar or identical mutations can be selected in vitro. The mechanism involves the enzyme becoming a stronger ceftazidime-destroying enzyme, not in it becoming avibactam resistant. The licensing of avibactam — a non-β-lactam — β-lactamase inhibitor with ceftazidime offers a new choice where organisms that produce both AmpC and an ESBL, or KPC2 carbapenemase cause systemic infection.
In phase II double-blind randomized trials, the efficacy of ceftazidime/avibactam was similar to imipenem/cilastatin in treatment of complicated urinary tract infection, (19/27) and (21/35) respectively. A Phase 3 RCT of doripenem versus ceftazidime avibactam in complicated UTI or pyelonephritis, with patients not selected for antibiotic resistance, showed equivalence with microbiological eradication in 304/393 (77.4%) in the ceftazidime/avibactam arm and 296/417 (71%) in the doripenem arm. Efficacy combined with metronidazole was similar to meropenem in a RCT of 203 patients with intra-abdominal infection. A Phase 3 RCT comparison of meropenem against ceftazidime/avibactam with metronidazole in 1066 complicated intra-abdominal infection, with the exclusion of a standardised set of highest mortality surgical indications, again showed equivalence. On intention to treat analysis response rates were 82.5% to the ceftazidime/avibactam-metronidazole combination and 84.9% to meropenem. There was no difference in patient outcome in the combination arm if a ceftazidime-resistant strain of Enterobacteriaceae was present or absent. Only 1 case of C. difficile was recognised in either arm of the study. A RCT of ceftazidime/avibactam and metronidazole against meropenem of 333 patients largely with patients with complicated UTI, but with some patients treated for intra-abdominal infections, all with infections with ceftazidime-resistant Enterobacteriaceae or P. aeruginosa showed 91% response rates at a test of cure visit. None of these patients were infected with carbapenemase-producing strains.

Evidence

Ceftazidime/avibactam has similar efficacy to carbapenems in abdominal and complicated UTI, the former requiring combination of ceftazidime/avibactam with metronidazole.

Evidence level: 1+
Although clinical experience is limited in MDR GNB largely to ceftazidime-resistant organisms in complicated urinary tract infection, it would be expected to be effective when OXA-48 producing MDR GNB cause infection.

Clinical experience against Klebsiella spp. producing KPC-carbapenemase is limited but ominously efficacy is only some 60% with resistance emerging in 10% of treated patients.

**Recommendations**

- Could use ceftazidime/avibactam as an alternative to carbapenems for infection with ESBL- and AmpC- producing Enterobacteriaceae but alternatives may be cheaper

Grading: Conditional recommendation for

- Evaluate further ceftazidime/avibactam use alone or in combination when non-MBL carbapenemase-producing organisms cause infection. KPC-3 producing Klebsiella spp. are vulnerable to mutations in the enzyme causing resistance

Grading: Recommendation for research and possibly conditional recommendation for use restricted to trials

- Do not use for treating infection with anaerobes or bacteria producing MBLs: these are resistant

Grading: Strong recommendation against
7.4 Ceftolozane/tazobactam

Ceftolozane is an oxyimino-cephalosporin that has been combined with tazobactam. Ceftolozane/tazobactam is active against many Gram-negative organisms, including Enterobacteriaceae and *P. aeruginosa*. It is active against *P. aeruginosa* isolates that are resistant to standard agents such as ceftazidime because of derepressed AmpC β-lactamases or upregulated efflux. In terms of MIC, ceftolozane is the most active β-lactam against *P. aeruginosa*, with resistance (MIC >4 mg/L EUCAST) largely confined to those with metallo-β-lactamas or unusual ESBLs such as VEB and GES types. MIC50/90 values against 310 multi-drug resistant isolates of *P. aeruginosa* were 2/8 mg/L. Activity against *Acinetobacter spp.* is variable. Ceftolozane/tazobactam has *in vitro* activity against Enterobacteriaceae producing ESBLs including most TEM, SHV, and CTX-M types. Since oxyimino-cephalosporins are stable to the inhibitor-resistant OXA-1 enzyme, ceftolozane is not compromised by co-production of this enzyme in CTX-M-15 producing Enterobacteriaceae as happens with piperacillin/tazobactam. Activity is less against ESBL-producing *Klebsiella spp.*, possibly owing to high ESBL levels arising from production of additional SHV enzymes. Activity against Enterobacteriaceae with copious AmpC enzyme is variable, but many *Enterobacter spp.* with derepressed AmpC are resistant. The combination has no activity against strains with MBLs (NDM-1, IMP, and VIM) or against those with KPC carbapenemases. Ceftazidime-resistant strains with OXA-48-like enzymes are mostly resistant: ceftazidime-susceptible OXA-48 producers are susceptible to ceftolozane/tazobactam (D.M. Livermore – unpublished data).

Ceftolozane/tazobactam therefore has potentially different uses from ceftazidime/avibactam and should not be used in infections due to AmpC- or KPC-producing Enterobacteriaceae. The absence of clinical comparisons of piperacillin/tazobactam and ceftolozane/tazobactam mean that choices must be made.
on *in vitro* grounds. The apparent enhanced activity of ceftolozane/tazobactam against
strains that co-produce the enzyme OXA-1, including the internationally prevalent *E.
coli* ST131 lineage, needs full laboratory and clinical verification but may make this drug
more likely to produce clinical cure. Caution on clinical outcome is necessary because of
the potential, as with ceftazidime/avibactam, for superinfection with *C. difficile*.

Ceftolozane activity against *P. aeruginosa* including ceftazidime-resistant strains *in vitro*
may offer clinical advantages where MDR Pseudomonas infections are a problem such
as in cystic fibrosis but this needs confirmation in a clinical trial. Optimal dosing in
cystic fibrosis needs to be established but the drug's pharmacokinetics appears to be the
same as in unaffected patients.

Ceftolozane/tazobactam is licensed, at present, for complicated intra-abdominal
infection and complicated urinary tract infection. In a prospective, randomised,
double-blind trial, 993 hospitalised patients with complicated intra-abdominal infection
received either ceftolozane/tazobactam (1.5g 8h IV) plus metronidazole, or meropenem
(1g 8h IV) for 4–14 days. Non-inferiority was demonstrated overall and MIC was not
related to outcome. In fifty patients an ESBL-producing organism was isolated. In these
patients, the clinical cure rate was 95.8% (23/24) in the ceftolozane/tazobactam plus
metronidazole group and 88.5% (23/26) in the meropenem group. In patients with
*CTX-M-14/15* ESBL-producing Enterobacteriaceae, clinical cure was observed in 13 of
13 (100%) and 8 of 11 (72.7%) patients, respectively. A double-dummy, double-blinded
RCT compared ceftolozane/tazobactam against levofloxacin in 1083 patients with
complicated UTI. Patients received ceftolozane/tazobactam (1.5g iv 8h) or
intravenous levofloxacin (750mg od iv). The majority of participants (82%) had
pyelonephritis. Overall, ceftolozane/tazobactam was found to be non-inferior in clinical,
and superior in microbiological, outcome to levofloxacin therapy. In the intention to
treat population, 20 (2.7%) of 731 Gram-negative pathogens were resistant to
ceftolozane/tazobactam at baseline, whereas 195 (26·7%) of 731 were resistant to
levofloxacin. Two (0·3%) of 594 of *E. coli* isolates were resistant to
ceftolozane/tazobactam and 144 (24·2%) of 594 were resistant to levofloxacin. For
patients with levofloxacin-resistant uropathogens (based on CLSI criteria) clinical cure
was seen in 90 (90·0%) of 100 patients in the ceftolozane/tazobactam group compared
(surprisingly) with 86 (76·8%) of 112 in the levofloxacin group. In patients with ESBL-
producing uropathogens, cure with ceftolozane/tazobactam was 55 (90·2%) of 61
compared with 42 (73·7%) of 57 for levofloxacin (95% CI 2·6–30·2). Treatment choice
in complicated UTI and pyelonephritis involving MDR GNB between
piperacillin/tazobactam, carbapenems, ceftolozane/tazobactam, temocillin or
ceftazidime-avibactam depends on the bacteria present and their patterns of
susceptibility.

**Evidence**

Ceftolozane/tazobactam is not active against CPE strains, excepting ceftazidime-
susceptible OXA-48-producers, but otherwise, when combined with metronidazole, is
non-inferior to meropenem in intra-abdominal infection

Evidence level: 1+

Ceftolozane/tazobactam is non-inferior to intravenous levofloxacin in complicated UTI
including those caused by ESBL-producing *E. coli* (most of which are resistant to
levofloxacin)

Evidence level: 2-Ceftolozane/tazobactam is the most active β-lactam *in vitro* against *P.
aeruginosa*

Evidence level: 4
Recommendations

- Use ceftolozane/tazobactam to treat susceptible *P. aeruginosa* infections resistant to ceftazidime

Grading: Conditional recommendation for

- Conduct clinical trials in *P. aeruginosa* infections in cystic fibrosis

Grading: Recommendation for research and possibly conditional recommendation for use restricted to trials

- Use ceftolozane-tazobactam as an alternative to carbapenems to treat urinary or intra-abdominal infection involving ESBL-producing *E. coli*. Caution may be needed when treating infection due to ESBL-producing *Klebsiella spp.* owing to a higher resistance rate.

Grading: Conditional recommendation for

- Do not use for infections due to AmpC- or carbapenemase-producing Enterobacteriaceae or MBL/ESBL-producing *P. aeruginosa*.

Grading: Strong recommendation against

7.5 Aztreonam

Aztreonam is labile to AmpC and ESBL enzymes. It is stable to MBLs and OXA-48-like carbapenemases but most Enterobacteriaceae with these enzymes also express ESBLs or AmpC which confer resistance. Isolates with MBLs or OXA 48 and no ESBL- or AmpC-production may be susceptible (those with OXA-48 alone are likely also to be susceptible to ceftazidime and ceftolozane/tazobactam). At EUCAST breakpoints (*S* <= 1, *R* > 16) most *P. aeruginosa* are intermediate in susceptibility and the drug is usually less
active than ceftazidime or ceftolozane/tazobactam except against MBL-producers
resistant to all other β-lactams which may be intermediate (rarely susceptible) to
aztreonam.

An aztreonam-avibactam combination is in Phase 11 development. This creates a
combination with very promising activity against Enterobacteriaceae with MBLs, OXA-
48, AmpC, ESBLs and other β-lactamases (including AmpC, OXA-1 and CTX-M class)\textsuperscript{214, 215, 216}.

**Evidence**

Aztreonam is not active against Gram-negative bacteria producing ESBLs, AmpC or KPC
carbapenemase; it is only moderately active against *P. aeruginosa*.

Evidence level: 4

It is stable to MBLs but strains possessing these often have ESBL or AmpC as well
resulting in resistance. Similar limitations apply to strains with OXA-48-like enzymes.

Evidence level: 3

Combination with a β-lactamase inhibitor such as avibactam would potentially make
aztreonam useful against MBLs (NDM, IMP and VIM)-producing bacteria that also have
ESBLs or AmpC enzymes.

Evidence level: 4

**Recommendations**

- Do not use aztreonam alone empirically if MDR GNB or Gram-positive or
  anaerobic pathogens are suspected.

Grading: Strong recommendation against
• Do not use aztreonam for CTX-M ESBL- or AmpC-producing bacteria even if these appear susceptible \textit{in vitro}

Grading: Strong recommendation against

• Use aztreonam for MBL- or OXA-48-producing strains if it is certain that they do not produce ESBLs or AmpC

Grading: Conditional recommendation for

• Research usefulness of aztreonam in combination with avibactam for bacteria producing MBLs with ESBL/AmpC enzymes and for those with other carbapenemases.

Grading: Recommendation for research

7.6 Cefepime

Cefepime is not available in the UK. It appeared to be active \textit{in vitro} against ESBL-producing \textit{Enterobacteriaceae} especially when the old NCCLS-CLSI breakpoint of $\leq 8\text{mg/l}$ was used. A retrospective, case-controlled study compared the clinical and microbiologic responses for 10 infections due to ESBL-producing \textit{Klebsiella spp.} and \textit{E. coli} from a non-urinary source with 20 matched controls receiving cefepime for non-ESBL strains. Four patients with ESBL-producers had strains that were resistant to cefepime by broth microdilution MIC, one of whom responded: Three of the remaining six with strains then regarded as susceptible (NCCLS-CLSI breakpoint MIC $\leq 8\text{mg/l}$), failed on treatment. Patients receiving cefepime for infection with ESBL-producing bacteria were 9.7 times more likely to have an unsuccessful clinical and microbiological response than those with non-ESBL-producing bacteria \cite{217}. A randomised evaluator-controlled trial of ICU patients compared cefepime with imipenem for the treatment of
hospital acquired pneumonia. The failure rate was 31% in the cefepime group compared with 0% in the imipenem group. Cefepime MICs of 2-4mg/l, then interpreted as susceptible by the NCCLS_(CLSI) breakpoint of <=8mg/l but now regarded as susceptible dose-dependent by CLSI and intermediate by EUCAST criteria were noted in strains from treatment failures 218. A retrospective case-control study of cefepime-susceptible bacteraemia caused by ESBL-producers in the period 20012-7 compared 30 day mortality amongst 17 patients treated with cefepime versus 161 cases treated with a carbapenem 219. Mortality in the cefepime group was 58.8% versus 16.8% for carbapenem treatment and, in multivariate analysis cefepime treatment was strongly associated with mortality (OR, 9.9; 95% CI, 2.8-319; p 0.001). Mortality with cefepime in definitive treatment also related to MIC being 16.7% (1/6) in those with an MIC=<1mg/l, 45% (5/11) in those with an MIC of 2-8mg/l and 100% (4/4) in those with an MIC of =>16mg/l. In a retrospective study of 305 adults with monomicrobial Enterobacter cloacae infections, those with MICs of 4-8mg/l (i.e. with CLSI dose-dependent susceptibility and straddling the EUCAST I/R breakpoint) had significantly higher mortality than those treated with carbapenem 71.4% vs. 18.2% (= 0.045) 14. Fifty eight percent of strains in the cefepime-treated group produced an ESBL in addition to AmpC. In those definitively treated with cefepime, ESBL-production (16/40 vs. 3/32 p=0.006) and susceptible dose-dependent strains (10/16 vs. 9/56 p=<0.001) were independently associated on multivariate analysis with increased mortality 14. ESBL production was more frequent in those strains with cefepime MICs of 4-8 mg/l (32/36 compared with 61/138 with MIC=<2mg/l p=<0.001). Mortality was not reduced even when high dose regimens (2g 8h iv) were used. Mortality in infections due to ESBL non-producers (with median MICs of 0.5mg/l) treated with definitive cefepime was similar to those who received definitive carbapenem therapy (9/56 vs. 16/72 p=0.5). This study demonstrates the efficacy of cefepime against the presumptive AmpC producer E.
Enterobacteriaceae was suggested by CLSI. In order to maximise cefepime use and spare carbapenems, but these findings suggest this is unwise. A recent systematic review did not support the use of cefepime in empirical therapy of critically-ill patients when ESBL-producing *E. coli* or *Klebsiella* sp. infection is suspected. Even in patients with ESBL strains susceptible to cefepime (≤2mg/l CLSI; < 1mg/L EUCAST), treatment failure can be seen.

**Evidence**

Cefepime has a higher failure rate in treatment of infections due to ESBL-producing GNB than carbapenems unless cefepime MICs were =<1mg/L.

Evidence level: 2+

Bacteraemias due to *E. cloacae* strains without ESBLs and with MIC =>2mg/l <8mg/L can be successfully treated with cefepime.

Evidence level 2+

**Recommendations**

- Could use cefepime to treat infection caused by ESBL- or Amp-C-producing bacteria if susceptible to the EUCAST breakpoint of MIC =<1mg/L.
Grading: Conditional recommendation for

- Do not use cefepime even at increased dose for isolates with i) MIC of 2-8 mg/l (CLSI “susceptible dose dependent”) or ii) MIC 2-4 mg/L (EUCAST intermediate, or iii) strains that produce both AmpC and ESBLs.

Grading: Strong recommendation against

- Do not use cefepime to treat infection caused by carbapenemase-producing Enterobacteriaceae.

Grading: Strong recommendation against

7.7 Cefoxitin

Cefoxitin, the original parenteral cephamycin, was developed by Merk and is now a generic. It is no longer available in Europe but has several suppliers in the USA.

Cefoxitin was licensed at the same time as second-generation cephalosporins like cefuroxime but differs in having activity against gut Bacteroides sp. but minimal activity against Haemophilus influenzae. Cefoxitin is on the list of forgotten antibiotics that may be useful against MDR GNB. It is active against ESBL-producing E. coli but is not active against AmpC-inducible species of Enterobacteriaceae e.g. Enterobacter spp., Citrobacter freundii, Serratia spp., Morganella morganii and Providencia stuartii, nor against P. aeruginosa. Cefoxitin differs from temocillin (which has a 6-alpha methoxy group corresponding to the 7-alpha methoxy group of cefoxitin) in having activity against Gram-positive bacteria including penicillin-susceptible Streptococcus pneumoniae and methicillin-susceptible Staphylococcus aureus, which may be advantageous if a urinary infection is diagnosed but the patient actually has infection due to these organisms elsewhere.
EUCAST no longer cites MIC breakpoints but BSAC had a breakpoint of S<8mg/L and resistant >8mg/L. Typical MICs for *E. coli* and *Klebsiella sp.* are slightly below this level meaning that small reductions in susceptibility can confer resistance. These can arise by reductions in permeability or, (in *E. coli* only) by mutation in promoter or attenuator sequences for *ampC*. Cefoxitin resistance is very common in the Middle East, India and China. In a multicentre study of 1762 isolates from urinary infection in the Asia-Pacific region 50.3% of strains were resistant to cefoxitin. Resistance also occurs in *E. coli* and *Klebsiella sp.*, from plasmid-mediated Amp-C production. Porin loss combined with other mechanisms of β-lactam resistance such as ESBL-production is described as emerging during treatment of some Klebsiella infections (See 6.3.3 & 6.7).

Cefoxitin is used in selective media for *C. difficile* and would be expected to trigger infection with this pathogen. In one recent study antibiotic prophylaxis with cefoxitin was an independent risk factor for *C. difficile* infection. The absolute frequency at which this will occur relative to other antibiotics is not known.

In murine models of pyelonephritis cefoxitin was effective against an OXA-1- and CTX-M-15- producing transconjugant *E. coli* and in combination with fosfomycin prevented selection for fosfomycin-resistant mutants. Only one human trial of cefoxitin against current ESBL-producers has been reported. In this 2015 French study largely of urinary and catheter-related bacteraemia 30/33 patients responded in the first 48 hours and 20/24 evaluable patients at follow-up. Six microbiological failures were documented with emergence of resistance in 2 patients with Klebsiella infection. A pharmacological model suggests 1 h. infusion of 2g four times daily would be effective.

Although cefoxitin appears active against CTXM-15-producing *E. coli* and *Klebsiella spp.*, it lacks temocillin’s activity against strains with copious inducible, derepressed or
plasmid-mediated, AmpC. Cefoxitin may be more prone than temocillin to select *C. difficile*. Temocillin unlike cefoxitin has no Gram-positive spectrum so in empirical use in the elderly where it is not clear if the urinary tract or the chest/skin is the source of infection, it may need supplementation with another antibiotic. It is not clear if cefoxitin’s reintroduction would offer any sustainable or competitive advantage apart from its carbapenem-sparing capacity as its four-times daily intravenous dosing makes it only usable in inpatient treatment not OPAT.

**Evidence:**

Cefoxitin is an intravenous cephemycin antibiotic, formerly licensed in the UK.

Inducible, derepressed or plasmid-mediated AmpC-production confers resistance as does porin loss, especially in association with ESBL-production. Nevertheless, in *vitro*, animal and human studies indicate activity against ESBL-producing strains of *E. coli* and *Klebsiella* spp. Treatment can be complicated by emergence of resistance due to porin loss.

**Grading:** Level 3.

**Recommendations**

- Could use as a carbapenem-sparing agent for infections caused by CTX-M-15-producing *E. coli* but is only suitable for inpatient use not OPAT because of the short serum half-life. Narrower Gram-negative spectrum than temocillin so less suitable for empirical use in UTI.

**Grading:** Recommendation for research and possibly conditional recommendation for use restricted to trials.
7.8 Temocillin

Temocillin is a semi-synthetic 6-alpha-methoxy derivative of ticarcillin that is highly stable to most β-lactamases except MBLs (e.g. IMP, NDM, and VIM) and OXA-48-like enzymes. It lacks activity against anaerobes, Gram positive bacteria and most Gram-negative non-fermenters such as *P. aeruginosa* and *Acinetobacter* spp. It retains *in vitro* activity against ESBL- and AmpC-producing Enterobacteriaceae\(^{231,232}\), and some KPC-producing *E. coli* and *Klebsiella pneumoniae* \(^{233}\), and *Burkholderia cepacia* complex \(^{234}\). It is active against Enterobacteriaceae strains whose AmpC production is stably derepressed \(^{235}\). No EUCAST breakpoint for susceptibility to the drug has yet been published but the BSAC had a systemic value of S <8, R>8mg/L MICs for temocillin of KPC-producing bacteria are in the range of 4-32mg/L (mode 16mg/L). In a lethal mouse model of intra-abdominal infection using strains of KPC producing *E. coli* temocillin was effective against KPC-2 \(^{236}\). Temocillin has poor activity against carbapenem-resistant isolates of Enterobacteriaceae lacking carbapenemases – presumptively due to porin loss \(^{237}\). This antibiotic has no activity against OXA-48 or MBL-producing strains \(^{238}\).

Caution is also needed in predicting results of treatment of systemic infections from *in vitro* susceptibility and further trials of temocillin alone at defined and possibly greater doses than the licensed 2g twice daily are necessary. Outcomes should be correlated with MIC.

At present, clinical studies are limited to non-comparative series. The largest multi-centre study (non-randomised retrospective case series) involved 92 patients who were treated with at least 3 days of therapy \(^{239}\). Urinary tract and bacteraemia (42 episodes each) were the most frequent indications followed by hospital acquired pneumonia.

Dosages of ≥4g/day, rather than 1 g twice daily, were associated with improved outcome. Patients with strains producing Amp C or ESBL enzymes responded microbiologically in 23/27 or 18/22 cases in respectively UTI or bacteraemia. Higher
dosage regimens, including 2g three times daily and 6g by continuous infusion and use in veno-venous haemofiltration are reported in the literature with suggestions that these improve efficacy\textsuperscript{240}. In a retrospective case review of bacteraemia caused by KPC producing Enterobacteriaceae, 14/14 patients treated either alone or in combination with temocillin survived, whereas 6/30 treated similarly with tigecycline died\textsuperscript{241}. Two studies have been published on the use of temocillin in cystic fibrosis patients with \textit{B. cepacia} complex and sometimes \textit{P. aeruginosa}. Both were retrospective non-randomised audits the first showing equivalence of combinations of temocillin with tobramycin versus other agents with tobramycin against \textit{B. cenoepacia} and the second showing that 18/32 courses of temocillin resulted in improvement in the patient’s infection\textsuperscript{242,243}.

\textbf{Evidence}

Temocillin at a dose of 2g twice daily is an effective and well tolerated drug for urinary tract infection with AmpC- or ESBL-producing bacterial infection.

Evidence Level: 3

Although \textit{in vitro} work suggests activity against many KPC-producing bacteria, there is little published clinical evidence to support this. Respiratory infections, including cystic fibrosis infections with \textit{Burkholderia cepacia}, and other sites of systemic infection requires further clinical trials.

Evidence Level: 4

\textbf{Recommendations}

- Use alone for UTIs and associated bacteraemia caused by AmpC- or ESBL-producing Enterobacteriaceae.
Grading: Conditional recommendation for

- Continuous infusion or thrice-daily dosing may be desirable for systemic infections with ESBL- or Amp-C producing bacteria

Grading: recommendation: for research and possible conditional recommendation for use restricted to trials

- Could use for UTIs with KPC-producing Enterobacteriaceae but not for OXA-48 or MBL-producers, on basis of published in-vitro data.

Grading: Recommendation for research and possible conditional recommendation for use restricted to trials

7.9 Ampicillin/sulbactam

Sulbactam has in vitro microbiological activity against some strains of *A. baumannii*, including some carbapenem-resistant lineages. Microbiological studies showed that sulbactam alone (without ampicillin) was active against these bacteria. In an uncontrolled study, forty-two patients with infections caused by multi-drug–resistant *A. baumannii* were treated with sulbactam or ampicillin/sulbactam. Eighteen received sulbactam alone and 24 received ampicillin/sulbactam; no difference in cure rate was observed between the two groups. Another study compared ampicillin/sulbactam to colistin therapy in a retrospective review of patients who had nosocomial infections caused by carbapenem-resistant *Acinetobacter spp.* from 1996 to 2004. Eighty-two patients received polymyxins and 85 were treated with ampicillin/sulbactam. The authors concluded that ampicillin/sulbactam appeared to be more efficacious than polymyxins. More generally, and predictably, multivariate analysis found that prognostic factors for in-hospital mortality were older age, septic shock and higher APACHE II score. A small retrospective non-blinded trial compared treatment with
ampicillin-sulbactam to imipenem and tried also to address the benefit of combining ampicillin/sulbactam with colistin. There was no difference in outcome\textsuperscript{246, 247}. Two small RCTs have tried to assess differences in dosing regimens and efficacy compared with colistin\textsuperscript{248, 249}. Overall the evidence base is poor and interpretation is difficult without consideration of the MIC for the organism. In context sulbactam MICs for most UK isolates of carbapenem-resistant \textit{A. baumannii} are 16-32mg/L implying poor rates of susceptibility (D.M. Livermore, unpublished data).

**Evidence**

Ampicillin/sulbactam appears effective in treating infections due to some carbapenem-resistant, \textit{Acinetobacter spp.} but many isolates in the UK have relatively high sulbactam MICs.

Evidence level: 3

**Recommendations**

- Could use against some carbapenem-resistant apparently sulbactam-susceptible \textit{A. baumannii} isolates, Caution needed in the UK because of a higher range of MICs. Absence of a breakpoint prevents categorisation as susceptible/resistant.

Grading: Conditional recommendation for

**7.10 Co-amoxiclav**

Co-amoxiclav is a combination of the broad-spectrum amoxicillin with the beta-lactamase inhibitor clavulanic acid. Co-amoxiclav is known to select for Enterobacteriaceae resistant to the clavulanate component as well as amoxicillin in the gastrointestinal flora\textsuperscript{250}. Co-amoxiclav has been successfully used to treat urinary tract infections due to ESBL-producers, as described in case reports and an observational
The cure rate among 37 patients with cystitis treated with co-amoxiclav was 93% for those with susceptible isolates (minimum inhibitory concentration ≤8 mg/L) and 56% for those with intermediate or resistant isolates (minimum inhibitory concentration ≥16 mg/L) (P=0.02). The study was performed in Spain, where many ESBL-producers have CTX-M-14 enzyme; in the UK more have CTX-M-15 and many of these co-produce OXA-1, an inhibitor-resistant penicillinase, raising co-amoxiclav MICs to the intermediate or resistant range. Furthermore MIC determinations were done with a β-lactam:β-lactamase inhibitor ratio of 2:1 and higher MICs would likely be obtained using the fixed clavulanate concentration of 2 mg/L now advocated by EUCAST. The outcomes for bacteraemias treated with co-amoxiclav or piperacillin/tazobactam have been reviewed and the findings are discussed in the section on piperacillin/tazobactam.

Evidence

These studies suggest that co-amoxiclav is effective in lower UTIs caused by ESBL-producing bacteria but efficacy was only reliably predicted in strains where these organisms were fully susceptible in vitro and lacked co-production of OXA-1 β-lactamase.

Evidence level: 3

Recommendations

- Use for lower UTI due to known ESBL-producing bacteria only if current isolates, or, if using empirically, recent isolates, are fully susceptible.

Grading: Conditional recommendation for
Different susceptibility standards are used worldwide and so correlations of mortality with in-vitro susceptibility cannot be reliably transferred between countries. EUCAST regards more isolates as resistant than CLSI. Some countries such as the UK have a higher prevalence of Enterobacteriaceae with CTX-M-15 and, in *E. coli*, OXA-1 β-lactamase and these are more resistant than the CTX-M-14 ESBL producers circulating, for example, in Spain. This may critically affect the validity of evidence collected from different laboratories and hospitals about the adequacy of these combinations against ESBL-producing bacteria.

The use of piperacillin/tazobactam for treating bacteraemias caused by ESBL-producing bacteria remains consequently contentious. One recent retrospective analysis of 331 patients in a US hospital with bacteraemia due to ESBL-producing bacteria suggested carbapenems were superior to piperacillin/tazobactam. One hundred three (48%) patients received piperacillin/tazobactam empirically and 110 (52%) received carbapenems empirically. The adjusted risk of death was 1.92 times higher for patients receiving empiric piperacillin/tazobactam compared with empiric carbapenem therapy. Another retrospective study of bacteraemic patients with ESBL-producing *P. mirabilis* compared the outcomes of patients treated by piperacillin/tazobactam or a carbapenem for at least 48 hours. Forty-seven patients with available clinical data were studied of whom 34 were included. Only 11% of strains were imipenem susceptible but MICs of the drug for Proteaeae typically cluster around the breakpoint. The overall 30-day mortality rate was 29.8%. 3/21 patients treated with carbapenems (all imipenem) died within 30 days (all in hospital) versus 4/13 treated with piperacillin-tazobactam – a non-significant difference. Furthermore, among those treated by piperacillin/tazobactam, the mortality rate was lower in those infected by the isolates with lower piperacillin/tazobactam MICs (≤0.5/4 mg/L) when compared with isolates.
with MICs of ≥1/4 mg/L (0/7 versus 3/5; P = 0.045). A study of 39 episodes of bacteraemia due to ESBL-producing *E. coli* from Spain found a statistically significant reduction in 30 day mortality in infections from non-urinary sources if the MIC ≤ 2 mg/L (0/11) compared with those strains with higher MIC (7/17). This suggests that even the current EUCAST breakpoints (S<8mg/L, R>16mg/L) are too high to give guidance on clinical response. An analysis of patients with bacteraemias due to ESBL-producing *E. coli* was performed to assess the efficacy of combinations of piperacillin/tazobactam or co-amoxiclav compared with carbapenems. Mortality in patients treated with such BL/BLI combinations or carbapenem was compared in two cohorts: empirical therapy and definitive therapy. Mortality rates at day 30 for those treated with BL/BLI versus carbapenems were 9.7% versus 19.4% for empirical therapy and 9.3% versus 16.7% for definitive therapy respectively. After adjustment for confounders, no association was found between either empirical therapy or definitive therapy and increased mortality. The study suggested that co-amoxiclav and piperacillin/tazobactam may be suitable alternatives to carbapenems for treating patients with bacteraemias due to ESBL-E coli but only in the minority that were susceptible in vitro. The study was not randomized, and confounding due to unmeasured variables may have occurred. This retrospective observational study has been repeated on a multi-national basis and extended to 627 patients with results that BL/BLI combinations were statistically as effective as carbapenems in empirical and directed therapy against ESBL-producing Gram-negative bacteraemia. A subset of 207 patients had their ESBL genes of their pathogens examined by PCR: 42 were identified as CTX-M-15, 27 as CTX-M-1, 31 CTX-M-14 and 18 as CTX-M-9. No details were given of response rates in relation to the presence of specific resistance genes and co-production of OXA enzymes was not sought. In another study co-amoxiclav and piperacillin/tazobactam susceptibility of the bacteria causing bacteraemia, particularly
for *E. coli* ST131, were not correlated: 51% of the isolates also had OXA-1 and 90% of isolates were reported susceptible to piperacillin/tazobactam versus 26% susceptible to co-amoxiclav by CLSI criteria. Such discrepancies with different BL/BLI may relate to whether the EUCAST or CLSI breakpoints are used as the MICs for many isolates with a combination of CTX-M-15 and OXA-1 enzymes cluster around 16mg/L. The relationship of the BL/BLI used and its MIC for infecting strain to efficacy in lower UTIs (where urinary concentrations are higher than in serum) or bacteraemia needs to be established. More generally, individual drug/inhibitor combinations must be separately studied for efficacy, and related to both the β-lactamase genes present and *in vitro* susceptibility. As American commentators have pointed out, it is important to note the dosing regimen when considering response to piperacillin-tazobactam of many ESBLs. Many Spanish studies used piperacillin-tazobactam at 4.5g 6-hourly not the usual licensed UK dose of 4.5g 8-hourly. With β-lactams increasing the time above the MIC substantially decreases mortality. It is possible that more frequent dosing would achieve this. More materially this can be achieved with continuous infusion, albeit with higher daily drug dosage (which might breach targets to reduce use) and could be considered to increase efficacy of piperacillin-tazobactam. It cannot be anticipated with biliary excretion whether this will change selection pressure for superinfecting organisms or *C. difficile* in the gastrointestinal flora.

A retrospective case review of empirical treatment of bacteraemia caused by ESBL-producing *E. coli* or ESBL-producing Klebsiella sp. showed a mortality rate of 18/70 (25.7%) when patients received carbapenems. If they received piperacillin/tazobactam 8/44 (18.2%) died if the strain retrospectively was susceptible by CLSI criteria but 3/6 died if the strain was resistant or intermediate. Similarly, if they received co-amoxiclav 3/40 (7.5%) died if the strain retrospectively was susceptible by CLSI criteria but 10/27...
(37%) died if the strain was resistant or intermediate\textsuperscript{261} piperacillin/tazobactam. Data on the genotypes of the ESBL producers present was not provided.

The findings of all these studies cannot be simply applied to the UK where many ESBL-producing strains are more resistant than CTX-M-14 as they co-produce CTX-M-15 and OXA-1 \(\beta\)-lactamases, with the latter enzyme compromising susceptibility to piperacillin/tazobactam. Variable dosing further complicates the picture.

Piperacillin/tazobactam is commonly used to treat infections caused by \textit{P. aeruginosa}. A retrospective cohort study of bacteraemic patients showed that in 34 episodes of bacteraemia caused by strains with a MIC of 32 or 64 mg/L to piperacillin/tazobactam, the 30-day mortality was significantly greater than controls given other appropriate therapy \textsuperscript{262}. At the time, CLSI defined strains as susceptible if they had an MIC of \(\leq\)64 mg/L whereas EUCAST, then as now, has a breakpoint for susceptibility of \(\leq\)16+4 mg/L and for resistance \(>\)16+4 mg/L.

**Evidence**

Could use piperacillin/tazobactam in some blood stream infections where ESBL-producers appear susceptible \textit{in vitro} but mortality may be higher than with carbapenems.

Evidence level 2-

Mortality when piperacillin/tazobactam is used in blood stream infection due to ESBL-producing Enterobacteriaceae without regard to \textit{in vitro} susceptibility appears higher than with carbapenems.

Evidence level 2+
In vitro susceptibilities by EUCAST and CLSI recommendations on what is a susceptible organism differ for Enterobacteriaceae but only two-fold. There is no good analysis of the impact of this difference in relation to i) strain MIC ii) clinical outcome of infections at different sites and iii) different ESBL genotypes.

Evidence level: 4.

Breakpoints for piperacillin/tazobactam against Enterobacteriaceae have changed with time. Better outcomes may be seen with isolates much more susceptible (MIC $\leq 2$ mg/L) than the currently agreed piperacillin/tazobactam Enterobacteriaceae breakpoints (EUCAST Sensitive if MIC $\leq 8$+4 mg/L resistant if MIC $> 16$+4 mg/L CLSI Sensitive if MIC $\leq 16$+4 mg/L, resistant if MIC $> 128$+4 mg/L.

Evidence level: 3

Recommendations

- Use for infections with known ESBL-producing bacteria only if current isolates, or, if using empirically, isolates from the recent past, are fully susceptible.

Grading: Conditional recommendation for

- Consider definitive use of piperacillin/tazobactam to treat infections caused by *P. aeruginosa* if susceptible by EUCAST standards.

Grading: Conditional recommendation for

7.12 Aminoglycosides

Parenteral broad-spectrum aminoglycosides are potentially important carbapenem-sparing drugs for infections due to MDR-GNB. Three such antibiotics, gentamicin, tobramycin and amikacin remain available in the UK following withdrawal of netilmicin.
and sisomicin. These antibiotics have intrinsic activity against all *P. aeruginosa*, *Acinetobacter spp.* and Enterobacteriaceae but plasmid-borne resistance (and chromosomal resistance in *Providencia spp.* and *Serratia spp.*) now limits their spectrum. Resistance is mostly due to i) bacterial aminoglycoside-modifying enzymes which acetylate, phosphorylate or adenylate vulnerable hydroxyl or amino groups or ii) to 16s ribosomal methyltransferases which alter the binding site for aminoglycosides. The latter mechanism produces pan-resistance to aminoglycosides except the veterinary product apramycin\textsuperscript{263}. By contrast, the vulnerability of aminoglycosides to modifying enzymes varies, with amikacin inactivated by fewer enzymes than gentamicin or tobramycin\textsuperscript{264}. Initially aminoglycoside-modifying enzymes were restricted to certain species but integron and transposon carriage have mediated their wide dissemination.

Amikacin evades AAC (3) and AAC (2') enzymes but remains vulnerable to AAC (6')-I as does tobramycin. AAC(6')-1b-cr arose from AAC(6')-1b by the substitutions Trp102Arg and Asp179Tyr and can acetylate ciprofloxacin (not levofloxacin) as well as aminoglycosides causing deactivation. This enzyme, formerly rare in the UK\textsuperscript{265} is commonly found in *E. coli* ST131. Amikacin MICs typically are raised to just below the susceptible breakpoint. Such reductions nevertheless may be important since efficacy of aminoglycosides is proportional to the ratio of peak concentration to MIC\textsuperscript{266}. EUCAST currently suggests that reports on isolates with this enzyme are edited to amikacin-resistant but this is under review. In contrast to other common aminoglycoside modifying enzymes AAC (6')-1 spares gentamicin. Aminoglycoside-nucleotidyl transferases (ANT-6, ANT-9, ANT-4', ANT-2", and ANT-3") do not confer amikacin resistance nor – except APH (3)-V1 which is mostly confined to *A. baumannii*, do aminoglycoside phospho-transferases in Gram-negative species.
Overall resistance rates to gentamicin in community-onset *E. coli* bacteraemia in 2012-2014 was 8.6%. This is a similar figure to the 8.7% resistance rate to piperacillin/tazobactam in community-onset cases. Such data must be considered when empirically treating probable Gram-negative bacteraemia of likely urinary or unknown origin. In the 1980s, parenteral aminoglycoside therapy rarely selected for resistant Enterobacteriaceae in the gut flora but oral aminoglycosides given for selective digestive decontamination in haematological malignancy frequently did so and continued to do so over a 20 year period once resistance emerged, even when combined with oral colistin.

There is limited surveillance of the genotypic distribution of aminoglycoside-modifying enzymes except in specific strains and in those with other resistances (e.g. ESBL-producers). Little is known of travel associations beyond those to gentamicin and tobramycin (but to a lesser extent amikacin) associated with acquisition of ESBL- or carbapenemase producers for which there are clear travel links.

Aminoglycoside activity against *P. aeruginosa* varies between patients with cystic fibrosis where aminoglycosides continue to be heavily used and patients with other comorbidities. Resistance due to efflux pumps and permeability defects are common, as well as aminoglycoside-modifying enzymes. Tobramycin which has greater intrinsic activity than gentamicin against this species (off-setting its lower activity against Enterobacteriaceae) and which causes less toxicity than gentamicin, continues to be the aminoglycoside most likely to remain active. A recent meta-analysis continues to suggest that use of β-lactam aminoglycoside combinations in the absence of cystic fibrosis offers no statistically significant advantage in terms of outcome compared with use of an active β-lactam alone.
A new aminoglycoside plazomicin (ACHN 490, Achaeogen)\textsuperscript{272,273,274} has completed clinical trials. This evades modification by almost all aminoglycoside modifying enzymes except the AAC(2') chromosomal enzymes of \textit{Providencia spp}. It is however compromised by the plasmid mediated ArmA and Rmt 16S ribosomal methyltransferases which are currently rare in UK MDR GNB except in Enterobacteriaceae strains producing NDM-1 carbapenemase\textsuperscript{263} or OXA-23 carbapenemase-producing \textit{A. baumannii} which have spread globally over the last 10 years.

Aminoglycosides have a narrow margin between being effective and toxic to the auditory and vestibular apparatus or to the kidneys. They fell from favour as broader – spectrum β-lactams were developed. For acceptably safe use, intervals between doses are increased usually to a minimum of once daily but with doses related to renal clearance and MIC and the presumption of a post-antibiotic effect. If the dosage is based on the patient’s weight it is possible, using a nomogram, to model the likely blood concentration at varying intervals after the dose. Measuring plasma levels between 6 and 14 hours after the dose, usually now by immunoassay, and relating these levels on to the nomogram permits more precise dosing intervals than by measuring renal function. Nomograms for gentamicin and tobramycin at doses of 7mg/kg\textsuperscript{275} and 5mg/Kg\textsuperscript{276} in adults have been constructed and their use is associated with a low incidence of detected ototoxicity (3/2184 cases in the former). The dosage recommendation for amikacin is 15mg/kg/day reflecting that, amikacin MICs are 2 to 4 fold higher than gentamicin MICs for susceptible strains. Much higher incidences of toxicity with all aminoglycosides are well recorded and it is still common to encounter in the UK deficiencies in i) weight-related dosage ii) dosage interval especially if there is renal impairment, iii) measuring levels in every case, and iv) taking blood for assay at the correct interval after dosage and recording both the time of administration and time...
of sample collection to enable later interpretation of assay results by other staff.

Validation of expected and achieved serum levels has been undertaken for 7mg/kg dose but not 5mg/kg doses which are based on exclusion of some patients considered in the former study. There is no validated nomogram for amikacin and immunoassays for this antibiotic are not widely available on automated immunoassay platforms. There are no trial data on amikacin use in *E. coli* ST131. Vestibular toxicity with all aminoglycosides commonly presents after the drug has stopped and the patient has left hospital. Toxicity can occur after normal courses of 5 daily doses or even a single dose. Auditory toxicity is initially often subclinical requiring audiograms to detect.

The true incidence of toxicity is difficult to determine. Renal toxicity can be measured by quantitative renal function tests or qualitative urinary renal tubular enzymes. These critical steps to safe use as determined by case follow-up after the patient has left hospital, have not yet been assessed for plazomicin although there are no described cases of toxicity yet in clinical trials. In older studies before the adoption of once daily regimens and weight-related dosage, auditory toxicity appears to have been commoner with amikacin than gentamicin whilst vestibular toxicity rates were not significantly different: toxicity was commoner with increasing age paralleling a decline in renal function. This creates an issue, insofar as infections with MDR GNB and ESBL-producers occur more frequently among those aged over 65 years and especially over 75 years of age. It is noteworthy that one recent Scottish national intervention in surgery as part of targeted antimicrobial stewardship measures to reduce the incidence of *C. difficile* by 30% in 2 years was to substitute use of gentamicin for cephalosporins in prophylaxis in surgery. In Tayside, an interrupted time series with segmented regression in 7666 patients undergoing orthopaedic surgery (excluding fractured neck of femur), where 2 doses of flucloxacillin 1G and one dose of 4mg/Kg gentamicin were substituted for cefuroxime was performed. An unacceptable 94% increase in acute
Kidney injury in gentamicin-treated patients occurred and the gentamicin use was stopped. Patients undergoing implant surgery had a mean age of 71 years and 36% had received non-steroidal anti-inflammatory drugs in the last year and 38% received a diuretic which are known cofactors for gentamicin nephrotoxicity but this was adjusted for in the study. One year mortality was higher in the acute kidney injury group (20.8% vs. 8.2%). There was no association of acute kidney injury in a further 4816 patients in other surgical specialties where gentamicin was substituted. It is not certain whether the effect was due to gentamicin, flucloxacillin, or the combination or whether all patients additionally received gentamicin bone cement.

Evidence:

Aminoglycosides retain activity against a similar proportion of Enterobacteriaceae to piperacillin/tazobactam (8.6-8.7%). However approximately 50% of ESBL-producing E. coli in the UK are resistant to gentamicin and more to tobramycin.

Evidence level: 3

Overall resistance rates to amikacin are lower than to gentamicin and tobramycin in the UK. However bacteria producing AAC(6') are usually amikacin resistant and bacteria producing the AAC(6')-1b-cr enzymes including many E. coli ST131 often have reduced amikacin susceptibility. Strains producing NDM-carbapenemase often carry 16S ribosomal methyltransferases which confer high-level pan-resistance to aminoglycosides including amikacin and plazomicin. 16S ribosomal methyltransferases are also frequent in UK A. baumannii.

Evidence level: 3

Plazomicin, a new aminoglycoside evades almost all aminoglycoside-modifying enzymes but is inactive if 16s ribosomal methyltransferases are present. It has recently
completed a phase 3 RCT with superiority to meropenem in complicated UTI so far
reported only in a press release.

Evidence level: 3

Historically parenteral aminoglycosides rarely proved selective for resistance among Enterobacteriaceae in the faecal flora. However, because of resistance linkage and carriage on transposons and integrons aminoglycoside resistance may be selected by use of other antibiotics.

Evidence level: 3

Evidence from travel-associated ESBL-producers suggests that aminoglycoside-resistance may also be travel-associated. The co-carriage of 16S ribosomal methyltransferases by strains with NDM-carbapenemase linked to the Indian subcontinent is noteworthy.

Evidence level: 3

The narrow therapeutic index of aminoglycosides demands attention to the detail of weight-related dosing and frequency of doses, collection of blood at an appropriate time for assays, and the careful interpretation of antibiotic assays by nomograms. These actions are essential for adequately safe management of patients treated with gentamicin and tobramycin. Similar modern safety measures are likely to be necessary for amikacin and plazomicin but nomograms are not, and assays may not be, widely available.

Evidence level: 4
When strains are susceptible and safety measures are well-organised and reviewed in hospitals, gentamicin and tobramycin are useful carbapenem-sparing agents for definitive treatment.

Evidence level: 4

**Recommendations**

- Could use gentamicin empirically in the UK if the likelihood of MDR GNB is low.

  Grading: Conditional recommendation for

- Could use gentamicin as a carbapenem sparing agent for urinary, intra-abdominal and bacteraemic infections due to ESBL-producing *E. coli* when susceptibility is confirmed but do not use empirically if the risk of MDR GNB is raised.

  Grading: Conditional recommendation for

- Could use gentamicin in combinations for urinary, intra-abdominal and bacteraemic infections due to gentamicin-susceptible KPC-producing *Klebsiella spp.* if strain is resistant to colistin and meropenem (See Section 7.18).

  Grading: Conditional recommendation for

- Use once daily dosage of gentamicin if no renal impairment followed by measurement of levels 6 to 14 hours post dose and adjust repeat dosage by reference to the appropriate 7mg/kg or 5mg/kg nomogram. Consider increased risks of toxicity if there is co-administration of nephrotoxic or ototoxic drugs.

  Grading: Strong recommendation for.
- Avoid tobramycin for MDR Enterobacteriaceae because of risk of resistance due to AAC (6′)1 and AAC (6′)-1b-cr

Grading: Conditional recommendation against

- Use tobramycin in preference to other aminoglycosides for susceptible Pseudomonas infection

Grading: Conditional recommendation for

- Use once daily dosage of tobramycin if no renal impairment followed by measurement of levels 6 to 14 hours post dose and adjust repeat dosage by reference to nomogram.

Grading: Strong recommendation for

- Modernise use of amikacin, which has improved activity, with development of validated nomograms. Ensure assays are readily available before repeat doses and consider, because of the risks of toxicity, the practicality of monitoring with audiograms.

Grading: Conditional recommendation for.

### 7.13 Polymyxins

The polymyxins are a group of five chemically different bactericidal antibiotics (polymyxins A to E). Only polymyxin B and polymyxin E (colistin) have been used in clinical practice. Intravenously administered colistin methane sulphonate is most widely used, and requires conversion in the body to the active colistin molecule.

Polymyxins have a wide spectrum of activity against Gram-negative organisms, including most Enterobacteriaceae, *A. baumannii, P. aeruginosa* and *S. maltophilia*, but
are inactive against *B. cepacia, Proteus spp., Providencia spp., Morganella spp.* and *Serratia marcescens*. Resistance to colistin occurs in some *P. aeruginosa* isolates but remains rare and almost exclusive to cystic fibrosis isolates. Acquired colistin resistance is generally rare but has become common in *K. pneumoniae* in Italy. Colistin heteroresistance is defined as the emergence of resistance to colistin in a subpopulation of an otherwise susceptible (MIC of ≤2 mg/L) population. This may be related to exposure to suboptimal polymyxin concentrations. Detection of resistance or heteroresistance is difficult and is reviewed elsewhere.

Etest®, disc diffusion, Microscan® and VITEK2® detections methods are currently unreliable and data for Phoenix® are only published for *Acinetobacter baumannii*. A comparison of BMD was made with VITEK2®, Sensititre™ and Etest® using a collection of 76 Enterobacteriaceae, including 21 MCR-1 positive strains. Both Etest® and VITEK2® performed poorly against BMD with very major error (VME) rates of 12% (Etest®) and 36% (VITEK2®) for colistin. Poor performance of both Phoenix® and VITEK2® with substantial under reporting of resistance has been reported when using these systems for testing *Acinetobacter baumannii*.

The difficulty of detecting colistin resistance in routine laboratories was evident in a recent US study. Resistance to gentamicin was rarer and tigecycline resistance commoner in colistin-resistant isolates. Colistin resistance was associated with increased hospital mortality. Most colistin resistance is chromosomally mediated, involving various mutations that modulate two component regulatory systems (*e.g.* pmrAB, phoPQ and its negative regulator mgrB in the case of *K. pneumoniae*), leading to modification of lipid A with moieties such as phosphoethanolamine or 4-amino-4-arabinose, or in rare instances to total loss of the lipopolysaccharide. Of concern is the recent reporting of plasmid-mediated polymyxin-resistance lipid A-modifying enzymes (MCR-1 and 2) that confer resistance in Enterobacteriaceae. MCR-1 was first...
found in China but is now being detected worldwide mainly in Enterobacteriaceae of animal origin but also in occasional human isolates. It remains much rarer than mutational resistance. China plans to stop use of 8000 tons of colistin in animal feed from April 2017. A recent study shows mcr-1 genes are very widespread (50-100%) in chicken in hatcheries, commercial farms and supermarkets and a slaughterhouse in Shandong. Although testing of hatcheries was negative, NDM-carbapenemase-producing E. coli were recovered from 21.8% of samples; 23% of carbapenem-resistant E. coli tested MCR-1 positive and multiple sequence types and NDM subtypes were found. There are widespread reports of MCR-1 in the European (including UK) food-chain. Synergy studies suggested many years ago that polymyxins, trimethoprim and sulphonamides might be useful together in therapy and these studies need repeating with other agents and newer strains. Pharmacokinetic and pharmacodynamic data have been limited, particularly in critically ill patients. Polymyxins were developed before the advent of contemporary drug evaluation. Colistin methanesulfonate is an inactive pro-drug converted in vivo to the active drug and different brands may produce different concentrations of active drug. Data suggested drug concentrations are very variable and dosing in excess of data-sheet recommendations may be required commonly on the basis of pharmacokinetic parameters. Recently the FDA and European medicines agency have made new, but different, recommendations for intravenous colistin in patients with various degrees of renal function. These have been assessed using data from 162 adult critically ill patients with varying renal function. A comparison showed that adequate serum levels with impaired renal function were more likely to be attained with European guidelines and a later paper suggests that in the critically ill target concentrations are difficult to achieve if creatinine clearance =>80ml/min/1.73m². Data are also now available on the
implications of haemodialysis. Therapeutic drug monitoring is advisable, if available and depends critically on maintaining stability of the drug in separated plasma.

Colistin can be given intravenously, or in respiratory infection via the aerosol route (typically in patients with CF; either alone or combined with IV administration), or intrathecal.

Polymyxin B or colistin sulphate can be given orally as a non-absorbed major component of selective digestive decontamination regimens. Selective digestive decontamination has been widely used for general infection prevention in neutropenia and intensive care. Polymyxins orally were widely added in haematology to aminoglycosides, trimethoprim-sulfamethoxazole or ciprofloxacin to prevent emergence of resistance and in intensive care units to parenteral cephalosporins and oral tobramycin. Recent findings that colistin resistance is difficult to detect accurately and it’s frequency is usually underestimated, the clear emergence in China and elsewhere of plasmid mediated resistance and the emergence of colistin resistance in KPC-producing Klebsiella spp. in Italy, China and the USA imply that it can no longer be relied on to prevent emergence of resistant strains in patients who have strains that are already frequently resistant to the drugs it was added to protect. Use of colistin in all patients in such a unit might well become a mechanism now for selection for XDR GNB or indeed pan-drug resistant MDR GNB in the critical care and haematology units where it is used. This is an enduringly controversial area which we do not have space to fully review but such selection of colistin resistance in ESBL-producing Klebsiella spp. in an ICU has already been reported. We consider continued use of colistin-containing decontamination regimens should be reviewed urgently within specialties and at the local level, and in our judgement is now unwise.
Clinical reports and reviews of experience with colistin are relatively encouraging, with side effects (principally nephrotoxicity and neurotoxicity) observed less often than expected from historical data. These studies are summarized in Table 6. In Italy strict rules for the use of colistin are advocated to stop the spread of colistin resistant KPC-producing Klebsiella spp., which have increased three fold in 4 years among bacteraemic patients. A case-control study of this guidance showed associations of resistance with previous colistin therapy, previous colonization or infection with KPC-producing Klebsiella spp., and a Charlson comorbidity score >3 (all of which were associated with mortality) and also with neutropenia and >3 hospitalisations.

The addition of aerosolized to IV colistin has been compared with IV colistin alone for the treatment of VAP in several studies. Korbila and colleagues demonstrated an improvement in outcome with the addition of aerosolized colistin but no benefit was demonstrated in another study. Both had methodological flaws. NICE has recently reviewed the usefulness of aerosolised colistin or tobramycin dry powders in patients with cystic fibrosis and concluded there were some patients who would benefit from colistin dry powder with cost reduction.

Polymyxin B is more toxic than colistin (polymyxin E) but has the advantage of not requiring subject-variable conversion to an active form. A recent retrospective cohort study compared 45 patients with P. aeruginosa bacteraemia treated with polymyxin B at a median dose of 141+/−54 mg/day usually in 2 divided doses: 11 received >200mg/day. Eighty eight patients were treated with a comparator (typically a β-lactam). The in-hospital mortality was 66% in the arm treated with polymyxin B versus 28% for those treated with a comparator, even when matched for mechanical ventilation and sepsis score suggesting polymyxin B was inferior. This was regardless of dosing regimens. A higher dose (≥200mg/day) of polymyxin B was found to be associated with reduced mortality but increased renal impairment in another
We do not recommend use of polymyxin B in the light of these results.

Combinations including colistin are more effective than monotherapy in treating *K. pneumoniae* carbapenemase (KPC) infections (See 7.18) \(^{316, 317}\).

Nephrotoxicity and neurotoxicity are the principal side effects associated with parenteral administration of polymyxins. The toxicity demonstrated in earlier studies was almost certainly related to lack of understanding of the drug's PK/PD and the use of inappropriate doses \(^{318}\). Studies now suggest that age, high doses, prolonged courses, concomitant vancomycin, hypoalbuminaemia and non-steroidal anti-inflammatory drugs, are independent risk factors for nephrotoxicity \(^{319, 320}\) and it is likely that other nephrotoxic drug are also associated. Monitoring renal function closely is essential for patients receiving colistin. Recent expert opinion suggests the risk benefit ratio should be carefully considered with strategies applied to reduce toxicity\(^ {321}\). There is no information on the dose-relationship of reversible neurotoxicity or encephalopathy: in a recent large paediatric series they occurred in 2% of patients \(^ {322}\).

There are gaps in our knowledge about these agents. Although they were developed some seventy years ago, they have only recently been used extensively. Much of the current knowledge is summarised in the Prato consensus report \(^ {323}\).

Dosing of intravenous colistin remains contentious. In adult cystic fibrosis (CF) patients, colistin is typically given at a standard dose of 2MU 8-hourly. However, evidence is emerging that higher-dose regimens may be more appropriate in the ICU setting (with therapeutic drug monitoring: to target a peak of 5-15mg/L and a trough of 2-6mg/L). A recent study of significant infections caused by a range of MDR GNB suggested that a loading dose of 9MU followed by 4.5MU twelve hourly reduced in renal impairment was effective (23/28 responses) and resulted in a reversible mild renal injury in only 5
Further clinical and PK/PD studies are required to confirm appropriate regimens including in relation to a loading dose, combination therapy and the need for monitoring. In the meantime European medicines agency guidance should be followed.

**Evidence**

Colistin is effective in treatment of infections caused by MDR GNB with low mortality at higher-than-previous, but well-controlled dosage.

Evidence level: 3

The role of loading doses of colistin, monitoring of serum levels and optimal combination therapy are inadequately researched.

Evidence level: 4

Use of aerosolized colistin dry powder has recently been accepted by NICE in cystic fibrosis.

Evidence level: 3

Use of aerosolized colistin dry powder in ventilator-associated pneumonia as an addition to intravenous chemotherapy appears useful.

Evidence level: 3

The dose-relationship of colistin nephrotoxicity and the rarer neurotoxicity and encephalopathy, require investigation.

Evidence level: 4

**Recommendations**

Accepted manuscript
- Reserve intravenous polymyxins for infections due to susceptible multi-resistant strains and preferably used in combination with other agents.

Grading: Conditional recommendation for

- Give careful consideration to use of higher dosage regimens in critically ill patients.

Grading: Conditional recommendation for

- Closely monitor renal function especially in the elderly, those receiving high intravenous doses for prolonged periods and those on concomitant nephrotoxic agents e.g. aminoglycosides.

Grading: Strong recommendation for

- Reconsider use of polymyxins in selective digestive decontamination regimens as these agents are now important last therapeutic options against carbapenemase-producing Enterobacteriaceae and are more threatened by resistance than previously appreciated.

Grading: Good practice point

- Need research on optimal rapid and practical methods of susceptibility testing outside intrinsically resistant groups such as Proteeeae and Serratia spp.

Grading: Recommendations for research

- Aerosolised colistin dry powder should be used in cystic fibrosis according to NICE guidelines. Use in combination in ventilator-associated pneumonia may be considered pending further trials without methodological flaws.

Grading: Conditional recommendation for
7.14 Fluoroquinolones

Fluoroquinolones suppress susceptible Enterobacteriaceae in the intestinal flora and also select for quinolone-resistant MDR GNB\textsuperscript{250,131}. Such suppression has been used in neutropaenic patients alone or with colistin\textsuperscript{269}. The continued efficacy of this combination in suppression and non-selection of resistance to either agent needs re-establishing, with the increasing recognition of colistin resistance which may well emerge alongside existing quinolone-resistance. Prophylaxis with quinolones alone in neutropenia against susceptible bacteraemia seems effective even when quinolone-resistance levels in the treated population reach a high level. Trials of withdrawing prophylaxis have been reported and show problematic increases in Gram-negative bacteraemia (See 6.5.)

Fluoroquinolones (intravenous and oral) may be suitable for complicated urinary tract infections due to ESBL-producing Enterobacteriaceae if there is no resistance \textit{in vitro}; however most ESBL-producing strains in the UK are resistant to fluoroquinolones including ciprofloxacin and levofloxacin. Furthermore quinolone resistance without ESBL production is now frequent, particularly in the multiple resistant if not MDR \textit{E. coli} ST131\textsuperscript{89}. Newer quinolones in development are unlikely to provide substantial additional benefits over ciprofloxacin for infections due to Gram-negative pathogens.

Three observational clinical studies have assessed the relative merits of quinolones and carbapenems for serious infections due to ESBL-producing organisms\textsuperscript{181,325,326}. Two of these found that carbapenems were superior to quinolones, although most strains were quinolone susceptible, whereas one study found equivalent effectiveness.

Fluoroquinolones have been used to treat infections caused by \textit{S. maltophilia}; however resistance is not uncommon so combination with one or more of: trimethoprim/sulfamethoxazole, ceftazidime, or tigecycline has been proposed\textsuperscript{327}.
These combinations have not been shown to offer any advantages over trimethoprim/sulfamethoxazole alone.

A wide range of resistance mechanisms exist: high-level resistance almost always involves mutations in the genes encoding subunits of the target-enzymes, DNA gyrase and topoisomerase 1V ($gyrA$ and $parC$ respectively), but reduced susceptibility can arise from plasmid-acquired genes e.g. $aac(6')-1b-cr$, $oqxAB$, $qnrA$, etc. or via up-regulation of outer-membrane efflux pumps and porin loss.

Evidences

Quinolones are effective in treatment of complicated urinary tract infection caused by susceptible ESBL-producing Gram-negative bacteria, but resistance is common limiting their usefulness.

Evidence level: 2+

Recommendations

- Could use orally to treat UTI caused by MDR GNB that are susceptible

Grading: Conditional recommendation for

7.15 Tigecycline and eravacycline

Tigecycline is a semisynthetic glycy glycine derivative of minocycline and like other tetracyclines is bacteriostatic. The main determinant of acquired plasmid-mediated, resistance to older tetracyclines in Gram-negative bacteria, namely active efflux by Tet pumps is overcome by steric hindrance by a large substituent group. Tigecycline has in vitro activity against most Enterobacteriaceae except Proteaeae i.e. Proteus spp., Providencia spp. and Morganella morganii. MICs for A. baumannii (including many carbapenem resistant strains) and S. maltophilia are low (mostly 0.25-2mg./L) but,
there are no break points or convincing efficacy studies. In common with other
tetracyclines, tigecycline lacks useful activity against \textit{P. aeruginosa}. Tigecycline is
vulnerable to the chromosomal resistance–nodulation–cell division (RND) multi-drug
efflux pumps, including \textit{MexXY–OprM} of \textit{P. aeruginosa}, and the AcrAB pump found in
\textit{Proteus mirabilis} which explains the intrinsic resistance of these species \cite{330,331}.

Whilst tigecycline-resistant isolates of Enterobacteriaceae have been described from
treatment naïve patients, another potential problem is the development of resistance
during treatment of infections with Enterobacteriaceae and \textit{Acinetobacter spp.} by the
mutational up-regulation of \textit{RND} pumps, but the frequency is unclear particularly when
used in combination \cite{332-336}. Use of tigecycline is an independent predictor of emergence
of tigecycline resistance when treating multi-resistant \textit{K. pneumoniae} infection \cite{337}.
Further studies are required, possibly including different dosing regimens and in
combination with other agents. Tigecycline has a potential to favour superinfections by
\textit{P. aeruginosa}, \textit{Proteae} \cite{338} and sometimes \textit{Klebsiella spp} \cite{339,337}; again, these aspects
require further investigation.

Subject to the earlier caveat about the lack of breakpoints, tigecycline has \textit{in vitro}
activity against \textit{S. maltophilia}, and susceptibility rates of >87\% have been reported \cite{340}.
However there is little clinical experience with the drug in treating infections caused by
\textit{this organism}.

Intravenous tigecycline is licensed for the treatment of complicated skin and soft tissue
infections and complicated intra-abdominal infections \cite{341,342}. However, the US FDA
issued a warning describing an increased mortality risk with its use when compared
with other drugs \cite{343,344}. The highest risk was in patients treated for ventilator-associated
pneumonia, which was not a licensed indication. However even in FDA approved uses
there was a higher risk of death among patients given tigecycline compared with those
given other antibacterial drugs. There are no RCTs comparing tigecycline with polymyxins, fosfomycin, sulbactam and other antibiotics against infections due to MDR GNB, alone or in combinations. Several meta-analyses examine the efficacy and safety of tigecycline in general (not just against MDR GNB) and these reported conflicting findings. One very recent analysis reviews the earlier studies and includes a number of new trials. Clinical success rates were lower than comparator for hospital-acquired pneumonia and diabetic foot infection, with increased gastrointestinal adverse events and higher all-cause mortality probably due to reduced efficacy.

Further work on tigecycline is needed, as its efficacy in ventilator associated pneumonia might be improved using higher doses (i.e. 200 mg initial and then 100 mg twice daily): an increase in adverse events was not seen with this regimen. Tigecycline in combination with other antibiotics (e.g. carbapenems and polymyxins) is a potentially valuable approach for infections caused by carbapenemase-producing Klebsiella spp., as shown by Tumbarello et al. (2012). In this retrospective cohort study largely of infections due to strains with KPC-3 carbapenemase 9/19 patients survived on tigecycline monotherapy, 0/11 on colistin monotherapy and 16/23 with tigecycline and colistin combinations. Two comparisons of monotherapy and combination therapy for infections with carbapenemase-producing Klebsiella spp. give further survival data on monotherapy: survival was respectively 71/116 for tigecycline and 70/132 for colistin and 16/27 for tigecycline and 12/22 for colistin.

Whilst the in vitro data supports use of tigecycline in respiratory infection there is poor correlation between the laboratory results and clinical outcome.

Eravacycline is a novel intravenous fluorocycline with a similar spectrum to tigecycline. It showed non-inferiority to ertapenem in a Phase 3 trial of complicated intra-
abdominal infection but failed to show non-inferiority to levofloxacin in an iv/oral switch Phase 3 trial of complicated UTI.\textsuperscript{354-356}

Evidence

The role of tigecycline remains uncertain in the treatment of infections due to MDR GNB.

Evidence level: 1-

Recommendations

- Could use tigecycline in combination in the treatment of multi-resistant soft tissue and intra-abdominal infections

    Grading: Conditional recommendation for

- Use alone in hospital-acquired respiratory infections is unlicensed and not advised with licensed dosing as outcomes are not clearly satisfactory in Acinetobacter and MDR GNB infections.

    Grading: Conditional recommendation against

- Use in combinations in hospital-acquired respiratory infections: precise combinations depend on the antibiotic-susceptibility of the MDR GNB causing the infection.

    Grading: Recommendation for research and possibly conditional recommendation for use restricted to trials

- Use higher-than licensed dosing such as 100mg twice daily for infections due to MDR GNB in critical care
Investigate if higher dosing counters the unexpectedly high mortality seen even in infections due to strains apparently susceptible in vitro.

Grading: Recommendation for research and possibly conditional recommendation for use restricted to trials

7.16 Fosfomycin

Fosfomycin, a strongly hydrophilic phosphonic acid (unrelated to aminoglycoside or macrolide antibiotics), inhibits the addition of phosphoenol-pyruvate to N-acetyl-glucosamine in synthesis of the bacterial cell wall. Fosfomycin MICs of *E. coli* vary from 1-4mg/L; those for *Klebsiella spp.* are higher at 2-64mg/L. EUCAST breakpoints for both IV and oral formulations are S ≤32mg/L, R >32mg/L, available for *E. coli* only.

*Morganella morganii* and *Bacteroides spp.* are inherently resistant and activity against *P. aeruginosa* is controversial, particularly in combination, although MICs=>128mg/L. The drug is otherwise very broad in its spectrum. Fosfomycin was active against 72% of Enterobacteriaceae resistant to carbapenems in a German study. *In vitro* testing with discs required the addition of Glucose-6-phosphate to the disc. In this study there were 22% major discrepancies between agar dilution in medium containing glucose-6-phosphate and disc or E-test testing and it is not clear if glucose-6-phosphate was present in discs and MIC gradient strips, an area for quality control development. There are similarly no published details on the reliability of automated susceptibility testing methods.

Fosfomycin trometamol is used as an oral treatment for patients with uncomplicated lower UTI due to fosfomycin-susceptible organisms resistant to first line agents. At the conventional dosage of 3g on a single occasion this oral formulation gives an adequate urinary concentration for 2 days (see 9.3.). An earlier oral product was a calcium salt.
only 30-40% of which was absorbed: this gave peak plasma levels of 7 to 9mg/L 4 hours
after a 3g dose. The trometamol salt which replaced this is better absorbed (60%
bioavailable) reaching peak plasma levels of 32mg/L 2 hours after a 3g dose."

Experience with IV fosfomycin disodium (not a trometamol formulation) is limited in
the UK where it has only recently been introduced specifically for treatment of infection
with multi-resistant bacteria. It has been more widely used elsewhere in Europe. The
intravenous sodium salt reaches levels of 25mg/L after a 1G dose. A very early single
open comparison of 38 patients with acute pyelonephritis showed that 7 days of
intravenous fosfomycin 2g six hourly achieved only a 44% response rate 358; the authors
therefore concluded the drug had no role in pyelonephritis: the oral trometamol salt has
never been examined for pyelonephritis. Intravenous dosage with MDR GNB is now
usually at 24g/day in 3 divided doses but dosage reduction is needed in renal
impairment as the drug is exclusively renally excreted, unchanged. The formulation has
a high sodium load and the most frequently encountered side effect is hypokalaemia
(26% patients) 359. Fosfomycin exhibits excellent penetration into tissue after an
intravenous dose as it is a small (138 Da), molecule with negligible protein binding; it
also has a long serum half-life of between 4 – 8 hours 360.

A prospective salvage study of 11 ICU patients with serious infections caused by
carbapenem-resistant K. pneumoniae reported an all-cause mortality of 2/11, although
analysis of the claimed successes is complicated because 6 patients were also treated
with colistin and 3 with gentamicin 361. A larger outcome study of 48 patients (mainly
VAP) infected with KPC-producing K. pneumoniae and to a lesser extent, VIM-producing
P. aeruginosa reported clinical success when fosfomycin was used mainly in
combination with colistin or tigecycline in 54.2% patients and 28-day all-cause
mortality of 37.5% 362. Of 15 patients with colistin-, tigecycline- aminoglycoside- and
carbapenem- resistant KPC-producing Klebsiella infection (one with an additional
carbapenem-resistant P. aeruginosa) 9 responded to fosfomycin combinations and in 8 microbiological eradication was achieved.

The use of intravenous fosfomycin has been reviewed extensively. Clinical cure was described in 1242 of 1529 (81.2%) of patients overall (for both Gram-positive and Gram-negative pathogens)\(^ {363}\). Most of the Gram-negative infections in this series were due to P. aeruginosa, (which most would regard as resistant), but also included infections due to Enterobacter spp., Klebsiella spp., E. coli, Proteus spp. and S. typhi. Most patients also received concomitant antibiotics, so again interpretation is difficult. A wide variety of infections were treated and fosfomycin was well tolerated. Despite in vitro resistance to fosfomycin, most patients with infections caused by P. aeruginosa improved although this may reflect concomitant antibiotics.

Further detailed studies of the parenteral form used alone in single indications (such as urinary tract infection, and ventilator-associated pneumonia are required to establish its relative efficacy and usefulness for specific MDR GNB. Similarly in combination therapy comparisons of specific combinations are required.

Evidence

Further details and regimens for the oral formulation are given in 9.6.3.

The parenteral formulation may be a valuable treatment alternative for infections due to MDR GNB including carbapenemase- and MBL-producing strains. However, further detailed comparative trial experience is necessary to determine its optimal use.

Evidence level: 3

Recommendations
Consider parenteral fosfomycin, probably in combination, as part of salvage treatment for susceptible MDR GNB: clear indications for use are not yet established.

Grading: Conditional recommendation for.

Need comparative clinical trials to establish optimal indications for, and optimal use of, parenteral fosfomycin, a potential drug of last resort against MDR GNB.

Grading: Recommendation for research and possibly conditional recommendation for use restricted to trials.

### 7.17 Trimethoprim/sulfamethoxazole

Trimethoprim/sulfamethoxazole (available as intravenous and oral formulations) has *in vitro* activity against *Stenotrophomonas maltophilia* and some less frequently encountered non-fermenting Gram-negative bacilli (e.g. *Achromobacter* spp., *Alcaligenes* spp., *Burkholderia* spp., *Chryseobacterium* spp. and *Elizabethkingia* spp.). These species have inherent resistance to most other antibiotics and often produce MBLs. *Stenotrophomonas* sp. typically have similar percentage susceptibility at the CLSI breakpoint to sulphonamides alone and trimethoprim/sulfamethoxazole but are resistant to trimethoprim alone. The combination has greater in-vitro potency than either trimethoprim or sulfamethoxazole. A similar comment applies to *Achromobacter* spp. and with few exceptions to *Alcaligenes* spp. *Chryseobacterium* spp. and *Elizabethkingia* spp.

These genera are susceptible to trimethoprim and more strains of these genera and *Burkholderia* spp. are more susceptible to trimethoprim/sulfamethoxazole than either component alone. The clinical use of sulphonamides alone against non-fermenters has not been explored and the combination of trimethoprim/sulfamethoxazole is usually used in *S. maltophilia* infections and for
simplicity, against those due to these other unusual species. Problems occur with disc
susceptibility testing of *S. maltophilia* and there are few data on the performance of
automated susceptibility systems. Trailing endpoints are frequent and results vary with,
the temperature of incubation and the susceptibility testing medium used. Occasional
resistance to trimethoprim/sulfamethoxazole is not well understood in these non-
fermenters but resistance to trimethoprim-sulfamethoxazole caused via the *sulI* gene
has been described repeatedly in *S. maltophilia*\(^{365}\). A recent systematic review suggested
that some strains of *Acinetobacter* *spp.* are susceptible to trimethoprim-
sulfamethoxazole and that use against this genus can be guided by *in vitro* testing\(^{366}\).
However over half the UK strains of *A. baumannii* show high level resistance \(^{364}\).

**Evidence**

Trimethoprim/sulfamethoxazole has wide *in vitro* activity against *S. maltophilia*,
*Achromobacter* *spp.*, *Alcaligenes* *spp.*, *Burkholderia* *spp.*, *Chryeobacterium* *spp.* and
*Elizabethkingia* *spp.* Susceptibility testing methods for these organisms are not well
established but some *S. maltophilia* have resistance to trimethoprim and
sulfamethoxazole. Carbapenem resistance is inherent to most of these species.

Evidence level: 3

**Recommendations**

- Use in treatment of infections due to susceptible *S. maltophilia* and consider in
infections due to *Achromobacter* *spp.*, *Alcaligenes* *spp.*, *Burkholderia* *spp.,
*Chryeobacterium* *spp.* and *Elizabethkingia* *spp.*

Grading: Conditional recommendation for
7.18 Intravenous combination therapy for infections due to carbapenemase-producers

Although results of RCTs will be available, most of the current evidence for advantage of combination therapy for carbapenem-resistant infections derives from observational studies and reports mainly focus on severely-ill patients or those where the pathogen has reduced sensitivity to colistin. An international working group report recommended combination including a carbapenem as optimal treatment but only in settings where NDM carbapenemases are infrequent. However, retrospective studies are liable to bias in that investigators have no control over antibiotic use.

Different studies and reviews of combination therapy have reached contradictory conclusions. One systematic review identified that evidence for combination treatment was poor quality and inherently biased, being based on small observational studies with heterogeneity of i) antibiotic choice and activity against responsible pathogens, ii) antibiotic dosage and iii) severity of illness. These authors concluded that any benefit in outcome between monotherapy with colistin and combination of colistin with other agents (aminoglycoside, tigecycline, carbapenem or rifampicin) was uncertain. There were methodological problems in the studies reviewed. Another systematic review which lacked quality assessments likewise found only observational studies with marked heterogeneity, and suggested no proven benefit in terms of mortality between combination treatment and monotherapy except for three more homogenous studies exclusively of bacteraemias due to KPC-producing Klebsiella spp. in critically ill patients which are worth detailed consideration.

Firstly, Tumbarello et al. (2012) in a 3-centre retrospective cohort study found 16/23 patients survived with tigecycline and colistin combinations and 12/14 with colistin-tigecycline-carbapenem combinations compared with 11/22 with colistin monotherapy and 10/19 with tigecycline monotherapy. Secondly, Qureshi et al. (2012) in a 2-
centre retrospective cohort study showed that 3/7 receiving polymyxin monotherapy, 1/5 receiving tigecycline monotherapy, 2/4 receiving carbapenem monotherapy and 2/3 other antibiotics as monotherapy survived 28 days compared with 5/6 receiving colistin combinations and 6/6 receiving tigecycline combinations. Thirdly, Zarkotou et al (2011) noted 3/7 survivals with colistin, 3/5 with tigecycline and 0/1 on carbapenem, all as monotherapy, compared with 9/9 receiving combined tigecycline and colistin, 3/3 receiving tigecycline and carbapenems and 8/8 among those treated with other combinations. Two studies of bacteraemias involving VIM-1-producers considered in this review produced even less interpretable results. A third systematic review of polymyxin treatment found mortality at 30 days was lower in patients given combination treatment. A 2017 systematic review and meta-analysis favours combination use of polymyxins.

Given this background, conclusions from further individual on-RCT studies must be interpreted with caution, but some support combination treatment. A larger retrospective cohort study of 661 infections caused by KPC-carbapenemase-producing strains of K. pneumoniae reported improved survival in patients treated with two or more active drugs versus those given monotherapy. Mortality at 14 days in bacteraemias with an unknown or non-urinary source was 52.8% with monotherapy and 34.1% with combination treatment. A similar result with 49.1% and 24.8% mortality respectively was seen with lower respiratory tract infection. There was no significant difference in bacteraemias from a known urinary source. Overall death rates on monotherapy were 62/132 (47%) with colistin, 45/116 (39%) with tigecycline, and 28/70 (40%) with gentamicin. With two drug therapy mortality was 38/134 (28%) and with three drug therapy 67/217 (31%). Only the use of meropenem in a combination produced a statistically significant improvement to 54/205 (26%). Use of meropenem was associated with lower mortality only if the MIC ≤8 mg/L as was the
case for 37% of the isolates. Colistin resistance was significantly associated with increased mortality. Overall combinations including tigecycline, colistin and meropenem were associated with the lowest mortality (12.5% OR 0.11 95% CI 0.02-0.69). Epidemiologically overall colistin, tigecycline and gentamicin resistance rates were 11%, 9% and 6% in 2010 but by 2014 were 21%, 27% and 25%.

A further review including some previously reviewed studies, suggested superiority of combination- over mono-therapy with mortality rates of 27.4% vs. 38.7% respectively. Again carbapenem-containing regimens had the lowest mortality (18.8%) and this was associated with isolates that were not resistant by the EUCAST breakpoint. Similar findings were reported in a retrospective observational study of 205 bacteraemias caused by carbapenemase-producing K. pneumoniae. Combination therapy was associated with a lower mortality rate of 27% compared with 44% for monotherapy, 11/27 with tigecycline, 10/22 with colistin, and 7/12 with carbapenems. The difference in mortality was most marked in the more severe cases. Furthermore, mortality with a carbapenem-containing combination was 19.3% (6/31) compared with 30.6% (22/72) without a carbapenem (5/16 in those treated with tigecycline and colistin alone).

Mortality on carbapenem-containing regimens in this study was lower only if the carbapenem MIC was <=8 mg/L. The authors comment that 40% of isolates with MICs by Etest <=8 were found resistant by automated machines. These studies suggest i) that KPC-carbapenemase –producing Klebsiella spp. commonly appear meropenem susceptible in vitro and ii) that treatment combinations containing conventionally-dosed carbapenems are advisable in such cases with lower MICs.

Much higher doses of meropenem by continuous infusion can also be used (See 7.1.). This extends the MIC range of strains that can be treated. Continuous infusion therapy of meropenem with doses up to 13.2G daily with levels optimised by therapeutic drug monitoring when used in combinations (mainly with colistin and tigecycline), were
associated with 73% clinical cures in patients with KPC-producing *K. pneumoniae* with MIC >16<64 mg/L. These are better outcomes in treatment of more-resistant KPC-producing Klebsiella than apparent in earlier studies of these more resistant KPC-producing Klebsiella. Direct comparisons have not been made including comparison with high-dose continuous infusion meropenem alone. The application of this approach to other carbapenem-resistant isolates with MICs within the attainable range has not been assessed.

Anecdotal reports suggest double carbapenem combinations of ertapenem plus either meropenem or doripenem can be effective as last resort treatment for infections due to *K. pneumoniae* producing KPC carbapenemase but not those with NDM enzymes. This is perhaps because ertapenem binds tightly to the KPC enzyme, acting as an inhibitory substrate and thereby protects the meropenem or doripenem. In cases where the *Klebsiella* *spp.* strain was resistant to colistin and carbapenems, the use of gentamicin in combination with various agents was independently associated with reduced mortality in a retrospective cohort study. However this was in the epidemiological context of a clonal *K. pneumoniae* ST512 (CC258) lineage with a KPC enzyme. This lineage commonly has the AAC (6')-1b enzyme; which confers resistance to amikacin but largely spares gentamicin; it is unlikely to be true for isolates with NDM carbapenemases, which mostly have Arm A or Rmt ribosomal methyltransferases, conferring high level resistance to all standard aminoglycosides, including gentamicin and plazomicin. Plazomicin might have a future role with non-NDM-producing, gentamicin-resistant strains.

Evidence for efficacy of tigecycline in combination largely derives from observational studies but microbiological cure rates with monotherapy are lower than clinical cure rates and mortality rates are high. Pooled results from 5 observational studies
suggested a clinical response rate of 77% (567/733) for all patients and 81% (329/408) for tigecycline monotherapy in the treatment of complicated intra-abdominal infection. Another review of five observational studies of uncomplicated soft tissue and intra-abdominal infection with tigecycline similarly found monotherapy was effective. These studies contain no data on response by resistances present and studies were with the licensed dose of 50mg twice daily.

In an open label RCT of treatment of ventilator-associated or hospital-acquired pneumonia caused by multi-drug-resistant Acinetobacter spp. addition of rifampicin to colistin did not affect 30-day mortality or length of hospital stay, but was associated with a higher rate of microbiological eradication. A retrospective observational study of 251 blood-stream infections treated with colistin or, colistin-sulbactam, colistin-carbapenem or another colistin combination reached the similar conclusion that mortality was not affected but microbiological eradication was higher with combination treatment. Another observational study of 101 patients with MDR Acinetobacter infections did not show any improvement in mortality rates for combination therapy (e.g. colistin plus tigecycline or carbapenem plus tigecycline) over a single agent (usually colistin) but the group size in this study was small.

In the case of multi-drug-resistant Pseudomonas infections a prospective cohort study showed no outcome advantage in combination versus monotherapy. Combination therapy with aminoglycosides did not reduce the development of resistance.

Fosfomycin in combination with tigecycline or colistin was effective in 54% of 48 patients with infections with MDR GNB, some of which had Pseudomonas infection.

The recent introduction of ceftazidime/avibactam and the possibilities of using this in treatment may change the need to use combination treatment for some KPC or ceftazidime-resistant OXA-48 carbapenemase-producing strains.
Evidence

Two of four systematic reviews do not show a benefit of combination therapy over monotherapy.

Evidence level 2++

In infections with KPC-carbapenemase producing *Klebsiella spp.*, combination therapy including meropenem is associated with lower mortality than colistin monotherapy if the meropenem MIC is <8mg/L but this was not the case with strains with higher MICs unless continuous infusion therapy with higher than licensed doses was used (see 7.1).

Combinations with other agents such as tigecycline or an aminoglycosides to which carbapenemase-producing strains are susceptible also seem advantageous but only the expected results of a new RCT will resolve this.

Evidence Level 3

Paul et al (2014)\(^{369}\) detail the hazards of bias in favour of combination therapy that arise without an RCT. Data from a subset with bacteraemia with *Klebsiella spp.* Producing KPC-carbapenemases in the second systematic review performed by Falagas et al (2014)\(^{370}\) suggests that in treatment of carbapenem-resistant Enterobacteriaceae infection, colistin used in combination with other agents is associated with a lower mortality than colistin alone and this is also a finding in the review of Ni et al (2015)\(^{373}\).

Evidence level: 1+

The evidence that tigecycline combinations, including other antibiotics active against Enterobacteriaceae, are more effective than tigecycline alone in intra-abdominal infections is poor

Evidence level: 1-
Ertapenem in combination with meropenem may be effective as salvage therapy for infections with KPC-carbapenemase-producers but the evidence is very weak.

Evidence level: 3

In treatment of multi-drug resistant Acinetobacter respiratory infections, addition of rifampicin to colistin does not affect 30 day mortality.

Evidence level: 1+

**Recommendations**

- Use colistin with meropenem to treat susceptible KPC-producing *Klebsiella* infection if the meropenem MIC is $\leq 8\text{mg/L}$ and consider higher meropenem dose by continuous infusion if the MIC is $>8$ and $\leq 32\text{mg/L}$

Grading: Conditional recommendation for

- Consider colistin with aminoglycosides or tigecycline in infections with strains producing other carbapenemases or KPC strains which are susceptible to these agents but resistant to meropenem

Grading: Conditional recommendation for

- Consider if ceftazidime/avibactam should be used with a carbapenem or colistin to treat infections with KPC3-producers based on latest evidence at the time of use.

Grading: recommendation for research and possibly conditional recommendation for use restricted to trials.
8 Oral agents for secondary/tertiary care treatment

8.1 Mecillinam and Pivmecillinam

Pivmecillinam (the oral form of mecillinam) can be considered alone as oral therapy for lower UTI caused by AmpC producing Enterobacteriaceae. The antibiotic is not active against carbapenemase producers. It has been suggested as active against ESBL-producing *E. coli*. Patients with infections with such strains referred from the community for intravenous treatment with carbapenems might be considered for oral follow-on therapy with pivmecillinam alone for UTI because of mecillinam’s apparent activity *in vitro*. However, additional measures are desirable and this oral treatment is dealt with under community use. (See 9.4 for more detail). Patients should be carefully monitored both clinically and microbiologically if pivmecillinam is prescribed alone in hospital for infections involving ESBL-producers as treatment failure is a risk.

8.2 Cefixime and oral cephalosporins

Cefixime is an oral third-generation cephalosporin, which has been used as an oral switch for patients with pyelonephritis. Among uropathogenic Enterobacteriaceae, it is not active alone against ESBL-producing *E. coli* because of their multiple resistances including quinolones but is useful if ESBL-producing organisms or CPE are not present. Cefixime could be used in combination with co-amoxiclav against ESBL-producing Enterobacteriaceae as supported by *in vitro* data. Data from transconjugant *E. coli* further suggests cefixime plus clavulanate is effective against strains producing CTX-M-15 enzyme which has higher cefixime MICs than strains producing CTX-M-9 enzyme. Other oral cephalosporins including cefdinir, cefpodoxime also showed synergy with clavulanate whereas sulbactam was less effective as a potentiator. Cefixime, with or without clavulanate, was not active against AmpC-producing organisms nor would it be expected to be active against CPE.
Consequently cefixime-co-amoxiclav combinations should not be used against cephalosporin-resistant organisms without tests to distinguish AmpC and ESBL production. No clinical trials of cefixime together with clavulanate or amoxicillin/clavulanate against ESBL-producing E. coli have been published. Cefixime is detectable in faeces after administration. Other cephalosporins e.g. cephalexin which are fully absorbed, are not detectable in faeces and less frequently provoke C. difficile may be better partners for clavulanate, although in vitro data to support this combination are lacking. Synergy in vitro between cephalosporins and mecillinam because of their different target penicillin-binding proteins is likely and synergy of cephalexin with fosfomycin (earlier known as alafosfalin or fosfonomycin), another cell-wall active antibiotic is also recorded.

Evidence

Cefixime with clavulanate, which is not available commercially, in vitro, has reliable activity against ESBL-producing E. coli and Klebsiella spp. (not Enterobacter spp. where AmpC will cause resistance). Cefixime is not useful alone against MDR GNB and no clinical studies with oral cephalosporins and clavulanate or amoxicillin/clavulanate have been published.

Evidence level: 3

Recommendations

- Do not use cefixime or other oral cephalosporins alone for treating infections caused by ESBL-, AmpC- or carbapenemase-producing Enterobacteriaceae.

Grading: Conditional recommendation against
Oral cephalosporins need clinical trials with clavulanate (alone or with amoxicillin) against ESBL-producing *E. coli* UTI.

Grading: Recommendation for research and possibly conditional recommendation for use restricted to trials

**8.3 What are the recommended antibiotics for community care, including care homes?**

Most MDR GNB infections encountered in the community involve the urinary tract. As described earlier, ESBL-producing isolates of Enterobacteriaceae are a significant and growing problem, whereas there are few community infections in the UK involving CPE. There are no published randomized controlled trials of antibiotic treatment of UTIs due to ESBL-producing organisms in the community or care homes. Recommendations must rely on observational studies of ESBL-producing GNB, or randomized controlled trials of effectiveness of antibiotics against UTIs caused by GNB lacking ESBLs.

**8.4 What are the risk factors for patients with UTIs caused by MDR GNB in the UK?**

In order to help the assessment of patients we review risk factors for MDR GNB and suitable oral agents for acute uncomplicated and complicated UTI. Prospective and retrospective epidemiological studies identified several risk factors for carriage of ESBL-producing *E. coli*[^99]^[136][184][391-393][394][395]. Patients are at increased risk if they have:

- recurrent UTI
- persistent urinary symptoms after an initial antibiotic,
- over 7 days hospital admission in the last 6 months,
- residence in a care home
• recent travel and especially healthcare in a country with increased antimicrobial resistance. Details of countries where prevalence is currently high are given in 8.5.

• previously known UTI (within a year) caused by bacteria resistant to amoxicillin-clavulanate, cephalosporins or quinolone or recent treatment with these agents.

There is no UK data validating an Italian scoring system devised and tested in 2009 for carriage of ESBL-producing bacteria on admission to hospital or incorporating information on travel, overseas healthcare in the previous 2 years or migration. The Italian scoring system identifies risk based on hospitalisation within the previous 12 months OR 5.69 (95% CI 2.94-10.99), transfer from another healthcare facility OR 5.61 (95% CI 1.65-19.08), Charlson comorbidity score >4 OR 3.80 (95% CI 1.90-7.59), β-lactam or fluoroquinolone prescription within the previous 3 months OR 3.68 (95% CI 1.96-6.91), recent urinary catheterization OR 3.52 (95% CI 1.96-6.91) and age >70 years OR 3.20 (95% CI 1.79-5.70). This model of risk factors has been re-assessed in the US to see if it can be used to realistically restrict the need for carbapenem treatment to an identifiable high risk subgroup. In the US evaluation, risk factors for community-onset clinical infection involving MDR GNB diagnosed within 48 h. of admission were: hospitalization OR 2.63 (95% CI 1.32-5.41), inter-hospital transfer OR 5.30 (95% CI 2.67-10.71), urinary catheterization OR 6.89 (95% CI 3.62-13.38), β-lactam or quinolone prescription OR 3.47 (95% CI 1.91-6.41) and additionally immunosuppression in the preceding 3 months OR 2.34 (95% CI 1.14-4.8). Age over 70 was not a risk factor but age was not examined as a continuous variable. In this model, the sensitivity and specificity were >=94% and <=65% for scores of 3 or below and <=58% and >=95% for scores of 8 or above. Urinary catheterization was also a risk factor in a Spanish study. A further paired US retrospective case-control studies compared infections with CTX-M ESBL.
producing *E. coli* infections with *E. coli* lacking CTX-M enzymes to uninfected controls; carbapenemase-producers were excluded. Patients with infections with CTX-M-producers were more likely to be male, have dementia or dependency, have higher Charlson median scores, receive H2 antagonists, and have exposure to health-care settings. Recent antibiotics did not differ between the two groups except that trimethoprim/sulfamethoxazole use was commoner in the non CTX-M-producing group. Exposure to immunosuppressives was also commoner in the CTX-M group. A similar 75-77% of strains were present within 48 h. of admission. When patients with strains producing CTX-M-ESBLs were compared with controls, the former had a higher incidence of comorbidity (Charlson score =>5), and were more often resident in nursing homes with greater exposure to healthcare and more indwelling urinary catheters. They were more likely to be receiving H2 antagonists or proton pump inhibitors and to have exposure to oxyimino cephalosporins within the last 3 months.

**Evidence**

Quoted rates of resistance in the community are biased to an unknown extent by infection occurring shortly after hospital discharge, care home cross-infection, an excess of treatment failures represented in the samples tested and an unknown proportion of patients with risk factors and recent antibiotic use.

Evidence level: 2-

UK surveillance suggests MDR GNB remain uncommon in community UTIs with few carbapenemase producers.

Evidence level: 3

Empirical antibiotic choice for lower urinary tract infection can be guided by the presence of established risk factors for a multi-resistant organism.
Predictive models have been established in Italy and the USA for ESBL-producing *E. coli* infections and colonisation on admission to hospital but these have not been validated in the UK nor do they consider travel-, migration-, or household-associated risks.

**Recommendations**

- In younger women with acute uncomplicated UTI, only consider MDR GNB in choosing empirical treatment if there are risk factors or recent foreign travel to countries where such strains are highly prevalent.

  Grading: Strong recommendation for

- If the defined risk factors for MDR GNB are present avoid cephalosporins, quinolones, trimethoprim and co-amoxiclav in treatment of lower UTIs unless the pathogens are confirmed to be susceptible.

  Grading: Strong recommendation against

- Building on previous work, predictive scoring should be developed in the UK for the presence of ESBL-producing *E. coli* in primary care and on admission to hospital to restrict the need to prescribe carbapenems and other antimicrobial agents generally active against ESBL-producing organisms.

  Grading: Strong recommendation for.
9 Which oral antibiotics are preferred for use in treating uncomplicated UTIs due to MDR GNB in the community?

9.1 Trimethoprim

Due to increasing resistance trimethoprim is no longer the suggested first-line empirical therapy for post menopausal women and older men in Public Health England guidance and nitrofurantoin is advised instead. In Wales trimethoprim remained until 2016 the suggested first-line empirical therapy for uncomplicated UTI in the community except for the elderly and for patients who have received antibiotics in the preceding 3 months. Following advice to decrease trimethoprim use, an 86% reduction in trimethoprim use was seen in a Swedish region (hospitals and community) from 2004-2006 with a compensatory increase in nitrofurantoin, pivmecillinam and ciprofloxacin use. This programme resulted in no overall change in trimethoprim resistance. Before the intervention trimethoprim resistance was more prevalent in *E. coli* phylogroups A, B1 and D than in phylogroup B2 strains, although rates were high in ST131 which belongs to phylogroup B2. There was a marked change after the intervention in the distribution of resistance between phylogroups and associated sequence types with an increase in the trimethoprim resistance in phylogroup B2 (including ST131) and a decrease in trimethoprim resistance in phylogroup A and B1 strains (which seldom cause extraintestinal infection) and to a lesser extent in phylogroup D. Trimethoprim resistance was associated with a change in prevalence of *dfrA1*. Resistance to other antibiotics, including those substituted for trimethoprim increased in phylogroup A and B1 strains. Amongst 273 urine isolates of *E. coli* collected in 2006 versus the same number collected in 2004, strains of ST69 (which includes the former clonal group A), ST12 and unusual strains became more prevalent increasing respectively from 4.8 to 8.1%, from 2.6 to 4.8% and from 42 to 51%. By contrast strains of ST131, ST127, and ST80 declined in prevalence from 4.8 to 2.2%, 8.1 to 3.7% and 5.1% to 1.1%. There
were statistically significant increases in trimethoprim resistance rates in the strains of ST131 and ST127. This would suggest that in types ST131 and ST127 susceptible strains were eliminated by the antibiotics substituted for trimethoprim (quinolones, pivmecillinam and nitrofurantoin) but because of resistance linkage trimethoprim resistance increased in these sequence types. Information is lacking on ST80. The increase in strains ST69 and ST12 suggests they may have been selected by the antibiotics substituted for trimethoprim but it is not clear which antibiotics would have this effect as these STs are usually only resistant to ampicillin and in the case of ST69 trimethoprim. In a structured survey of extraintestinal strains from US veterans in 2011 quinolone-resistant ST131 accounted for 78% of quinolone resistant strains which comprised 29% of reported strains overall. It accounted for 56% of trimethoprim resistant strains and 52% of quinolone and trimethoprim resistant strains. This suggests that quinolones have the potential to select against trimethoprim susceptible ST131 strains, decreasing in the Swedish intervention study the overall prevalence at that time but potentially selecting for later increased prevalence of the ST131. Thus, because of resistance linkage, community-wide change in use of a single antibiotic may unpredictably change the epidemiology and the prevalence of antibiotic resistance in more pathogenic phylogroups. It cannot be assumed that risk factors for multi-resistance, or the likelihood of success with an antibiotic in reinfection or recurrent infection will stay the same after abandonment of trimethoprim as a first line agent. This aspect of change needs urgent study.

Trimethoprim-resistant strains are much more frequently resistant to amoxicillin than trimethoprim-susceptible strains and this is a feature of ST69. Trimethoprim resistance rates in ESBL-producing *E. coli* in 2010 in the West Midlands were between 86% and 92% depending on whether the strain was not, or was, ST131. Ciprofloxacin resistance is also usual in these strains. Trimethoprim consequently is a poor choice for patients
with treatment failures on amoxicillin with, or without, clavulanate, cephalosporins or quinolones who require an urgent prescription before samples can be tested for antibiotic susceptibilities.

More generally, trimethoprim should not be used as empirical treatment for UTI if there are risk factors for an antibiotic resistant bacterium unless i) susceptibility has been confirmed in the previous month ii) there are no new risk factors for resistance, and iii) there have been no treatment failure with trimethoprim. In the absence of resistance, trimethoprim attains excellent bacteriological cure, two-weeks after completion of treatment, 94% of women using a 3-day course achieved bacteriological cure compared with 97% of those using a 10-day course (n = 135).

Evidence:

Trimethoprim use has not been explored as a risk factor for MDR GNB infection but resistance is common generally and very common in ESBL-producing bacteria. Trimethoprim is no longer recommended as a first line antibiotic choice for post menopausal women and older men with UTI and has little place in treatment of infection due to MDR GNB.

Evidence level: 3

3 day courses are almost as effective as longer courses in bacteriological cure of susceptible infections.

Evidence level: 1+

Recommendations:

- Do not use trimethoprim in treating MDR GNB or treatment failures with other agents unless in vitro-susceptibility has been demonstrated.
Grading: Strong recommendation against

- Do not use trimethoprim to treat lower UTIs as a first line agent if ≥ 50 years old.

Only consider use if there are no risk factors for resistance, or confirmed, in vitro susceptibility

Grading: Conditional recommendation against

9.2 Nitrofurantoin

Nitrofurantoin is widely used for acute uncomplicated UTI in the community, and is now the recommended first line treatment in England. It attains only low concentrations in renal tissue and the blood stream and should not be used if pyelonephritis or bacteraemia is suspected: treatment may fail if used for ascending infection. Nitrofurantoin resistance is inherent in *Proteus spp.*, *Morganella morganii*, *Providencia spp.* and *Serratia spp.* and the drug may not be effective in the alkaline urine produced by urease-producing bacteria such as these and possibly *Staph saprophyticus*, which is apparently susceptible in vitro but also produces large amounts of urease. Nitrofurantoin resistance is very common in CPE.

In early studies nitrofurantoin had a minimal effect on rectal flora and a recent metagenomics study supports this. Resistant strains of *E. coli* and increased numbers of Proteaeae may be detected in the faecal flora but UTIs breaking through prophylaxis in recurrent infection are usually due to strains that remain susceptible unlike the situation with trimethoprim. Recurrent UTIs after nitrofurantoin treatment of ESBL-producing *E. coli* may reflect relapse or recurrent infection arising from persistent carriage in the gastrointestinal flora: these possibilities cannot easily be distinguished. Frequent recurrence of UTI due to ESBL strains may justify using an alternative antibiotic regimen such as fosfomycin, or amoxicillin-
clavulanate with pivmecillinam, with a greater theoretical chance of changing the gastrointestinal flora, which may act as the source for reinfection.

If a patient has a reduced glomerular filtration rate, urinary concentrations of nitrofurantoin may be too low to be effective. eGFR frequently declines with age, on average by between 6 and 9ml/min/1.73m² per decade. Around half of women over 75 years and men over 85 years have an eGFR under 60mL/min/1.73m² which used to be the lower limit for use of nitrofurantoin \textsuperscript{401}. In a cohort study of lower UTI in 21,317 women treated with nitrofurantoin and 7926 treated with trimethoprim, there was no greater risk of nitrofurantoin treatment failure in patients with creatinine clearance of 30-50ml/min; however the risk of pulmonary adverse events was significantly increased with creatinine clearance <50ml/min (HR 4.1, 95% of CI.31-13.09) \textsuperscript{406}. In 2014, and in the context of increasing antibiotic resistance to trimethoprim the UK, the Medicine and Healthcare Regulatory Agency reviewed the evidence for use of nitrofurantoin in reduced renal function\textsuperscript{407}. They concluded on evidence \textsuperscript{401, 406} that the eGFR below which nitrofurantoin should not be used could be lowered to 45 ml/min/1.73m². The MHRA further stated that a short course (3 to 7 days) may be used with caution in patients with an eGFR of 30 to 44 ml/min/1.73m²; but only advocates prescribing in such patients for lower UTIs with suspected or proven multi-drug resistant pathogens when the benefits of nitrofurantoin are considered to outweigh the risks of side effects. Long term or repeated courses of nitrofurantoin are associated with severe pulmonary fibrosis \textsuperscript{408}. Nevertheless 219 courses of prophylaxis for one year for recurrent UTI in normal patients were not associated with a single case so this unwanted effect may be rare under controlled conditions where the drug is very effective \textsuperscript{405}. Nitrofurantoin is poorly tolerated by some patients, but the modified release form has fewer side effects \textsuperscript{409}. When used in this formulation an open RCT over 20 years ago (n = 538) found that nitrofurantoin had equivalent clinical cure rates to...
trimethoprim/sulfamethoxazole and trimethoprim (both given for 7 days) in a group of patients with acute uncomplicated lower UTI\textsuperscript{409}. The rate of gastrointestinal adverse effects was similar between groups (7-8\%). At this time the rates of nitrofurantoin resistance across all pathogens isolated was 3.9\% whereas the rate of trimethoprim resistance was 12.5\%. Trimethoprim- but not nitrofurantoin-resistance is now far commoner.

A recent review and meta-analysis suggested nitrofurantoin had a similar clinical cure rate to comparators but with a 5- rather than 3-day course for nitrofurantoin apparently producing better cure rates \textsuperscript{410}. However 5 day and 3 day courses have not been directly compared in adequate numbers and Public Health England has not recommended 5 day courses. We consider in MDR GNB UTI that course lengths should be those that produce the best rates of bacteriological cure. There is no convincing evidence that shorter courses are equivalent to longer courses specifically in MDR GNB infections nor that the risk of serious unwanted effects is increased with longer courses. Whether such longer course lengths should be used more generally for nitrofurantoin is therefore unresolved. Unwanted effects in the systematic review were mainly gastrointestinal and no pulmonary events were reported although this may reflect short follow up periods \textsuperscript{410}. There are no specific studies of nitrofurantoin in UTI caused by ESBL-producing organisms, but UTIs that are susceptible to nitrofurantoin have a similar response rate irrespective of ESBL-production. However ESBL-producing members of the \textit{E coli} ST131 clone which are common in the UK and elsewhere often have urinary virulence factors that are associated with recurrence, infection of the upper urinary tract and bacteraemia \textsuperscript{411} and when infection reaches the upper tract nitrofurantoin is ineffective. Nitrofurantoin resistance has appeared in this sequence type (See 6.3.4). Further comparative studies in UTIs due to ESBL-producing \textit{E. coli} are needed.
Evidence:

Nitrofurantoin is effective in lower, uncomplicated UTI and resistance rates remains low in E. coli although new plasmid-mediated mechanisms of resistance are now described. Mechanisms of acquired resistance in the UK, including in travellers, have not been recently studied. Resistance is intrinsic in Proteus spp. and Serratia spp.

Evidence level: 1+

There is usually no change in faecal Enterobacteriaceae during or immediately after use. Breakthrough infection, when the drug is used prophylactically, remains susceptible unlike with trimethoprim.

Evidence level: 3

Nitrofurantoin’s activity is reduced in alkaline urine.

Evidence level: 4

Use of nitrofurantoin in moderate renal impairment, as seen with increasing age, has been controversial, but unrestricted use down to an eGFR of >45mL/min may be acceptable.

Evidence level: 1+

Use in moderate renal impairment or in long term/repeated courses may be associated, albeit rarely with serious pulmonary unwanted effects.

Evidence level: 3

Five-day not 3-day courses are recommended for susceptible ESBL-producing E. coli.

Evidence level: 1+
Recommendations:

- Could use nitrofurantoin for 5 days to treat uncomplicated, lower urinary tract infections with nitrofurantoin-susceptible MDR *E. coli* (not Proteae or *P. aeruginosa*).

  Grading: Strong recommendation for

- Do not use repeatedly if there is moderate renal impairment, or in long-term courses, as these are associated with rare unwanted pulmonary effects.

  Grading: Conditional recommendation against

- Use alternative agents if there are repeated recurrences with MDR GNB but do not anticipate the emergence of resistance in *E. coli* infections on a single recurrence as selection for resistant strains in the urine or faecal flora is rare.

  Grading: Conditional recommendation for

- Need comparative studies of nitrofurantoin and other active antimicrobials in patients with ESBL-producing *E. coli* and *Klebsiella spp.*

  Grading: Recommendation for research and possibly conditional recommendation for use restricted to trials.

9.3 Fosfomycin trometamol

Fosfomycin has not been widely used in the UK, where the oral form was available between Feb 1994 and 1996 was thereafter withdrawn and not marketed for nearly two decades until 2013. Its use elsewhere in Europe has been associated with clinical success in lower UTIs. Fosfomycin suppresses Enterobacteriaceae in the faecal flora of
60% of patients by day 3 after a single dose but this rapidly drops to 30% at days 10 to 14: in contrast, nitrofurantoin does not suppress these organisms. Oral fosfomycin should be administered while fasting or 2 or 3 hours before meals, as food can slow its absorption, leading to lower concentrations in the urine. Oral fosfomycin is licensed solely for the treatment of uncomplicated cystitis. A single oral dose of 3 grams results in a plasma $C_{\text{max}}$ of 22-32 mg/L and a urine maximum concentration ($U_{\text{max}}$) of 1053-445 mg/L. The urinary concentration remains inhibitory for *E. coli* for at least 48 hours. In elderly patients with a mean GFR of 40 mL/min concentrations after 24 hours exceeded those reported for healthy young subjects but there was considerable variation in excretion rates.

Treatment with a 3g single dose of fosfomycin trometamol was associated with clinical success rates (defined as the resolution of symptoms after treatment) between 77.8% and 94.2% in four observational studies (some complicated and some receiving >1 dose) of treatment of lower UTI due to multi-resistant bacteria. Oral fosfomycin trometamol has been used successfully for prophylaxis of pyelonephritis in patients with asymptomatic bacteriuria (ASB) in pregnancy, and there are reports of its use, sometimes in combination, in chronic prostatitis. The use and kinetics of fosfomycin has recently been extensively reviewed following its re-introduction to Canada.

**Evidence**

Fosfomycin is effective and well tolerated in treatment of UTI but the oral drug has only been studied in lower UTI.

Evidence level: 2++

Plasmid- and chromosomally-mediated resistance has emerged in populations where fosfomycin is widely used.
Recommendations

- Use in the treatment of lower UTI due to MDR Enterobacteriaceae. Oral formulation available. Useful for infections with ESBL-producers or carbapenemase producers. No trials of oral formulation for upper UTI.

Grading: Strong recommendation for

- Carry out ongoing local and national surveillance of use and resistance because of previous emergence of bacterial resistance in populations and the drug's potential as an important parenteral agent.

Grading: Strong recommendation for

9.4 Mecillinam and Pivmecillinam

Pivmecillinam is an oral inactive ester and prodrug that is converted to microbiologically active mecillinam, penicillin, after intestinal absorption. Mecillinam has in vitro activity against most Enterobacteriaceae (including those with copious AmpC and some with ESBLs), but innate resistance occurs in Proteus spp., Morganella morganii, Providencia spp., some Serratia spp., and most non-fermenters including Acinetobacter spp. and P. aeruginosa. Mecillinam has no activity against enterococci or S. saprophyticus.

Some TEM and SHV ESBLs confer clear resistance and an inoculum effect on testing is common for other ESBL producers. In one study of ESBL-producing E. coli the MIC$_{50}$ by agar dilution was 1mg/L with an inoculum of $10^4$ cfu/spot but the MIC$_{90}$ was 4mg/L. Experiments with E. coli transconjugants showed that mecillinam MICs rose to 8mg/L when CTX-M-15 or -3 were present but only to 0.25-0.5mg/L with CTX-
M-9 or -14. Combination with clavulanate reduced all mecillinam MICs for ESBL producers (except SHV-4) to \(\leq 4\) mg/L at high inocula and \(\leq 2\) mg/L with usual light inocula\(^{389}\). In another study of combination with clavulanate \(^{418}\) 47/48 ESBL producers, were susceptible to mecillinam. Most of these produced CTX-M-3 (found in N. Ireland) not the commoner CTX-M-15 enzymes usual in England, Wales, and Scotland. There was no difference between the MICs for transconjugants producing CTX-M-3 and -15 in the earlier study. Synergy with clavulanate was detected in 40-60.4% of ESBL-producing isolates depending on the method of assessment. When a high inoculum was used, there was a marked inoculum effect raising the MIC of mecillinam alone but not mecillinam plus clavulanate. This study needs to be repeated with \textit{E. coli} ST131 strains producing CTX-M-15 enzyme and also often OXA-1 which is not inhibited by clavulanate but said to have little activity against mecillinam.

Mutants resistant to mecillinam by non-ESBL mechanisms can readily be obtained by laboratory selection. These show mutations in many different cellular functions \(^{68}\). However, a recent study of mecillinam-resistant clinical isolates found them all to have mutations leading to inactivation of the \textit{cysB} gene. Reduced cysteine biosynthesis results in accumulation of the transcriptional regulator guanosine 3’-diphosphate 5’-diphosphate (ppGpp) so that the mecillinam targeted PBP2 becomes non-essential \(^{419}\). Addition of cysteine to the growth medium \textit{in vitro} reversed the resistance to mecillinam for such mutants raising possible issues with regard to current \textit{in vitro} testing media.

Mecillinam is inactive against Enterobacteriaceae with KPC enzymes but some published data suggest \textit{in vitro} activity against isolates with OXA-48-like enzymes \(^{68,389}\) and even some with NDM-1 enzymes, as reflected in an MIC\(_{50}\) of 4 mg/L for NDM carbapenemase-producing \textit{E. coli} \(^{420}\) although this low value is disputed by others (D.M. Livermore, unpublished data).
Pivmecillinam at 200mg three time daily only produces sustained inhibition in Monte Carlo simulations if the mecillinam MIC is $\leq 0.25$mg/l suggesting a higher dose or lower EUCAST breakpoint may be required respectively to produce and predict clinical response \(^{421}\).

Pivmecillinam is used mainly for lower urinary tract infection, where it has similar short-term symptomatic efficacy to amoxicillin and trimethoprim/sulfamethoxazole if organisms are susceptible \(^{422, 423}\) and also to norfloxacin in 3- or 7-day regimens \(^{424}\). Seven-day pivmecillinam regimens are associated with more frequent clinical success than 3-day regimens \(^{425}\). Pivmecillinam prophylaxis in children with vesicoureteric reflux markedly reduced faecal *E. coli* and urinary breakthrough with *E. coli*; unlike nitrofurantoin, breakthrough infection with enterococci was common, reflecting different *in vitro* resistance \(^{426}\). Urinary concentrations are very high \(^{427}\).

Clinical trials of pivmecillinam against ESBL-producing Enterobacteriaceae are limited to case series. In one small trial pivmecillinam was used alone with 30/39 patients receiving 400mg three times daily and 9/39 receiving 200mg three times daily. Dosage did not affect clearly the cure rates regardless of whether the UTI was complicated. Twenty eight patients were noted to have calculi, prostatic hypertrophy or urinary catheters (i.e. complicated UTI) and 6 of these were bacteriological failures. Two other bacteriological failures were seen among the remaining 11 patient. Bacteriological cure was attained in 31/39 (79% overall), but five relapsed; clinical cure was attained in 16/19 patients but the rest were lost to follow-up \(^{428}\). There is no theoretical, trial or practise evidence to support a regimen with a loading dose of 400mg followed by 200mg three times daily which has been recommended in the UK as a compromise \(^{429}\). A population-based Norwegian study of pivmecillinam treatment of community-acquired UTIs examined the impact of MICs and ESBL-production in *E. coli*: it is not clear this was restricted to uncomplicated lower UTIs for which, alone, pivmecillinam is licensed \(^{430}\).
total of 343 patients were included, of whom 158 (46%) were treated with pivmecillinam. Eighty-one patients had infections caused by ESBL producing E. coli, and 41 (51%) received pivmecillinam as the primary treatment usually at a dose of 200mg three times daily for at least 7 days. Mecillinam MICs were higher for ESBL-producers than non-producers: 68% of strains had CTX-M Group 1 enzymes (including CTX-M-15) and 28% had Group 9 enzymes (including CTX-M-9 and -14). Treatment failure was (atypically) defined as a new antibiotic prescription appropriate for UTI within two weeks of the initial therapy or failure to clinically improve. Clinical treatment failure with pivmecillinam was observed in 18 (44%) of patients infected by ESBL-producing strains and in 16 (14%) of patients with ESBL non-producing strains. Mecillinam MICs for isolates from treatment failures (n=34, 18 ESBLs) averaged 2mg/L (range 1-4mg/L) compared with MICs of <1mg/L for all isolates from treatment successes (n=124, 23 ESBLs). Treatment failures occurred in 50% of cases with mecillinam MICs of 2mg/L rising to 63% at MICs of 4mg/L. This compares with a EUCAST breakpoint of S=<8mg/L, R>8mg/L for mecillinam, again suggesting inadequate levels or too high a breakpoint. Multivariate analysis showed that ESBL status (odds ratio (OR) 3.2, 95% confidence interval (CI) 1.3-7.8, p = 0.009) and increased MIC of mecillinam (OR 2.0 for each doubling value of MIC, CI 1.4-3.0, p<0.001) were associated with pivmecillinam treatment failure. Treatment failure rates above 25% were associated with mecillinam MICs >=2mg/L for ESBL-producers and >4mg/L for isolates lacking ESBL. From the transconjugant study cited earlier it is likely that UK CTX-M-15 producing isolates will be in this more resistant category and will respond poorly if pivmecillinam is used alone. This study must be seen also in the context of the earlier studies on the doses necessary to achieve adequate urinary concentrations.

There has been controversy over whether studies should be repeated with higher doses such as 400mg three times daily but a more effective action to improve cure rates may
be combined use of a 200mg three times daily regimen together with amoxicillin/clavulanate at 375mg three times daily. We recommend this combination if oral pivmecillinam follow-on therapy is prescribed following hospital or OPAT iv treatment for UTI involving an ESBL-producer. Co-administration of amoxicillin/clavulanate may not only provide efficacy via inhibition of ESBL but also 10- to 100- fold bactericidal synergy by combining amoxicillin’s action on PBP1 and 3 and mecillinam’s action on PBP2.  

Future use of co-amoxiclav, rather than clavulanate without amoxicillin, in combination with mecillinam is partly supported by a high quality double-blind multicentre RCT of mecillinam and ampicillin-congeners without clavulanate in pyelonephritis in 1995, in the era before CTX-M enzymes. Equivalent results to cefotaxime/cefadroxil were achieved with an oral switch from parenteral mecillinam (no longer available) and ampicillin to pivmecillinam (at 400 mg three times daily) plus an oral ampicillin prodrug, suggesting that synergy of amoxicillin and pivmecillinam potentially would be clinically useful in follow-on therapy for pyelonephritis. In modern circumstances, including against ESBL-producers, this efficacy might be restored by protecting both mecillinam and amoxicillin by using them with clavulanate. A clinical success rates of 93% for pivmecillinam as against 53% with pivampicillin in a study in 1986 of pyelonephritis suggests the drug has activity in the upper urinary tract. However, it is important to note that clinical trials of the combination of amoxicillin/clavulanate with pivmecillinam have never been undertaken in pyelonephritis, and pivmecillinam has no license for pyelonephritis. Further clinical comparative studies with outcome data are urgently required for pivmecillinam, with and without clavulanate (probably administered as amoxicillin/clavulanate), for both complicated (including upper urinary tract) and lower urinary tract infection against ESBL producers. Amoxicillin/clavulanate unlike
clavulanate alone is available and licensed for upper UTI. These trials would determine pivmecillinam’s role and its potential to reduce the need for hospitalisation or OPAT admissions to administer IV agents active against ESBL-producers.

Pivmecillinam is claimed to have a minimal effect on the intestinal and vaginal flora of the host with little selection for resistant bacteria, vaginal Candida or *C. difficile*. However, the earlier study of suggests it markedly reduces faecal *E. coli* at least in children. In an *in vitro* human gut model, it did not elicit *C. difficile* germination, proliferation or toxin production; suggesting that superinfection with this pathogen should be rare if the drug is used alone. Clinical studies with pivmecillinam-amoxicillin/clavulanate regimens should include studies on persistence of ESBL-producing *E. coli* gut colonisation and new infections with *C. difficile*.

Overall there are uncertainties about how pivmecillinam should best be used in the modern era. The drug has very valuable potential and these uncertainties need resolution by large clinical trials which are now urgent. Selection for resistant strains (such as SHV-producers) in the interim would be unfortunate and for this reason we await further substantive trials and action and do not include its use alone in our general recommendations.

**Evidence:**

Pivmecillinam is a prodrug for mecillinam and is the sole oral β-lactam (excluding tebipenem and faropenem which are available only in Asia) with some activity against ESBL- and AmpC-producing organisms. It has a European license, and is widely and effectively used for lower UTI in some countries. Parenteral mecillinam has been manufactured in the past but is now unavailable.

Evidence level: 2++
Pivmecillinam has no published clinical trials against CPE and *in vitro* activity appears poor or non-existent.

Evidence level: 4

Urinary levels following doses of 200mg three times daily are inadequate to inhibit some ESBL-producing MDR GNB including some with CTX-M-15 considered susceptible by the current EUCAST breakpoint (S=<8mg/L).

Evidence level: 3

Failure rates with 200mg three times daily pivmecillinam used alone against lower UTIs due to ESBL-producing *E. coli* are too high to recommend regular use in such infections.

A higher dose, 400mg three times daily, has been proposed but there is no convincing evidence to show it is more effective Comparative studies with fosfomycin have not been reported but there are no suggestions of such ESBL-related failures in existing fosfomycin studies in the absence of resistance.

Evidence level: 3

There are inadequate trial data to support the use of pivmecillinam in *Klebsiella* infection especially where the strain responsible produces ESBLs

Evidence level: 4

*In vitro* evidence and early trials of combination with ampicillin or pivampicillin suggest that a useful measure to increase efficacy would be combination with amoxicillin as well as clavulanate (See below).

Evidence level: 2+

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In vitro studies suggest that clavulanate (available clinically only as amoxicillin/clavulanate) would protect mecillinam from destruction by ESBLs and lower its MICs for Enterobacteriaceae. If pivmecillinam is prescribed as follow-on to OPAT or in-patient treatment, use of the combination is recommended.

Evidence level: 3

Clinical trials of pivmecillinam alone versus pivmecillinam with amoxicillin/clavulanate in lower UTI would be in the public interest. These should be sized to give information on efficacy against ESBL-producing bacteria and should include studies on the bowel-flora and associated recurrence rates and C. difficile. If results of combination treatment are satisfactory consideration should be given to trials in upper UTI including economic assessment against OPAT treatment. Comparative trials with nitrofurantoin or fosfomycin trometamol for MDR GNB lower UTI are also required.

Evidence level: 4

**Recommendations**

- Consideration should be given to reducing the mecillinam EUCAST breakpoint for classification of susceptibility

  Grading: Conditional recommendation for

- Treat lower UTI due to ESBL-negative *E. coli* with pivmecillinam at 200mg three times daily: do not use for infections caused by Proteae, *Klebsiella* or *Pseudomonas*. Some ESBL-producing *E. coli* respond, but efficacy is poor against CTX-M-15 enzyme producers: dosing at 400mg three times daily may be no more effective. Consider combination of the 200mg dose with 375mg
amoxicillin/clavulanate for follow on to parenteral therapy for such infections in hospital or OPAT.

Grading: Conditional recommendation for

- Requires clinical comparative trials in UTI in the public interest in i) alone or together with amoxicillin/clavulanate for UTI involving ESBL-producing organisms including particularly those producing CTX-M-15 enzymes ii) in uncomplicated lower UTI generally compared with fosfomycin trometamol and nitrofurantoin as the relative advantages of these drugs have not been directly compared by industry over the least 10 years as MDR GNB have become more problematic.

Grading: Recommendation for research and possibly conditional recommendation for use restricted to trials

10 Managing urinary tract infection

10.1 Diagnosis and the need for treatment or prophylaxis

Because UTIs are the major group of infections due to antibiotic-resistant Gram negative infections in primary care, we have chosen to make specific recommendations about their diagnosis and about specific antibiotic stewardship.

Good practice in differentiating urinary infections from other infections and asymptomatic bacteriuria is vital to reduce the unnecessary use of antibiotics. When clinical variables were examined in a validation study of a previously derived predictive dipstick rule-based on having nitrite or both leucocytes and blood, the positive predictive value for urinary infection was 82% for women with all three of cloudy urine, dysuria, and nocturia. The negative predictive value for urinary infection was 67% when none of these three features was present. When individual clinical features
were considered alone, cloudy urine or dysuria was predictive of UTI, but nocturia or
smelly urine was not \(^{335}\), which brings into question its value in the assessment above of
the combination of cloudy urine, dysuria and nocturia. In women aged 17-70 years with
uncomplicated UTI, the negative predictive value when nitrite, leucocytes, and blood are
ALL negative was 76\% \(^{335}\). The positive predictive value for having nitrite alone or
nitrite together with either blood or leucocytes was 92\% \(^{335}\). A systematic review of
diagnostic studies found that the presence of vaginal discharge or vaginal irritation
reduced the probability of urinary infection to 20-30\% \(^{337}\).

Several different studies have shown the prevalence of asymptomatic bacteriuria is
about 6\% in men and 16\% of women aged over 65 years \(^{338}\) and is higher in older age
groups and in the institutionalized elderly. In a cohort study, 1173 elderly female
residents without catheters in care homes were followed for 9 years with urine cultures
every six months\(^{339}\). No relationship was found between ever having had asymptomatic
bacteriuria and death after adjusting for covariates (hazard ratio, 1.10; CI, 0.78 to 1.55).
The death rate in the group who never had asymptomatic bacteriuria was similar to
those who had bacteriuria but either received no treatment or were treated (\(P > 0.2\)) \(^{339}\).
The lack of benefit in treating asymptomatic bacteriuria was confirmed in another
smaller study: neither mortality nor the frequency of symptomatic episodes was
reduced, but for every three women with asymptomatic bacteriuria in a care home
given antibiotics (the type was not specified in this study), one experienced adverse
effects (such as rash or GI symptoms) \(^{340}\). Cumulatively, 3-6\% of people acquire
bacteriuria per day of urinary catheterisation even with best practice for insertion and
care of the catheter, and therefore many older people with long term catheters have
bacteriuria \(^{341,342}\). Intermittent catheterisation is associated with a lower incidence of
asymptomatic bacteriuria than long-term catheterisation \(^{343}\). Catheterised patients
should only receive antibiotic treatment when they are systemically symptomatic to
reduce the risk of colonisation by antibiotic resistant bacteria. Differentiating urinary tract infection from asymptomatic bacteriuria can be particularly challenging in elderly patients with dementia as they cannot always describe their symptoms. A positive urine culture or dipstick test will not differentiate between UTI and ASB.

Patients with asymptomatic bacteriuria may have white blood cells in the urine just as in true infection. In older patients including those with dementia, diagnosis should be based on a full clinical assessment, including vital signs.

A Canadian randomized controlled trial of a diagnostic and treatment algorithm for UTI implemented in care homes, using a multifaceted approach, reduced antibiotics for urinary indications by 31%, compared with control care homes, with no increase in hospital admissions or mortality. Patients were considered for antibiotic treatment based primarily on presence of fever greater than 37.9˚C or 1.5˚C increase above baseline on at least two occasions over last 12 hours and one or more signs of UTI.

The full algorithm used is shown in Figure 5. Fewer courses of antibiotics for suspected urinary tract infections per 1000 resident days were prescribed in the intervention nursing homes than in control care homes (1.17 versus 1.59 courses per 1000 resident days). Antimicrobials for suspected UTI represented 28.4% of all courses of drugs prescribed in the intervention nursing homes compared with 38.6% prescribed in the control care homes (weighted mean difference – 9.6%, − 16.9% to −2.4%). No significant difference was found in admissions to hospital or mortality between the study arms.

In recurrent UTI, deciding whether to give prophylaxis is a balance between the benefits of reducing symptomatic relapse and pyelonephritis versus side effects and the risks of selecting antibiotic resistance. Guidance is based on a systematic review of 19 trials. Nightly prophylaxis in non-pregnant women with recurrent urinary infection showed that prophylaxis reduced the relative risk of having one microbiological recurrence by
five-fold (0.21) (95% CI 0.13 to 0.34), giving number needed to treat of 1.85 over 6–12 months. However, adverse effects occurred, particularly following nitrofurantoin, and 30% of women did not adhere to treatment. Any benefit was lost as soon as the prophylaxis stopped. Post-coital antibiotics were equally effective to nightly prophylaxis. Previous studies before the rise in resistance showed the same effect with postcoital single-dose cephalexin when used for recurrent urinary infection in pregnancy. If recurrence is not too frequent it may be better to provide the patient with standby nitrofurantoin, to take as soon as symptoms occur; this approach was shown to result in less use of antibiotics and intuitively should result in less antibiotic resistance. Studies with cephalexin before the rise of ESBLs showed a slight increase in use with post coital cephalexin offset considerably by antibiotics used in treatment of UTI recurrences. The offset needs to be taken into account in individual patients if standby nitrofurantoin is used. Prophylaxis, if used, can usually be stopped after a year without a resumption of the recurrences and there are now European guidelines that this review should be made at 6 months. The increase in trimethoprim resistance makes prophylaxis with this drug less suitable than it was and prolonged nitrofurantoin is associated with an increased risk of unwanted pulmonary damage, although this is rare. Patients on prophylaxis for >6 months should be reviewed. If the patient wishes to continue with a prophylactic regimen, consideration should be given in advance as to which antibiotic would be appropriately substituted for trimethoprim, nitrofurantoin or indeed ciprofloxacin (which can also be used in prophylaxis), if resistance develops or a breakthrough infection occurs. Persisting with an agent where breakthrough with a resistant strain has occurred will be ineffective. Cranberry juice prophylaxis is less effective in preventing breakthrough infection but cotrimoxazole generates more multiple resistance in breakthrough strains. Prophylaxis with beta-lactam antibiotics commonly selects for resistant Enterobacteriaceae in the faecal flora and is not
There are relevant studies of prophylaxis after symptomatic UTI in infants which show similar problems with emergence of resistance on continuous prophylactic antibiotics, including resistance to cephalosporins due to ESBL-production.

NICE notes that prophylactic antibiotics given at catheter change or insertion do not reduce infections in those with neurological conditions and recommends that they should not be used: such use for any indication contributes to pressure on emergence of resistance and should be avoided. NICE recommends that clinicians should consider antibiotic prophylaxis at change of catheter for patients who:

i) have a history of symptomatic urinary tract infection after catheter change or

ii) experience trauma during catheterisation (frank haematuria after catheterisation or two or more attempts of catheterisation). Placement of an incontinence implant is also an indication for short term prophylaxis but the recent insertion of an orthopaedic implant is not.

**Evidence**

Specific symptoms and signs hitherto accepted as characteristic of urinary infection have different predictive values.

In women with uncomplicated urinary infection the highest positive predictive value for strip testing was for having nitrite alone or nitrite with either positive leucocyte esterase or blood.

There is no patient benefit in treating asymptomatic bacteriuria.
Using an algorithm based on fever and at least one sign of urinary infection reduces the number of antibiotic prescriptions in nursing homes.

Evidence level: 3

Treatment or prophylaxis with antibiotics in catheterised patients increases colonisation by antibiotic-resistant strains.

Evidence level: 1+

Prophylactic antibiotics given short-term at catheter change or insertion do not reduce infections but are indicated with specific criteria of i) traumatic catheterisation, ii) previous severe symptomatic infection on catheter change, or iii) to cover placement of a urinary continence implant.

Evidence level: 4.

In recurrent UTI, antibiotic prophylaxis is very effective whether given daily (Evidence level 1++) or post coitally (Evidence level 1+) but an alternative is to consider pre-prescribed standby antibiotics to take at the onset of symptoms.

Evidence level 4.

If prophylaxis is used and effective it should be usually restricted to six-months prescription,

Evidence level 3

Previous resistances, or breakthrough of resistant isolates on prophylaxis should preclude use of an agent and consideration should be given to unwanted effects with long courses and what antibiotic would be chosen for breakthroughs.

Evidence level 4

**Recommendations**

- Always consider the positive and negative predictive value of specific symptoms before sending urine for culture or starting antibiotics for a UTI. Use dipstick
tests, if no catheter is present, to confirm the diagnosis, before prescribing especially when symptoms are mild or not localized.

Grading: Strong recommendation for

- For an elderly patient, do NOT send urine for culture or start empirical antibiotics unless there are specific symptoms or signs of UTI and none elsewhere. Use the algorithm in Figure 5 to decide whether to do this in elderly patients especially in those with dementia

Grading: Conditional recommendation for

- Do not prescribe antibiotics in asymptomatic bacteriuria (ASB) in the elderly with, or without, an indwelling catheter.

Grading: Strong recommendation for

- Avoid antibiotic prophylaxis for urinary catheter insertion or changes unless there is previous history of symptomatic UTI with the procedure, insertion of incontinence implant, or trauma at catheterization.

Grading: Conditional recommendation for

- To reduce recurrent UTI, consider firstly, the option of pre-prescribed standby antibiotics to take when symptoms begin, rather than daily or post-coital antibiotic prophylaxis.

Grading: Conditional recommendation for

- Where prophylaxis is used successfully for recurrent infection in adults limit use to six months.

Grading: conditional recommendation for
10.2 Choosing a suitable antibiotic

Choosing an antibiotic to which an uropathogen is susceptible, is important as UTI symptoms resolve more slowly when an inappropriate antibiotic is given\textsuperscript{454}. All patients should be given advice on when to seek further medical advice, i.e. if their symptoms worsen (even if, after taking antibiotics, on the same day) or do not improve after several days. Treating patients with infections due to MDR GNB in the community is a challenge as oral antimicrobial treatment is preferred. ESBL-producing bacteria are generally resistant to trimethoprim, ciprofloxacin, amoxicillin and cephalosporins; susceptibility to amoxicillin/clavulanate is variable and interpretation by the laboratory is affected by different breakpoints used formerly by BSAC, and currently by EUCAST, or CLSI.

Local community antibiotic guidance should be informed by national and local surveillance data. An algorithm on choices based on the individual agents discussed is given in Figure 4. Choosing between fosfomycin, pivmecillinam and nitrofurantoin is difficult as there are no direct comparisons of these three antibiotics in infections due to ESBL-producing organisms. High failure rates with pivmecillinam may be due to the precise ESBL present and not using the drug in combination with amoxicillin/clavulanate, or possibly inadequate dosage: optimal ways to use the drug now in the UK have not been proven. In urinary infections due to non-ESBL-producing organisms nitrofurantoin for 3, or 5 days (or 7 days, which is not significantly different from the results of a 5 day course)\textsuperscript{410} and a single dose of fosfomycin have similar efficacy\textsuperscript{455,456}.

In a systematic review of the length of antibiotic treatment for acute uncomplicated urinary infection before the rise in prevalence of ESBL-producing Enterobacteriaceae, therapy for 3 days, delivered in the case of fosfomycin trometamol by a single 3g dose,
was similarly effective to prolonged therapy in achieving symptomatic cure for
cystitis. However, in this systematic review, bacteriological failure rates in the
subgroup of trials where the same antibiotic was used in both short and long treatment
arms of the trial, were higher in the short duration arms (RR 1.37, 95% CI 1.07 to 1.74, P
= 0.01). After a single dose of fosfomycin high concentrations are usually maintained in
the urine for 2 days. This is usually curative in uncomplicated UTI in women, but for
infection due to confirmed ESBL-producers, or in males, a second dose on the third day
has been suggested to promote bacteriological cure. On the same basis 5 not 3 days
nitrofurantoin would be recommended for confirmed ESBL-producing bacteria and 7
days for pivmecillinam regimens. Although frequently used as an end-point in
regulatory trials, it is uncertain if bacteriological cure immediately after treatment is of
any long term clinical or bacteriological significance in patients with UTIs involving
MDR GNB but the precautionary principle of adequate elimination of infections with
MDR GNB would suggest regimens for best bacteriological cure should be followed in
such cases. Eight studies in the systematic review included pivmecillinam at various
doses and durations. An analysis of E. coli strains from persistent or relapsed infection
after pivmecillinam showed an increased frequency of phylogenetic group B2 (which
includes ST131) and showed that when matched by virulence factors 7 days treatment
was preferable to 3 days therapy because it was less likely to be followed by persistence
or relapse. Studies of urinary infection with strains producing the CTX-M-15-ESBL
suggest that pivmecillinam alone at 200mg three times daily is inadequate treatment. In
vitro studies suggesting use with amoxicillin/clavulanate have not been followed by
clinical trials.

Based on evidence collected before the spread of ESBL-producing strains nitrofurantoin
(100mg twice daily) should be given for 3 or 5, not 7, days for fully susceptible strains.
No trials of nitrofurantoin 100mg twice daily with ESBL-producing strains have been
published although the antibiotic is widely used. Efficacy, relapse/recurrence rates or incidence of spread to the upper urinary tract or blood stream are all uncertain and no studies have been published on the emergence of resistance during or after treatment or in relapses. MDR Klebsiella spp., but not E. coli, are commonly resistant to nitrofurantoin but the mechanisms for resistance in the UK have not been investigated recently.

**Evidence**

Local community antibiotic guidance on empirical treatment of urinary infection should be informed by national and local surveillance data.

Evidence level: 4

In lower uncomplicated UTI where risk factors for MDR GNB are present these four treatment options can be used rather than trimethoprim:

- Fosfomycin trometamol

Evidence level: 2+

- Nitrofurantoin (unless patients eGFR is less than 45 ml/min/1.73m$^2$).

Evidence level: 2+

- Pivmecillinam but *in vitro* and clinical data suggest this is less successful than a) and b) for ESBL-producing bacteria likely to be present in the UK.

Evidence level: 3

Another other relevant antibiotic if the causative organism is confirmed as susceptible.

Evidence level: 4
Recommendations

- Inspect up-to-date national and local antibiotic surveillance when compiling local antibiotic guidelines on treatment of UTI.

  Grading: Strong recommendation for

- If there are risk factors for MDR GNB or previous presence of MDR GNB and the patient is symptomatic, send a urine specimen for culture and susceptibility testing

  Grading: Strong recommendation for

- Always inform the patient or their carer(s) on what to look out for and how to re-consult if symptoms worsen or do not improve as community-onset *E. coli* bacteraemias of urinary origin are increasing

  Grading: Strong recommendation for

- Use fosfomycin, or nitrofurantoin or as third-line choice pivmecillinam, guided where possible i) by susceptibility testing and ii) by this guideline's recommendation on choice, combinations, dosing and duration, for uncomplicated lower urinary tract infection where MDR GNB are suspected.

  Grading: Strong recommendation for

- Use nitrofurantoin for 5 days with MDR GNB. Alternatively use fosfomycin trometamol 3g orally as single dose, and repeat on third day only if MDR GNB are confirmed to improve bacteriological cure. Pivmecillinam at 200mg three times daily for 7 days may be a third line choice but consider combination use with
amoxicillin/clavulanate. Clinical trial results on pivmecillinam for MDR GNB in the UK are urgently required.

Grading: Conditional recommendation for

10.3 Treatment of pyelonephritis and complicated UTI caused by MDR Gram-negative bacteria

Whenever resistant pathogens are anticipated, it is essential to send a urine specimen for culture and susceptibility testing before empirical treatment and such specimens will be useful in this condition even if resistant pathogens are not anticipated. As nitrofurantoin, pivmecillinam and oral fosfomycin are currently considered inappropriate in suspected or confirmed pyelonephritis, intravenous ertapenem (unlicensed in Europe for this indication) should be given in an Outpatient Parenteral Antibiotic Therapy setting to treat patients with pyelonephritis confirmed or suspected to be caused by ESBL-producing pathogens that are resistant to trimethoprim and quinolones. If the patient requires admission to hospital meropenem or, depending on costs and local policy, ceftolozane/tazobactam or temocillin should be given for infection due to ESBL-producing strains. Piperacillin/tazobactam may be considered if the isolate has been shown to be susceptible. Amikacin might be considered but activity may be impaired if AAC (6’)-1b-cr is produced. In practise strains with this enzyme may be reported as either susceptible or resistant and the enzyme cannot easily be detected: no trials of amikacin use against such strains have been reported. Measuring amikacin levels promptly and adjusting doses is less likely to be easily supportable than use of gentamicin but the latter is unsuitable for infection with ESBL-producers unless susceptibility is known.

Ceftazidime/avibactam or non-β-lactam agents in combination perhaps with meropenem should be considered for infections with CPE- See Figure 4. Temocillin may
have a place for more susceptible strains with KPC-carbapenemases but this has not been established by trials: it does not have a role against strains with MBLs or OXA-48 like carbapenemases. Such factors and choices are important when empirically treating pyelonephritis caused by probable or confirmed MDR GNB as this may be complicated by bacteraemia. If a patient with pyelonephritis due to ESBL-producing bacteria has penicillin or cephalosporin-hypersensitivity, there are two alternative strategies. Firstly meropenem can be given despite a risk of cross-allergenicity that is now thought to be largely hypothetical. In this case caution must be exercised with appropriate drugs ready to treat any severe acute reaction. This seems to be safe. Alternatively urgent susceptibility tests by automated methods should be performed. Depending on any previous results for the patient’s isolates, intravenous gentamicin or amikacin (which has more auditory than vestibular toxicity but a lower resistance rate than gentamicin) may initially be used until a less toxic antibiotic can be identified from the concurrent susceptibility testing. Trimethoprim, ciprofloxacin or co-amoxiclav can be used in pyelonephritis if the pathogen is known to be susceptible (or a susceptible organism has been isolated in the preceding month with a satisfactory therapeutic response). A retrospective cohort study of community onset acute pyelonephritis due to ESBL-producing E. coli compared 85 patients receiving carbapenems with 67 receiving other agents to which the infecting bacterium was susceptible in vitro. There was no difference in rates of clinical or microbiological failure. A randomized double-blind controlled trial showed that 7 days of ciprofloxacin 500 mg twice daily was as effective as 14 days trimethoprim/sulfamethoxazole against susceptible organisms. However, trimethoprim and quinolone resistance are now common and therefore none of these agents remain suitable for empirical use in pyelonephritis. The substitution of OPAT
therapy for oral antibiotic use in early pyelonephritis has not been costed in its effects on services.

Evidence

Pending antibiotic susceptibility testing, patients at increased risk of MDR GNB and suspected of pyelonephritis or complicated UTIs (i.e. indwelling catheter, recent urinary instrumentation, renal stones, prostatic obstruction, diabetes, immunosuppression, pregnancy, functional or anatomical urological abnormality) can be treated empirically with:

a) outpatient intravenous therapy with ertapenem.

Evidence level: 2+

b) admission for i) intravenous meropenem, temocillin, or ceftolozane/tazobactam if infected by ESBL-producing E. coli or Klebsiella spp., ii) intravenous fosfomycin and colistin with or without meropenem, or ceftazidime/avibactam therapy if infected by a susceptible carbapenemase-producer.

Evidence level: 1+

If hypersensitive to penicillin treat with meropenem with caution or gentamicin (if no past evidence of resistance) or amikacin

Evidence level: 4

c. Trimethoprim, ciprofloxacin or co-amoxiclav if urine testing shows an organism that was susceptible in the preceding month and there has been no history of clinical failure.

Evidence level: 1+
Recommendations

- In pyelonephritis always collect a urine sample before treatment. MDR GNB are unlikely to respond to oral treatment so consider risk factors for an MDR isolate including travel. Use an active oral agent only if the patient is well enough and if known to have had ciprofloxacin-, trimethoprim-, or co-amoxiclav-susceptible MDR GNB in last month.

Grading: Conditional recommendation for

- If the patient has pyelonephritis and risk factors for MDR GNB, start, if hospitalisation not required, empirical intravenous therapy with ertapenem if OPAT therapy available. This will treat ESBL and Amp-C producing Enterobacteriaceae. If the patient needs hospitalisation, or OPAT is not available, admit for meropenem, temocillin or ceftolozane/tazobactam if no evidence of CPE organism. If the patient is penicillin-hypersensitive then the hospital may use amikacin or meropenem, or if only susceptible isolates in the past, gentamicin. If carbapenem-resistant bacteria are, or have been, present, base treatment on susceptibility testing of recent or current isolates.

Grading: Strong recommendation for

10.4 What is the threshold level of resistance for changing the choice of empirical treatment for urinary tract infections?

Most patients with UTI are treated empirically, particularly in a first episode of lower UTI. Failure of empirical therapy particularly in complicated UTI (e.g., pyelonephritis) is a common source of Gram-negative bacteraemia where increased 30-day mortality is associated with ineffective empirical therapy though maybe only in patients with sepsis syndrome. The probability of ineffective empirical therapy would be predicted to increase as the proportion of ESBL-producing, or carbapenem-resistant, bacteria rise.
Older narrower spectrum antibiotics may be recommended for empirical use in order to slow the emergence of resistance. One group of authors asserts that the right of future patients to come to less harm outweighs the right of the present patient to share in decisions on antibiotic treatment but this is a view many do not share. There is no agreement within the Working Party on the threshold resistance rate to an antibiotic that would justify substitution of other agents, nor on the degree to which routine laboratory testing of submitted samples overestimates the “true” resistance rate.

Rates of 20% have been suggested as justifying a change of empirical treatment in UTI. Confounders are i) that resistance rates are affected by duplicates within the series including when infection control sampling is intensive, ii) a bias towards performing culture and susceptibility only for difficult/unresponsive cases iii) by sequential testing second-line agents only for resistant strains according to local laboratory policy and iv) differences in breakpoints between laboratories. These sources of variation may justify central susceptibility testing of all UTI from sentinel groups of GPs in regions for national surveillance purposes or requirements for national notification and annual updating of method changes and assessment of their effects. Local and regional and variations exist in resistance rates for ESBLs as demonstrated by regional and national surveys. Quinolone resistance rates in *E. coli* are below 20% in most reported susceptibility surveys but resistance in bacteraemia is associated with increased mortality and with the ST131 group of strains which have an unrivalled ability to acquire other resistances. The risk of selection for resistance with a switch from trimethoprim leads us not to recommend their widespread use.

When the probability of bacteraemia associated arising from UTI rises, a lower threshold for altering normal treatment to cover a resistant strain is needed owing to the greater risk to the individual patient. A threshold of <5% resistance may be appropriate for higher risk situations.
Evidence

There are no accurate current figures on the prevalence of antibiotic resistance in UTI. Routine clinical data are subject to sample bias. These probably lead to overestimated resistance.

Evidence level: 2-

A threshold of 20% true resistance has been suggested as an indication to change “first line” empirical treatment of lower UTI. A lower threshold of, perhaps, 5% is appropriate when the risk of the patient becoming bacteraemic is increased. The Working Party consider that, in the absence of accurate national resistance surveillance these, or similar thresholds, presently can only be applied at a local laboratory level with i) careful de-duplication ii) precisely understood testing policies and iii) consistent local methodology.

Evidence level: 4

Recommendations

- Locally assess the true rate of resistance and determine from this when changes to guideline recommendations for empirical therapy in UTI are necessary including recommendations where the risk of antibiotic-resistant bacteraemia is high.

Grading: Conditional recommendation for

- Personalise empirical chemotherapy for each patient by considering current features of bacteraemia, risk factors for antibiotic resistance and past susceptibility testing including the presence of MDR GNB in the patient or unit.

Grading: Conditional recommendation for

Accepted manuscript
What effect does good antibiotic stewardship have on rates of MDR GNB?

The impact of good antibiotic stewardship in secondary/tertiary care facilities

The evidence base and practice of antibiotic stewardship in the UK has been recently promulgated in the Public Health England “Guidelines for Antimicrobial Prescribing and Stewardship Competencies” and the guidance from NICE (National Institute for Health and are excellence) Guideline 15: Antimicrobial stewardship: systems and processes for effective antimicrobial medicine use. This report will focus on aspects of stewardship that pertain to MDR GNB: more general aspects can be found also in the above sources. A Cochrane systematic review showed that interventions to reduce excessive antibiotic prescribing to hospital inpatients might reduce antimicrobial resistance and that interventions to increase effective prescribing can improve clinical outcome. Of the 89 studies cited to 2009 (reporting 95 interventions), 56 were interrupted time series (ITS), 25 were RCTs, 5 were controlled before-after studies (CBAs) and three were controlled clinical trials (CCTs). The reporting of outcomes was very variable (only 13/25 RCTs reported on mortality and only 5 on readmissions) complicating comparative assessment of studies. Interventions that enhanced the quality of prescribing in patients (defined softly as prescribing in accordance with guidelines) with any infection had no effect on mortality whereas interventions to increase compliance with evidence-based guidelines in community-acquired pneumonia, usually due to Gram-positive Streptococcus pneumoniae, was associated with reduced mortality. Reducing prescribing for all indications, determined as excessive by reference to evidence-based guidelines, was associated with increased re-admission but not with increased mortality or length of stay. Restrictive and persuasive interventions were associated with improved prescribing outcomes based on median outcome effect (proportion of subjects with an improvement or change in antibiotic selection, dose, route or duration versus control). Multifaceted interventions were
common but not necessarily more effective than simple interactions. Most (80/95, 84%) of the interventions targeted the antibiotic prescribed (choice of antibiotic, timing of first dose and route of administration). The remaining 15/95 interventions aimed to change exposure of patients to antibiotics by targeting the decision to treat or the duration of treatment. Only nine studies reported the effect of interventions on colonization or infection with antibiotic-resistant Gram-negative bacteria. Seven of these were ITSs, with a median effect size of 47%.

Although most studies reported >25% reduction in colonisation/infection with resistant Gram-negative bacteria, the confidence intervals were wide and in two studies the effects were not statistically significant and one crossover study of cycling empirical gentamicin, ceftazidime, and piperacillin/tazobactam showed an unintended increase of 39% in colonization with GNB resistant to any of the target drugs. One cluster CCT in neonatal units, showed, as intended, a reduction from baseline in colonization/infection of 68% by cefotaxime-resistant organisms, predominantly E. cloacae, when the initial empirical treatment was penicillin and tobramycin rather than ampicillin-cefotaxime. This study, the only one of the nine to report on mortality, showed a small increase in mortality when penicillin and tobramycin was substituted for cefotaxime ampicillin in matched neonatal units. A 2017 update of this Cochrane review concluded that there was still no statistically significant evidence that antibiotic stewardship reduced multiple antibiotic resistance although the impact on C. difficile is undoubted. Additionally this updated unwanted effects from stewardship interventions including an aminoglycoside substitution producing acute kidney injury (See 7.12) and studies where there was consequent delay in instituting antibiotics. Furthermore some studies reported a disruption of interaction between physicians and infection specialists as guidelines were used more frequently. Nevertheless an editorial on this review called for stewardship to be adopted in every
health care institution. One must now consider the homogeneity and quality of local hospital guidelines given guideline compliance is being used as a criterion of good stewardship.

In the 2013 Cochrane review, 11 studies of attempts to reduce excessive prescribing, reported data on mortality with no significant overall effect seen (and this continued to be the case in the 2017 revision). Interestingly one of the interrupted time-series studies examined the impact of a switch from penicillin and gentamicin to penicillin and amikacin in a neonatal unit with gentamicin-resistant E. cloacae infections and showed a reduction in gentamicin-resistant E. cloacae but an increase in E. aerogenes and enterococci.

Kaki et al. produced another systematic review of antibiotic stewardship programmes, limited to the critical care unit. These included three RCTs, three ITSs, and 18 uncontrolled before-and-after studies. Introduction of various antibiotic stewardship interventions led to 11% to 38% reductions in antimicrobial defined daily doses/1000 patient-days (except in a single study that found an increase of 6%), and lower total antimicrobial costs. Stewardship programmes led to shorter average duration of antibiotic therapy, less inappropriate use and fewer antibiotic-related adverse events. They also found some reductions in antimicrobial resistance rates extending beyond six months.

A meta-analysis of 52 ITS was used to compare restrictive versus persuasive interventions. Restrictive interventions had significantly greater impact on prescribing outcomes at one month (32%), 95% CI 2-61%, P=0.03) and on microbial outcomes at 6 months (53%, 95% CI 31-75%, P=0.001) but there were no significant differences at 12 or 24 months. Clinical outcome data were limited with 11 studies reporting on all-cause mortality but with no defined time-boundary, 4 studies showed
increased mortality, 7 found decreased mortality giving a non-significant overall effect \((0.92, 95\% \text{CI} 0.81-1.06, P=0.25)\).

In the USA, the Department of Veterans Affairs recently commissioned a systematic review of antimicrobial stewardship programmes (ASP) \(^{481, 482}\). The key findings have been published and the reader is referred to these publications for details \(^{483, 484}\). To avoid duplication, the VA systematic review only included papers meeting their eligibility criteria but not included in the 2013 Cochrane review. The review reported mixed results for clinical/microbial outcomes and overall improvement in prescribing. Because (i) few studies of different interventions reported each outcome, (ii) of inconsistency across studies and (iii) medium/high risk of bias, the strength of evidence for all clinical outcomes was low: no single antimicrobial stewardship programme was found to be superior but amongst studies since 2000 the greatest body of evidence of effectiveness was for decreasing inappropriate or increasing appropriate antibiotic use. Effects were seen across all species of Gram-negative bacteria and broad-spectrum antimicrobials.

There are individual studies of high quality. Introduction of a stewardship programme in one US hospital reduced the use of broad spectrum agents, and was associated with a reduction in hospital-acquired infections caused by MDR GNB from 37% to 8% over 6 years \(^{485}\). Similarly resistance in \(P. \text{aeruginosa}\) declined when state guidelines on stewardship were implemented using a computerized programme in an Australian ICU \(^{486}\). In another study in Israel, a carbapenem-restriction policy was used as part of a successful infection control strategy also including emergency department flagging of colonized or infected patients, building an isolation facility, eradication of clusters, environmental and personnel hand cultures, with rectal screening of 8376 patients. This was effective in controlling an outbreak of carbapenem-resistant \(K. \text{pneumoniae}\).

Although there was a significant reduction in meropenem use, prescription of colistin...
rose\textsuperscript{487}. Restriction of use of some antibiotics may need, or lead to, use of a diversity of other agents and even introduction of newly available antibiotics or appropriate use of older agents. These aspects also need to be subject to stewardship with appropriate actions in responsible bodies within hospitals and reporting to users. This can be complex and time-consuming. Some effective interventions are simple, for example, a high-quality study compared 8- and 15-day antibiotic treatment of ventilator-associated pneumonia (n=401) and did not find any difference in mortality or unfavourable outcome. Patients who received 8 day treatment had significantly less emergence of MDR pathogens (42\% versus 62\% p=0.04) but had a higher recurrence rate if they initially had non-fermenting organisms as pathogen (40.6\% versus 25.4\% risk difference 15.2\% (CI 3.9\%-26.6\%).\textsuperscript{488} Effective antibiotic stewardship requires the use of timely bacterial antimicrobial susceptibility testing. Relatively simple phenotypic tests, such as a comprehensive antibiogram by automated methods, screening for resistance in bacteraemia isolates by direct disc testing,\textsuperscript{514} double disc diffusion tests for ESBL, and biochemical carbapenemase detection can provide useful information for treatment and infection control purposes.\textsuperscript{515} Automated diagnostic tests for bacterial identification (e.g. MALDI-ToF) and PCR-based resistance gene detection (e.g. Cepheid\textregistered for carbapenemase and ESBL detection) can provide even more detailed information within the same day for MDR GNB. More rapid susceptibility methods for resistance detection are being developed. Further information may be found in recent reviews.\textsuperscript{515-518} This information together with promptly administered appropriate antibiotics is likely to improve prognosis. All UK laboratories should have access to phenotypic and basic genotypic methods described above within their resources. As a performance measure, overall time elapsed from sample collection to administration of treatment appropriate to the bacterial susceptibility can and should be assessed and repeatedly audited.
against what could best be achieved with modern methods. Particular attention should be paid to MDR GNB as defined either for community or hospital originating strains. Audit of outcomes associated with bacteraemia provides an objective measure of the appropriateness of antimicrobial treatment, particularly for MDR GNB.

The deployment of antibiotic stewardship programmes is variable, as shown by a survey of 660 hospitals in 67 countries. This study included the first data from sites in Asia, Africa and South America, many with considerable problems with MDR GNB. There is an urgent need for the adoption of an international antibiotic stewardship timetable.

**Evidence**

Up-to-date local resistance and outcome surveillance data are needed to inform guidelines on empirical antibiotic advice and must be persuasive to medical and nursing staff, to all prescribers and to pharmacists advising on guidelines.

Evidence level: 4

Interventions intended to decrease prescribing that is excessive (by reference to guidelines) for specific antibiotics have been associated with reductions in both colonisation and infections caused by carbapenem, aminoglycoside or cephalosporin-resistant bacteria but this is not a consistent finding across all stewardship initiatives.

Evidence level: 2++

Restrictive rather than persuasive prescribing interventions cause a significant short-term change in prescribing and there is scanty evidence that they may contribute to reductions in the prevalence of resistant GNB. Persuasive prescribing interventions should also be used and are as effective over a 1- to 2-year period.

Evidence level: 2++

Clinical outcome data on infections that is linked to antibiotic prescribing should be collected as well as data on resistance and prescriptions of antimicrobials to ensure
stewardship approaches do not degrade outcomes, and ensure high and consistent standards between hospitals.

Evidence level: 2++

Audit and feedback should be used to reduce antimicrobial use in hospitals. Local and national advice on which antibiotics to prescribe are a useful standard against which to conduct audit and to explore clinical and microbiological outcomes

Evidence level: 4

**Recommendations**

- Provide an on-going antimicrobial stewardship programme in all care settings, based on resistance rates, with audit of compliance with guidelines, surveillance of outcomes, and active feedback.

  Grading: Strong recommendation for

- Use restrictive prescribing policies to acutely reduce the incidence of infection, or colonization, with MDR GNB; thereafter, maintain persuasive and restrictive approaches and monitor that gains persist.

  Grading: Strong recommendation for

- Identify through horizon scanning, and make available, new antimicrobials that may be required to treat MDR GNB. Monitor their use through formulary/drug and therapeutics committees.

  Grading: Conditional recommendation for
11.2 The national monitoring of good antibiotic stewardship in secondary/tertiary care facilities

Antibiotic therapy differs from other treatment in man in being directed against diverse and frequently unknown organisms and in exercising selection for resistant organisms, these change the potential target for drug action and may then cause infection either in the same or other patients. Treatment options for infections due to MDR GNB are restricted and failure to deploy appropriate treatment in these infections may be associated with a poor outcome whereas excessive use of a single agent in a hospital or unit is more likely to select for superinfection caused by resistant organisms. The clinical governance of antibiotic policies therefore is a balance between treatment of the individual and management of the community’s antibiotic armamentarium.

Antibiotic use and the prevalence of MDR GNB are now widely monitored in communities and hospitals but (i) monitoring use does not indicate whether use was appropriate, and (ii) monitoring the accumulative prevalence of resistant strains is no guide to the incidence rate of new cases caused by MDR GNB. Root cause analysis of individual cases is burdensome and very complex if it is intended to relate to outcome. It also runs the risk of bias with regard to outcome unless the proportions of resistant or susceptible organisms that are examined match the overall population. It does not produce reliable statistically comparable data between institutions to support good practice. Nevertheless, such comparisons were used with MRSA bacteraemia and *C. difficile* in the past in the UK but these are acute events unlike chronic prevalence of antibiotic resistant strains.

Clinical trials early in a product’s availability offer guidance on efficacy against susceptible organisms and with some agents, an indication of potential for selection for resistance. However, antibiotic efficacy is not usually sustained as resistance emerges,
and unlike other classes of drug, early clinical trials become less relevant with the passage of time. Anticipating when empirical therapy should include coverage against MDR GNB is difficult but is a key part of local guidelines. Recommendations that i) limit use of broad spectrum drugs such as carbapenems, or ii) which reserve particular agents for patients with MDR GNB present in infections that have a potential high mortality, need also to consider the potential hazard of poor clinical outcomes. Despite assistance from other professions, deployment of infection and microbiology specialists into surveillance and away from patient care is frequent, and mundane tasks in surveillance employing specialists should be reduced to a minimum, without compromising excessively data quality. Routine national reporting systems on bacteraemia in the UK should be routinely linked to public health date of death data held nationally for each person by the Office for National Statistics as has been described in one study restricted to E. coli bacteraemia¹⁰². Such linked information should be fed back annually to, and within, individual hospitals and summarized findings provided to hospitals to enable comparisons of performance. Incidence and mortality rates in bacteraemia at the local level would provide key assurance on the prevention of systemic infections and the quality of outcomes. If these data on outcome were provided by patient, it would provide a focus to examine and attempt to reduce, the increasing incidence of bacteraemias and their associated mortality. Further these data would ensure locally that overall and specific audit could be made of the antibiotic resistance in organisms and the antibiotics actually deployed to treat serious infections that they caused. Added to existing data, such audit and source information could nationally and locally identify locations where there is high mortality either in primary or secondary/tertiary care enabling appropriate investigation and action to be taken locally. A crucial foundation has already been organized in England and Scotland via mandatory reporting of bacteraemia data for E. coli which specifically includes, inter
alia, data on community or hospital onset, and nursing home residency entered locally by laboratories. In England laboratories voluntarily and automatically (via computer links) submit antibiotic susceptibility data for 82% (54,301/66,512 over 2 years) of cases of *E. coli* bacteraemia reported by the mandatory programme, which does not, itself capture susceptibility data. This could be built upon to deliver local and nationally useful data on outcome by antibiotic resistance. Furthermore, this process should be expanded to capture mortality information on other important bacteraemias e.g. *Klebsiella spp.* where prevalence is increasing and resistance is a major global threat or indeed to all bacteraemias. Reduction in the absolute number of associated deaths from bacteraemia may well involve changes other than in chemotherapy provided audit suggests chemotherapy is actively employed and appropriate. This requires multidisciplinary joint engagement and clinical management expertise in the community quite as much as in hospital to avoid sepsis and improve its management. A decrease in prevalence of bacteraemia and multi-drug-resistance within such infections is one aspect of this. Quantitative reduction in the number of deaths, and not changes in the comparative position of hospitals and communities in their respective peer groups should be the focus.

Bacteraemias should be, assigned reliably as being of community, wider healthcare or hospital onset so that responsibility can be assigned and accepted for performance by relevant commissioning groups, public health services and hospitals. Whilst the date of sampling of bacteraemia can be recorded, patients may become colonized by the causative bacterium much earlier and the exact timing of acquisition usually cannot be proven from existing laboratory records. IT coordination and shared responsibility across the health economy is needed to access the last date of discharge from hospital, which may be a practical proxy for date of colonization in cases of apparent community acquisition that are actually hospital-acquired. Where care does not involve transfer to a
tertiary centre and the patient is not being admitted to multiple hospitals in a conurbation, such information should already be available in many localities but non-automated extraction is time consuming. It is important for securing improvement that the bacteria isolated from bacteraemias can be related to likely acquisition in hospital, wider healthcare or community and not simply to onset in hospital or community and that responsibility for resistant strains falls accurately on hospitals or community commissioners of healthcare. Targeting reductions in MDR GNB in potentially life-threatening infection is problematic because of variations between community populations in ethnic origin associated apparently with antibiotic resistance such as ESBL-production\(^4,137\). For this reason a simple process of commissioned reduction in resistance may be unachievable in some communities and their associated hospitals.

Residence in a nursing home is a marker of healthcare acquisition, not general community acquisition, and nursing-home patients should be separately and reliably categorized. Dates of hospital discharge of patients admitted from nursing homes may be relevant to intervention if the patient has moved between the nursing home and hospital recently – say within the last 2 years.

Tertiary and international referral in some hospitals (including referrals from armed forces deployed overseas\(^490\)) even if the hospitals are not formally categorized as specialist hospitals may also skew their resistance profile towards multiple resistance\(^491,492\) so it is important to keep a balance between recognizing that this may be a reason for high resistance rates and ensuring that such resistant strains should be, as they always have been, a target for effective infection control. Again for this reason targeting antibiotic resistance reduction appropriately within a national context, may be more straightforward if it is directed at a local level.
Dates of collection of blood cultures, as recorded in laboratory computer systems, may be distorted by entry of default dates of registration on Monday mornings after submission of samples from Friday night on wards. There is no information on the frequency of this problem but it is time-consuming to retrospectively correct or prospectively avoid. An interval of <3 days since admission, is recommended for defining ‘community onset’ as more practical than the 48 hour limit suggested internationally and probably without important consequence, if permitted. This should be investigated if the mandatory programme is expanded as recommended. Laboratory data should not be reported multiple times and should utilize as little manual entry as possible and hospital trusts should ensure the automated transfer of data from laboratory systems to monitoring bodies. Information transfer should be frequent.

However in the presence of good infection control and absence of an ongoing MDR GNB outbreak, annual batch processing of mortality linkage and annual central audit should be adequate in most hospitals for governance monitoring of hospitals and this would be adequate to support changes to infection management including antibiotic policy (which are seldom made more frequently). Not only good performance in reducing antibiotic use but also in better-than-average performance in bacteraemia reduction and better outcomes in bacteraemia (including that which is antibiotic resistant) should be rewarded.

Such laboratory-based extended surveillance of all bacteraemias would address (i) the diversity of organisms and, at a local level, the match to antibiotics prescribed (which itself could be centrally reported, if pharmacy systems and laboratory systems are linked by patient/NHS number and then ordered by concatenated patient/NHS number and reversed Julian date) ii) the usual, but not invariable, progression in antibiotic resistance rates. (iii) the need for organisations to make changes to prescribing policy with document control, feedback to clinicians and corporate responsibility of CCGs and
hospitals for infection management. To address bacterial species- and resistance-
specific aspects in any locality, analysis (including trend analysis) of data cumulated
over 5 years may be needed to avoid problems with small numbers of some pathogens.
Individual hospitals need more local as well as the existing national data to
systematically analyse, explain and address unsatisfactory outcomes. The already
striking increase in incidence of *E. coli* bacteraemia often in patients being admitted
from the community will probably increase further, with better ascertainment of sepsis.
Commissioning attention needs to be paid to the appropriateness of prior
chemotherapy (i.e. for UTIs in the community) to attempt to reduce such rising
incidence and associated mortality. Owing to the rise of MDR GNB, central monitoring
of, and action on, informatics is required in all hospitals. Collation of information is
required to explain clinical and resistance outcomes by patients and to plan action in
hospital and community onset cases. Early Warning Scores, which are required for such
analysis, are frequently now available on computerised systems to monitor vital signs.
Separate patient-based prescribing systems record the date of prescription and
antibiotics given. Laboratory data systems record (i) the date of collection of the first
positive blood culture for an organism-episode from a patient, and (ii) the organism and
its antimicrobial susceptibilities. These data sets should be linked electronically along
with, from hospital patient administration systems, the admission date, the date of last
hospital discharge and place of residence (i.e. home or residential care). Early Warning
Scores of 6 or more within 3 days of the bacteraemia indicate a poorer prognosis in
bacteraemia but this data is continuously collected and may be difficult to link as single
values. The most difficult area to address is usually the unequivocal assessment of
outcome. Mortality is associated with poor functional state and co-morbidities, which
may link to age and have been assessed automatically from computerized discharge
records of diagnoses (ICD or Diagnosis-related group codes) in the US 493 and France 494.
Defining mortality at a point less than 30 days after bacteraemia could tighten linkages to resistance and inappropriate prescribing, and should be studied. Acute renal injury is also a useful outcome measure as is subsequent development of *C. difficile* infection within 28 days. Sometimes these linkages can be made expediently without linking systems by exporting data and linking it in data bases or spreadsheets but the mechanics of this should not be dependent directly and solely on infection specialists, although they must advise on what should be done.

Quality and commissioning organisations should ensure hospitals are collecting and analysing all such data to explain and improve their results in the treatment of serious infections such as bacteraemias not just those with MDR GNB. Particular scrutiny of year-on-year improvement in outcome of bacteraemia and reduction in prevalence according to onset in hospital or the community is needed both in CCGs and hospitals. Application of enhanced definitions of place of likely acquisition together with the working party’s definitions of multi-resistance as applied to hospitals and the community and within the context of the local communities population make-up, may explain the reasons for, and sometimes enable multi-faceted action on, problematic multiple resistance as a whole health economy approach. Hospital-, community-healthcare and community-onset bacteraemia therefore require separate analysis.

**Evidence**

Key components of an effective antimicrobial stewardship programme are consistent effort and audit of outcome by specialists with full communication and support from electronic prescribing/laboratory and clinical records. Computerised systems can and should be integrated. Also required are full accountability of responsible organisations for occurrence of serious infections, and the outcomes of treating them.
information is required on serious infections with MDR GNB but must not be assessed in isolation. Evidence level: 2+

Hospital or community antibiotic use (by DDDs, or perhaps better in the context of resistance selection, number of patients exposed to each agent), should be reviewed locally together with antibiotic resistance data. These data sets are available from pharmacy and microbiology systems respectively. Audit on compliance with local guidelines can be undertaken, but this provides no assurance on clinical outcome in severe infections: these require comparison with performance of other similar institutions and analysis to ensure the quality of care. Evidence level: 2++

Extended surveillance of bacteraemia with appropriate record linkage both centrally and in the hospital would provide clinical outcome assurance in the most severe infections and also a means of comparing improvement in hospitals and communities. Further this would lead to a sharp focus on improvements to antibiotic guidance, usage and infection control Evidence level: 2+

Recommendations

- Ensure production of local guidelines for empirical and definitive antibiotic use, regularly updated for community-, wider healthcare-, and hospital- onset infections, and audit compliance with these.

Grading: Conditional recommendation for
• Integrate hospital IT to deliver annually linked data for each bacteraemia,
  including patient demographics, whether the bacteraemias onset was in the
  community, wider healthcare or hospital, antibiotic resistances of isolates,
  antibiotics prescribed, and maximum early warning score or occurrence of septic
  shock, and, if possible, defined time-limited (not admission-limited) mortality.

  Use these integrated data to review the adequacy of treatment of infection in
  communities and hospitals

  Grading: Good practice recommendation

• Central public health departments or the Chief Medical Officers should receive
  bacteraemia data from the jurisdictions of trusts and CCGs or equivalent primary
  care organisations. Annually, either peripherally or centrally they should ensure
  computerized record linkage to give dates of death to be added to, organism,
  specific antibiotic resistance and pattern, date of collection, nursing home
  residency, optionally local records on last hospital discharge before bacteraemia.

  This data should be made available, for open interrogation and downloading,
  with rolling cumulative data within the health service. They should ensure
  information findings on mortality rate are categorized by locality (separately for
  hospitals and for community with associated separate wider healthcare data).

  Grading: Strong recommendation for

• Make publicly available tabulated incidence and outcome data for bacteraemia
  giving hospital onset data by region and hospital, and for community and wider
  healthcare outcome data by CCG or equivalent primary care organisation.

  Correlate this data with similar analysed and tabulated annual data on total
  antibiotic use and organism and antibiotic resistance in clinical infections.
Continuously monitor bacteraemia outcomes and antibiotic resistance by organism and devise improvement programmes to both, locally and appropriately within health economies.

Consider central production of unbiased national or regional data on true resistance rates in community-onset localized or systemic infections to guide national community antibiotic recommendations.

11.3 Antibiotic stewardship in the community and care homes to reduce MDR

Gram-negative infections

Several RCTs in the UK communities have shown that multifaceted interventions that included i) general practice staff education and ii) education of the patient through improving communication during the doctor-patient consultation have improved prescribing. There have also been several Cochrane reviews that included studies in hospitals, but which should be transferable to the community and care homes, aiming to improve antibiotic prescribing. In one Cochrane review, restrictive interventions (selective reporting of laboratory susceptibilities, formulary restriction, and antibiotic policy change strategies) had a greater effect in the short term in reducing use of broad spectrum antibiotics than persuasive interventions (distribution of educational materials; educational meetings; local consensus processes; educational outreach visits; local opinion leaders; reminders provided verbally, on paper or on computer; audit and feedback). However both were equally effective in controlling antibiotic use and antimicrobial resistance after 6 months. In a separate Cochrane review, printed
educational materials alone had an effect on the practice of healthcare professionals and patient health outcomes. Based on seven RCTs and 54 outcomes, the median absolute risk difference in categorical practice outcomes was 0.02 when printed educational materials were compared with no intervention (range from 0 to +0.11). Other Cochrane reviews show multifaceted interventions are more effective. Moreover, interventions that are based on cognitive theories and consider personal attitudes, subjective norms and perceived behavioural controls (confidence and other barriers) are more likely to be successful, e.g., posters raise awareness and change subjective norms but are ineffective when used alone.

In an audit and feedback process, an individual’s professional practice or performance is measured and then compared with professional standards or targets. The results of this comparison are then fed back to the individual. In general practices this will probably be via the medicine manager, local GP prescribing champions or in collaboration with local microbiologists. The aim is to encourage the individual to follow professional standards. A Cochrane review considered 82 comparisons from 49 studies of any health care interventions in which audit and feedback was core and evaluated effects on professional practice. There was a median 4.3% increase in healthcare professionals’ compliance with desired practice (interquartile range (IQR) 0.5% to 16%) when i) baseline performance was low, ii) the source was a supervisor or colleague iii) it was provided more than once, iv) it was delivered in both verbal and written formats, and v) when it included both explicit targets and an action plan. In addition, the effect size varied based on the clinical behaviour targeted by the intervention. An RCT evaluating a multifaceted intervention in English general practice aimed at improving antibiotic prescribing included feedback of practice level data on antibiotic prescribing and resistance: this led to a 4.2% fall in total antibiotic use. In some parts of the UK, audit with action plans, and intense infection control measures, have been associated
with falls in quinolones and cephalosporin use and resistance. Incentives attached
to action plans can be very effective but, without personal attitude changes, the change
may reverse when the incentive is reduced. Any audit indicators need to be well
monitored, as implementation of an effective multiple-intervention strategy achieved no
reduction of antibiotic prescription rates when deployed at a larger scale in general
practice: the authors attributed the failure to a less tight monitoring of the intervention
and audit. It is necessary to demonstrate by further study, that such interventions
can be effective at practice or hospital unit/hospital level.

Relevant outcomes, which should be monitored, include mortality from systemic
infections such as bacteraemia, hospital admission, emergency room attendance,
requirement for outpatient parenteral antibiotic therapy, re-consultation in person or
by telephone, time-limited re-prescription of antibiotics and microbiological and clinical
persistence of infection.

Evidence
Restrictive and persuasive interventions are equally effective in controlling antibiotic
use and antimicrobial resistance and a multi-faceted approach is most effective
Evidence level: 1+
Audit and feedback interventions result in an increase in healthcare professionals’
compliance with desired practice
Evidence level: 1++
Local and national surveillance data are needed to determine appropriate empirical
antibiotic guidelines.
Evidence level: 3
Collection and analysis of outcome data is important in assessment of measures needed
to improve the management of infection and to reduce the increase in antibiotic use and
resistance.
Recommendations

- Use persuasive and restrictive interventions to reduce the total antibiotic consumption, particularly broad-spectrum antibiotics in the community and care homes.

  Grading: Strong recommendation for

- Provide and use active feedback of monitoring to prescribers, and nursing staff ensuring optimization of clinical, microbiological, and antimicrobial prescribing outcomes. Use audit and feedback to reduce inappropriate antimicrobial use in the community and wider healthcare.

  Grading: Strong recommendation for

- Review outcome data linked to antibiotic prescribing to improve quality of care in the community and care homes.

  Grading: Conditional recommendation for

12 Conclusions

The selection of antibiotics for the treatment of infections caused by Gram-negative bacteria (GNB) has always been difficult. Following the introduction of the first antibiotics with activity against GNB such as tetracycline, chloramphenicol and streptomycin, introduced in the late 1940’s, resistance in *E. coli* causing urinary tract infection was observed at rates of 5-10% as early as 1953. Subsequently it emerged that Enterobacteriaceae can exchange and re-assort antibiotic resistance genes with great ease via plasmids, transposons, integrons and other mobile, or potentially mobile, genetic elements. This meant that resistances to antimicrobials no longer being used
were easily and stably maintained as the relevant resistance genes commonly become linked to, and compromise, antibiotics that remain in use. These linked resistances became transferable to a wider and more versatile range of strains.

As each class of new agent was introduced so resistance negated its reliable empirical use for the treatment of serious sepsis and also undermined any future reliance on the older agents. This is exemplified in the UK by the rise of plasmid mediated TEM beta-lactamase conferring resistance to ampicillin in the 1960's, aminoglycoside modifying enzymes conferring gentamicin resistance in the 1970's, extended spectrum TEM and SHV beta-lactamases conferring cephalosporin resistance in the 1980's and beginning in the 1990s CTX-M ESBLs, DNA gyrase mutations, and dihydrofolate reductases conferring resistance to third generation cephalosporins, fluoroquinolones and trimethoprim, respectively. We are now facing a similar process with carbapenems and polymyxins.

The bacterial ability to maintain older resistances may undermine any benefit from the introduction of more resolute antibiotic stewardship. Over-reliance on stewardship as the sole strategy for reducing MDR GNB may not be productive although reductions in antibiotic use if they are substantial enough to reduce selection in the human microflora for resistant strains are welcome. Use of a diversity of agents focused to proven bacterial infection may be more important than restricting entirely the use of certain antibiotics and classes. Empirical prescribing based on generic clinical diagnoses will also need to be safely reduced.

Because of widely differing usage of antibiotics active against GNB in both medicine and agriculture in different parts of the globe since the 1980's we have created widely differing rates of occurrence of MDR GNB in these different locations and in some cases between food animals and man. Furthermore the increasing recognition of restricted
extraintestinal pathogens in different species suggests that animal husbandry quality and control of these strains may be variable. Higher rates of MDR GNB pose therapeutic problems for those countries. In addition over the last decade the movement of people, goods and food has resulted in countries such as the UK meeting unpredictable and alarming appearances of MDR GNB by importation. Imported food-producing animals from overseas founder stock, and foodstuffs, need to be free of important antibiotic resistance in Gram negative bacilli to just as great an extent as returned travellers for biosecurity and as a foundation for enhanced antimicrobial stewardship.

In order to produce relevant guidelines for the empirical treatment of infections caused by MDR GNB an understanding of the local epidemiology and susceptibility patterns is essential. The unpredictability of horizontal gene transfer and nosocomial spread may necessitate specific guidelines being produced for individual hospitals/communities. The present guideline has attempted to assess the relative clinical efficacy of different agents. We have found very few good quality clinical trials to support treatment regimens, particularly for licensed older agents, formerly little-used, that have been re-introduced into regular use. Finding much more rapidly a mechanism to address this deficit in trials is an important overarching research objective as the existing pattern of industry-sponsored initial regulatory trials fails to address the need.

It is self-evident that selection of antibiotic treatment based on susceptibility testing is the optimum strategy for treating infections caused by MDR GNB. The initiative to develop and deploy molecular and rapid phenotypic susceptibility testing methods will help refine antibiotic usage. Any additional expense must be funded within the healthcare system for these to be introduced. Risk factor, rule-based prescribing for MDR GNB is unlikely to be sufficiently predictive alone for the reasons outlined above but risk-assessment of travel, household spread, and screening on admission to hospitals needs urgent improvement. However we have attempted to present an
evidence base and suggestions to support the development of local prescribing policies and possibly for the future application of such technologies and overall improvement in outcomes.

Over-reliance on empirical piperacillin/tazobactam, and for treatment failure meropenem, has and will drive selection for resistance to these agents, and UK health policy is attempting to contain this upsurge in usage. For patients presenting with serious sepsis convincingly caused by GNB and in the absence of prior exposure to healthcare in countries/hospitals with endemic carbapenemase producing Enterobacteriaceae, carbapenems remain the best empirical therapy with early and embedded shift to alternative definitive treatment. The overall prevalence of resistance in *E. coli* alone to piperacillin-tazobactam or gentamicin (approximately 10%) is the basis for this superiority of carbapenems although factors such as aminoglycoside toxicity and *C. difficile* risk must be considered. Combinations of these agents or cephalosporins without β-lactamase inhibitors increase antibiotic use and are unlikely to produce adequate activity against ESBLs because of resistance linkage. Algorithms for predicting accurately presence of ESBLs need urgent validation in the UK health service so piperacillin/tazobactam or gentamicin can be safely used to provide Gram-negative cover in their absence, and cephalosporin-BLI combinations in their presence thus diversify antibiotic use in serious infections within a stewardship framework. Use of piperacillin/tazobactam or existing licensed aminoglycosides as empirical therapy where ESBL-producing strains are prevalent such as after overseas travel or hospitalisation, in communities where such travel has been frequent, and hospital or nursing home exposure is unwise. Historical evidence suggests these agents continue to be appropriate for sepsis if these risk factors are not implicated.

In England, use of the Commissioning for Quality and Innovation (CQUIN) payments framework (or public health control of institutions and community healthcare) needs to
be sensitive to the requirement to have safe effective antibiotics to use in sepsis caused
by non-MDR GNB which remain the majority of GNB causing serious infections in UK
hospitals. The role and utility of the latest generation of BL/BLI combinations is yet to
fully emerge. The early reports of emergence of resistance to ceftazidime-avibactam in
KPC-3-producing carbapenem resistant Enterobacteriaceae is extremely ominous.
Nevertheless, at the moment new BL/BLIs and fosfomycin offer the only immediate new
help to treat the latest MDR GNB particularly for carbapenemase producers and ESBL-
producing GNB. Further development of BLI combinations for oral use is an urgent need
in primary care.

Initiatives are being put in place to address the paucity of new agents but they will take
time to give results which are by no means inevitable. A greater emphasis in
communities should be given to the better use of existing treatments for effective
treatment of complicated and upper UTI with prevention of bacteraemia and in
hospitals to an auditable improved outcome in well-defined groups of patients with life-
threatening Gram-negative infections such as bacteraemia. This effort should match the
attention given to reducing inappropriate use of wide-spectrum agents for less
important infections and should ensure that reductions in antibiotic use are appropriate
and do not adversely affect patients. Computerised support to spare infection
professional time is necessary locally for surveillance of bacteraemia to focus attention
on improvements in performance in life-threatening infection.

Greater research and deployment efforts in the area of very rapid diagnostics to guide
immediate prescribing are needed. In the healthcare environment stopping spread of
infection with MDR GNBs is of paramount importance and such infection control
measures have been dealt with comprehensively in another working-party publication.

Accepted manuscript
The greatest long-term threat arises from the fundamental epidemiology of GNB, with their large faecal reservoirs in both humans and food animals leading to dissemination into the environment. This leads to unpredictable acquisition by individuals with high rates of commensal carriage and subsequent infection. Not only antibiotic control in man but parallel control of use of the same agents in food animals is important. This is exemplified by use of colistin, mequindox and fosfomycin in food animals in China and other parts of the world, and consequent emergence of plasmid-mediated colistin, nitrofurantoin and fosfomycin resistance mediated by \textit{mcr-1}, \textit{oqxAB} and modified nitroreductases, and \textit{fosA} as discussed previously (See 6.3.4). The close association of NDM MBL with connections with the Indian sub-continent is likely to change with the demonstration of this carbapenemase in poultry, farm workers, flies and wild birds in Shandong, China. Practical measures to contain human importations of carbapenemases but also assessment and potentially prevention of any spread in foodstuffs are urgent at this early stage. Variations in the prevalence of MDR GNB in different localities and cultural backgrounds even within the UK need to be further explored and considered in empirical therapy. Separate effects of migration, travel, household cross- colonization/infection and food consumption need to be rapidly studied to make risk assessments practical and effective.

Internationally, public health hygiene measures to reduce faecal oral transmission such as clean water initiatives and sewerage and irrigation systems to prevent transmission are of major importance. Food stuffs including imports should be regulated for the presence of GNB resistant to third-generation cephalosporins, quinolones and possibly in the future carbapenems. Failure to address these under-recognised threats will undo our ability to treat infections caused by MDR GNB. If we do not control human and agricultural use of antibiotics and the spread of MDR GNB from faeces back into humans.
and food animals as a consistent multi-faceted, global-scale, public-health programme, we will suffer greatly.

13 Further research and development

Without consideration of the research needed for new compounds and formulations in the antibiotic pipeline, there are numerous areas which require research with a 5 year horizon for completion.

- Diagnostic tests and or serum markers should be formally and comprehensively assessed for safety and efficacy as aids in deciding when to start and stop antimicrobial treatment, particularly in critically ill patients and those with haematological malignancies.

- Develop and introduce new cheap, rapid, and preferably bedside, diagnostic tests for important multiple antibiotic resistant organisms in urine and blood.

- Undertake RCT studies of antimicrobial agents (both new and old) in the treatment of Gram-negative infection in areas where multi-resistance is likely e.g. admissions unit, critical care and urology in hospitals and in treatment of infections due to ESBL-producing bacteria in the community. Identified research areas in this guideline include

  a. Use of continuous infusion meropenem at dose determined by nomogram if infection with KPC-carbapenemase –producing Klebsiella with MIC of >8<64mg/L.

  b. Use of temocillin for non-urinary infections with trials to establish their optimal dosage.
c. Use of temocillin alone, or in combination, in UTIs caused by Enterobacteriaceae with KPC-enzyme.

d. Use of ceftazidime/avibactam alone when non-MBL carbapenemase-producing organisms cause infection in comparison with alternatives, including combination therapy.

e. Use of ceftolozane/tazobactam in *P. aeruginosa* infections in cystic fibrosis.

f. *In vitro* and *in vivo* research to identify the usefulness of aztreonam in combination with avibactam for infections due to Enterobacteriaceae with MBLs and other carbapenemases.

g. Research into the role of loading doses of colistin, monitoring of serum levels and optimal combination therapy.

h. Research into use of polymyxin-containing and non-containing selective digestive decontamination regimens and the prevalence of newly identified polymyxin resistance mechanisms.

i. Optimal rapid and practical methods of colistin susceptibility testing outside intrinsically resistant species such as Proteaeae and *Serratia* spp.

j. Higher dosing studies with tigecycline to investigate if the unexpectedly high mortality in infections with strains that are apparently susceptible *in vitro*, can be reduced.

k. Optimal use of high dose tigecycline in combinations in hospital-acquired respiratory infections.
l. Specific system-based and resistance-mechanism-based indications for use of parenteral fosfomycin, in infections due to MDR GNB.

m. Cefixime (or other oral cephalosporin) with clavulanate (alone or with amoxicillin) against ESBL-producing *E. coli* UTI.

n. Nitrofurantoin versus fosfomycin trometamol versus pivmecillinam (with or without amoxicillin/clavulanate) in patients with ESBL-producing *E. coli* and *Klebsiella spp.*

o. Use of meropenem, or temocillin or ceftolozane/tazobactam in community onset pyelonephritis where hospitalisation is required and where MDR GNB excluding CPE are, or are likely to be, present. These studies should include assessment of meropenem or aminoglycosides if the patient describes penicillin-hypersensitivity.

- Undertake surveillance in both the hospital and community populations, and households of newly detected colonised individuals, for incidence of known mechanisms of resistance and the emergence of novel resistance mechanisms to currently used antimicrobials. Link this surveillance to travel, prior hospitalisation as in-patient, or residential healthcare.

- Develop new models of licensing and funding of antimicrobials for treating MDR GNB infections. Develop non-microbial therapies for MRGNB (e.g. phage, antibacterial peptides, etc.)

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16 Transparency declarations

The BSAC, BIA and HIS commissioned the authors to undertake the Working Party
Report. All authors but not the members of the patient advisory panel are, or have been,
members of one or more of these societies.

PH: Consultancy: BioMerieux, Becton-Dickinson, Eumedica, Merck, Novartis,
MagusCommunications, Pfizer, Wyeth; director of ModusMedica (medical education
company); Funded research: Astra-Zeneca, Merck, Novartis, and Pfizer.
REW: family shareholdings in Astra Zeneca, Bayer, GSK, Johnson & Johnson, Merck,
Pfizer and Roche amounting to approx. 15% of portfolio value.
CM: Travel expenses Merieux Diagnostics

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DML: Advisory Boards or ad-hoc consultancy Accelerate, Achaogen, Adenium, Allegra, AstraZeneca, Auspherix, Basilea, BioVersys, Centauri, Discuva, Meiji, Merck, Pfizer, Roche, Shionogi, Tetraphase, VenatoRx, Wockhardt, Zealand, Paid lectures – AstraZeneca, Beckman-Coulter, Cardiome, Merck and Nordic. Relevant shareholdings in Dechra, GSK, Merck, Perkin Elmer, Pfizer amounting to <10% of portfolio. Contract research: Achaogen, Allegra, AstraZeneca, Melinta, Meiji, Merck, Roche, Wockhardt.

DAE: Received funding to attend conferences from MSD, Eumedica, Gilead and Astellas.

JAO: Was employed part-time by Bioquell Ltd. during the preparation of this manuscript. He is now a consultant to Gama Healthcare and Pfizer Ltd. These consultancies began after this working party report was written.

APRW: Consultant on Drug Safety Monitoring Boards for Roche and Genentech.

Advisory Panel for 3M.

All other authors no conflicts declared.
### Table 1. Summary of recommendations for stakeholders including prescribers

<table>
<thead>
<tr>
<th>Organisation</th>
<th>Recommendation</th>
<th>Strength</th>
</tr>
</thead>
<tbody>
<tr>
<td>Central public health authorities</td>
<td>Central public health departments or the Chief Medical Officers should receive bacteraemia data from the jurisdictions of trusts and CCGs or equivalent primary care organisations bacteraemia data in their localities. Annually, either peripherally or centrally they should ensure computerized record linkage to give dates of death. They should ensure information is categorized by locality (separately for hospitals and for community with associated separate wider healthcare data), date of onset or acquisition, organism, specific antibiotic resistance and pattern, the mortality rate. This data should be made available, for open interrogation, with rolling cumulative data within the health service.</td>
<td>Strong for</td>
</tr>
<tr>
<td></td>
<td>Make publicly available tabulated incidence and outcome data for bacteraemia giving hospital onset data by region and hospital, and for community and wider healthcare onset data by CCG or equivalent primary care organisations. Correlate this data with similar analysed and tabulated annual data on total antibiotic use and organisms and antibiotic resistance in clinical infections.</td>
<td>Good practise</td>
</tr>
<tr>
<td></td>
<td>Consider central production of unbiased national or regional data on true resistance rates in community-onset localized or systemic infections to guide national community antibiotic recommendations.</td>
<td>Strong for</td>
</tr>
<tr>
<td>Commissioning and quality organisations</td>
<td>Continuously monitor bacteraemia outcomes and antibiotic resistance by organism and devise improvement programmes to both, locally and appropriately within health economies.</td>
<td>Good practise</td>
</tr>
<tr>
<td></td>
<td>Provide and use active feedback of monitoring to prescribers, and nursing staff ensuring optimization of clinical, microbiological, and antimicrobial prescribing outcomes. Use audit and feedback to reduce inappropriate antimicrobial use in the community and wider healthcare.</td>
<td>Conditional for</td>
</tr>
<tr>
<td></td>
<td>Use persuasive and restrictive interventions to reduce the total antibiotic consumption, particularly broad-spectrum antibiotics in the, community and care home setting.</td>
<td>Strong</td>
</tr>
<tr>
<td>Hospital and primary care: general</td>
<td>Ensure production of local guidelines for empirical and definitive antibiotic use, regularly updated for community-, wider healthcare-, and hospital-onset infections and audit compliance with these.</td>
<td>Conditional for</td>
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<td>-------------------------------</td>
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</tr>
<tr>
<td>Provide an on-going antimicrobial stewardship programme in all care settings, based on resistance rates, with audit of compliance with guidelines, surveillance of outcome, and active feedback.</td>
<td>Strong</td>
<td></td>
</tr>
<tr>
<td>Identify through horizon scanning, and make available, and make available new antimicrobials that may be required to treat MDR GNB. Monitor use through formulary/drug and therapeutics committees.</td>
<td>Conditional for</td>
<td></td>
</tr>
<tr>
<td>Use restrictive prescribing policies to acutely reduce the incidence of infection or colonisation with MDR GNB; thereafter, maintain persuasive and restrictive approaches and monitor that gains persist.</td>
<td>Strong for</td>
<td></td>
</tr>
<tr>
<td>Integrate hospital IT to deliver annually linked data for each bacteraemia, including patient demographics, whether the bacteraemias onset was in the community, wider healthcare or hospital, antibiotic resistances of isolate, antibiotics prescribed, and maximum early warning score or occurrence of septic shock, and if possible defined time-limited (not admission-limited) mortality. Use these integrated data to review the adequacy of treatment of infection in communities and hospitals</td>
<td>Good practise</td>
<td></td>
</tr>
<tr>
<td>Hospital &amp; primary care treatment of UTI</td>
<td>Inspect up-to-date national and local antibiotic surveillance when compiling local antibiotic guidelines on treatment of UTI. Follow local guidance on what antibiotics to prescribe,</td>
<td>Strong for</td>
</tr>
<tr>
<td>For an elderly patient, do NOT send urine for culture or start empirical antibiotics unless there are specific symptoms or signs of UTI and none elsewhere. Use the algorithm in Figure 5 to decide whether to do this in elderly patients especially in those with dementia</td>
<td>Conditional for</td>
<td></td>
</tr>
<tr>
<td><strong>Do not prescribe antibiotics in asymptomatic bacteriuria (ASB) in the elderly with, or without, an indwelling catheter.</strong></td>
<td>Strong for</td>
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<tr>
<td><strong>Always consider the positive and negative predictive value of specific symptoms before sending urine for culture or starting antibiotics for a UTI. Base decision on when to prescribe (whatever the age) primarily on symptoms. Use dipstick tests, if no catheter is present, to confirm the diagnosis, before prescribing especially when symptoms are mild or not localized.</strong></td>
<td>Strong for</td>
<td></td>
</tr>
<tr>
<td><strong>If there are risk factors for MDR GNB or previous presence of MDR GNB and the patient is symptomatic, send a urine specimen for culture and susceptibility</strong></td>
<td>Strong for</td>
<td></td>
</tr>
<tr>
<td><strong>Building on previous work, predictive scoring should be developed for the presence of ESBL-producing <em>E. coli</em> in primary care and on admission to hospital to restrict the need to prescribe carbapenems and other antimicrobial agents generally active against ESBLs</strong></td>
<td>Strong for</td>
<td></td>
</tr>
<tr>
<td><strong>Need to quantify risks of infection with/ carriage of, extraintestinal pathogenic <em>E. coli</em> and of <em>Klebsiella sp.</em> resistant to all antibiotics and relate to time since travel to countries with high prevalence of MDR GNB and incorporate in risk assessments for clinical infection with MDR GNB in the community and on admission to hospital to guide therapy</strong></td>
<td>Strong for</td>
<td></td>
</tr>
<tr>
<td><strong>If defined risk factors for MDR GNB are present avoid cephalosporins, quinolones, trimethoprim and co-amoxiclav in treatment of lower UTIs unless the pathogens are confirmed to be susceptible.</strong></td>
<td>Strong for</td>
<td></td>
</tr>
<tr>
<td><strong>Personalise empirical chemotherapy for each patient by considering current features of bacteraemia, risk factors for antibiotic resistance and past susceptibility testing including the presence of MDR GNB in the patient, hospital unit, nursing home, or community.</strong></td>
<td>Conditional for</td>
<td></td>
</tr>
<tr>
<td><strong>In pyelonephritis always collect a urine sample before treatment. MDR GNB are unlikely to respond to oral treatment so consider risk factors for MDR GNB including travel. Use an active oral agent only if patient is well enough and if known to have had ciprofloxacin-, trimethoprim-, or co-amoxiclav-susceptible MDR GNB in last month.</strong></td>
<td>Conditional for</td>
<td></td>
</tr>
<tr>
<td>If the patient has pyelonephritis and risk factors for MDR GNB, start, if hospitalisation not required, empirical intravenous therapy with ertapenem if OPAT therapy available. This will treat ESBL and Amp-C producing Enterobacteriaceae. If hospitalisation required for this or OPAT not available, admit for meropenem, temocillin or ceftolozane/tazobactam if no evidence of CPE organism. If the patient is penicillin-hypersensitive then the hospital may use amikacin or meropenem, or if only susceptible isolates in the past, gentamicin. If carbapenem-resistant bacteria are, or have been, present, base treatment on susceptibility testing of recent or current isolates.</td>
<td>Strong for</td>
<td></td>
</tr>
<tr>
<td>---</td>
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<td></td>
</tr>
<tr>
<td>Locally assess the true rate of resistance and determine from this when changes to guideline recommendations for empirical therapy for UTI in guidelines are necessary including recommendations where the risk of antibiotic-resistant bacteraemia is high.</td>
<td>Conditional for</td>
<td></td>
</tr>
<tr>
<td>Primary care prescriber for UTI</td>
<td>Always inform the patient or their carer(s) on what to look out for and how to reconsult if symptoms worsen or do not improve as community-onset <em>E. coli</em> bacteraemias of urinary origin are increasing</td>
<td>Strong for</td>
</tr>
<tr>
<td>In younger women with acute uncomplicated UTI, only consider MDR GNB in choosing empirical treatment if there are risk factors See Section 9.3.1. or recent foreign travel to countries where such strains are highly prevalent.</td>
<td>Strong for</td>
<td></td>
</tr>
<tr>
<td>Use fosfomycin, nitrofurantoin or pivmecillinam, guided where possible i) by susceptibility testing and ii) by this guideline's recommendation on choice, dosing and duration, for uncomplicated lower urinary tract infection where MDR GNB are suspected.</td>
<td>Strong for</td>
<td></td>
</tr>
<tr>
<td>Use nitrofurantoin for 5 days with MDR GNB. Alternatively use fosfomycin trometamol 3g orally as single dose, and repeat on third day only if MDR GNB confirmed to improve bacteriological cure. Pivmecillinam alone at 200mg three times daily for 7 days may be a third line choice but consider combination use with amoxicillin/clavulanate depending on clinical trial results at the time.</td>
<td>Conditional for</td>
<td></td>
</tr>
<tr>
<td>Review outcome data linked to antibiotic prescribing to improve quality of care in the community and care homes</td>
<td>Conditional for</td>
<td></td>
</tr>
<tr>
<td>---</td>
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<td></td>
</tr>
<tr>
<td>To reduce recurrent UTI, consider firstly, the option of pre-prescribed standby antibiotics to take when symptoms begin, rather than daily or post-coital antibiotic prophylaxis. Where prophylaxis is used successfully for recurrent infection in adults limit use to six months.</td>
<td>Conditional for</td>
<td></td>
</tr>
<tr>
<td>Avoid antibiotic prophylaxis for urinary catheter insertion or changes unless there is previous history of symptomatic UTI with the procedure, insertion of incontinence implant, or trauma at catheterization.</td>
<td>Conditional for</td>
<td></td>
</tr>
</tbody>
</table>
Table 2 Summary recommendations for specific antibiotics
<table>
<thead>
<tr>
<th>Antibiotic</th>
<th>Guidance</th>
<th>Strength</th>
</tr>
</thead>
<tbody>
<tr>
<td>Amikacin</td>
<td>Modernise use of amikacin, which has improved activity, with development of validated nomograms. Ensure assays are readily available before repeat doses and consider, because of the risks of toxicity, the practicality of monitoring with audiograms.</td>
<td>Conditional for</td>
</tr>
<tr>
<td>Amoxicillin/clavulanate</td>
<td>Use for lower UTI due to known ESBL-producing bacteria only if current isolates, or if using empirically, recent isolates, are fully susceptible.</td>
<td>Conditional for</td>
</tr>
<tr>
<td>Ampicillin/sulbactam</td>
<td>Could use against some carbapenem-resistant apparently sulbactam-susceptible A. baumannii isolates, Caution needed in the UK because of a higher range of MICs. Absence of a breakpoint prevents categorisation as susceptible/resistant.</td>
<td>Conditional for</td>
</tr>
<tr>
<td>Aztreonam</td>
<td>Do not use aztreonam alone empirically if MDR GNB or Gram-positive or anaerobic pathogens are suspected.</td>
<td>Strong against</td>
</tr>
<tr>
<td></td>
<td>Do not use aztreonam for CTX-M ESBL- or AmpC- producing bacteria even if these appear susceptible in vitro</td>
<td>Strong against</td>
</tr>
<tr>
<td></td>
<td>Use aztreonam for MBL- or OXA-48- producing strains if it is certain that they do not produce ESBLs or AmpC</td>
<td>Strong for</td>
</tr>
<tr>
<td></td>
<td>Research usefulness of aztreonam in combination with avibactam for bacteria producing MBLs with ESBL/AmpC enzymes and for those with other carbapenemases.</td>
<td>Conditional for Research</td>
</tr>
<tr>
<td>Cefepime</td>
<td>Could use cefepime to treat infection caused by ESBL- or Amp-C-producing bacteria if susceptible to the EUCAST breakpoint of MIC &lt;=1 mg/L</td>
<td>Conditional for</td>
</tr>
<tr>
<td></td>
<td>Do not use cefepime even at increased dose for isolates with i) MIC of 2-8 mg/l (CLSI “susceptible dose dependent”) or ii) MIC 2-4mg/L (EUCAST intermediate, or iii) strains with stable derepression of AmpC or iv) strains that produce both AmpC and ESBLs.</td>
<td>Strong against</td>
</tr>
<tr>
<td>Medication</td>
<td>Recommendation</td>
<td>Notes</td>
</tr>
<tr>
<td>------------------------------------</td>
<td>---------------------------------------------------------------------------------------------------</td>
<td>---------------------------</td>
</tr>
<tr>
<td>Cefepime</td>
<td>Do not use cefepime to treat infection caused by carbapenemase-producing Enterobacteriaceae</td>
<td>Strong against</td>
</tr>
<tr>
<td>Cefixime and other oral cephalosporins</td>
<td>Do not use cefixime and other oral cephalosporins to treat infection caused by ESBL, AmpC and carbapenemase-producing Enterobacteriaceae</td>
<td>Conditional</td>
</tr>
<tr>
<td>Cefoxitin</td>
<td>Confirmation needed of its usefulness as a carbapenem-sparing agent for in-patients to empirically treat urinary infection or use definitively for infections caused by CTX-M-15-producing <em>E. coli</em>: its short serum half-life means it is unsuitable for OPAT and probably it has insufficient advantage to displace existing agents.</td>
<td>Research and trials</td>
</tr>
<tr>
<td>Ceftazidime</td>
<td>Use ceftazidime for susceptible infections with <em>P. aeruginosa</em> including quinolone- or some imipenem- resistant strains</td>
<td>Strong for</td>
</tr>
<tr>
<td></td>
<td>Do not use ceftazidime to treat infections due to ESBL- or AmpC-producing Enterobacteriaceae or CPE (other than OXA-48 producers), even if <em>in vitro</em> tests suggest the isolate is susceptible</td>
<td>Conditional against</td>
</tr>
<tr>
<td>Ceftazidime/avibactam</td>
<td>Could use ceftazidime/avibactam as an alternative to carbapenems for infection with ESBL- and AmpC- producing Enterobacteriaceae but alternatives may be cheaper</td>
<td>Conditional for</td>
</tr>
<tr>
<td></td>
<td>Evaluate further ceftazidime/avibactam use alone or in combination when non-MBL carbapenemase-producing organisms cause infection. KPC-3 producing Klebsiella are vulnerable to mutations in the enzyme causing resistance</td>
<td>Research and trials</td>
</tr>
<tr>
<td></td>
<td>Consider if ceftazidime/avibactam should be used with a carbapenem or colistin to treat infections with KPC3-producers based on latest evidence at the time of use</td>
<td>Research and trials</td>
</tr>
<tr>
<td>Antibiotic</td>
<td>Indications</td>
<td>Notes</td>
</tr>
<tr>
<td>------------</td>
<td>-------------</td>
<td>-------</td>
</tr>
<tr>
<td>Ceftolozane/tazobactam</td>
<td>Do not use for treating infection with anaerobes or bacteria producing MBLs: these are resistant</td>
<td>Strong against</td>
</tr>
<tr>
<td>Ceftolozane/tazobactam</td>
<td>Use ceftolozane/tazobactam to treat susceptible infections with <em>P. aeruginosa</em> resistant to ceftazidime</td>
<td>Conditional for</td>
</tr>
<tr>
<td>Ceftolozane/tazobactam</td>
<td>Conduct clinical trials in <em>P. aeruginosa</em> infections in cystic fibrosis</td>
<td>Research and trials</td>
</tr>
<tr>
<td>Ceftolozane/tazobactam</td>
<td>Use ceftolozane- tazobactam as an alternative to carbapenems to treat urinary or intra-abdominal infection involving ESBL-producing <em>E. coli</em>. Caution may be needed when treating infections with ESBL-producing <em>Klebsiella spp.</em> owing to a higher resistance rate.</td>
<td>Conditional for</td>
</tr>
<tr>
<td>Ceftolozane/tazobactam</td>
<td>Do not use for infections due to AmpC- or carbapenemase- producing <em>Enterobacteriaceae</em> or MBL/ESBL- producing <em>P. aeruginosa</em>.</td>
<td>Strong against</td>
</tr>
<tr>
<td>Ertapenem</td>
<td>Use ertapenem to treat serious infections with ESBL and AmpC-producing <em>Enterobacteriaceae</em>.</td>
<td>Strong for</td>
</tr>
<tr>
<td>Ertapenem</td>
<td>Apply antibiotic stewardship to use of all carbapenems to minimize the risk of developing resistance either by acquisition of carbapenemase-producing strains or by porin loss.</td>
<td>Strong for</td>
</tr>
<tr>
<td>Ertapenem</td>
<td>Preferred carbapenem for outpatient antibiotic treatment (OPAT) of susceptible infections in view of the once daily dosing regimen</td>
<td>Conditional for</td>
</tr>
<tr>
<td>Fluoroquinolones</td>
<td>Could use orally to treat UTI caused by MDR GNB that are susceptible</td>
<td>Conditional for</td>
</tr>
<tr>
<td><strong>Fosfomycin</strong></td>
<td>Use in the treatment of lower UTI due to MDR Enterobacteriaceae. Oral formulation available is useful for ESBL producers after repeated recurrence after nitrofurantoin and potentially for carbapenemase-producers</td>
<td>Conditional for</td>
</tr>
<tr>
<td></td>
<td>Consider dosage and trials of oral formulation for upper UTI</td>
<td>Research and trials</td>
</tr>
<tr>
<td></td>
<td>Consider parenteral fosfomycin, probably in combination, as part of salvage treatment for susceptible MDR GNB: clear indications for use are not yet established. Potential drug of last resort</td>
<td>Research and trials</td>
</tr>
<tr>
<td></td>
<td>Need comparative clinical trials to establish optimal indications for, and optimal use of, oral and parenteral drug.</td>
<td>Research and trials</td>
</tr>
<tr>
<td></td>
<td>Carry out ongoing local and national surveillance of use and resistance because of previous emergence of bacterial resistance in populations and the drug’s potential as an important parenteral agent.</td>
<td>Strong for</td>
</tr>
<tr>
<td><strong>Gentamicin</strong></td>
<td>Could use gentamicin empirically in the UK if the likelihood of MDR GNB is low.</td>
<td>Conditional for</td>
</tr>
<tr>
<td></td>
<td>Could use gentamicin as a carbapenem sparing agent for urinary, intra-abdominal and bacteraemic infections due to ESBL-producing <em>E. coli</em> when susceptibility is confirmed but do not use empirically if the risk of MDR GNB is raised</td>
<td>Conditional for</td>
</tr>
<tr>
<td></td>
<td>Could use gentamicin in combinations for urinary, intra-abdominal and bacteraemic infections due to gentamicin-susceptible KPC-producing <em>Klebsiella spp.</em> if strain is resistant to colistin and meropenem (See Section 7.18).</td>
<td>Conditional for</td>
</tr>
<tr>
<td></td>
<td>Use once daily dosage of gentamicin or tobramycin if no renal impairment, followed by measurement of levels 6 to 14 hours post dose and adjust repeat dosage by reference to the appropriate 7mg/kg or 5mg/kg nomogram. Consider</td>
<td>Strong for</td>
</tr>
<tr>
<td>Drug</td>
<td>Recommendation</td>
<td>Strength</td>
</tr>
<tr>
<td>-----------------------</td>
<td>-----------------------------------------------------------------------------------------------------</td>
<td>----------</td>
</tr>
<tr>
<td>Imipenem &amp; Meropenem</td>
<td>Use meropenem or imipenem or ertapenem to treat serious infections with ESBL and AmpC-producing Enterobacteriaceae.</td>
<td>Strong for</td>
</tr>
<tr>
<td></td>
<td>Apply antibiotic stewardship to use of all carbapenems to minimize the risk of developing resistance either by acquisition of carbapenemase-producing strains or, with ertapenem, by porin loss.</td>
<td>Strong for</td>
</tr>
<tr>
<td></td>
<td>Do not use imipenem to treat susceptible Pseudomonas infections</td>
<td>Conditional for</td>
</tr>
<tr>
<td></td>
<td>Introduce in the UK mandatory reporting of meropenem- or imipenem- resistant Enterobacteriaceae from all anatomical sites and specimens.</td>
<td>Strong for</td>
</tr>
<tr>
<td></td>
<td>Test all meropenem- or imipenem- resistant isolates of Enterobacteriaceae immediately for the precise level of resistance and for an indication of the responsible class of carbapenemase. Submit to agreed reference laboratories to determine susceptibility to a wide range of potentially active agents including, as appropriate, colistin, ceftazidime/avibactam, temocillin, aminoglycosides, fosfomycin and tigecycline.</td>
<td>Strong for</td>
</tr>
<tr>
<td></td>
<td>Consider use of continuous infusion meropenem in combination at dose determined by nomogram if infection with KPC-carbapenemase –producing Klebsiella with MIC of &gt;8 &amp; &lt;64mg/L.</td>
<td>Research and trials</td>
</tr>
<tr>
<td>Nitrofurantoin</td>
<td>Could use nitrofurantoin for 5 days to treat uncomplicated, lower urinary tract infections with nitrofurantoin-susceptible MDR E. coli (not Proteeeae or P. aeruginosa).</td>
<td>Strong for</td>
</tr>
<tr>
<td>Condition</td>
<td>Action</td>
<td>Notes</td>
</tr>
<tr>
<td>-----------</td>
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</tr>
<tr>
<td>Do not use repeatedly if there is moderate renal impairment (eGFR&lt;45mks/min/1.73m²), or in long-term courses, as these are associated with rare unwanted pulmonary effects.</td>
<td>Conditional against</td>
<td></td>
</tr>
<tr>
<td>Use alternative agents if there are repeated recurrences with MDR GNB but do not anticipate the emergence of resistance in E. coli infections on a single recurrence as selection for resistant strains in the urine or faecal flora is rare.</td>
<td>Conditional for</td>
<td></td>
</tr>
<tr>
<td>Need comparative studies of nitrofurantoin and other active antimicrobials in patients with ESBL-producing E. coli and Klebsiella spp.</td>
<td>Research and trials</td>
<td></td>
</tr>
<tr>
<td><strong>Piperacillin/tazobactam</strong></td>
<td>Use for infections with known ESBL-producing bacteria only if current isolates, or, if using empirically, isolates from the recent past, are fully susceptible by EUCAST criteria.</td>
<td>Conditional for</td>
</tr>
<tr>
<td>Consider definitive use of piperacillin/tazobactam to treat infections caused by <em>P. aeruginosa</em> if susceptible by EUCAST criteria.</td>
<td>Conditional for</td>
<td></td>
</tr>
<tr>
<td><strong>Pivmecillinam</strong></td>
<td>Consideration should be given to reducing the mecillinam EUCAST breakpoint for classification of susceptibility.</td>
<td>Conditional for</td>
</tr>
<tr>
<td>Treat lower UTI due to ESBL-negative <em>E. coli</em> with pivmecillinam at 200mg three times daily: do not use for infections caused by Proteeeae, Klebsiella or <em>Pseudomonas</em>.</td>
<td>Conditional for</td>
<td></td>
</tr>
<tr>
<td>Some ESBL-producing <em>E. coli</em> respond, but efficacy is poor against CTX-M-15 &amp; OXA-1 enzyme producers: dosing at 400mg three times daily may be no more effective. Consider combination of the lower dose with 375mg three times daily amoxicillin/clavulanate for follow on to parenteral therapy for such infections in hospital or OPAT.</td>
<td>Conditional for</td>
<td></td>
</tr>
</tbody>
</table>
Requires clinical comparative trials in the public interest i) alone or together with amoxicillin/clavulanate for UTIs due to ESBL-producing organisms including particularly those producing CTX-M-15 enzymes ii) in uncomplicated lower UTI generally against fosfomycin trometamol and nitrofurantoin as the relative advantages of these drugs have not been directly compared over the last 10 years as MDR GNB have become more problematic.

<table>
<thead>
<tr>
<th>Polymyxins(including colistin)</th>
<th>Reserve intravenous colistin for infections due to polymyxin susceptible but multiresistant bacteria and preferably use in combination with other agents. Conditional for</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Give careful consideration to use of higher dosage regimens in critically ill patients Conditional for</td>
</tr>
<tr>
<td></td>
<td>Use colistin with meropenem to treat susceptible KPC-producing <em>Klebsiella spp.</em> if the meropenem MIC is ( \leq 8) mg/L and consider higher meropenem dose by continuous infusion if the MIC is ( &gt;8 ) and ( \leq 32) mg/L. Conditional for</td>
</tr>
<tr>
<td></td>
<td>Consider colistin with aminoglycosides or tigecycline in infections with strains producing KPC or other carbapenemases, which are susceptible to these but resistant to meropenem with MIC&gt;32mg/L. Conditional for</td>
</tr>
<tr>
<td></td>
<td>Closely monitor renal function especially in the elderly, those receiving high intravenous doses for prolonged periods and those on concomitant nephrotoxic agents e.g. aminoglycosides Strong for</td>
</tr>
<tr>
<td></td>
<td>Reconsider use of polymyxins in selective digestive decontamination regimens as these agents are now important last therapeutic options against carbapenemase-producing Enterobacteriaceae and are more threatened by resistance than previously appreciated Good practise</td>
</tr>
<tr>
<td><strong>Need research on optimal rapid and practical methods of susceptibility testing outside intrinsically resistant groups such as <em>Proteae</em> and <em>Serratia spp.</em></strong></td>
<td></td>
</tr>
<tr>
<td>Research and trials</td>
<td></td>
</tr>
<tr>
<td><strong>Aerosolised colistin dry powder should be used in cystic fibrosis according to NICE guidelines</strong> Use in combination in ventilator-associated pneumonia may be considered pending further trials without methodological flaws.</td>
<td></td>
</tr>
<tr>
<td>Conditional for</td>
<td></td>
</tr>
<tr>
<td><strong>Temocillin</strong></td>
<td></td>
</tr>
<tr>
<td>Use alone for UTIs and associated bacteraemia caused by AmpC- or ESBL-producing Enterobacteriaceae.</td>
<td></td>
</tr>
<tr>
<td>Conditional for</td>
<td></td>
</tr>
<tr>
<td>Continuous infusion or thrice-daily dosing may be desirable for systemic infections with ESBL- or Amp-C producing bacteria.</td>
<td></td>
</tr>
<tr>
<td>Research and trials</td>
<td></td>
</tr>
<tr>
<td>Could use for UTIs with KPC-producing Enterobacteriaceae but not for OXA-48 or MBL-producers, on basis of published in-vitro data.</td>
<td></td>
</tr>
<tr>
<td>Research and trials</td>
<td></td>
</tr>
<tr>
<td><strong>Tigecycline</strong></td>
<td></td>
</tr>
<tr>
<td>Could use tigecycline in combination in the treatment of multiresistant soft tissue and intra-abdominal infections</td>
<td></td>
</tr>
<tr>
<td>Conditional for</td>
<td></td>
</tr>
<tr>
<td>Use alone in hospital-acquired respiratory infections is unlicensed and not advised as outcomes with current dosing are not clearly satisfactory in Acinetobacter and MDR GNB infections.</td>
<td></td>
</tr>
<tr>
<td>Conditional against</td>
<td></td>
</tr>
<tr>
<td>Use in combinations in hospital-acquired respiratory infections: precise combinations depend on the antibiotic-susceptibility of the MDR GNB causing the infection.</td>
<td></td>
</tr>
<tr>
<td>Research and trials</td>
<td></td>
</tr>
<tr>
<td>Use higher-than licensed dosing such as 100mg twice daily for infections due to MDR GNB in critical care</td>
<td></td>
</tr>
<tr>
<td>Conditional for</td>
<td></td>
</tr>
<tr>
<td>Investigate if higher dosing counters the unexpectedly high mortality seen even in infections due to strains apparently susceptible <em>in vitro.</em></td>
<td></td>
</tr>
<tr>
<td>Research and trials</td>
<td></td>
</tr>
<tr>
<td>Medicine</td>
<td>Recommendation</td>
</tr>
<tr>
<td>--------------------------</td>
<td>-------------------------------------------------------------------------------</td>
</tr>
<tr>
<td>Tobramycin</td>
<td>Avoid tobramycin for MDR Enterobacteriaceae because of risk of resistance due to AAC (6')1 and AAC (6')-1b-cr</td>
</tr>
<tr>
<td></td>
<td>Use tobramycin in preference to other aminoglycosides for susceptible Pseudomonas infection</td>
</tr>
<tr>
<td></td>
<td>Use once daily dosage of tobramycin if no renal impairment followed by measurement of levels 6 to 14 hours post dose and adjust repeat dosage by reference to nomogram.</td>
</tr>
<tr>
<td>Trimethoprim</td>
<td>Do not use trimethoprim in treating MDR GNB or treatment failures with other agents unless <em>in vitro</em>-susceptibility has been demonstrated.</td>
</tr>
<tr>
<td></td>
<td>Do not use trimethoprim to treat lower UTIs as a first line agent. Only consider use if there are no risk factors for resistance, or confirmed, <em>in vitro</em> susceptibility</td>
</tr>
<tr>
<td>Trimethoprim/sulfamethoxazole</td>
<td>Use in treatment of infections due to susceptible <em>S. maltophilia</em> and consider in infections due to <em>Achromobacter spp.</em>, <em>Alcaligenes spp.</em>, <em>Burkholderi spp.</em>, <em>Chryeobacterium spp.</em> and <em>Elizabethkingia spp.</em></td>
</tr>
</tbody>
</table>
## Table 3 Levels of evidence for intervention studies

<table>
<thead>
<tr>
<th>Level</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>1++</td>
<td>High-quality meta-analyses, systematic reviews of RCTs or RCTs with a very low risk of bias</td>
</tr>
<tr>
<td>1+</td>
<td>Well-conducted meta-analyses, systematic reviews or RCTs with a low risk of bias</td>
</tr>
<tr>
<td>1-</td>
<td>Meta-analyses, systematic reviews or RCTs with a high risk of bias*</td>
</tr>
<tr>
<td>2+++</td>
<td>High-quality systematic reviews of case–control or cohort studies. High-quality case–control or cohort studies with a very low risk of confounding or bias and a high probability that the relationship is causal. Interrupted time series with a control group: (i) there is a clearly defined point in time when the intervention occurred; and (ii) at least three data points before and three data points after the intervention</td>
</tr>
<tr>
<td>2++</td>
<td>Well-conducted case–control or cohort studies with a low risk of confounding or bias and a moderate probability that the relationship is causal OR Controlled before–after studies with two or more intervention and control sites</td>
</tr>
<tr>
<td>2-</td>
<td>Case–control or cohort studies with a high risk of confounding or bias and a significant risk that the relationship is not causal. Interrupted time series without a parallel control group: (i) There is a clearly defined point in time when the intervention occurred; and (ii) at least three data points before and three data points after the intervention. Controlled before–after studies with one intervention and one control site</td>
</tr>
<tr>
<td>3</td>
<td>Non-analytic studies (e.g. uncontrolled before–after studies, case reports, case series)</td>
</tr>
<tr>
<td>4</td>
<td>Expert opinion. Legislation</td>
</tr>
</tbody>
</table>

*Studies with an evidence level of ‘1-’ and ‘2-’ should not be used as a basis for making a
recommendation.

RCT randomised controlled trial.
Table 4 Grading of Recommendations

<table>
<thead>
<tr>
<th>Recommendation</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Undesirable consequences clearly outweigh desirable consequences</td>
<td>Strong recommendation against</td>
</tr>
<tr>
<td>Undesirable consequences probably outweigh desirable consequences</td>
<td>Conditional recommendation against</td>
</tr>
<tr>
<td>Balance between desirable and undesirable consequences is closely balanced or uncertain.</td>
<td>Recommendation for research and possibly conditional recommendation for use restricted to trials</td>
</tr>
<tr>
<td>Desirable consequences probably outweigh undesirable consequences</td>
<td>Conditional recommendation for</td>
</tr>
<tr>
<td>Desirable consequences clearly outweigh undesirable consequences</td>
<td>Strong recommendation for</td>
</tr>
</tbody>
</table>
Table 5 Stability of various β-lactam antibiotics and different inhibitor activities against important β-lactamases found in MDR GNB

<table>
<thead>
<tr>
<th>Inhibitor</th>
<th>Enterobacteriaceae</th>
<th>Acinetobacter</th>
<th>Burkholderia</th>
<th>Pseudomonas</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>AmpC</td>
<td>TEM ESBL</td>
<td>SHV ESBL</td>
<td>CTX-M ESBL</td>
</tr>
<tr>
<td>clavulanate</td>
<td>Not inhibited</td>
<td>Inhibited</td>
<td>Inhibited</td>
<td>Inhibited</td>
</tr>
<tr>
<td>sulbactam</td>
<td>Not inhibited</td>
<td>Inhibited</td>
<td>Inhibited</td>
<td>Inhibited</td>
</tr>
<tr>
<td>tazobactam</td>
<td>Not inhibited+</td>
<td>Inhibited</td>
<td>Inhibited</td>
<td>Inhibited</td>
</tr>
<tr>
<td>avibactam</td>
<td>Inhibited</td>
<td>Inhibited</td>
<td>Inhibited</td>
<td>Inhibited</td>
</tr>
<tr>
<td>β-lactam</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>temocillin</td>
<td>Stable</td>
<td>Stable</td>
<td>Stable</td>
<td>Stable</td>
</tr>
<tr>
<td>piperacillin</td>
<td>Labile*</td>
<td>Labile</td>
<td>Labile</td>
<td>Labile</td>
</tr>
<tr>
<td>ceftazidime</td>
<td>Labile*</td>
<td>Labile</td>
<td>Labile</td>
<td>Labile</td>
</tr>
<tr>
<td>meropenem/imipenem</td>
<td>Stable</td>
<td>Stable</td>
<td>Stable</td>
<td>Stable</td>
</tr>
<tr>
<td>ertapenem</td>
<td>Moderately stable*</td>
<td>Stable</td>
<td>Stable</td>
<td>Stable</td>
</tr>
<tr>
<td>aztreonam</td>
<td>Labile*</td>
<td>Labile</td>
<td>Labile</td>
<td>Labile</td>
</tr>
<tr>
<td>mecillinam</td>
<td>Stable</td>
<td>Moderately stable</td>
<td>Labile</td>
<td>Moderately stable</td>
</tr>
</tbody>
</table>
except Morganella morganii

*May appear active if AmpC is inducible, as induce weakly

x Inhibition not reliable with KPC3
**Table 6 Studies of the efficacy of Colistin**

<table>
<thead>
<tr>
<th>Study</th>
<th>No of patients</th>
<th>Conditions treated</th>
<th>Pathogens</th>
<th>Duration (mean)</th>
<th>Outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td>Levin 1999\textsuperscript{305}</td>
<td>59</td>
<td>VAP 33%; UTI 20%; BSI 15%; CNS 8%</td>
<td>A. baumannii 65%; P. aeruginosa 35%</td>
<td>12 days</td>
<td>58% success overall. Worst in pneumonia group (25%)</td>
</tr>
<tr>
<td>Garnacho-Montero et al. 2003\textsuperscript{304}</td>
<td>21</td>
<td>VAP 100%</td>
<td>A. baumannii 100%</td>
<td>14 days</td>
<td>57% success</td>
</tr>
<tr>
<td>Linden et al. 2003\textsuperscript{306}</td>
<td>23</td>
<td>VAP 78%; BSI 35%; Intra-abdominal 26%</td>
<td>P. aeruginosa 100%</td>
<td>17 days</td>
<td>61% favourable</td>
</tr>
<tr>
<td>Markou et al. 2003\textsuperscript{307}</td>
<td>24</td>
<td>VAP 63%; Catheter related 12%; Meningitis 4%</td>
<td>A. baumannii 24%; P. aeruginosa 76%</td>
<td>13.5 days</td>
<td>73% success</td>
</tr>
<tr>
<td>Michalopoulos et al. 2005\textsuperscript{308}</td>
<td>43</td>
<td>VAP 73%; BSI 33%</td>
<td>A. baumannii 19%; P. aeruginosa 81%</td>
<td>18.6 days</td>
<td>69% clinical cure</td>
</tr>
<tr>
<td>Reina et al. 2005\textsuperscript{309}</td>
<td>55</td>
<td>VAP 53%; UTI 18%; BSI 16%</td>
<td>A. baumannii 65%; P. aeruginosa 35%</td>
<td>13 days</td>
<td>15% cure on day 6 of treatment</td>
</tr>
<tr>
<td>Koomanachaie et al 2007\textsuperscript{505}</td>
<td>78</td>
<td>VAP 58%; BSI 10%</td>
<td>A. baumannii 91%; P. aeruginosa 9%</td>
<td>12 days</td>
<td>81% clinical response</td>
</tr>
</tbody>
</table>

VAP ventilator associated pneumonia

UTI urinary tract infection

BSI bloodstream infection

CNS central nervous system
Figure 1 Flow chart of systematic review

Records identified through database searching
N = 2398

Additional records identified through other sources
N = 1

Records after duplicates removed
N = 2385

Records screened
N = 2385

Records excluded
N = 1902

Full-text articles assessed for eligibility
N = 2523

Full-text articles assessed for eligibility
N = 440

Full-text articles not retrieved
N = 6

Studies included in qualitative synthesis
N = 49

Studies included in qualitative synthesis
N = 16

Studies included in quantitative synthesis (meta-analysis)
N = 0

Full-text articles assessed for eligibility in September 2014
N = 114

Studies included in qualitative synthesis in September 2014
N = 16
Figure 2 – Carbapenemase-producing Enterobacteriaceae submitted to and confirmed by PHE-AMRHAI-Colindale from Laboratories in England.

Courtesy of Dr Katie Hopkins, Public Health England

In a national context, a regional non PHE centre in an area of KPC endemicity became active in 2014 and did not submit or report isolates
Figure 3 Suggested algorithm for the treatment of MDR Gram negative bacteria admitted to UK hospitals

Figure 4 Suggested algorithm for the treatment of UTI in the UK community likely to be due to MDR GNB.

Avoid cephalosporins without BLI, trimethoprim, quinolones

No past carbapenem-resistance

Resistance to carbapenem in past or past healthcare in high risk country according to local/national policy for resistance

**KPC-carbapenemase**
Colistin & meropenem (2G three times daily if past S or unknown)
Add 100mg twice daily tigecycline if unknown. **IF R** consider adding continuous meropenem or use ceftazidime avibactam with meropenem

**OXA-48**
Aztreonam or Ceftazidime or Ceftolozane/tazobactam
**IF R or unknown**
Ceftazidime/avibactam

**Metallo-B-carbapenemase**
Fosfomycin and colistin
Consider tigecycline
Use cotrimoxazole for Stenotrophomonas

**Outpatient:** Ertapenem
**Inpatient:** Meropenem or Meropenem-sparing:
Patient requires hospital admission

**Oral follow-on if mecillinam S**
Pivmecillinum with amoxicillin/clavulanate

**Outpatient:** Ertapenem
**eGFR >45ml/min/1.73 m²**
Piperacillin/tazobactam or Gentamicin or Amikacin
**Oral follow on**
Nitrofurantoin
Amoxicillin/clavulanate
Fosfomycin
1 Not nitrofurantoin if pyelonephritis or eGFR <45ml/min. or Age <50 years
2 Caution re prolonged/frequently repeated courses
3 Not fosfomycin if pyelonephritis
4 Unlike co-amoxiclav, 1st gen cephalosporins, fosfomycin, and pivmecillinam
ciprofloxacin is generally active against *Proteus vulgaris, Morganella and Providencia.*
Figure 5: Diagnostic algorithm for ordering urine cultures and starting antibiotics if positive for nursing home residents in the intervention arm in the Loeb trial. (Loeb 2005)

- Fever of >37.9°C (100°F) or 1.5°C (2.4°F) increase above baseline on at least two occasions over last 12 hours?
  - Yes
  - 2 or more symptoms or signs of non-urinary tract infection?
    - Yes
      - Order urine culture for one or more of following: Dysuria, Urinary catheter, Urgency, Flank pain, Shaking chills, Urinary incontinence, Frequency, Gross haematuria, Suprapubic pain
    - No
      - Urinary catheter?
        - Yes
          - Order urine culture for new onset burning urination or for two or more of following: Urgency, Flank pain, Shaking chills, Urinary incontinence, Frequency, Gross haematuria, Suprapubic pain
        - No
          - Order urine culture for one or more of following: New costovertebral tenderness, Rigors, New onset of delirium
  - No

- Results of urine culture?
  - >10^5 CFU/mL (positive) or pending
    - Urinary catheter?
      - Yes
        - Is there one or more of following? New costovertebral tenderness, Rigors, New onset of delirium, Fever*
      - No
        - Is there dysuria or two or more of following? Fever, Urgency, Flank pain, Urinary incontinence, Shaking chills, Frequency, Gross haematuria, Suprapubic pain
  - Negative (no growth or mixed)
    - No urinary tract infection?
      - Yes
        - If yes begin antibiotics*. If no, do not treat for UTI
      - No

*Respiratory symptoms include increased shortness of breath, increased cough, increased sputum production, new pleuritic chest pain. Gastrointestinal symptoms include nausea or vomiting, new abdominal pain, new onset of diarrhoea. Skin and soft tissue symptoms include new redness, warmth, swelling, purulent drainage.

¥ >37.9°C (100°F) or 1.5°C (2.4°F) above baseline on two occasions over last 12 hours

B Stop antibiotics if urine culture is negative or no pyuria is present
References


166. Tangden T, Adler M, Cars O, et al. Frequent emergence of porin-deficient subpopulations with reduced carbapenem susceptibility in ESBL-


Accepted manuscript 240


Accepted manuscript


334. Gordon NC, Wareham DW. A review of clinical and microbiological outcomes following treatment of infections involving multidrug-resistant


479. Plachouras D, Hopkins S. Antimicrobial stewardship: we know it works; time to make sure it is in place everywhere. *Cochrane Database Syst Rev* 2017; **2**: ED000119.


Appendix 1 – Glossary

AmpC \(\beta\)-lactamases: clinically important cephalosporinases encoded by the chromosomes of many Enterobacteriaceae or (less often) by plasmids. High-level expression confers resistance to penicillins (except temocillin), cephalosporins (except cefepime), aztreonam and penicillin- \(\beta\)-lactamase inhibitor combinations.

Antimicrobial: A substance that kills or inhibits the growth of microorganisms. This includes antibiotics and totally synthetic compounds.

Bacteraemia: The presence of micro-organisms in the blood stream

\(\beta\)-lactamases: Enzymes produced by some bacteria that confer resistance to \(\beta\)-lactam antibiotics such as penicillins and cephalosporins, by breaking down the central structure of the antibiotic.

Carbapenemases: These are \(\beta\)-lactamases that inactivate carbapenems such as meropenem; most also attack and confer resistance to penicillins and cephalosporins

CBA – (Controlled before and after study) is a more limited assessment than interrupted time series because it does not contain an initial pre-study period to examine underlying trends not a post-study period to assess the sustainability of trend, A cross-over study design may exclude bias due to sequential change,

CCG: Clinical Commissioning Group. This is a locality based authority in England responsible for primary care services and placing financial contracts with local hospitals for specific services
CQUIN: NHS England Commissioning for Quality and Innovation payments framework, to encourage care providers to share and continually improve how care is delivered and to achieve transparency and overall improvement in healthcare.

Cluster randomized controlled clinical trial. This is a trial where groups of individuals rather than individuals are randomized to treatment. This complex study design may reduce the chances of one patient’s treatment having an effect on detection of effects in a patient randomized to a different treatment in the same environment.

Colonization: Situation whereby microorganisms establish themselves in a particular environment, such as a body surface, without producing disease.

Community-acquired: infection that is acquired outside of hospitals.

Community-onset or community-associated: usually defined as infection or colonization detected in an outpatient or within 48 hours of hospital admission. Recommended to permit extension to 72 hours.

CCT – (Controlled clinical trial) A clinical trial where there is a comparative arm that is not randomized.

ESBL (extended-spectrum β-lactamase): β-Lactamases that attack cephalosporins with an oxyimino side chain, for example, cefotaxime, ceftriaxone, ceftazidime, ceftolozane as well as the oxyimino-monobactam aztreonam. Unlike AmpC β-lactamases (q.v.) they are inhibited by clavulanic acid and tazobactam and unlike carbapenemases (q.v.) they do not attack carbapenems. Avibactam inhibits them and AmpC β-lactamases.

Healthcare – associated (acquired): infection or colonization detected in an in-patient more than 48 hours after hospital admission or in a resident of a nursing (or residential) home. Recommended to permit extension to 72 hours.
Hospital-onset or Hospital-associated (-acquired): infection or colonization detected in an inpatient more than 48 hours after hospital admission. Recommended to permit extension to 72 hours.

IMP carbapenemase (of MBL class) prevalent particularly in Asia and Australia sometimes in association with a second carbapenemase (bla\textsubscript{KPC}) gene

Infection: Invasion by and multiplication of pathogenic microorganisms in the body, producing tissue injury and disease, requiring treatment.

ITS – (Interrupted time series). A series of sequential cases where an intervention is made in the middle of the study as in before and after studies but additional time periods before and after the two comparative periods are included to give information on prior trends and sustainability. There may be further interventions in the series similarly studied.

KPC Klebsiella pneumoniae carbapenemase-producing bacteria are drug-resistant Gram negative bacilli which spread rapidly and cause significant morbidity and mortality. They are the most prevalent carbapenemase producers encoded by the bla\textsubscript{KPC} gene, which can be found in other Gram negative species.

MBL (Metallo β-lactamase) producing Gram negative bacteria use a Zn\textsuperscript{2+} ion in expressing resistance to carbapenems and other B-lactams

MDR GNB – (Multi-drug resistant Gram-negative bacteria) are defined as bacteria resistant to at least three different antibiotic classes or susceptible to only one or two classes.
NDM New Delhi metallo β-lactamase is a carbapenemase located on a mobile genetic element \( \text{bla}_{\text{NDM-1}} \) and is found on plasmids of various sizes. It is found in various species making outbreaks more difficult to identify.

OXA-48 carbapenemases hydrolyze penicillins at a high level but carbapenems at a low level sparing broad spectrum cephalosporins and are no susceptible to β-lactamase inhibitors. Recognition in the laboratory can be difficult. The gene \( \text{bla}_{\text{OXA-48}} \) is carried on a transposon and can be in a plasmid or chromosome.

Outbreak: at least two similar (i.e. not distinct) cases related in time and place

Porins: These are proteins that span the outer membrane of Gram-negative bacteria and mycobacteria forming pores that allow the entry of small water-soluble molecules, including antibiotics.

RCT (randomised controlled trial). Trials where patient allocation to the control and test arms of the study are allocated at random. They can be open label where treating physicians know which arm a patient has been allocated to or blinded where this is not the case. The latter is less likely to be subject to bias.

VIM MBL is a carbapenemase predominantly found in \textit{Pseudomonas aeruginosa} but found in Enterobacteriaceae as well. The genes \( \text{bla}_{\text{VIM}} \) are located on mobile integrons.
Appendix 2 Remit scope and related NICE guidelines

Joint BSAC/HIS/BIA Working Party on Multi-resistant Gram-negative bacteria

2.1. Guideline title

Treatment of MDR Gram-negative bacteria – report from a Joint Working Party

Short title: Treatment of Multi-Drug-Resistant Gram negative bacteria

2.2. Clinical need for the guideline

Epidemiology

There are a rising number of MDR Gram-negative infections across community and hospital care and the dual problems of finding an appropriate antibiotic and preventing spread.

APRHAI has recently produced brief guidelines on infection control and treatment options for these infections.

There is significant interest attracted by the May 2010 BSAC conference examining the dearth of new antibiotics effective against Gram-negative bacteria.

The Department of Health’s recognised that whilst control of MRSA and C difficile has been relatively successful, Gram-negative infections have continued to increase.

Consequent to this is the surveillance subcommittee of APRHAI recommendation that E. coli bacteraemia be included in mandatory surveillance.

Current practice

Members of BSAC and HIS, with the knowledge of the Councils of each, have been discussing the issues surrounding the recent increase in infections with multi-resistant Gram-negative bacteria in UK hospitals.

Following discussions and consideration of the forthcoming APRHAI report we now believe it an appropriate time to set up a Joint Working Party to look at making authoritative recommendations both for treatment and prevention of transmission of these infections.

2.3. The remit

To examine and make recommendations both for treatment and prevention of transmission of multi-drug-resistant (MDR) Gram-negative infections, resulting in the publication of guidelines on:

- current epidemiology and infection control issues; and
therapeutic issues and antibiotic guidance for treating infections caused by MDR Gram-negative bacteria.

For the purposes of this Working Party, the remit will mainly include infections in critical and non-critical care patients in secondary care. However, the same general principles would apply in community settings, particularly in areas where inappropriate treatment is encouraging selection. Consideration will be given to laboratory testing and susceptibility testing, although only screening and confirmatory tests available in a general microbiology laboratory. The use of antibiotic combinations in the therapy of infections will be considered, both parenteral and oral agents.

2.4. The Guideline

The guideline development process is described on the NICE website and reproduced in Appendix 3. The Working Party will follow the SIGN process when developing guidance including the hosting of a national stakeholder meeting as part of the national stakeholder consultation process.

2.5. The Scope

Defines what the guideline will and will not examine and what the guideline developers will consider. The scope is based on the referral from the three Societies and is the final scope.

2.5.1. Population Groups that will be covered

a) Adults
   Particular consideration given to patients of 65 years and older, and people at high risk of acquiring multi-resistant bacteria such as those requiring care in hospital settings

b) Children over 1 month old

2.5.2. Key clinical issues that will be covered

a) Antimicrobial treatment of MDR Gram-negative infections

b) Antimicrobial stewardship

c) Epidemiology

d) Surveillance
e) Infection prevention: standards, hand and environmental hygiene, organizational structures

Clinical situations that will not be covered include:

- Cystic fibrosis
- Community outbreaks

2.5.3. Infections that will be covered

Those caused by the following organisms

*Escherichia coli*, *Klebsiella* spp. including *Klebsiella pneumoniae*, *Enterobacter* spp., *Pseudomonas aeruginosa*, *Acinetobacter* spp., *Proteus* spp., *Serratia* spp., *Citrobacter freundii*, *Morganella morgani*

Sexually transmitted infections, *Helicobacter* spp. *Salmonella* spp. and some anaerobes are Gram-negative and are increasingly resistant, but were excluded because relevant public health control actions are substantially different or they have not been researched.

2.5.4. Antibiotics that will be considered

*Standard antibiotics currently in use* such as most cephalosporins, coamoxiclav, piperacillin/tazobactam quinolones, temocillin (pivmecillinam is the oral formulation of mecillinam)

*Old antibiotics that have been re-introduced*: such as aminoglycosides (including gentamicin and amikacin), colistin, fosfomycin, nitrofurantoin

*Recently developed antibiotics*: tigecycline, cefepime, new B-lactam-B-lactamase inhibitor combinations and carbapenems or those new agents at preliminary stages of testing.

2.5.5. Healthcare settings

All settings in which NHS care is received

2.6. Main outcomes

Outputs will be the production of guidelines, which will be approved via a process of national consultation. The intention is to inform and guide practice but also to highlight areas where more research is needed. The following will be produced and published as indicated:

- Current epidemiology and infection control issues – *Journal of Hospital Infection*
Therapeutic issues and antibiotic guidance for treating infections caused by multi-resistant Gram-negatives – Journal of Antimicrobial Chemotherapy
In addition, it is expected that each Journal will carry a leading article or review article on the guidance that is published by the joint societies.

2.7. Recommendations for practice

Treatment
Surveillance
Screening
Prevention of transmission
Cleaning and environment

2.8. Economic aspects

Developers will take into account both clinical and cost effectiveness when making recommendations involving a choice between alternative interventions. Failure to implement the recommendations would result in greater costs in terms of life expectancy or quality. Screening and isolation will result in significant cost pressures where this is not currently practised, but these costs are set against reduced transmission and fewer cases needing antibiotic treatment. Prolonged isolation can have adverse effects on a patient’s psychological health, so may have additional unexpected costs.

2.9. Patient Representation and Equality

Patient representatives are invited to all meetings and involved in the writing and drafting of the guidelines. As part of these discussions potential impacts on equality of groups sharing protected characteristics are considered and incorporated into the guidelines. Health inequalities associated with socioeconomic factors and with inequities in access for groups to healthcare and social care are considered and opportunities identified to improve health.

2.10. Status

2.10.1 Scope

This is the final scope.
2.10.2 Timing

The development of the guideline recommendation began in July 2011.
Appendix 3 Guideline development process

3.1. Guidance document

3.2. Related NICE guidance


National Institute for Health and Care Excellence. Urinary Tract Infection in Adults. London: NICE; Quality standard [QS90] Published date: June 2015. Available at: https://www.nice.org.uk/guidance/qs90/chapter/introduction


3.3. Process followed
The subject was identified by the Scientific Development Committee of the Healthcare Infection Society in February 2011 and approved by HIS in May 2011. The BSAC Council agreed a similar proposal at the same time. BIA Council agreed to join in September 2011. The members were chosen to reflect the range of stakeholders and not limited to members of the three Societies. The questions were decided at the first meeting of the
Group in November 2011 from issues presented to the members and patient representatives by staff and patients in the preceding months. Each was debated by the Group before adoption. Enhance Reviews was paid for the search and data extraction. Working Party members were not paid except for travel expenses.

3.4. Conflict of Interests

Conflicts of interest were registered at the outset and renewed during the process. They are stated in the Transparency declaration of the Report. In the event of a potential conflict being identified, the Working Party agreed that the member should not contribute to the section affected. With one exception, no interests were declared that required any actions and this related to the infection control paper produced by the working party.

3.5. PICO

**Patients:** All patient groups were included. The guideline is careful not to make recommendations which may prejudice clinical care based on gender, age, ethnicity or socio-economic status.

**Interventions:** interventions were identified in the literature to generate intervention specific recommendations

**Comparisons:** comparisons between intervention and standard management were used;

**Outcomes** were objective referring to length of hospital stay, mortality, rate of acquisition or infection.

3.6. Systematic Review Questions: Infection Control

1. What is the definition of Multidrug Resistant Gram-negative bacilli?
2. What Gram-negative bacilli cause infection control problems?
3. What are the relative contributions of community and hospital acquisition?
4. What is the evidence for reservoir and spread of multidrug-resistant Gram-negatives in Care Homes and secondary care?

5. What is the role of agricultural use of sewage and antibiotic treatment in veterinary practice in spreading ESBL?

6. What insights has national *E. coli* bacteraemia surveillance provided?

7. What is the role for screening in patients and staff?

8. What organisms should screening include?

9. Who, how and when to screen patients for Multidrug Resistant Gram-negative bacilli?

10. What can be done concerning patients unable to consent to a rectal swab?

11. How frequently does screening need to be performed?

12. Is there evidence for effective interventions on positive patients i.e. can carriage be cleared?

13. Selective decontamination: Why is it not used? Is there a role?

14. When should the environment be sampled?

15. What is the evidence that respiratory equipment contributes to transmission?

16. What national surveillance is performed and how should it be developed?

17. What is the evidence that sensor taps contribute to transmission?

18. Is there any cleaning method more effective than others at removing the Multidrug Resistant Gram-negative bacilli from the environment?

19. What is the evidence that infection control precautions prevent transmission?

20. Are standard infection control measures sufficient to stop transmission?

21. What are the minimum standards to stop spread in public areas, primary care or care homes?

22. Is there evidence for high/low risk areas within a healthcare facility?

23. Are there any organisational structures within a healthcare facility that play a role in the successful control of multi-resistant Gram-negative bacilli?

24. How should we undertake local screening, why is it important and how should it be interpreted?

25. At what point should passive surveillance switch to active surveillance i.e. screening?

26. What is the role of isolation in the care home/hospital settings?
Is there evidence of differences between organisms in respect of transmission, morbidity and mortality:

3.7. Antimicrobial Chemotherapy - Systematic Review Questions

1. What is the clinical importance of carbapenemases versus AmpC and CTX-M strains?
2. What impact have returning travellers made on UK epidemiology?
3. What is the global epidemiology of MDR-GNR?
4. How do Multidrug Resistant Enterobacteriaceae differ from the non-fermenters in terms of their prevalence and associated resistance genes?
5. What is the efficacy of carbapenems, mecillinam, temocillin, fosfomycin and colistin against specific pathogens?
6. What are the recommended antibiotics for community/secondary/tertiary care?
7. What is the threshold level of resistance for changing choice of empirical treatment for urinary infection?

Appendix 4 Systematic Review

4.1. Databases and Search terms Used 23/5/14

4.1.1. Databases

The Cochrane Library; MEDLINE; EMBASE; CINAHL

MeSH Terms See 4.2.

Free text terms. See 4.2.

Search Date: Medline 1946-2014; Embase 1980-2012; CINAHL (1984-2012)

Search Results (Figure 1)

Total number of articles located after duplicates removed = 2523
Sift 1 Criteria
Abstract screening: Systematic review, primary research, infection relates to MDR Gram-negative infection, informs one or more review question

Articles Retrieved
Total number of studies selected = 597

Sift 2 Criteria

Full text confirms that the article is primary research (randomised controlled trial, non-randomised controlled trials, controlled before and after studies, interrupted time series, case control study, case series, prospective cohort, systematic review; informs one or more of the review questions.

Articles selected for appraisal (10 full text publications could not be retrieved)
Total number of studies selected = 49

Critical appraisal

Articles presenting primary research or a systematic review and meeting the sift criteria were critically appraised by two reviewers using SIGN and EPOC criteria. Consensus was achieved through discussion

Accepted and Rejected Evidence

No meta analyses were available

Accepted after critical appraisal 49
Rejected after critical appraisal 0
4.2. Search

4.2.1. CINAHL (January 1984-December 2012)

<table>
<thead>
<tr>
<th>#</th>
<th>Query</th>
<th>Results</th>
</tr>
</thead>
<tbody>
<tr>
<td>S83</td>
<td>S48 AND S82</td>
<td>275</td>
</tr>
<tr>
<td>S82</td>
<td>S55 OR S56 OR S81</td>
<td>515,966</td>
</tr>
<tr>
<td>S81</td>
<td>S57 or S58 or S59 or S60 or S61 or S62 or S63 or S64 or S65 or S66 or S67 or S68 or S69 or S70 or S71 or S72 or S73 or S74 or S75 or S76 or S77 or S78 or S79 or S80</td>
<td>471,263</td>
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<tr>
<td>S80</td>
<td>TI ( (time points n3 over) or (time points n3 multiple) or (time points n3 three) or (time points n3 four) or (time points n3 five) or (time points n3 six) or (time points n3 seven) or (time points n3 eight) or (time points n3 nine) or (time points n3 ten) or (time points n3 eleven) or (time points n3 twelve) or (time points n3 month*) or (time points n3 hour*) or (time points n3 day*) or (time points n3 ‘more than’) ) or AB ( (time points n3 over) or (time points n3 multiple) or (time points n3 three) or (time points n3 four) or (time points n3 five) or (time points n3 six) or (time points n3 seven) or (time points n3 eight) or (time points n3 nine) or (time points n3 ten) or (time points n3 eleven) or (time points n3 twelve) or (time points n3 month*) or (time points n3 hour*) or (time points n3 day*) or (time points n3 ‘more than’) )</td>
<td>1,527</td>
</tr>
<tr>
<td>S78</td>
<td>TI ( multicentre or multicenter or multi-centre or multi-center ) or AB random*</td>
<td>101,899</td>
</tr>
<tr>
<td>S77</td>
<td>TI random* OR controlled</td>
<td>94,669</td>
</tr>
<tr>
<td>S76</td>
<td>TI ( trial or (study n3 aim) or ‘our study’ ) or AB ( (study n3 aim) or ‘our study’ )</td>
<td>87,121</td>
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<tr>
<td>S75</td>
<td>TI ( pre-workshop or preworkshop or post-workshop or postworkshop or (before n3 workshop) ) or AB ( pre-workshop or preworkshop or post-workshop or postworkshop or (before n3 workshop) )</td>
<td>283</td>
</tr>
<tr>
<td>S74</td>
<td>TI ( demonstration project OR demonstration projects OR preimplement* or pre-implement* or post-implement* or postimplement* ) or AB ( demonstration project OR demonstration projects OR preimplement* or pre-implement* or post-implement* or postimplement* )</td>
<td>1,290</td>
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<tr>
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<td>Query</td>
<td>Results</td>
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</tr>
<tr>
<td>S73</td>
<td>(intervention n6 clinician*) or (intervention n6 community) or (intervention n6 complex) or (intervention n6 design*) or (intervention n6 doctor*) or (intervention n6 educational) or (intervention n6 family doctor*) or (intervention n6 family physician*) or (intervention n6 family practitioner*) or (intervention n6 financial) or (intervention n6 GP) or (intervention n6 general practice*) Or (intervention n6 hospital*) or (intervention n6 impact*) Or (intervention n6 improv*) or (intervention n6 individualize*) Or (intervention n6 individualise*) or (intervention n6 individualizing) or (intervention n6 individualising) or (intervention n6 interdisciplin*) or (intervention n6 multicomponent) or (intervention n6 multi-component) or (intervention n6 multidisciplin*) or (intervention n6 multi-disciplin*) or (intervention n6 multifacet*) or (intervention n6 multi-facet*) or (intervention n6 multimodal*) or (intervention n6 multi-modal*) or (intervention n6 personalize*) or (intervention n6 personalisation) or (intervention n6 personalising) or (intervention n6 pharmacist*) or (intervention n6 pharmacy) or (intervention n6 physician*) or (intervention n6 practitioner*) Or (intervention n6 prescrib*) or (intervention n6 prescription*) or (intervention n6 provider*) or (intervention* n6 regulatory) or (intervention n6 tailor*) or (intervention n6 target*) or (intervention n6 team*) or (intervention n6 usual care)</td>
<td>23,198</td>
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<tr>
<td>S72</td>
<td>TI ( collaborativ* or collaboration* or tailored or personalised or personalized ) or AB ( collaborativ* or collaboration* or tailored or personalised or personalized )</td>
<td>38,021</td>
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<tr>
<td>S71</td>
<td>TI pilot</td>
<td>13,958</td>
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<tr>
<td>S70</td>
<td>(MH ‘Pilot Studies’)</td>
<td>36,433</td>
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<tr>
<td>S69</td>
<td>AB ‘before-and-after’</td>
<td>17,437</td>
</tr>
<tr>
<td>S68</td>
<td>AB time series</td>
<td>1,670</td>
</tr>
<tr>
<td>S67</td>
<td>TI time series</td>
<td>359</td>
</tr>
<tr>
<td>S66</td>
<td>AB ( before* n10 during or before n10 after ) or AU ( before* n10 during or before n10 after )</td>
<td>32,982</td>
</tr>
<tr>
<td>S65</td>
<td>TI ( (time point*) or (period* n4 interrupted) or (period* n4 multiple) or (period* n4 time) or (period* n4 various) or (period* n4 varying) or (period* n4 week*) or (period* n4 month*) or (period* n4 year*) ) or AB ( (time point*) or (period* n4 interrupted) or (period* n4 multiple) or (period* n4 time) or (period* n4 various) or (period* n4 varying) or (period* n4 week*) or (period* n4 month*) or (period* n4 year*) )</td>
<td>51,050</td>
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<tr>
<td>#</td>
<td>Query</td>
<td>Results</td>
</tr>
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<td>----</td>
<td>---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------</td>
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</tr>
<tr>
<td>S64</td>
<td><strong>TI</strong> ( ( quasi-experiment* or quasixperiment* or quasi-random* or quasirandom* or quasi control* or quasicontrol* or quasi* W3 method* or quasi* W3 study or quasi* W3 studies or quasi* W3 trial or quasi* W3 design* or experimental W3 method* or experimental W3 study or experimental W3 studies or experimental W3 trial or experimental W3 design* ) ) or <strong>AB</strong> ( ( quasi-experiment* or quasixperiment* or quasi-random* or quasirandom* or quasi control* or quasicontrol* or quasi* W3 method* or quasi* W3 study or quasi* W3 studies or quasi* W3 trial or quasi* W3 design* or experimental W3 method* or experimental W3 study or experimental W3 studies or experimental W3 trial or experimental W3 design* ) )</td>
<td>12,758</td>
</tr>
<tr>
<td>S63</td>
<td><strong>TI</strong> pre w7 post or <strong>AB</strong> pre w7 post</td>
<td>9,367</td>
</tr>
<tr>
<td>S62</td>
<td><strong>MH</strong> 'Multiple Time Series' or <strong>MH</strong> 'Time Series'</td>
<td>1,312</td>
</tr>
<tr>
<td>S61</td>
<td><strong>TI</strong> ( (comparative N2 study) or (comparative N2 studies) or evaluation study or evaluation studies ) or <strong>AB</strong> ( (comparative N2 study) or (comparative N2 studies) or evaluation study or evaluation studies )</td>
<td>11,680</td>
</tr>
<tr>
<td>S60</td>
<td><strong>MH</strong> Experimental Studies or Community Trials or Community Trials or Pretest-Posttest Design + or Quasi-Experimental Studies + Pilot Studies or Policy Studies + Multicenter Studies</td>
<td>34,567</td>
</tr>
<tr>
<td>S59</td>
<td><strong>TI</strong> ( pre-test* or pretest* or posttest* or post-test* ) or <strong>AB</strong> ( pre-test* or pretest* or posttest* or post-test* ) OR <strong>TI</strong> ( preimplement* or pre-implement* ) or <strong>AB</strong> ( pre-implement* or preimplement* )</td>
<td>6,868</td>
</tr>
<tr>
<td>S58</td>
<td><strong>TI</strong> ( intervention* or multiintervention* or multi-intervention* or postintervention* or post-intervention* or preintervention* or pre-intervention* ) or <strong>AB</strong> ( intervention* or multiintervention* or multi-intervention* or postintervention* or post-intervention* or preintervention* or pre-intervention* )</td>
<td>151,748</td>
</tr>
<tr>
<td>S57</td>
<td><strong>(MH</strong> 'Quasi-Experimental Studies')</td>
<td>5,747</td>
</tr>
<tr>
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<td>Query</td>
<td>Results</td>
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<tr>
<td>S56</td>
<td>(TI (systematic* n3 review*)) or (AB (systematic* n3 review*)) or (TI (systematic* n3 bibliographic*)) or (AB (systematic* n3 bibliographic*)) or (TI (systematic* n3 literature)) or (AB (systematic* n3 literature)) or (TI (systematic* n3 review*)) or (AB (systematic* n3 review*)) or (TI (comprehensive* n3 literature)) or (AB (comprehensive* n3 literature)) or (TI (comprehensive* n3 bibliographic*)) or (AB (comprehensive* n3 bibliographic*)) or (JN 'Cochrane Database of Systematic Reviews') or (TI (information n2 synthesis)) or (TI (data n2 synthesis)) or (AB (information n2 synthesis)) or (AB (data n2 synthesis)) or (TI (data n2 extract*)) or (AB (data n2 extract*)) or (TI (medline or pubmed or psyclit or cinahl or (psycinfo not 'psycinfo database') or 'web of science' or scopus or embase)) or (AB (medline or pubmed or psyclit or cinahl or (psycinfo not 'psycinfo database') or 'web of science' or scopus or embase)) or (MH 'Systematic Review') or (MH 'Meta Analysis') or (TI (meta-analy* or metaanaly*)) or (AB (meta-analy* or metaanaly*))</td>
<td>59,817</td>
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<tr>
<td>S55</td>
<td>S49 OR S50 OR S51 OR S52 OR S53 OR S54</td>
<td>158,596</td>
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<tr>
<td>S54</td>
<td>TI (‘control* N1 clinical’ or ‘control* N1 group*’ or ‘control* N1 trial*’ or ‘control* N1 study’ or ‘control* N1 studies’ or ‘control* N1 design*’ or ‘control* N1 method*’ ) or AB (‘control* N1 clinical’ or ‘control* N1 group*’ or ‘control* N1 trial*’ or ‘control* N1 study’ or ‘control* N1 studies’ or ‘control* N1 design*’ or ‘control* N1 method*’ )</td>
<td>1</td>
</tr>
<tr>
<td>S53</td>
<td>TI controlled or AB controlled</td>
<td>68,638</td>
</tr>
<tr>
<td>S52</td>
<td>TI random* or AB random*</td>
<td>117,418</td>
</tr>
<tr>
<td>S51</td>
<td>TI (‘clinical study’ or ‘clinical studies’ ) or AB (‘clinical study’ or ‘clinical studies’ )</td>
<td>7,969</td>
</tr>
<tr>
<td>S50</td>
<td>(MM ‘Clinical Trials+’)</td>
<td>10,670</td>
</tr>
<tr>
<td>S49</td>
<td>TI ( (multicent* n2 design*) or (multicent* n2 study) or (multicent* n2 studies) or (multicent* n2 trial*) ) or AB ( (multicent* n2 design*) or (multicent* n2 study) or (multicent* n2 studies) or (multicent* n2 trial*) )</td>
<td>8,917</td>
</tr>
<tr>
<td>S48</td>
<td>S18 AND S21 AND S47</td>
<td>917</td>
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<tr>
<td>S47</td>
<td>S22 OR S23 OR S24 OR S25 OR S26 OR S27 OR S28 OR S29 OR S30 OR S31 OR S32 OR S33 OR S34 OR S35 OR S36 OR S37 OR S38 OR S39 OR S40 OR S41 OR S42 OR S43 OR S44 OR S45 OR S46</td>
<td>16,726</td>
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<td>Results</td>
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</tr>
<tr>
<td>S46</td>
<td>TI ( (belcomycin or colicort or colimycin* or colisitin or colisticin or Colistin or colomycin or (coly n1 mycin) or colymicin or colymycin or coly-mycin or multimycin or (Polymyxin n1 E) or totazina) ) OR AB ( (belcomycin or colicort or colimycin* or colisitin or colisticin or Colistin or colistine or colomycin or (coly n1 mycin) or colymicin or colymycin or coly-mycin or multimycin or (Polymyxin n1 E) or totazina) )</td>
<td>171</td>
</tr>
<tr>
<td>S45</td>
<td>(MH 'Colistin')</td>
<td>134</td>
</tr>
<tr>
<td>S44</td>
<td>TI ( ((amdinocillin n1 pivoxil) or (FL n1 ‘1039’) or FL1039 or fl1039 or FL-1039 or pivaminocillin or Pivmecillinam or Selexid or coactabs or (ro n1 ‘109071’) or (ro10 n1 ‘9071’) or ro109071) ) OR AB ( ((amdinocillin n1 pivoxil) or (FL n1 ‘1039’) or FL1039 or fl1039 or FL-1039 or pivaminocillin or Pivmecillinam or Selexid or coactabs or (ro n1 ‘109071’) or (ro10 n1 ‘9071’) or ro109071) )</td>
<td>13</td>
</tr>
<tr>
<td>S43</td>
<td>TI ( ((Cephalosporanic n1 Acid*) or Cephalosporin* or Cefamandole or Cefoperazone or Cefazolin or Cefonicid or Cefsulodin or Cephacetrile or Cefotaxime or Cephalothin or Cefapirin or Cephalexin or Cefaclor or Cefadroxil or Cephaloglycin or Cephradine or Cephaloridine or Ceftazidime or Cephamycins or Cefmetazole or Cefotetan or Cefoxitin) ) OR AB ( ((Cephalosporanic n1 Acid*) or Cephalosporin* or Cefamandole or Cefoperazone or Cefazolin or Cefonicid or Cefsulodin or Cephacetrile or Cefotaxime or Cephalothin or Cefapirin or Cephalexin or Cefaclor or Cefadroxil or Cephaloglycin or Cephradine or Cephaloridine or Ceftazidime or Cephamycins or Cefmetazole or Cefotetan or Cefoxitin) )</td>
<td>1,569</td>
</tr>
<tr>
<td>S42</td>
<td>TI ( (Axepim* or bmy 28142 or bmy28142 or BMY-28142 or Cefepim* or cefepitax or ceficad or cepimax or forzyn beta or maxcef or maxfrom or maxipime or Quadrocef) ) OR AB ( (Axepim* or bmy 28142 or bmy28142 or BMY-28142 or Cefepim* or cefepitax or ceficad or cepimax or forzyn beta or maxcef or maxfrom or maxipime or Quadrocef) )</td>
<td>171</td>
</tr>
<tr>
<td>S41</td>
<td>(MH 'Cephalosporins+')</td>
<td>2,105</td>
</tr>
<tr>
<td>#</td>
<td>Query</td>
<td>Results</td>
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</tr>
<tr>
<td>S40</td>
<td>TI ( (berkfurin or biofurin or chemiofuran or dantafur or f 30 or f30 or fua-med or furaben or furadantin* or furadantoin or furadina or furadoine or furadonin or furadonine or furalan or furanpur or furantocompren or furantoin* or furobactina or furofen or furophen or infurin or ituran or ivadantin or macrobid or macrodantin* or macrofurin or macrofuradantin* or macrofuradantoin or macrofurantin* or nitrofuracin or nitrofuradantoin or nitrofurantine or nitrofurantoin* or nitrofurin or novofuran or nsc 2107 or nsc2107 or orafuran or parfuran or phenurin or (potassium n1 furagin) or ralodantin or trocurine or urantin or (uro n1 tablinen) or urodil or urodin or urofuran or urolong or urotabliden or uro-tabliden or urotoina or uvamin ) ) OR AB ( berkfurin or biofurin or chemiofuran or dantafur or f 30 or f30 or fua-med or furaben or furadantin* or furadantoin or furadina or furadoine or furadonin or furadonine or furalan or furanpur or furantocompren or furantoin* or furobactina or furofen or furophen or infurin or ituran or ivadantin or macrobid or macrodantin* or macrofurin or macrofuradantin* or macrofuradantoin or macrofurantin* or nitrofuracin or nitrofuradantoin or nitrofurantine or nitrofurantoin* or nitrofurin or novofuran or nsc 2107 or nsc2107 or orafuran or parfuran or phenurin or (potassium n1 furagin) or ralodantin or trocurine or urantin or (uro n1 tablinen) or urodil or urodin or urofuran or urolong or urotabliden or uro-tabliden or urotoina or uvamin ) )</td>
<td>325</td>
</tr>
<tr>
<td>S39</td>
<td>TI ( ((az n1 threonam) or azactam or azenan or azthreonam or aztreonam or (corus n1 ‘1020’) or dynabiotic or primbactam or SQ 26,776 or sq 26,776 or sq 26776 or SQ-26,776 or sq26776 or sq-26776 or urobactam ) ) OR AB ( ((az n1 threonam) or azactam or azenan or azthreonam or aztreonam or (corus n1 ‘1020’) or dynabiotic or primbactam or SQ 26,776 or sq 26,776 or sq 26776 or SQ-26,776 or sq26776 or sq-26776 or urobactam ) )</td>
<td>96</td>
</tr>
<tr>
<td>S38</td>
<td>(MH ‘Aztreonam’)</td>
<td>54</td>
</tr>
<tr>
<td>S37</td>
<td>TI ( (fosfocil or fosfocin or fosfocina or fosfomicin or fosfomycin or fosfonomycin or ‘mk 0955’ or mk 955 or mk0955 or mk955 or monuril or phosphomycin or phosphonomycin) ) OR AB ( (fosfocil or fosfocin or fosfocina or fosfomicin or fosfomycin or fosfonomycin or ‘mk 0955’ or mk 955 or mk0955 or mk955 or monuril or phosphomycin or phosphonomycin) )</td>
<td>57</td>
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<td>Query</td>
<td>Results</td>
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</tr>
<tr>
<td>S36</td>
<td>TI ( (akacin or akicin or amicacina or amicasil or amicin or amiglymide v or amikacin* or amikafur or amikalem or amikan or amikayect or amikin or amiklin or amikozit or amiktam or amitracin or amixin or amukin or apalin or bk 8 or bb k8 or bbk 8 or bbk 8 or bbk8 or bbk-8 or bb-k8 or bbk8 or bbk-8 or bbk 8 or biclin or biklin or biokacin or briclin or briiklin or chemacin or cinmik or fabianol or gamikal or glukamin or kacinth-a or kanbine or kormakin or likacin or lukadin or miacin or mikasome or onikin or oprad or orlobin or pediakin or pierami or riklinak or savox or selaxa or selemycin or sulfate amikacin or tybikin or vs 107 or vs107 or yectamid) ) OR AB ( (akacin or akicin or amicacina or amicasil or amicin or amiglymide v or amikacin* or amikafur or amikalem or amikan or amikayect or amikin or amiklin or amikozit or amiktam or amitracin or amixin or amukin or apalin or bk 8 or bb k8 or bbk 8 or bbk 8 or bbk8 or bbk-8 or bb-k8 or bbk8 or bbk-8 or biclin or biklin or biokacin or briclin or briiklin or chemacin or cinmik or fabianol or gamikal or glukamin or kacinth-a or kanbine or kormakin or likacin or lukadin or miacin or mikasome or onikin or oprad or orlobin or pediakin or pierami or riklinak or savox or selaxa or selemycin or sulfate amikacin or tybikin or vs 107 or vs107 or yectamid) )</td>
<td>342</td>
</tr>
<tr>
<td>S35</td>
<td>(MH ‘Amikacin’)</td>
<td>140</td>
</tr>
<tr>
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<td>Query</td>
<td>Results</td>
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<tr>
<td>----</td>
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</tr>
<tr>
<td>S34</td>
<td>TI ( (adelanin or alcomicin or apigent or apogen or apoten or azupel or bactiderm or biogaracin or cidomycin or danigen or dermogen or dianfarma or dispensent or duragentam* or epigent or (frieso n1 gent) or garabiotic or garalone or garamicin* or garamycin or garbilocin or gencin or gendril or genoptic or genrex or gensumycin or gentabiotic or gentabiox or gentac or gentacidin or gentacin or gentacor or gentacycol or gentacyl or gentafair or gentagram or gentak or gental or gentalone or gentalline or gentalol or gentalyn or gentamax or gentame* or gentamicin* or gentamina or gentamycin* or gentamyl or gentamytrex or gentaplus or gentarad or gentasil or gentasol or gentasone or gentasporin or gentatrim or gentavet or genticin* or genticyn or gentiderm or gentimycin or gentocin or gentogram or gentomyacin or genum or geomyicine or gevramycin or g-mycin or gmyticin or g-mythicin or grammicin or hexamycin or jenamicin or konigen or lacromycin or lisagent or martigenta or migenta or miragenta or miramycin or nichogencin or nsc 82261 or nsc82261 or obogen or ocugenta or ocu-mycin or oftagen or ophtagram or optagam or optigen or opti-genta or ottoagenta or pyogenta or refobacin or ribomicin or rigaminol or rocy gen or rovixida or rupegen or sagestam or sch 9724 or sch9724 or sedanazin or servigenta or skinfect or sulmycin or tangyn or u-gencin or versigen or yectamicina) ) OR AB ( (adelanin or alcomicin or apigent or apogen or apoten or azupel or bactiderm or biogaracin or bristagen or cidomycin or danigen or dermogen or dianfarma or dispensent or duragentam* or epigent or (frieso n1 gent) or garabiotic or garalone or garamicin* or garamycin or garbilocin or gencin or gendril or genoptic or genrex or gensumycin or gentabiotic or gentabiox or gentac or gentacidin or gentacin or gentacor or gentacycol or gentacyl or gentafair or gentagram or gentak or gental or gentalone or gentalline or gentalol or gentalyn or gentamax or gentame* or gentamicin* or gentamina or gentamycin* or gentamyl or gentamytrex or gentaplus or gentarad or gentasil or gentasol or gentasone or gentasporin or gentatrim or gentavet or gentricin* or genticyn or gentiderm or gentimycin or gentocin or gentogram or gentomyacin or genum or geomyicine or gevramycin or g-mycin or gmyticin or g-mythicin or grammicin or hexamycin or jenamicin or konigen or lacromycin or lisagent or martigenta or migenta or miragenta or miramycin or nichogencin or nsc 82261 or nsc82261 or obogen or ocugenta or ocu-mycin or oftagen or ophtagram or optagam or optigen or opti-genta or ottoagenta or pyogenta or refobacin or ribomicin or rigaminol or rocy gen or rovixida or rupegen or sagestam or sch 9724 or sch9724 or sedanazin or servigenta or skinfect or sulmycin or tangyn or u-gencin or versigen or yectamicina) )</td>
<td>993</td>
</tr>
<tr>
<td>S33</td>
<td>(MH ‘Gentamicins’)</td>
<td>808</td>
</tr>
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<td>#</td>
<td>Query</td>
<td>Results</td>
</tr>
<tr>
<td>---</td>
<td>-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------</td>
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</tr>
<tr>
<td>S32</td>
<td>TI ( (Aminoglycosides or Anthracyclines or Aclarubicin or Daunorubicin or Plicamycin or Butirosin Sulfate or Sisomicin or Hygromycin B or Kanamycin or Dibekacin or Nebramycin or Metrizamide or Neomycin or Framycetin or Paromomycin or Ribostamycin or Puromycin or Spectinomycin or Streptomycin or Dihydrostreptomycin Sulfate or Streptothricins or Streptozocin) ) OR AB ( (Aminoglycosides or Anthracyclines or Aclarubicin or Daunorubicin or Plicamycin or Butirosin Sulfate or Sisomicin or Hygromycin B or Kanamycin or Dibekacin or Nebramycin or Metrizamide or Neomycin or Framycetin or Paromomycin or Ribostamycin or Puromycin or Spectinomycin or Streptomycin or Dihydrostreptomycin Sulfate or Streptothricins or Streptozocin) )</td>
<td>1,269</td>
</tr>
<tr>
<td>S31</td>
<td>(MH 'Aminoglycosides+')</td>
<td>6,215</td>
</tr>
<tr>
<td>S30</td>
<td>TI ( ((chinolone n1 derivative) or fluoroquinolones or (haloquinolone n1 derivative) or ketoquinolines or oxoquinolines or quinolinones or quinolones ) ) OR AB ( ((chinolone n1 derivative) or fluoroquinolones or (haloquinolone n1 derivative) or ketoquinolines or oxoquinolines or quinolinones or quinolones ) )</td>
<td>834</td>
</tr>
<tr>
<td>S29</td>
<td>(MH 'Quinolines+') OR (MH 'Antiinfective Agents, Quinolone+')</td>
<td>4,842</td>
</tr>
<tr>
<td>S28</td>
<td>TI ( (tigecycline or (tbg n1 mino) or tygacil or gar 936 or gar936 or (tert n1 butylglycinamido*)) ) OR AB ( (tigecycline or (tbg n1 mino) or tygacil or gar 936 or gar936 or (tert n1 butylglycinamido*)) )</td>
<td>208</td>
</tr>
<tr>
<td>S27</td>
<td>TI ( ((brl n1 ‘17421’) or brl17421 or (thiophenemalonamic n1 acid) or negaban or temocillin or temopen ) ) OR AB ( ((brl n1 ‘17421’) or brl17421 or (thiophenemalonamic n1 acid) or negaban or temocillin or temopen ) )</td>
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<tr>
<td>#</td>
<td>Query</td>
<td>Results</td>
</tr>
<tr>
<td>-----</td>
<td>----------------------------------------------------------------------</td>
<td>---------</td>
</tr>
<tr>
<td>S26</td>
<td>TI ((aclam or aktil or ambilan or amoca or amoclan or amoclav or</td>
<td>805</td>
</tr>
<tr>
<td></td>
<td>amoksiklav or amolanic or amometin or (amox n1 clav) or amox-clav or</td>
<td></td>
</tr>
<tr>
<td></td>
<td>(amoxi n1 plus) or (amoxNear/3clavulan*) or amoxiclav or amoxiclav-</td>
<td></td>
</tr>
<tr>
<td></td>
<td>bid or amoxiclav-teva or amoksiklav or amoxslox or (amoxicillin-</td>
<td></td>
</tr>
<tr>
<td></td>
<td>clavulanic n1 acid) or ancla or (auclatin n1 duo) or augamox or</td>
<td></td>
</tr>
<tr>
<td></td>
<td>augmaxcil or augmentan or augmentin* or augmex or augpen or</td>
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<tr>
<td></td>
<td>(augucillin n1 duo) or augurcin or ausclav or auspicil or bactiv or</td>
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<tr>
<td></td>
<td>bactoclav or bioclavid or (brl n1 '25000') or brl25000 or brl-25000</td>
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<td></td>
<td>or cavumox or ciblor or (clacillin n1 duo) or clamax or clamentin or</td>
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<td></td>
<td>clamobit or clamonex or clamovid or clamoxin or (clamoxyl n1 duo*)</td>
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<tr>
<td></td>
<td>or clarin-duo or clavamox or clavar or clavifar or clavoxil or</td>
<td></td>
</tr>
<tr>
<td></td>
<td>(clavoxilin n1 plus) or clavubactin or clavudale or clavulanate-</td>
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<tr>
<td></td>
<td>amoxicillin or clavulin or (clavulox n1 duo) or clavumox or (co n1</td>
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<td>amoxiclav) or (co n1 amoxyclav) or coamoxiclav or co-amoxiclav or</td>
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<td></td>
<td>coamoxiclav or (cramon n1 duo) or (croanan n1 duo) or curam or</td>
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<td>danoclav or (darzitil n1 plus) or e-moxclav or enhancin or fleming</td>
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<td>or fugentin or (fullicilina n1 plus) or gumentin or hibiotic or</td>
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<td>incilav or klamonex or kmoxilin or lactamox or lansiclav or moxiclav</td>
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<td>or moxicle or moxyclav or nufaclav or palentin or quali-mentin or</td>
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<td>ranclav or spektramox or statillin or suplentin or synermox or</td>
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<td>synulox or (velamox n1 cl) or vestaclav or viaclov or vulamox or</td>
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<td></td>
<td>xiclav or (zami n1 '8503')) ) OR AB ((aclam or aktil or ambilan or</td>
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<td></td>
<td>amoca or amoclan or amoclav or amoxiclav or amoksiklav or amolanic</td>
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<td></td>
<td>or cavumox or ciblor or (clacillin n1 duo) or clamax or clamentin or</td>
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<td>clamobit or clamonex or clamovid or clamoxin or (clamoxyl n1 duo*)</td>
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<td>or clarin-duo or clavamox or clavar or clavifar or clavoxil or</td>
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<td>amoxicillin or clavulin or (clavulox n1 duo) or clavumox or (co n1</td>
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<td>amoxiclav) or (co n1 amoxyclav) or coamoxiclav or co-amoxiclav or</td>
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<td>coamoxiclav or (cramon n1 duo) or (croanan n1 duo) or curam or</td>
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<td>or moxicle or moxyclav or nufaclav or palentin or quali-mentin or</td>
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<td>xiclav or (zami n1 '8503')) )</td>
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<td>tazocel or tazocillin* or tazocin or tazomax or tazonam or tazopril or</td>
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<td>yp 14 or yp14 or ytr 830 or ytr 830h or ytr830 or ytr830h or zosyn) ) OR</td>
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<td></td>
<td>AB ( (cl 307579 or cl298741 or cl307579 or tazabactam or tazobac* or</td>
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<td></td>
<td>tazocel or tazocillin* or tazocin or tazomax or tazonam or tazopril or</td>
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<td></td>
<td>yp 14 or yp14 or ytr 830 or ytr 830h or ytr830 or ytr830h or zosyn) )</td>
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<td>or cypercil or hishiyaclorin or ivacin or pentcillin or pentocillin or</td>
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<td>picillin* or pipcil or pipera hameln or pipercil or pipercillin* or</td>
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<td>piperasin or pipera-hameln or pipercillin or piperilline or pipriaci*</td>
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<td>or pipraks or pipril or piprilin or pitamycin or t 1220 or t1220 or</td>
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<td>t-1220 or t-1220 or taiperacillin) ) OR AB ( (acopex or avocin or cl</td>
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<td>227,193 or Cl 227193 or cl 227193 or Cl227193 or cl227193 or Cl-227193</td>
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<td>t-1220 or t-1220 or taiperacillin) )</td>
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<td>Ronem or S 4661 or S-4661 or SM 7338 or SM-7338 or Thienamycin*) ) OR AB</td>
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</tr>
<tr>
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<td>( (Carbapenem* or doripenem or ertapenem or Imipemide or Imipenem or</td>
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<tr>
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<td>Invancor Invancor or meropenem or Merrem or ‘MK 0787’ or MK0787 or MK-</td>
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<td>0787 or N Formimidoylthienamycin or N-Formimidoylthienamycin or Penem or</td>
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</tr>
<tr>
<td></td>
<td>Ronem or S 4661 or S-4661 or SM 7338 or SM-7338 or Thienamycin*) )</td>
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<td>(MH ‘Carbapenems+’)</td>
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<td>S19 OR S20</td>
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<td>(MH ‘Drug Resistance, Microbial+’)</td>
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<td>TI ((multiresistant or (multi n1 resistant*)) ) OR AB ((multiresistant</td>
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<td>or (multi n1 resistant*)) )</td>
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<td>OR S12 OR S13 OR S14 OR S15 OR S16 OR S17</td>
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<td>Query</td>
<td>Results</td>
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<td>S17</td>
<td>TI ( ((bacillus n1 morgen*) or (bacterium n1 morgana) or (morganella n1 morgagni*) or (morganella n1 morganii) or (proteus n1 morgagni) or (proteus n1 morgana*) or (salmonella n1 morgana)) ) OR AB ( ((bacillus n1 morgan*) or (bacterium n1 morgana) or (morganella n1 morgagni*) or (morganella n1 morganii) or (proteus n1 morgagni) or (proteus n1 morgana*) or (salmonella n1 morgana)))</td>
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<td>S16</td>
<td>TI ( ((Citrobacter n1 freundii) or (bacterium n1 freundii) or (Escherichia n1 freundii)) ) OR AB ( ((Citrobacter n1 freundii) or (bacterium n1 freundii) or (Escherichia n1 freundii)))</td>
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<td>S15</td>
<td>(MH ‘Citrobacter’)</td>
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<td>S14</td>
<td>TI Serratia OR AB Serratia</td>
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<td>S13</td>
<td>(MH ‘Serratia’) OR (MH ‘Serratia Infections’)</td>
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<td>S12</td>
<td>TI Proteus OR AB Proteus</td>
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<td>S11</td>
<td>(MH ‘Proteus’) OR (MH ‘Proteus Infections’)</td>
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<td>S10</td>
<td>TI ( (Acinetobacter or mima or mimae or herellea or acinetobacterium) ) OR AB ( (Acinetobacter or mima or mimae or herellea or acinetobacterium) )</td>
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<td>S9</td>
<td>(MH ‘Acinetobacter Infections’)</td>
<td>581</td>
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<td>S8</td>
<td>TI ‘p. aeruginosa’ OR AB ‘p. aeruginosa’</td>
<td>610</td>
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<td>S7</td>
<td>TI ( ((bacillus n1 pyocyanus) or (bacterium n1 (aeruginosum or pyocyaneum)) or (blue n1 apus) or (Pseudomonas n1 (aeruginosa or aureofaciens or pyoceaneus or pyocyanea or pyocyaneus))) ) OR AB ( ((bacillus n1 pyocyanus) or (bacterium n1 (aeruginosum or pyocyaneum)) or (blue n1 apus) or (Pseudomonas n1 (aeruginosa or aureofaciens or pyoceaneus or pyocyanea or pyocyaneus))) )</td>
<td>1,855</td>
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<td>S6</td>
<td>TI ( (enterobacter or aerobacter) ) OR AB ( (enterobacter or aerobacter) )</td>
<td>370</td>
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<td>S5</td>
<td>TI ( ('k. pneumoniae’ or ‘b. friedlander’) ) OR AB ( ('k. pneumoniae’ or ‘b. friedlander’) )</td>
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</table>
### Results

<table>
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<th>Query</th>
<th>Results</th>
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<td>S4</td>
<td>TI ( (klebsiella or Calymmatobacterium or (aerobacter n1 aerogenes) or ((bacillus or bacterium) n1 pneumonia) or ((friedlaender or Friedlander) n1 bacillus) or (Hyalococcus n1 pneumonia) or Pneumobacillus)) OR AB ((klebsiella or Calymmatobacterium or (aerobacter n1 aerogenes) or ((bacillus or bacterium) n1 pneumonia) or ((friedlaender or Friedlander) n1 bacillus) or (Hyalococcus n1 pneumonia) or Pneumobacillus))</td>
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<td>S3</td>
<td>(MH 'Klebsiella') OR (MH 'Klebsiella Infections')</td>
<td>835</td>
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<td>S2</td>
<td>TI ((Eaggec or (escherichia n1 coli) or (e n1 coli) or (alkalescens-dispar n1 group) or (bacillus n1 escherichii) or (Coli n1 bacillus) or (Coli n1 bacterium) or colibacillus or (colon n1 bacillus)) OR AB ((Eaggec or (escherichia n1 coli) or (e n1 coli) or (alkalescens-dispar n1 group) or (bacillus n1 escherichii) or (Coli n1 bacillus) or (Coli n1 bacterium) or colibacillus or (colon n1 bacillus))</td>
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<td>S1</td>
<td>(MH 'Escherichia Coli') OR (MH 'Escherichia Coli Infections')</td>
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#### 4.2.2. Cochrane Library (Issue 11, 2012)

**ID**  
**Search**

#1  MeSH descriptor: [Escherichia coli] explode all trees

#2  (Eaggec or (escherichia near/1 coli) or (e near/1 coli) or (alkalescens-dispar near/1 group) or (bacillus near/1 escherichii) or (Coli near/1 bacillus) or (Coli near/1 bacterium) or colibacillus or (colon near/1 bacillus)):ti,ab,kw (Word variations have been searched)

#3  MeSH descriptor: [Klebsiella] explode all trees

#4  (klebsiella or Calymmatobacterium or (aerobacter near/1 aerogenes) or ((bacillus or bacterium) near/1 pneumonia) or ((friedlaender or Friedlander) near/1 bacillus) or (Hyalococcus near/1 pneumonia) or Pneumobacillus):ti,ab,kw (Word variations have been searched)

#5  k. pneumoniae or b. friedlander:ti,ab,kw (Word variations have been searched)

#6  MeSH descriptor: [Enterobacter] explode all trees

#7  (enterobacter or aerobacter):ti,ab,kw (Word variations have been searched)

#8  MeSH descriptor: [Pseudomonas aeruginosa] explode all trees

#9  ((bacillus near/1 pyocyaneus) or (bacterium near/1 (aeruginosum or pyocyaneum)) or (blue near/1 apus) or (Pseudomonas near/1 (aeruginosa or aureofaciens or pyoceaneus or pyocyanea or pyocyaneus)):ti,ab,kw (Word variations have been searched)

#10  p. aeruginosa:ti,ab,kw (Word variations have been searched)

#11  MeSH descriptor: [Acinetobacter] explode all trees

Accepted manuscript 291
(Acinetobacter or mima or mimae or herellea or acinetobacterium):ti,ab,kw (Word variations have been searched)

MeSH descriptor: [Proteus] explode all trees

Proteus:ti,ab,kw (Word variations have been searched)

MeSH descriptor: [Serratia] explode all trees

Serratia:ti,ab,kw (Word variations have been searched)

MeSH descriptor: [Citrobacter freundii] explode all trees

(Citrobacter near/1 freundii) or (bacterium near/1 freundii) or (Escherichia near/1 freundii)):ti,ab,kw (Word variations have been searched)

MeSH descriptor: [Morganella morganii] explode all trees

((bacillus near/1 morgan$) or (bacterium near/1 morgana) or (morganella near/1 morgagni$) or (morganella near/1 morganii) or (proteus near/1 morgagni) or (proteus near/1 morgana$) or (salmonella near/1 morgana)):ti,ab,kw (Word variations have been searched)

(meSH descriptor: [Drug Resistance, Multiple] explode all trees

Multi resistant or (multi near/1 resistan$)):ti,ab,kw (Word variations have been searched)

MeSH descriptor: [Colistin] explode all trees

(belcomycin or colicort or colimycin$: or colisitin or colisticin or Colistin or colistine or colomycin or (coli near/1 mycin) or colymcin or colymycin or coly-mycin or multimycin or (Polymyxin near/1 E) or totazina):ti,ab,kw (Word variations have been searched)

MeSH descriptor: [Carbapenems] explode all trees

(Carbapenem$: or doripenem or ertapenem or Imipemide or Imipenem or Invanoz or Invanz or meropenem or Merrem or 'MK 0787' or MK0787 or MK-0787 or N Formimidoylthienamycin or N-Formimidoylthienamycin or Penem or Ronem or S 4661 or S-4661 or SM 7338 or SM-7338 or Thienamycin$):ti,ab,kw (Word variations have been searched)

MeSH descriptor: [Piperacillin] explode all trees

(acopex or avocin or cl 227,193 or Cl 227193 or cl 227193 or cl227,193 or Cl227193 or cl227193 or Cl-227193 or Cl-227193 or cypercil or hishiyadorin or ivacin or pentcillin or pentocillin or pentocillin$ or pipcil or pipera hameln or piperacll or piperaclina$ or piperacin or pipera-hameln or pipercill or pipiiline or pipraci$ or pipraks or pipril or piprilin or pitamycin or t 1220 or t1220 or t-1220 or taiperacillin):ti,ab,kw (Word variations have been searched)

MeSH descriptor: [Tazobactam] explode all trees

ci 307579 or cl298741 or cl307579 or tazabactam or tazobac$: or tazocel or tazocillin$: or tazocin or tazomax or tazonam or tazopril or yp 14 or yp14 or ytr 830 or ytr 830h or ytr830 or ytr830h or zosyn):ti,ab,kw (Word variations have been searched)
MeSH descriptor: [Amoxicillin-Potassium Clavulanate Combination] explode all trees

(aclam or akti1 or ambilan or amocla or amoclan or amoclav or amoksiklav or amolanic or amometin or (amox near/1 clav) or amox-clav or (amoxi near/1 plus) or (amoxNear/3clavulan$) or amoxiclav or amoxiclav-bid or amoxiclav-teva or amoxiklav or amoxulin or (amoxycillin-clavulanic near/1 acid) or ancla or (auclatin near/1 duo) or augamox or augmaxcil or augmentan or augmentin$ or augmex or augpen or (augucillin near/1 duo) or auguric or ausclav or auspilic or bactiv or bactoclav or bioclavid or (brl near/1 '25000') or brl25000 or brl-25000 or cavumox or ciblor or (clacillin near/1 duo) or clamax or clamentin or clamobit or clamonex or clamovid or clamoxin or (clamoxyl near/1 duo$) or clarin-duo or clavamox or clav or clavina or clavolin or (clavoxin near/1 plus) or clavubactin or clavudale or clavulanate-amoxicillin or clavulin or (clavulox near/1 duo) or clavumox or (co near/1 amoxiclav) or (co near/1 amoxyclav) or coamoxiclav or co-amoxiclav or coamoxyclav or (cronon near/1 duo) or (croanan near/1 duo) or curam or danoclav or (darzitil near/1 plus) or e-moxclav or enhancin or fleming or fANGUAGE or (fullicilina near/1 plus) or gumentin or hibiotic or inciclav or klamonex or kloxin or lactamox or lansiclav or moxiclav or moxyclyclor natravox or nufaclav or palentin or quali-mentin or ranclav or spektramox or stacillin or suplentin or synelodex or synulox or (velamox near/1 cl) or vestaclav or viadclav or vulamox or xicl or (zami near/1 '8503')":ti,ab,kw (Word variations have been searched)

(brl near/1 '17421') or brl17421 or (thiophenemalonamic near/1 acid) or negaban or temocillin or temopen":ti,ab,kw (Word variations have been searched)

tigecycline or (tbg near/1 mino) or tygacil or gar 936 or gar936 or (tert near/1 butylglycinamido$)":ti,ab,kw (Word variations have been searched)

MeSH descriptor: [Quinolones] explode all trees

((chinolone near/1 derivative) or fluoroquinolones or (haloquinolone near/1 derivative) or ketoquinolines or oxoquinolines or quinolinones or quinolones):ti,ab,kw (Word variations have been searched)

MeSH descriptor: [Aminoglycosides] explode all trees

(Aminoglycosides or Anthracyclines or Aclarubicin or Daunorubicin or Plicamycin or Butirosin Sulfate or Sisomicin or Hygromycin B or Kanamycin or Dibekacin or Nebramycin or Metrizamide or Neomycin or Frumycin or Paromomycin or Ribostamycin or Puromycin or Spectinomycin or Streptomycin or Dihydrotreptomycin Sulfate or Streptothricins or Streptozocin):ti,ab,kw (Word variations have been searched)

MeSH descriptor: [Gentamicins] explode all trees

(adelanin or alcomicin or apigent or apogen or apoten or azupel or bactiderm or biogaracin or bristlina or cidomyin or danigen or dermogen or dianfarma or dispagent or duragentam$ or epignet or (friese near/1 gent) or garabiotic or garalone or garamicin$ or garitucou or garbiloci or gencin or gendril or genoptico or genrex or gensumycin or gentabiotic or gentabio or gentac or gentacidin or gentacin or gentacor or gentacycol or gentacyl or gentafair or gentagram or gentak or gental or gentaline or gentalline or gentalol or gentalyn or gentamax or gentame$ or gentamicin$ or gentamina or gentamycin$ or gentamyl or gentamytrex or gentaplus or gentarad or
gentasil or gentasol or gentasone or gentasporin or gentatrim or gentavet or genticin or genticyan or gentiderm or gentimycin or gentocin or gentogram or gentomycin or genum or georgycin or gevramycin or g-mycin or gmyticin or g-mytycin or grammicin or hexamycin or jenamicin or konigen or lacremycin or lisagent or martigenta or migenta or miragenta or miramycin or nichogencin or nsc 82261 or nsc82261 or obogen or ocugenta or ocut-mycin or oftogen or ophtagram or opthagen or optigen or opti-genta or ottogenta or pyogenta or refobacin or ribomicin or rigaminol or rocy gen or rovixida or rupegen or sagenstam or sch 9724 or sch9724 or sedanazin or servigenta or skinfect or sulmycin or tangyn or u-gencin or versigen or yectamicina):ti,ab,kw
(Word variations have been searched)

#42 MeSH descriptor: [Amikacin] explode all trees

#43 (akacin or akicin or amicacina or amicasil or amicin or amiglymid v or amikacin$ or amikafur or amikalem or amikan or amikayect or amikin or amiklin or amikozit or amikatam or amitracin or amixin or amukin or apalin or bb k 8 or bb k 8 or bbk 8 or bb-k 8 or bbk8 or bbk-8 or bb-k8 or biclin or biklin or biokcin or briclin or briklin or chemacin or cinmik or fabianol or gamikal or glukamin or kacinth-a or kanbine or kormakin or likacin or lukadin or miacin or mikasome or onikin or oprad or orlabin or pediakin or pieramik or riklinak or savox or selemycin or sulfate amikacin or tybikin or vs 107 or vs107 or yectamid):ti,ab,kw (Word variations have been searched)

#44 MeSH descriptor: [Fosfomycin] explode all trees

#45 (fosfocil or fosfocin or fosfocina or fosfomicin or fosfonomyein or 'mk 0955' or mk 955 or mk0955 or mk955 or monuril or phosphomycin or phosphonomyein):ti,ab,kw (Word variations have been searched)

#46 MeSH descriptor: [Aztreonam] explode all trees

#47 ((az near/1 threonam) or azactam or azenan or azthreonam or aztreonam or (corus near/1 '1020') or dynabiotic or primbactam or sq 26,776 or sq 26,776 or sq 26776 or sq-26,776 or sq26776 or sq-26776 or urobactam):ti,ab,kw (Word variations have been searched)

#48 MeSH descriptor: [Nitrofurantoin] explode all trees

#49 (berkurfin or biofurin or chemofuran or dantafur or f 30 or f30 or fua-med or furabren or furadantin$ or furadantoin or furadina or furadoine or furadonin or furadonine or furalan or furanpur or furantocompren or furantoin$: or furobactina or furofen or furophen or infurin or ituran or ivadantin or macrobid or macrodantin$ or macrofurin or macrofurin or micofurantin$: or nitrofuratoind or nephronex or nieruf or nifurantin or nifuryl or (nitro near/1 macro) or nitrofuracin or nitrofurandantoin or nitrofurantitoin or nitrofurin or novofuran or nsc 2107 or nsc2107 or orafuran or parfurin or phenurin or (potassium near/1 furagin) or ralodantin or trocurine or urantin or (uro near/1 tablinen) or urodi or urofin or urofuran or urolong or urotablunen or uro-tablunen or urotoïna or uva`:ti,ab,kw (Word variations have been searched)

#50 MeSH descriptor: [Cephalosporins] explode all trees

#51 ((Cephalosporanic near/1 Acid$) or Cephalosporin$ or Cefamandole or Cefoperazone or Cefazolin or Cefonicid or Cefsulodin or Cephalactril or Cefotaxime or Cephalexin or Cephapirin or Cephalexin or Cefaclor or Cefadroxil or Cephaloglycin or
Cephradine or Cephaloridine or Ceftazidime or Cephamycins or Cefmetazole or Cefotetan or Cefoxitin):ti,ab,kw (Word variations have been searched)

#52 MeSH descriptor: [Amdinocillin Pivoxil] explode all trees

#53 ((amdinocillin near/1 pivoxil) or (FL near/1 ‘1039’) or FL1039 or fl1039 or FL-1039 or pivamdinocillin or Pivmecillnam or Selexid or coactabs or (ro near/1 ‘109071’) or (ro10 near/1 ‘9071’) or ro109071):ti,ab,kw (Word variations have been searched)

#54 #25 or #26 or #27 or #28 or #29 or #30 or #31 or #32 or #33 or #34 or #35 or #36 or #37 or #38 or #39 or #40 or #41 or #42 or #43 or #44 or #45 or #46 or #47 or #48 or #49 or #50 or #51 or #52 or #53

#55 #21 and #24 and #54 (21)

4.2.3. Embase (January 1980 to December 1012)

1 exp Escherichia coli/ (255846)
2 (Eaggec or (escherichia adj coli) or (e adj coli) or (alkalescens-dispar adj group) or (bacillus adj escherichii) or (Coli adj bacillus) or (Coli adj bacterium) or colibacillus or (colon adj bacillus)).ti,ab. (240749)
3 exp Klebsiella/ (30199)
4 (klebsiella or Calymmatobacterium or (aerobacter adj aerogenes) or ((bacillus or bacterium) adj pneumonia) or ((friedlaender or Friedlander) adj bacillus) or (Hyalococcus adj pneumonia) or Pneumobacillus).ti,ab. (22836)
5 ('k. pneumoniae' or 'b. friedlander').ti,ab. (5513)
6 exp Enterobacter/ (12784)
7 (enterobacter or aerobacter).ti,ab. (9700)
8 exp Pseudomonas aerugiosa/ (55073)
9 ((bacillus adj pyocyaneus) or (bacterium adj (aeruginosum or pyocyaneum)) or (blue adj apus) or (Pseudomonas adj (aeruginosa or aureofaciens or pyoceaneus or pyocyanea or pyocyaneus))).ti,ab. (43474)
10 'p. aeruginosa'.ti,ab. (17572)
11 exp Acinetobacter/ (12028)
12 (Acinetobacter or mima or mimae or herellea or acinetobacterium).ti,ab. (10917)
13 exp Proteus/ (14447)
14 Proteus.ti,ab. (10461)
15 exp Serratia/ (9507)
16 Serratia.ti,ab. (7407)
17 exp Citrobacter freundii/ (1778)
18 ((Citrobacter adj freundii) or (bacterium adj freundii) or (Escherichia adj freundii)).ti,ab. (1675)
19 exp Morganella morganii/ (1134)
20 ((bacillus adj morgan$) or (bacterium adj morgana) or (morganella adj morgagni$) or (morganella adj morganii) or (proteus adj morgagni) or (proteus adj morgana$) or (salmonella adj morgana)).ti,ab. (804)
21 or/1-20 (396800)
22 (multiresistant or (multi adj resistan$)).ti,ab. (5599)
23 exp multidrug resistance/ (29629)
24 22 or 23 (33705)
25 exp Colistin/ (8049)
26 (belcomycin or colicort or colimycin$ or colisitin or colisticin or Colistin or colistine or colomycin or (coly adj mycin) or colymicin or colymycin or coly-mycin or multimycin or (Polymyxin adj E) or totazina).ti,ab. (3104)
27 exp Carbapenems/ (4745)
28 (Carbapenem$ or doripenem or ertapenem or Imipemide or Imipenem or Invanz or Invanz or meropenem or Merrem or ‘MK 0787’ or MK0787 or MK-0787 or N Formimidoylthienamycin or N-Formimidoylthienamycin or Penem or Ronem or S 4661 or S-4661 or SM 7338 or SM-7338 or Thienamycin$).ti,ab. (18086)
29 exp Piperacillin/ (14822)
30 (acopex or avocin or cl 227,193 or Cl 227193 or cl 227193 or cl227,193 or Cl227193 or cl227193 or Cl-227193 or cl-227193 or cypercil or hishiyacolin or ivacin or pentcillin or pentocillin or picillin$ or pipcil or pipera hameln or pipercil or pipercillin$ or piperacin or pipera-hameln or pipercillin or piperrline or pipraci$ or pipraks or pipril or piprilin or pitamycin or t 1220 or t1220 or t-1220 or taiperacillin).ti,ab. (6462)
31 exp Amoxicillin-Potassium Clavulanate Combination/ (23616)
32 (aclam or aktil or ambilan or amocla or amoclav or amoksiklav or amolanic or amometin or (amox adj clav) or amox-clav or (amoxi adj plus) or (amox adj3 clavalan$) or amoxiclav or amoxiclav-bid or amoxiclav-teva or amoxiklav or amoxindin or (amoxicillin-clavulanic adj acid) or ancla or (auclatin adj duo) or augamox or augmaxcil or augmentan or augmentin$ or augmex or augpen or (augucillin adj duo) or augurcin or ausclav or ausplici or bactiv or bactoclav or bioclavid or (brl adj ‘25000’) or brl25000 or brl-25000 or cavumox or ciblor or (clacillin adj duo) or clamax or clamentin or clamobit or clamonex or clamoid or clamoxin or (clamoxyl adj duo$) or clarin-duo or clavamox or clavar or clavinex or clavodar or clavoxil or (clavoxilin adj plus) or clavubactin or clavudale or clavulanate-amoxicillin or clavulin or (clavulox adj duo) or clavumox or (co adj amoxiclav) or (co adj amoxyclav) or coamoxiclav or coamoxyclov or (cramon adj duo) or (croanan adj duo) or curam or danoclav or (darzitil adj plus) or e-moxclav or enhancin or fleming or fugentin or (fullicilina adj plus) or gumentin or hibiotic or incidclav or klanonex or kmoxinil or lactamox or lansiclav or moxiclav or moxicle or moxyclav or natravax or nufaclav or palentin or quali-mentin or ranclav or spektromax or stacillin or suplentin or synermox or synulox or (velamox adj cl) or vestaclav or viaclav or vulamox or xiclav or (zami adj ‘8503’)).ti,ab. (11598)
33 exp Quinolones/ (101072)
34 ((chinolone adj derivative) or fluoroquinolones or (haloquinolone adj derivative) or ketoquinolines or oxoquinolines or quinolinones or quinolones).ti,ab. (15677)

35 exp Aminoglycosides/ (10599)

36 (Aminoglycosides or Anthracyclines or Aclarubicin or Daunorubicin or Plicamycin or Butirosin Sulfate or Sisomicin or Hygromycin B or Kanamycin or Dibekacin or Nebramycin + or Metrizamide or Neomycin or Framycetin or Paromomycin or Ribostamycin or Puromycin or Spectinomycin or Streptomycin or Dihydrostreptomycin Sulfate or Streptothricins or Streptozocin).ti,ab. (56708)

37 exp Gentamicins/ (70647)

38 (adelanin or alcomicin or apigent or apogen or apoten or azupel or bactiderm or biogaracin or bristagen or cidomycin or danigen or dermogen or dianfarma or disagent or duragentam$ or epigent or (frieso adj gent) or garabiotic or garalone or garamicin$ or garamycin or garbilocin or gencin or gendril or genoptic or genrex or gensumycin or gentabiotic or gentabiox or gentac or gentacidin or gentacin or gentacor or gentacycol or gentacly or gentafair or gentagram or gentak or gental or gentaline or gentalline or gentalol or gentalyn or gentamax or gentame$ or gentamicin$ or gentamina or gentamytrex or gentaplus or gentarad or gentasil or gentasol or gentasone or gentasporin or gentatrium or gentavet or genticin$ or genticyn or gentiderm or gentimycin or gentocin or gentogram or gentonmycin or genum or gevromycin or g-mycin or gmyticin or grammicin or hexamycin or jenamicin or konigen or lacromycin or lisagent or martigenta or migenta or miragenta or miramycin or nichogencin or nsc 82261 or nsc82261 or obogen or ocugenta or ocu-mycin or oftagen or optagram or opthagen or optigen or opti-genta or ottogenta or pyogenta or refobacin or ribomicin or rovixida or rupegen or sagentom or sch 9724 or sch9724 or sedanazin or servigenta or skinfect or sulmycin or tangyn or u-gencin or versigen or yectamicina).ti,ab. (23700)

39 exp Amikacin/ (28644)

40 (akacin or akicin or amicacin or amicasil or amicin or amiglymide v or amikacin$ or amikafur or amikan or amikayect or amikin or amikozit or amiktam or amitracin or amixin or amunin or apalin or bb k 8 or bb k8 or bbk 8 or bbbk 8 or bbbk8 or bbk-8 or bb-k8 or bicklin or biclin or bioclin or biokcin or briklin or chemacin or cinmik or fabianol or gamikal or gluakamin or kacinth-a or kanbine or kormakin or likacin or lukadlon or miacin or mikasome or onikin or oprad or orlobin or pediakin or pierami or riklinak or savox or selaxa or selemycin or sulfate amikacin or tybikin or vs 107 or vs107 or yectamid).ti,ab. (9841)

41 exp Fosfomycin/ (5561)

42 ([fosfocil or fosfocin or fosfocina or fosfomicin or fosfomycin or fosfonomycin or ‘mk 0955’ or mk 955 or mk0955 or mk955 or monuril or phosphomycin or phosphonomycin).ti,ab. (2386)

43 exp Aztreonam/ (10567)

44 ((az adj threonam) or azactam or azenam or azthreonam or aztreonam or (corus adj ‘1020’) or dynabiotic or primbactam or SQ 26,776 or sq 26,776 or sq 26776 or SQ-26,776 or sq26776 or sq-26776 or urobactam).ti,ab. (3245)

45 exp Nitrofurantoin/ (9724)
46 (berkfurin or biofurin or chemiofuran or dantafur or f 30 or f30 or fua-med or furaben or furadantin$ or furadantoin or furadina or furadoine or furadonin or furadonine or furalan or furanpur or furantocompren or furantoin$ or furobactina or furofen or furophen or infurin or ituran or ivadantin or macrobid or macrodantin$ or macrofurin or macrofurin or micofurantin$ or nitrofuratin or nephronex or niero6 or nifurantin or nifuryl or (nitro adj macro) or nitrofuracin or nitrofuradantoin or nitrofurantoin or nitrofurantoin$ or nitrofurin or novofuran or nsc 2107 or nsc2107 or orafuran or parfuran or phenurin or (potassium adj furagin) or ralodantin or trocurine or urantin or (uro adj tablinen) or urodi6 or urodon or urofurin or urolong or urotablinen or uro-tablinen or urotoline or uvi6min).ti,ab. (3412)

47 exp Cephalosporins/ (150937)

48 (Axepim$ or bmy 28142 or bmy28142 or BMY-28142 or Cefepim$ or cefepitax or ceficad or cepimax or forzyn beta or maxcef or maxfrom or maxipime or Quadrocef).ti,ab. (2995)

49 exp tazobactam/ (3045)

50 (cl 307579 or cl298741 or cl307579 or tazabactam or tazobac$ or tazocel or tazocillin$ or tazocin or tazomax or tazonam or tazopril or yp 14 or yp14 or ytr 830 or ytr 830h or ytr830 or ytr830h or zosyn).ti,ab. (3809)

51 exp temocillin/ (499)

52 ((brl adj ‘17421’) or brl17421 or (thiophenemalonamic adj acid) or negaban or temocillin or temopen).ti,ab. (236)

53 exp tigecycline/ (3876)

54 (tigecycline or (tbg adj mino) or tygacil or gar 936 or gar936 or (tert adj butylglycinamido$)).ti,ab. (1970)

55 exp cefepime/ (9948)

56 ((Cephalosporanic adj Acid$) or Cephalosporin$ or Cefamandole or Cefoperazone or Cefazolin or Cefonicid or Cefsulodin or Cephacetrile or Cefotaxime or Cephapirin or Cephalexin or Cefaclor or Cefadroxil or Cephaloglycin or Cephadrine or Cephaloridine or Ceftazidine or Cephemycins or Cefmetazole or Cefotetan or Cefoxitin).ti,ab. (45983)

57 exp pivmecillinam/ (685)

58 ((amdinocillin adj pivoxil) or (FL adj ‘1039’) or FL1039 or fl1039 or FL-1039 or pivamdinocillin or Pivmecillinam or Selexid or coactabs or (ro adj ‘109071’) or (ro10 adj ‘9071’) or ro109071).ti,ab. (280)

59 or/25-58 (349366)

60 21 and 24 and 59 (4969)

61 (review or review,tutorial or review, academic).pt. (1901059)

62 (systematic$ adj5 review$).tw,sh. (70959)

63 (systematic$ adj5 overview$).tw,sh. (869)

64 (quantitativ$ adj5 review$).tw,sh. (15516)

65 (quantitativ$ adj5 overview$).tw,sh. (203)
(quantitative adj5 synthesis).tw,sh. (2716)
(methodologic adj5 review).tw,sh. (3414)
(methodologic adj5 overview).tw,sh. (238)
(integrative research review or research integration).tw. (94)
(meta-analysis or meta analys or metaanalys).tw,sh. (96394)
(meta synthesis or meta synthesis or metasynthesis).tw,sh. (238)
(meta-regression or meta regression or metaregression).tw,sh. (2242)
(synthes$ adj3 literature).tw. (1448)
(synthes$ adj3 evidence).tw. (3583)
(integrative review.tw. (604)
data synthesis.tw. (8747)
(research synthesis or narrative synthesis).tw. (547)
(systematic study or systematic studies).tw. (7413)
(systematic comparison$.tw. (1183)
(comprehensive review$.tw. (6873)
critical review.tw. (11216)
(quantitative review.tw. (488)
(structured review.tw. (492)
(realist review.tw. (34)
(realist synthesis.tw. (12)
(review.ti. (264011)
(systematic$ literature review$.tw. (3464)
('systematic review'/ (55637)
('systematic review (topic)'/ (2885)
(meta analysis/ (67746)
('meta analysis (topic)'/ (5552)
(synthes$ adj2 qualitative).tw. (428)
(systematic adj2 search$).tw. (7848)
(systematic$ literature research$.tw. (102)
(review adj3 scientific literature).tw. (833)
(literature review adj2 side effect$).tw. (10)
(literature review adj2 adverse effect$).tw. (2)
(literature review adj2 adverse event$).tw. (6)
(evidence-based adj2 review).tw. (1915)
critical analysis.tw. (5559)
101 (review$ adj10 (papers or trials or trial data or studies or evidence or intervention$ or evaluation$ or outcome$ or findings)).tw. (248295)
102 review.ti. (264011)
103 metanaly$.tw. (316)
104 letter.pt. (800258)
105 editorial.pt. (417835)
106 104 or 105 (1218093)
107 or/61 - 103 (2212977)
108 107 not 106 (2200787)
109 (clin$ adj2 trial).mp. (968683)
110 ((singl$ or doubl$ or trebl$ or tripl$) adj (blind$ or mask$)).mp. (190403)
111 (random$ adj5 (assign$ or allocat$)).mp. (101920)
112 randomi$.mp. (613392)
113 crossover.mp. (59181)
114 exp randomized-controlled-trial/ (334017)
115 exp double-blind-procedure/ (112280)
116 exp crossover-procedure/ (35737)
117 exp single-blind-procedure/ (16758)
118 exp randomization/ (60197)
119 or/109-118 (1282139)
120 intervention?.ti. or (intervention? adj6 (clinician? or collaborat$ or community or complex or DESIGN$ or doctor? or educational or family doctor? or family physician? or family practitioner? or financial or GP or general practice? or hospital? or impact? or improv$ or individuali?e? or individuali?ing or interdisciplin$ or multicomponent or multi-component or multidisciplin$ or multi-disciplin$ or multifacet$ or multi-facet$ or multimodal$ or multi-modal$ or personali?e? or personali?ing or pharmacies or pharmacist? or pharmacy or physician? or practitioner? or prescrib$ or prescription? or primary care or professional$ or provider? or regulatory or regulatory or tailor$ or target$ or team$ or usual care)).ab. (175033)
121 (hospital$ or patient?).hw. and (study or studies or care or health$ or practitioner? or provider? or physician? or nurse? or nursing or doctor?).ti,hw. (1363115)
122 demonstration project?.ti,ab. (2081)
123 (pre-post or ‘pre test$’ or pretest$ or posttest$ or ‘post test$’ or (pre adj5 post)).ti,ab. (78013)
124 (pre-workshop or post-workshop or (before adj3 workshop) or (after adj3 workshop)).ti,ab. (673)
125 trial.ti. or ((study adj3 aim?) or ‘our study’).ab. (724065)
126 (before adj10 (after or during)).ti,ab. (394152)
127 (time points adj3 (over or multiple or three or four or five or six or seven or eight or nine or ten or eleven or twelve or month$ or hour? or day? or 'more than')).ab. (10006)
128 pilot.ti. (43036)
129 (multicentre or multicenter or multi-centre or multi-center).ti. (34428)
130 random$.ti,ab. or controlled.ti. (819713)
131 review.ti. (264011)
132 *experimental design/ or *pilot study/ or quasi experimental study/ (5205)
133 ('quasi-experiment$' or quasiexperiment$ or 'quasi random$' or quasirandom$ or 'quasi control$' or quasicontrol$ or ((quasi$ or experimental) adj3 (method$ or study or trial or design$))).ti,ab. (105122)
134 or/120-133 (3341084)
135 exp animals/ or exp invertebrate/ or animal experiment/ or animal model/ or animal tissue/ or animal cell/ or nonhuman/ (18985259)
136 human/ or normal human/ or human cell/ (14037258)
137 135 and 136 (14004971)
138 135 not 137 (4980288)
139 ('time series' adj2 interrupt$).ti,ab. (922)
140 134 not (138 or 139) (2996658)
141 108 or 119 or 140 (5157863)
142 and 141 (1860)

4.2.4. Medline (January 1946 to December 2012)

1 exp Escherichia coli/ (224545)
2 (Eaggec or (escherichia adj coli) or (e adj coli) or (alkalescens-dispar adj group) or (bacillus adj escherichii) or (Coli adj bacillus) or (Coli adj bacterium) or colibacillus or (colon adj bacillus)).ti,ab. (226847)
3 exp Klebsiella/ (13720)
4 (klebsiella or Calymmatobacterium or (aerobacter adj aerogenes) or ((bacillus or bacterium) adj pneumonia) or ((friedlaender or Friedlander) adj bacillus) or (Hyalococcus adj pneumonia) or Pneumobacillus).ti,ab. (18345)
5 ('k. pneumoniae' or 'b. friedlander').ti,ab. (3902)
6 exp Enterobacter/ (5504)
7 (enterobacter or aerobacter).ti,ab. (8130)
8 exp Pseudomonas aeruginosa/ (30232)
9 ((bacillus adj pyocyaneus) or (bacterium adj (aeruginosum or pyocyaneum)) or (blue adj apus) or (Pseudomonas adj (aeruginosa or aueofaciens or pyoceanus or pyocyanea or pyocyaneus))).ti,ab. (35984)
10 'p. aeruginosa'.ti,ab. (14103)
11 exp Acinetobacter/ (5262)
12 (Acinetobacter or mima or mima$ or herellea or acinetobacterium).ti,ab. (8005)
13 exp Proteus/ (8091)
14 Proteus.ti,ab. (9496)
15 exp Serratia/ (5505)
16 Serratia.ti,ab. (6720)
17 exp Citrobacter freundii/ (438)
18 ((Citrobacter adj freundii) or (bacterium adj freundii) or (Escherichia adj freundii)).ti,ab. (1361)
19 exp Morganella morganii/ (133)
20 ((bacillus adj morgan$) or (bacterium adj morgana) or (morganella adj morgagni$) or (morganella adj morganii$) or (proteus adj morgagni) or (proteus adj morgana$) or (salmonella adj morgana)).ti,ab. (601)
21 or/1-20 (360253)
22 (multiresistant or (multi adj resistan$)).ti,ab. (3949)
23 exp drug resistance, multiple/ (21763)
24 22 or 23 (24405)
25 exp Colistin/ (2107)
26 (belcomycin or colicort or colimycin$ or colisitin or colisticin or Colistin or colistine or (coly adj mycin) or colymicin or colymycin or coly-mycin or multymycin or (Polymyxin adj E) or totazina).ti,ab. (2346)
27 exp Carbapenems/ (6668)
28 (Carbapenem$ or doripenem or ertapenem or Imipemide or Imipenem or Invanz or Merrem or ‘MK 0787’ or MK0787 or MK-0787 or N Formimidoylthienamycin or N-Formimidoylthienamycin or Penem or Ronem or S 4661 or S-4661 or SM 7338 or SM-7338 or Thienamycin$).ti,ab. (11771)
29 exp Piperacillin/ (2035)
30 (acopex or avocin or cl 227,193 or Cl 227193 or cl 227193 or cl 227193 or cl227,193 or Cl227193 or cl227193 or Cl-227193 or cl-227193 or cypercil or hishiyadorin or ivacin or penticillin or pentocillin or picillin$ or pipcil or pipera hameln or piperacil or piperacillin$ or piperacin or pipera-hameln or pipercillin or piperilline or pipraci$ or pipraks or pipril or piprilin or pitamycin or t 1220 or t1220 or t-1220 or taiperacillin).ti,ab. (4319)
31 (cl 307579 or cl298741 or cl307579 or tazabactam or tazobac$ or tazocel or tazocillin$ or tazocin or tazomax or tazonam or tazopril or yp 14 or yp14 or yp 830 or yp 830h or ytr830 or ytr830h or zosyn).ti,ab. (2217)
32 exp Amoxicillin-Potassium Clavulanate Combination/ (1914)
33 (aclam or aktil or ambilan or amoca or amoclav or amokslav or amolanic or amometin or (amox adj clav) or amox-clav or (amoxi adj plus) or (amox adj3 clavulan$) or amoxiclav or amoxiclav-bid or amoxiclav-teva or amoxsiklav or
amoxxlin or (amoxycillin-clavulanic adj acid) or ancla or (auclatin adj duo) or augamox or augmaxcil or augmentan or augmentin$ or augmex or augpen or (augucillin adj duo) or clavamox or clavamox or clavavil or clavavil or clavulanate-amoxicillin or clavulin or (clavulox adj duo) or clarimox or (co adj amoxiclav) or (co adj amoxyclav) or coamoxiclav or coamoxiclav or coamoxiclav or coamoxiclav or (crandoff adj duo) or (crandoff adj duo) or curam or danoclav or (darzitil adj plus) or e-moxclav or enhancin or fleming or fustin or (fullicilina adj plus) or gumentin or hibiopic or inciclav or klamonex or lactamox or lansiclav or moxiclav or moxicle or moxyclav or nufaclav or palentin or quali-mentin or ranclav or spectramol or stacillin or suplentin or synermox or synulox or (velamox adj cl) or vestaclave or viaclav or vulamox or xiclav or (zami adj \'8503\')).ti,ab. (9184)

34 ((brl adj \'17421\') or brl17421 or (thiophenemalonamic adj acid) or negaban or temocillin or temopen).ti,ab. (179)

35 (tigecycline or (tbg adj mino) or tygacil or gar 936 or gar936 or (tert adj butylglycinamido$)).ab,ti. (1161)

36 exp Quinolones/ (33277)

37 ((chinolone adj derivative) or fluoroquinolones or (haloquinolone adj derivative) or ketoquinolones or oxoquinolines or quinolinones or quinolones).ti,ab. (11055)

38 exp Aminoglycosides/ (122582)

39 (Aminoglycosides or Anthracyclines or Aclarubicin or Daunorubicin or Plicamycin or Butirosin Sulfate or Sisomicin or Hygromycin B or Kanamycin or Dibekacin or Nebramycin + or Metrizamide or Neomycin or Framycetin or Paromomycin or Ribostamycin or Puromycin or Spectinomycin or Streptomycin or Dihydrostreptomycin Sulfate or Streptothricins or Streptozocin).ti,ab. (52288)

40 exp Gentamicins/ (16678)

41 (adelanin or alcomicin or apigent or apigen or apogen or apoten or azupel or bactiderm or biogaracin or bristan or cidomycin or danigen or dermogen or dianfarma or dispagent or duragentam$ or epigent or (frieso adj gent) or garbiopic or garalone or garmicit$ or garamycin or garbilocin or gencin or gendril or genciopic or genoptic or genrex or gensumycin or gentabiotic or gentabiox or gentac or gentacidin or gentacin or gentacor or gentacycol or gentacyl or gentafair or gentagram or gentak or gental or gentaline or gentalline or gentamo or gentamyl or gentamycin or gentamytrex or gentaplast or gentarad or gentasil or gentasol or gentasone or gentasporin or gentatrim or gentame$ or gentamicin$ or gentamin$ or gentamycin or gentamycin or gentalol or gentalyn or gentamax or gentame$ or gentamin$ or gentamycin or gentamycin or gentamyl or gentamyn$ or gentamyl or gentamytrex or gentaplast or gentarad or gentasil or gentasol or gentasone or gentasporin or gentatrim or gentavet or gentacin$ or genticyn or gentiderm or gentimycin or gentoxin or gentogram or gentomyacin or genuum or geomyicine or gevramycin or g-mycin or gmyticin or g-myctin or grammicin or hexamycin or jenamicin or konigen or lacromycin or lisagent or martigena or migenta or miragent a or miragmcin or nichogencin or nsc 82261 or nsc82261 or obogen or ocugen or ocu-mycin or oftagen or ophagam or optagen or optigen or opti-genta or ottogenta or pyogenta or refobacin or ribomicin or rigaminol or rocy gen or rovixida or rupegen or sagetam or sch 9724 or sch9724 or sedanazin or servigenta or skinfect or sulmymcin or tangyn or u-gencin or versigen or yectamicina).ti,ab. (19829)
42 exp Amikacin/ (3372)
43 (akacin or akcin or amicacina or amicasil or amicin or amiglymide v or amikacin$ or
amikafur or amikalem or amikan or amikayect or amikin or amiklin or amikozit or
amiktam or amitracin or amixin or amukin or apalin or bb k 8 or bb k8 or bbb k 8 or bbb-k
8 or bbb-8 or bb-k8 or bclin or bclin or biocacin or brcilin or briklin or chemacin or cinnik
or fabianol or gamikal or glukamin or kacinth-a or kanbine or kormakin or likacin or
lukad in or miacin or mikasome or onikin or oprad or orlobin or pediakin or pierami or
riklinak or savox or selaxa or selemycin or sulfate amikacin or
tybikin or vs 107 or vs107 or yectamid).ti,ab. (7140)
44 exp Fosfomycin/ (1378)
45 (fosfocil or fosfocin or fosfocina or fosfomicin or fosfomyacin or ‘mk
0955’ or mk 955 or mk0955 or mk955 or monuril or phosphomycin or
phosphonomycin).ti,ab. (1779)
46 exp Aztreonam/ (1233)
47 ((az adj threonam) or azactam or azenam or azthreonam or aztreonam or (corus adj
'1020’) or dynabiotic or primbactam or SQ 26,776 or sq 26,776 or sq 26776 or SQ-
26,776 or sq26776 or sq-26776 or urobactam).ti,ab. (2333)
48 exp Nitrofurantoin/ (2253)
49 (berkfurin or biofurin or chemiofuran or dantafur or f 30 or f30 or fua-med or
furab en or furadantin$ or furudantoin or furadina or furadoine or furodonin or
furadonine or furalan or furanpur or furantocompren or furantoin$ or furobactina or
furofen or furophen or infurin or ituran or ivadantin or macrobid or macrodantin$ or
macrofurin or macrofurin or micofurantin$ or nitrofuratoin or nephronex or nierofu or
nifurantin or nifuryl or (nitro adj macro) or nitrofuracin or nitrofurantoin or
nitrofurantaine or nitrofurantoin$ or nitrofurin or novofuran or nsc 2107 or nsc2107 or
orafuran or parfur an or phenurin or (potassium adj furagin) or ralodantin or trocurine
or urantin or (uro adj tablinen) or urodil or urodin or urofur an or urolong or
urotabl inen or uro-tablinen or urotoidina or uvamin).ti,ab. (2721)
50 exp Cephalosporins/ (35352)
51 (Axepim$ or bmy 28142 or bmy28142 or BMY-28142 or Cefepim$ or cefepitax or
ceficad or cepimax or forzyn beta or maxcef or maxfrom or maxipime or
Quadrocef).ti,ab. (1916)
52 ((Cephalosporanic adj Acid$) or Cephalosporin$ or Cefamandole or Cefoperazone or
Cefazolin or Cefonicid or Cefsulodin or Cephacetrile or Cefotaxime or Cephalothin or
Cephapirin or Cephalexin or Cefaclor or Cefadroxil or Cephaloglycin or Cephadrine or
Cephalexidine or Ceftazidime or Cephamycins or Cefmetazole or Cefotetan or
Cefoxitin).ti,ab. (35099)
53 exp Amdinocillin Pivoxil/ (199)
54 ((am dinocillin adj pivoxil) or (FL adj ‘1039’) or FL1039 or fl1039 or FL-1039 or
pivam dinocillin or Pivmecillinam or Selexid or coactabs or (ro adj ‘109071’) or (ro10
adj ‘9071’) or ro109071).ti,ab. (237)
55 or/25-54 (246506)
56 21 and 24 and 55 (3195)
57 exp clinical trial/ (706293)
58 exp randomized controlled trials/ (85563)
59 exp double-blind method/ (118498)
60 exp single-blind method/ (17086)
61 exp cross-over studies/ (30990)
62 randomized controlled trial.pt. (342334)
63 clinical trial.pt. (476450)
64 controlled clinical trial.pt. (85694)
65 (clinic$ adj2 trial).mp. (552367)
66 (random$ adj5 control$ adj5 trial$).mp. (443104)
67 (crossover or cross-over).mp. (59003)
68 ((singl$ or double$ or trebl$ or tripl$) adj (blind$ or mask$)).mp. (162179)
69 randomi$.mp. (509202)
70 (random$ adj5 (assign$ or allocat$ or assort$ or reciev$)).mp. (150717)
71 or/57-70 (968331)
72 (review or review,tutorial or review, academic).pt. (1758734)
73 (systematic$ adj5 review$).tw,sh. (40365)
74 (systematic$ adj5 overview$).tw,sh. (663)
75 (quantitativ$ adj5 review$).tw,sh. (3684)
76 (quantitativ$ adj5 overview$).tw,sh. (153)
77 (quantitativ$ adj5 synthesis$).tw,sh. (1107)
78 (methodologic$ adj5 review$).tw,sh. (2696)
79 (methodologic$ adj5 overview$).tw,sh. (180)
80 (integrative research review$ or research integration).tw. (78)
81 meta-analysis as topic/ (12608)
82 (meta-analys$ or meta analys$ or metaanalys$).tw,sh. (62359)
83 (meta synthesis or meta synthesis or metasynthesis).tw,sh. (215)
84 (meta-regression or meta regression or metaregression).tw,sh. (1650)
85 meta-analysis.pt. (37918)
86 (synthes$ adj3 literature).tw. (1070)
87 (synthes$ adj3 evidence).tw. (2956)
88 integrative review.tw. (583)
89 data synthesis.tw. (6328)
90 (research synthesis or narrative synthesis).tw. (463)
91 (systematic study or systematic studies).tw. (5679)
92 systematic comparison$.tw. (953)
93 systematic comparison$.tw. (953)
94 evidence based review.tw. (965)
95 comprehensive review$.tw. (5290)
96 critical review.tw. (9227)
97 quantitative review.tw. (382)
98 structured review.tw. (376)
99 realist review.tw. (24)
100 realist synthesis.tw. (11)
101 review.ti. (212126)
102 (review$ adj4 (papers or trials or studies or evidence or intervention$ or evaluation$)).tw. (80949)
103 metanaly$.tw. (137)
104 letter.pt. (766872)
105 editorial.pt. (310993)
106 comment.pt. (493546)
107 or/104-106 (1166749)
108 or/72-103 (1897061)
109 108 not 107 (1860495)
110 intervention?.ti. or (intervention? adj6 (clinician? or collaborat$ or community or complex or DESIGN$ or doctor? or educational or family doctor? or family physician? or family practitioner? or financial or GP or general practice? or hospital? or impact? or improv$ or individuali?e? or individuali?ing or interdisciplin$ or multicomponent or multi-component or multidisciplin$ or multi-disciplin$ or multifacet$ or multi-facet$ or multimodal$ or multi-modal$ or personali?e? or personali?ing or pharmacies or pharmacist? or pharmacy or physician? or practitioner? or prescrib$ or prescription? or primary care or professional$ or provider? or regulatory or regulatory or tailor$ or target$ or team$ or usual care)).ab. (128957)
111 (pre-intervention? or preintervention? or 'pre intervention?' or post-intervention? or postintervention? or 'post intervention?').ti,ab. (7451)
112 demonstration project?.ti,ab. (1742)
113 (pre-post or ‘pre test$’ or pretest$ or posttest$ or ‘post test$’ or (pre adj5 post)).ti,ab. (52427)
114 (pre-workshop or post-workshop or (before adj3 workshop) or (after adj3 workshop)).ti,ab. (472)
115 trial.ti. or ((study adj3 aim?) or ‘our study’).ab. (500725)
116 (before adj10 (after or during)).ti,ab. (314768)
117 ('quasi-experiment$' or quasiexperiment$ or 'quasi random$' or quasirandom$ or 'quasi control$' or quasicontrol$ or ((quasi$ or experimental) adj3 (method$ or study or trial or design$))).ti,ab,hw. (84783)
118 ('time series' adj2 interrupt$).ti,ab,hw. (744)
119 (time points adj3 (over or multiple or three or four or five or six or seven or eight or nine or ten or eleven or twelve or month$ or hour? or day? or 'more than')).ab. (7043)
120 pilot.ti. (32084)
121 Pilot projects/ (74648)
122 (clinical trial or controlled clinical trial or multicenter study).pt. (595489)
123 (multicentre or multicenter or multi-centre or multi-center).ti. (24301)
124 random$.ti,ab. or controlled.ti. (624993)
125 (control adj3 (area or cohort? or compare? or condition or design or group? or intervention? or participant? or study)).ab. not (controlled clinical trial or randomized controlled trial).pt. (342332)
126 'comment on'.cm. or review.ti,pt. or randomized controlled trial.pt. (2652864)
127 (rat or rats or cow or cows or chicken? or horse or horses or mice or mouse or bovine or animal?).ti. (1254855)
128 exp animals/ not humans.sh. (3812817)
129 (or/110-126) not (or/127-128) (3811646)
130 71 or 109 or 129 (4107075)
131 and 130 (822)
### 4.3. Clinical Review Tables

#### 4.3.1. Antibiotic stewardship

<table>
<thead>
<tr>
<th>Study</th>
<th>Objective and participants</th>
<th>MDR Gram-negative bacteria</th>
<th>Intervention, control and follow-up</th>
<th>Results</th>
<th>Quality assessment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ben-David 2010 ITS</td>
<td>To assess the effect of an intensified intervention, that included active surveillance, on the incidence of infection with carbapenem-resistant <em>K. pneumoniae</em></td>
<td><strong>Bacteria</strong>: <em>K. pneumoniae</em> <strong>Resistant to</strong>: carbapenems, cephalosporins, fluoroquinolones, trimethoprim-sulfamethoxazole <strong>Mechanism of resistance</strong>: not reported</td>
<td>1. Enhanced national infection control programme: contact precautions were used for the care of all patients with CRKP colonization or infection; the prevalence of colonization or infection was reported daily, and this information was mailed to the hospital management and the national coordinator; and patients infected with CRKP had their names entered into a database so that they could be identified at hospital readmission 2. Active surveillance programme: obtaining rectal culture samples from patients hospitalized in ICUs and in step-down units, at admission to the unit and once weekly until the patient was discharged</td>
<td>Infection control Before the intervention, the incidence of clinical infection with CRKP had increased 6.42-fold to 6.93 cases per 10,000 patient-days After an enhanced infection control and active surveillance programme was introduced, the incidence of clinical infection reduced to 1.8 cases per 10,000 patient-days (<em>P</em>&lt;0.001). The slope significantly changed with the introduction of the intervention from 0.12 to -0.07 (<em>P</em>&lt;0.001)</td>
<td>ITS Protection against secular changes (high quality) Protection against detection bias (acceptable quality)</td>
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<tr>
<td>Setting Tertiary (one hospital) Israel</td>
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<tr>
<td>January 2006–December 2008</td>
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</table>

<table>
<thead>
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<th>MDR Gram-negative bacteria</th>
<th>Intervention</th>
<th>Results</th>
<th>Quality assessment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Borer 2011 ITS</td>
<td>To devise a local strategy for eradication of a hospital-wide outbreak caused by CRKP</td>
<td><strong>Bacteria</strong>: <em>K. pneumoniae</em></td>
<td>1. Emergency department flagging system</td>
<td>Bacterial colonization and infection</td>
<td>ITS Protection against secular changes (high quality)</td>
</tr>
</tbody>
</table>
### Objective and participants

**Setting**
Tertiary (one hospital)
Israel
May 2006–May 2010

**Participants**
N=803
Adolescents 13–18 years, adults 19–45 years, middle aged 46–64 years, elderly 65–79 years, elderly 80+ years
Male: 410, female: 393

Inclusion criteria: data from medical records of patients with CRKP infection
Exclusion criteria: not reported

**MDR Gram-negative bacteria**
Resistant to: carbapenems
Mechanism of resistance: not reported

**Intervention, control and follow-up**
2. Building of a cohort space or ward
3. Intensive active surveillance in high-risk wards
4. Epidemiological investigations
5. Carbapenem-restriction policy

**Length of pre-intervention**: 11 months prior
**Length of post-intervention**: 36 months following

**Results**
During the intervention, the CRKP undetected ratio showed a significant increase from 55.7% for June–December 2007 to 71.2% in 2008, 78.9% in 2009 and 92.5% for February–May 2010 (P≤0.001).

From May 2006 through April 2007 (pre-intervention), the CRKP-IN incidence density per 10,000 patient-days was 5.26. After the intervention programme was introduced, the incidence of clinical CRPK infection reduced to 2.91 cases per 10,000 patient-days (P<0.001) in 12/2007, 1.91 in 12/2008 and 1.28 in 12/2009. The slope changed significantly with the introduction of the intervention (P=0.004).

**Antibiotic use**
Meropenem use showed a statistically significant decrease from 2007 to 2010 (P≤0.001); colistin use increased significantly during the same period (P≤0.001)

---

### Church 2011

**ITS Setting**
Secondary (one hospital)

**Participants**
N: not reported

**Bacteria**: *P. aeruginosa*
Resistant to: aminoglycosides (tobramycin), cephalosporins (cefepime), piperacillin/tazobactam

**Intervention**
1. Levofloxacin replaced with gatifloxacin in 2001
2. Gatifloxacin replaced with moxifloxacin in 2006

**Antibiotic resistance and susceptibility**
No association between the susceptibility of *P. aeruginosa* isolates to tobramycin and formulary changes was noted. With cefepime, a significant change in susceptibility was detected after the introduction of gatifloxacin (P=0.0099) and

**ITS** Protection against secular changes (low quality)
Protection against detection bias (low quality)
<table>
<thead>
<tr>
<th>Objective and participants</th>
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<th>Intervention, control and follow-up</th>
<th>Results</th>
<th>Quality assessment</th>
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<tbody>
<tr>
<td>USA</td>
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<tr>
<td>Age: not reported</td>
<td>Mechanism of resistance: not reported</td>
<td>Length of pre-intervention: 15 months prior</td>
<td>moxifloxacin ($P=0.0571$). In the case of piperacillin/tazobactam, a positive change in susceptibility over time was detected after introduction of moxifloxacin ($P=0.0589$). In each analysis, the effect of total fluoroquinolone usage was not significant</td>
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<tr>
<td>Male: not reported, female: not reported</td>
<td></td>
<td>Length of post-intervention 1: 60 months</td>
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<tr>
<td>Inclusion criteria: data from clinical microbiology and pharmacy databases of the Medical University of South Carolina Medical Centre</td>
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<td>Length of post-intervention 2: 30 months following</td>
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<tr>
<td>Exclusion criteria: not reported</td>
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<tr>
<td></td>
<td>Bacteria: <em>K. pneumoniae</em></td>
<td>Intervention</td>
<td>Bacterial colonization and infection</td>
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<tr>
<td></td>
<td>Resistant to: carbapenems</td>
<td>1. Single-room isolation and contact precautions</td>
<td>The incidence (total number of cases of in-hospital CRKP acquisition detected by clinical cultures) and weekly point prevalence were reported as the number of cases per 1000 hospital beds</td>
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<td></td>
<td>Mechanism of resistance: not reported</td>
<td>2. Cohorting of patients and nursing staff, screening of patients in the same room as newly identified carriers of CRKP, and local protocol for continued cohorting of returning patients</td>
<td>Incidence was found to change significantly after intervention 2 (06/2007) and 3 (10/2008). Prevalence was found to change significantly only in September 2009 (after intervention 4)</td>
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<td>3. Weekly active surveillance in the ICU</td>
<td>In the emergency department, the mean rate of compliance with the active surveillance protocol (± SD) was 43% ± 10%</td>
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<td>4. Active surveillance of patients on admission to the emergency department</td>
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<td></td>
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<td>Length of pre-intervention: not reported</td>
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<td>Length of post-intervention 1: 14 months</td>
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<td>Length of post-intervention 2: 39 months</td>
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<td>Cohen 2011</td>
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<td>ITS Protection against secular changes (high quality)</td>
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<td>ITS</td>
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<td></td>
<td>Protection against detection bias (acceptable to low quality)</td>
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<tr>
<td>Setting</td>
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<tr>
<td>Tertiary (one hospital)</td>
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<td>Israel</td>
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<td>March 2006–August 2010</td>
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<tr>
<td>To describe the implementation of an institution-wide, multiple-step intervention to curtail the epidemic spread of CRKP</td>
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<tr>
<td>Participants N=33,570</td>
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<tr>
<td>Age: not reported</td>
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<tr>
<td>Male: not reported, female: not reported</td>
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<tr>
<td>Inclusion criteria: all patients affected by CRKP</td>
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<tr>
<td>Exclusion criteria: not reported</td>
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<tr>
<td></td>
<td>Bacteria: <em>K. pneumoniae</em></td>
<td>Intervention</td>
<td>Bacterial colonization and infection</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Resistant to: carbapenems</td>
<td>1. Single-room isolation and contact precautions</td>
<td>The incidence (total number of cases of in-hospital CRKP acquisition detected by clinical cultures) and weekly point prevalence were reported as the number of cases per 1000 hospital beds</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Mechanism of resistance: not reported</td>
<td>2. Cohorting of patients and nursing staff, screening of patients in the same room as newly identified carriers of CRKP, and local protocol for continued cohorting of returning patients</td>
<td>Incidence was found to change significantly after intervention 2 (06/2007) and 3 (10/2008). Prevalence was found to change significantly only in September 2009 (after intervention 4)</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>3. Weekly active surveillance in the ICU</td>
<td>In the emergency department, the mean rate of compliance with the active surveillance protocol (± SD) was 43% ± 10%</td>
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<tr>
<td></td>
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<td>4. Active surveillance of patients on admission to the emergency department</td>
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<tr>
<td></td>
<td></td>
<td>Length of pre-intervention: not reported</td>
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<tr>
<td></td>
<td></td>
<td>Length of post-intervention 1: 14 months</td>
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<tr>
<td></td>
<td></td>
<td>Length of post-intervention 2: 39 months</td>
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<tr>
<td>Objective and participants</td>
<td>MDR Gram-negative bacteria</td>
<td>Intervention, control and follow-up</td>
<td>Results</td>
<td>Quality assessment</td>
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<tr>
<td><strong>Dortch 2011</strong></td>
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<tr>
<td><strong>ITS</strong></td>
<td></td>
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<tr>
<td><strong>Setting</strong></td>
<td>Tertiary (one TICU, one SICU)</td>
<td></td>
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<tr>
<td><strong>USA</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>January 2001–December 2008</strong></td>
<td></td>
<td></td>
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</tr>
<tr>
<td>To examine the effect of the antibiotic stewardship programme on the incidence of resistant Gram-negative HAI</td>
<td>Bacteria: <em>P. aeruginosa</em>, <em>Acinetobacter</em> spp.</td>
<td></td>
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</tr>
<tr>
<td></td>
<td>Resistant to: aminoglycosides, carbapenems, cephalosporins (third- and fourth-generation), fluoroquinolones</td>
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</tr>
<tr>
<td></td>
<td>Mechanism of resistance: not reported</td>
<td></td>
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</tr>
<tr>
<td><strong>Intervention</strong></td>
<td>1. Antibiotic stewardship: April 2002, guidelines for prophylactic antibiotics were devised for select procedures</td>
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<tr>
<td></td>
<td>2. Antibiotic rotation: January 2005, institution-wide initiative for surgical prophylaxis based on the Surgical Care Improvement Project</td>
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<tr>
<td><strong>Length of post-intervention 3</strong>: 2 years</td>
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<td><strong>Length of post-intervention 4</strong>: 15 months</td>
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<tr>
<td><strong>Lewis 2012</strong></td>
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<tr>
<td><strong>ITS</strong></td>
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<tr>
<td><strong>Setting</strong></td>
<td>Tertiary (11 ICUs and intermediate care units)</td>
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<tr>
<td>To examine the effect of restricting ciprofloxacin use on the resistance of nosocomial Gram-negative bacilli, including <em>P. aeruginosa</em>, to group 2 carbapenems in a hospital’s ICUs</td>
<td>Bacteria: <em>E. aerogenes</em>, <em>E. cloacae</em>, <em>P. aeruginosa</em>, <em>A. baumannii</em></td>
<td></td>
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<tr>
<td></td>
<td>Resistant to: carbapenems (imipenem,</td>
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<tr>
<td></td>
<td>Intervention</td>
<td>Restriction of ciprofloxacin: ciprofloxacin use was restricted hospital wide in July 2007; after this restriction, pre-approval by the on-call infectious diseases fellow was required for its use</td>
<td></td>
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</tr>
<tr>
<td></td>
<td><strong>Antibiotic use</strong></td>
<td>Both in the SICU and TICU and there was a significant decrease in the utilization of total broad-spectrum antibiotics (BLIC, carbapenems, fluoroquinolones, third- and fourth-generation cephalosporins) targeting Gram-negative pathogens over the observation period (<em>P</em>&lt;0.001)</td>
<td></td>
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<tr>
<td></td>
<td></td>
<td><strong>Infection</strong></td>
<td>During the 8-year observation period, the proportion of healthcare-associated infections caused by MDR Gram-negative pathogens decreased from 37.4% (2001) to 8.5% (2008), whereas the proportion of healthcare-associated infections caused by pan-sensitive pathogens increased from 34.1% to 53.2%</td>
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<tr>
<td></td>
<td></td>
<td><strong>ITS</strong></td>
<td>Protection against secular changes (high quality)</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>Protection against detection bias (acceptable to low quality)</td>
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</tr>
</tbody>
</table>
Objective and participants

**MDR Gram-negative bacteria**

Intervention, control and follow-up

Results

Quality assessment

<table>
<thead>
<tr>
<th>Objective and participants</th>
<th>MDR Gram-negative bacteria</th>
<th>Intervention, control and follow-up</th>
<th>Results</th>
<th>Quality assessment</th>
</tr>
</thead>
<tbody>
<tr>
<td>immediate care units) USA</td>
<td>N: not reported</td>
<td>meropenem, doripenem, cephalosporins (cefepime), pipercillin/tazobactam, fluoroquinolones (ciprofloxacin)</td>
<td>Length of pre-intervention: 42 months</td>
<td>significantly (P=0.0134) from 11.96 DDD/1000 patient-days in 2004 to 28.19 DDD/1000 patient-days in 2010. Overall, there was a hospital-wide decrease of 18.4% (P&lt;0.0001) in the use of antibacterials during the study time</td>
</tr>
<tr>
<td>January 2004– December 2010</td>
<td>Age: not reported</td>
<td>Mechanism of resistance: not reported</td>
<td>Length of post-intervention: 42 months</td>
<td>Infection There were no changes observed in the number of nosocomial S. maltophilia isolates per 10,000 patient-days following the restriction of ciprofloxacin</td>
</tr>
<tr>
<td></td>
<td>Male: not reported, female: not reported</td>
<td></td>
<td></td>
<td>Antibiotic resistance Over the seven-year time period, there was a decrease of 13.7% in the percentage of ciprofloxacin-resistant P. aeruginosa isolates that were collected, which equates to a decrease of 3.9% per year (P=0.0017). No significant changes was observed in the susceptibilities to the group II carbapenems of nosocomial Enterobacteriaceae or A. baumannii isolates</td>
</tr>
<tr>
<td></td>
<td>Inclusion criteria: all clinical ICU and intermediate care unit specimens (blood, sterile fluid, sputum, urine, wounds and anaerobic specimens) with test results that were positive for P. aeruginosa, E. aerogenes, E. cloacae, A. baumannii and S. maltophilia. Only nosocomial cases, defined as involving patients who had a hospital length of stay exceeding two days</td>
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<td></td>
<td>Exclusion criteria: results of surveillance and environmental sample cultures.</td>
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<tr>
<td>Meyer 2009</td>
<td>To test whether reduction of third-generation cephalosporin use has a sustainable positive impact on the high endemic prevalence of third generation cephalosporin-resistant K. pneumoniae and E. coli in an ICU</td>
<td>Bacteria: E. coli, K. pneumoniae, P. aeruginosa</td>
<td>Antibiotic use Following the implementation of guidelines in a surgical ICU, a significant and sustainable decrease in the use of third-generation cephalosporins of -110.2 DDD/1000 patient-days (95% CI -140.0 to -80.4, R²=0.468) was observed. There was</td>
<td></td>
</tr>
<tr>
<td>ITS</td>
<td>Setting Tertiary (one ICU) Germany</td>
<td>Resistant to: cephalosporins (third-generation), pipercillin</td>
<td>Intervention 1. Education programmes for professionals and patients in July 2004 2. Education sessions on antibiotic guidelines were</td>
<td></td>
</tr>
<tr>
<td>Protection against secular changes (high quality)</td>
<td>Participants</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Protection against detection bias (acceptable quality)</td>
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</tbody>
</table>
**Objective and participants**

**MDR Gram-negative bacteria**

**Intervention, control and follow-up**

**Results**

**Quality assessment**

<table>
<thead>
<tr>
<th><strong>January 2002–December 2006</strong></th>
<th><strong>N=3758</strong></th>
<th><strong>Age: not reported</strong></th>
<th><strong>Male: not reported, female: not reported</strong></th>
<th><strong>Mechanism of resistance: ESBL</strong></th>
<th><strong>held in the departments of surgery and anaesthesiology</strong></th>
<th><strong>Intervention, control and follow-up</strong></th>
<th><strong>a significant reduction in the use of ampicillins (-167.4 DDD/1000, 95% CI -223.8 to -110.9, R^2=0.378) and in the use of imidazoles (-94.5 DDD/1000, 95% CI -121.2 to -67.7, R^2=0.463)</strong></th>
<th><strong>detection bias (high quality)</strong></th>
</tr>
</thead>
</table>

**Mechanism of resistance:**

- **ESBL**

**Intervention, control and follow-up**

- **Empiric standard therapy for peritonitis and other intra-abdominal infections was switched from third-generation cephalosporins to piperacillin in combination with a beta-lactamase inhibitor. The duration of antibiotic therapy for open fractures was shortened to single-shot pre-operative prophylaxis**

**Length of pre-intervention:** 30 months

**Length of post-intervention:** 30 months

**Antibiotic use**

Following the implementation of a comprehensive teaching session on antibiotic prophylaxis in cerebrospinal shunts in a surgical ICU, pre-operative prophylaxis for shunt catheters was changed into single-shot prophylaxis, and total antibiotic use decreased (-147.3 DDD/1000 patient-days, P=0.052). This corresponded to a decrease of 15% in the use of cefuroxime.

**Quality assessment**

- **ITS Protection against secular changes (high quality)**
- **Protection against detection bias (acceptable quality)**

**Meyer 2010**

**ITS**

**Setting**

- **Tertiary (one ICU)**
- **Germany**

**January 2002–December 2006**

**Participants**

- **N=11,887**
- **Age: not reported**
- **Male: not reported, female: not reported**

**Bacteria:**

- **E. coli, K. pneumoniae, P. aeruginosa**

**Resistant to:**

- carbapenems (imipenem), cephalosporins (third-generation)

**Mechanism of resistance:**

- not reported

**Intervention**

Change in antibiotic prophylaxis:

- Revised recommendation of single-shot prophylaxis with cefuroxime for shunt catheters, beginning in January 2004

**Length of pre-intervention:** 24 months prior

**Length of post-intervention:** 36 months following

**Antibiotic use**

- Following the implementation of a comprehensive teaching session on antibiotic prophylaxis in cerebrospinal shunts in a surgical ICU, pre-operative prophylaxis for shunt catheters was changed into single-shot prophylaxis, and total antibiotic use decreased (-147.3 DDD/1000 patient-days, P=0.052). This corresponded to a decrease of 15% in the use of cefuroxime.

**Quality assessment**

- **ITS Protection against secular changes (high quality)**
- **Protection against detection bias (acceptable quality)**

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<table>
<thead>
<tr>
<th><strong>Objective and participants</strong></th>
<th><strong>MDR Gram-negative bacteria</strong></th>
<th><strong>Intervention, control and follow-up</strong></th>
<th><strong>Results</strong></th>
<th><strong>Quality assessment</strong></th>
</tr>
</thead>
<tbody>
<tr>
<td>Inclusion criteria: monthly data on antimicrobial use obtained from the computerized pharmacy database. Monthly resistance data collected from the microbiology laboratory. Only samples taken in the ICU were considered. Exclusion criteria: copy strains – defined as an isolate of the same species showing the same susceptibility pattern throughout a 1-month period in the same patient, no matter what the site of isolation.</td>
<td><strong>Bacteria</strong>: <em>E. coli</em>, <em>Klebsiella</em> spp., <em>Enterobacter</em> spp., <em>P. aeruginosa</em>, <em>Acinetobacter</em> spp.</td>
<td><strong>Intervention</strong> National guidelines on antimicrobial prescribing; antibiotic stewardship via computerized decision support systems. In 2001, one system guiding antibiotic use outside the ICU – a web-based antimicrobial approval system for third-generation cephalosporins (cefotaxime and ceftriaxone). In 2002, targeting the ICU specifically – computerized decision support system for antibiotic prescribing.</td>
<td>The reduction in total antibiotic consumption was sustainable and did not increase over the next 36 months.</td>
<td></td>
</tr>
<tr>
<td><strong>Yong 2010</strong></td>
<td><strong>Mechanism of resistance</strong>: not reported</td>
<td><strong>Length of pre-intervention</strong>: 30 months <strong>Length of post-intervention</strong>: 54 months</td>
<td><strong>Antibiotic use</strong> Following the implementation of national guidelines on antimicrobial prescribing and antibiotic stewardship, there was a significant reduction in the number of imipenem-resistant <em>E. coli</em> and <em>Klebsiella</em> spp. isolates observed in the ICU. A small but significant improvement in the number of imipenem-resistant <em>Acinetobacter</em> spp. isolates was also observed. For Enterobacteriaceae with potentially inducible beta-lactamases, no significant changes was observed in imipenem susceptibility, although gentamicin susceptibility increased at a rate of 2.1%/year (95% CI 0.7–3.4), and</td>
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<tr>
<td><strong>ITS</strong></td>
<td><strong>Protection against secular changes (high quality)</strong></td>
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<tr>
<td><strong>Setting</strong></td>
<td><strong>Protection against detection bias (acceptable to low quality)</strong></td>
<td></td>
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<tr>
<td>Tertiary (one ICU) Australia</td>
<td></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>January 2000–December 2006</td>
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<tr>
<td>Objective and participants</td>
<td>MDR Gram-negative bacteria</td>
<td>Intervention, control and follow-up</td>
<td>Results</td>
<td>Quality assessment</td>
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<tr>
<td>To determine the relation of carbapenem restriction with the incidence of MDR <em>A. baumannii</em> in VAP</td>
<td>Bacteria: <em>A. baumannii</em></td>
<td>Carbapenem restriction policy limiting the use of third-generation carbapenems. Only used when severe sepsis and after consultation with a physician from the Department of Infectious Diseases. <em>N</em>=12</td>
<td>Mortality rates did not differ significantly between the treatment groups (RR 0.78; 95% CI 0.29–2.12).</td>
<td>RCT Low methodological quality (0)</td>
</tr>
<tr>
<td>Participants</td>
<td>Resistant to: carbapenems</td>
<td>Control group</td>
<td>Antibiotic resistance</td>
<td>Small sample size</td>
</tr>
<tr>
<td><em>N</em>=26</td>
<td>Mechanism of resistance: ESBL</td>
<td>Conventional treatment: no restrictions of carbapenem (doctors were able to prescribe if necessary). <em>N</em>=15</td>
<td>More patients in the conventional group developed a carbapenem-resistant strain of <em>A. baumannii</em>, although the difference was not statistically significant (RR 0.63; 95% CI 0.38–1.04)</td>
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<tr>
<td>Adults 19–45 years, middle aged 46–64 years, aged 65–79 years</td>
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<tr>
<td>Male: 15, female: 11</td>
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</tr>
<tr>
<td>Inclusion criteria: Patients receiving mechanical ventilation for more than five days and diagnosed with VAP</td>
<td></td>
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</tr>
<tr>
<td>Exclusion criteria: not reported</td>
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</table>

*K. pneumoniae, Klebsiella pneumonia; P. aeruginosa, Pseudomonas aeruginosa; A. baumannii, Acinetobacter baumannii; E. coli, Escherichia coli; E. aerogenes, Enterobacter aerogenes; E. cloacae, Enterobacter cloacae; S. maltophilia, Stenotrophomonas maltophilia; CRKP, carbapenem-resistant K. pneumoniae; SICU, surgical intensive care unit; TICU, trauma intensive care unit; VAP, ventilator-associated pneumonia; MDR, multi-drug resistant; ESBL, ciprofloxacin susceptibility increased at a rate of 0.9%/year (95% CI 0.1–1.7).*
extended-spectrum beta-lactamase; BLIC, beta-lactam/beta-lactamase inhibitor combinations; ITS, interrupted time series; RCT, randomized controlled trial; ICU, intensive care unit; FQ, fluoroquinolones; 3/4CEPH, third- and fourth-generation cephalosporins; HAI, healthcare-associated infection; CI, confidence interval; RR, risk ratio; DDD, defined daily dose; SD, standard deviation.
4.3.2. Other infection control measures

<table>
<thead>
<tr>
<th>Study details</th>
<th>Objective and participants</th>
<th>MDR Gram-negative bacteria</th>
<th>Intervention, control and follow-up</th>
<th>Results</th>
<th>Quality assessment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Levin 2010 CBA</td>
<td>To analyse whether single patient rooms in the ICU decreased bacterial transmission between ICU patients</td>
<td><strong>Bacteria:</strong> Acinetobacter spp., other Gram-negative bacteria</td>
<td><strong>Intervention</strong> ICU A converted to single patient rooms. Old ICU A N=64, new ICU A N=62</td>
<td>Infection control The single-room ICU A had a significantly lower ICU acquisition of resistant organisms when compared with ICU B during the same period [3/62 (5%) vs 7/39 (18%), respectively, ( P=0.043 )], which was confirmed using survival analysis (( P=0.011 )). ICU B showed no changes over the study</td>
<td>CBA Low methodological quality (0)</td>
</tr>
<tr>
<td><strong>Setting</strong></td>
<td><strong>Participants</strong> N=207</td>
<td><strong>Resistant to:</strong> carbapenems</td>
<td><strong>Control group</strong> ICU B remained open plan. Old ICU B N=44, new ICU B N=39</td>
<td><strong>Length of follow-up:</strong> not reported</td>
<td></td>
</tr>
<tr>
<td><strong>Tertiary (two ICUs)</strong> Israel</td>
<td>Age: not reported Male: not reported, female: not reported</td>
<td><strong>Mechanism of resistance:</strong> ESBL</td>
<td></td>
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<tr>
<td>Dates not reported</td>
<td>Inclusion criteria: not reported</td>
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<tr>
<td></td>
<td>Exclusion criteria: not reported</td>
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ICU, intensive care unit; ESBL, extended-spectrum beta-lactamase; CBA, controlled before–after study.

4.3.3. Selective decontamination

<table>
<thead>
<tr>
<th>Study details</th>
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<th>MDR Gram-negative bacteria</th>
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<th>Quality assessment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Agusti 2002 Quasi-randomized</td>
<td>To determine the efficacy of SDD in patients with multi-drug-resistant A. baumannii intestinal colonization</td>
<td><strong>Bacteria:</strong> A. baumannii</td>
<td><strong>Intervention</strong> SDD: a combination of polymyxin E (colistin) (150 mg) and tobramycine (80 mg) administered in 20-mL liquid form x 4/day (orally or through</td>
<td>Bacterial colonization Rates of faecal, pharyngeal and axillary colonization did not significantly reduce during ICU stay in the control group (( P ) value not reported). In the SDD group, the rate</td>
<td>Quasi-randomized Low methodological quality (0)</td>
</tr>
<tr>
<td><strong>Setting</strong></td>
<td><strong>Participants</strong> N=54</td>
<td><strong>Resistant to:</strong> aminoglycosides (tobramycine)</td>
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<table>
<thead>
<tr>
<th>Study details</th>
<th>Objective and participants</th>
<th>MDR Gram-negative bacteria</th>
<th>Intervention, control and follow-up</th>
<th>Results</th>
<th>Quality assessment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tertiary (one ICU) Spain October 1998–June 1999</td>
<td>Adults 19–45 years, middle aged 46–64 years, aged 65–79 years Male: 16, female: 5</td>
<td>Mechanism of resistance: not reported</td>
<td>nasogastric tube), and 0.5 g of gel containing 2% of colistin and tobramycine applied round the gum margins and oropharynx x 4/day. Duration of treatment from detection of <em>A. baumannii</em> to discharge from ICU. N=21</td>
<td>of faecal and pharyngeal carriage was reduced significantly (<em>P</em>&lt;0.001 and <em>P</em>=0.003, respectively), but not the rate of cutaneous carriage</td>
<td>Small sample size</td>
</tr>
<tr>
<td></td>
<td>Inclusion criteria: Intervention group 1. All patients with <em>A. baumannii</em> fecal colonization 2. An expected ICU stay exceeding five days</td>
<td></td>
<td></td>
<td>Antibiotic resistance</td>
<td></td>
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<tr>
<td></td>
<td>Control group 1. All patients admitted 1 October–30 November 1998 with <em>A. baumannii</em> faecal colonization 2. At least one series of axillary-pharyngeal-rectal swab performed</td>
<td></td>
<td></td>
<td>MDR <em>A. baumannii</em> had not been detected at the time of faecal carriage in 21 of 33 (63.6%) of the control group and 11 of 21 (52.3%) of the SDD group. In the SDD group, all <em>A. baumannii</em> strains were tobramycin resistant and susceptible to colistin at the beginning of the study. No resistance to colistin developed during the study</td>
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<tr>
<td></td>
<td>Exclusion criteria: not reported</td>
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<tr>
<td>Brun-Buisson 1989</td>
<td>To study the efficacy of intestinal decontamination by oral non-absorbable antibiotic agents to control a nosocomial outbreak of intestinal colonization and infection with MDR <em>Enterobacteriaceae</em>, and to examine its effects on endemic nosocomial infection rates.</td>
<td><em>Bacteria</em>: <em>Enterobacter</em> spp., <em>P. aeruginosa</em></td>
<td>Intervention  SDD: a combination of polymyxin E (colistin), 50 mg; neomycin, 1 g; and nalidixic acid (quinolone), 1 g administered in liquid form x 4/day either orally or through a nasogastric tube, starting within 24 h of admission and continuing until discharge from the unit. N=36</td>
<td>Mortality</td>
<td>Quasi-randomized</td>
</tr>
<tr>
<td>Quasi-randomized Setting</td>
<td></td>
<td><em>Resistant to</em>: aminoglycosides (amikacin), third-generation cephalosporins</td>
<td></td>
<td>All-cause mortality and mortality from nosocomial infections did not differ significantly between patients receiving SDD or no prophylaxis</td>
<td>Low methodological quality (0)</td>
</tr>
<tr>
<td>Tertiary (one ICU) France January 1987-May 1987</td>
<td>Participants  N=86 Adults 19–45 years, middle aged 46–64 years, aged 65–79 years Male: not reported, female: not reported</td>
<td><em>Mechanism of resistance</em>: ESBL</td>
<td>Control group No prophylaxis. N=50</td>
<td>Clinical success/improvement</td>
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<td></td>
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<td></td>
<td></td>
<td>There was no significant difference between patients receiving SDD or no prophylaxis in:</td>
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<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>– the incidence of any nosocomial infection</td>
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<td></td>
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<td></td>
<td></td>
<td></td>
<td>– the infections caused by Gram-negative bacteria</td>
</tr>
<tr>
<td>Study details</td>
<td>Objective and participants</td>
<td>MDR Gram-negative bacteria</td>
<td>Intervention, control and follow-up</td>
<td>Results</td>
<td>Quality assessment</td>
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<tr>
<td>Inclusion criteria: 1. Consecutive patients with unit stay exceeding two days 2. Severity score at admission &gt;2</td>
<td></td>
<td></td>
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<td>the number of nosocomial infections that needed antibiotic treatment There was no significant difference in the number of patients staying on ICU longer than seven or 15 days</td>
<td></td>
</tr>
<tr>
<td>Exclusion criteria: 1. Severe neutropenia routinely receiving oral antibiotic prophylaxis</td>
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</tbody>
</table>
| **Saidel-Odes 2012** | **RCT** | **Setting** Tertiary (one internal medicine ward) Israel | **Participants** N=40 Middle aged 46–64 years, aged 65–79 years, elderly 80+ years Male: 26, female: 14 | **Intervention** SDD: topical application in the oropharynx of colistin sulfomethate sodium 100,000 U per g and gentamicin sulfate 1.6 mg per g incorporated into the gel. Dose of 0.5 g x 4/day for seven days. Plus an oral solution of 80 mg of gentamicin and 1x10 U of polymyxin E (colistine), given orally or through a nasogastric | **Results** Mortality The rate of mortality did not differ significantly between the SDD group and the placebo group. The causes of mortality were not reported. No adverse events were reported | **Quality assessment** RCT High methodological quality (+++) Small sample size
<p>| <strong>Bacteria</strong>: <em>K. pneumoniae</em> | <strong>Resistant to</strong>: carbapenems | <strong>Mechanism of resistance</strong>: not reported | <strong>Antibiotic susceptibility</strong> CRKP isolates from patients in the SDD arm remained susceptible to gentamicin and polymyxin E | | |</p>
<table>
<thead>
<tr>
<th>Study details</th>
<th>Objective and participants</th>
<th>MDR Gram-negative bacteria</th>
<th>Intervention, control and follow-up</th>
</tr>
</thead>
<tbody>
<tr>
<td>November 2008–June 2010</td>
<td>1. Hospitalized patients with CRKP colonization with or without infection</td>
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<td>tube X 4/day for seven days. N=20</td>
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<tr>
<td></td>
<td>2. &gt;18 years of age</td>
<td></td>
<td><strong>Control group</strong></td>
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<td></td>
<td>3. Available for a follow-up period (while hospitalized or as outpatients) of at least seven weeks</td>
<td></td>
<td>Placebo: topical application in the oropharynx of the placebo gel, which was compounded from carboxymethyl cellulose. Dose of 0.5 g x 4/day for seven days. Plus two oral solutions, one containing sodium chloride 0.45% and the other containing pulverized sacarin, given orally or through a nasogastric tube X 4/day for seven days. N=20</td>
</tr>
<tr>
<td></td>
<td>Exclusion criteria: &lt;18 years of age, pregnancy, lactation, a known allergy to one of the study drugs, renal failure with creatinine clearance less than 50 mL/min, treatment with intravenous gentamicin or intravenous polymyxin E at the time of randomization</td>
<td></td>
<td><strong>Bacterial colonization</strong></td>
</tr>
<tr>
<td></td>
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<td></td>
<td>At the end of treatment, the number of participants in the SDD group that had a throat culture that was CRKP positive reduced from 30% to 0%, whereas in the placebo group, this reduced from 35% to 30% (P&lt;0.0001)</td>
</tr>
</tbody>
</table>

**Length of follow-up:** six weeks

**Quality assessment** throughout the study (MIC ≤2 mg/mL and ≤0.094 mg/mL, respectively)

*A. baumannii, Acinetobacter baumannii; K. pneumoniae, Klebsiella pneumoniae; MDR, multi-drug resistant; SDD, selective digestive decontamination; RR, risk ratio, CI, confidence interval; CRKP, carbapenem-resistant K. pneumonia; MIC, minimum inhibitory concentration; RCT, randomized controlled trial; ICU, intensive care unit.*
### 4.3.4. Systematic reviews

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<tr>
<th>Study details</th>
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<th>Results</th>
<th>Quality assessment</th>
</tr>
</thead>
</table>
| Falagas 2009* | To assess the clinical and microbiological effectiveness of fosfomycin in the treatment of MDR, XDR or PDR non-fermenting Gram-negative bacterial infections | **Bacteria:** *Pseudomonas* spp., *Acinetobacter* spp., *Stenotrophomonas* spp. and *Burkholderia* spp. | Intervention **Fosfomycin**  
Control group **Combination of fosfomycin with other antimicrobial agents** | **Microbiological:** a total of 1859 MDR non-fermenting Gram-negative isolates. Susceptibility rate to fosfomycin of MDR *P. aeruginosa* isolates was ≥90% and 50–90% in 7/19 and 4/19 relevant studies, respectively. 30.2% isolates of MDR *P. aeruginosa*, 3.5% MDR *A. baumannii* isolates were found to be susceptible to fosfomycin  
**Clinical:** 91% of the patients clinically improved (treatment of infections caused by MDR *P. aeruginosa*) | Low methodological quality (0) |
<p>| Setting International | | | | | |
| Search up to January 2009 | | | | | |
| <strong>Participants</strong> | | | | | |
| N=33 | | | | | |
| Studies: 23 microbiological, one animal and three cohort studies and three case reports | | | | | |
| Inclusion criteria: microbiological, animal experimental or clinical data on the effect of fosfomycin against MDR non-fermenting Gram-negative pathogens such as <em>Pseudomonas</em> spp., <em>Acinetobacter</em> spp., <em>Stenotrophomonas</em> spp. and <em>Burkholderia</em> spp. MDR, XDR or PDR non-fermenting Gram-negative bacilli or to Gram-negative bacilli with resistance to two or more classes of potentially effective antimicrobial agents | | | | | |
| Exclusion criteria: studies written in languages other than English, French, German, Italian or Spanish. | | | | | |</p>
<table>
<thead>
<tr>
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<tbody>
<tr>
<td></td>
<td>Studies representing abstracts in scientific conferences</td>
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<tr>
<td>Falagas 2009²</td>
<td>To evaluate the available clinical evidence regarding the effectiveness and safety of systemic colistin in children without cystic fibrosis</td>
<td><strong>Bacteria</strong>: <em>P. aeruginosa</em>, <em>A. baumannii</em>, <em>K. aerogenes</em>, <em>H. influenza</em>, <em>P. pyocyanin</em>, <em>P. aeruginosa</em>, <em>K. pneumoniae</em> and <em>A. aerogenes</em></td>
<td><strong>Intervention</strong> Colistin for the treatment of infections (<em>N</em>=326)</td>
<td><strong>Case series treatment</strong>: 271 evaluable subjects Cure: 235/271 Improvement: 10/271 Deterioration: 6/271 Death: 20/271 Adverse effects (included in safety assessment <em>N</em>=311) 1. Nephrotoxicity: 33/311 had cylindruria or haematuria, 8/311 had a blood urea nitrogen elevation of &gt;10% (in one child owing to an overdosage of colistin), 5/311 had renal tubular cells in the urine, 3/311 had proteinuria and 2/311 had a significant increase in serum creatinine levels during intravenous colistin treatment. Data regarding adverse events not provided for two children 2. Neurotoxicity: 0/311 3. Other: 8/311</td>
<td>Acceptable methodological quality (+)</td>
</tr>
<tr>
<td>Setting</td>
<td>Not reported</td>
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<tr>
<td>Participants</td>
<td><em>N</em>=370</td>
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<td>Studies: 10 case series and 15 case reports</td>
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<td>Inclusion criteria: studies with data regarding the use of intravenous, intrathecal, intramuscular or intraventricular colistin in paediatric patients for the treatment of infections caused by colistin-susceptible pathogens or for prophylaxis. All or the majority of patients involved in each individual study should not have cystic fibrosis</td>
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<tr>
<td>Exclusion criteria: studies that focused on colistin use in paediatric patients with cystic fibrosis, or reporting the use of oral colistin or the use of colistin for topical treatment in paediatric patients. Abstracts in scientific conferences or studies published in languages other than English, Spanish, French, German, Italian or Greek</td>
<td><strong>Control group</strong> Colistin for surgical prophylaxis or prophylaxis of infections in burns patients (<em>N</em>=44)</td>
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</table>

This review was included because it is on the topic; however, the conclusions reached are not supported by the study design.
<table>
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</thead>
<tbody>
<tr>
<td>Falagas 2010³</td>
<td>To the evidence on fosfomycin as a treatment option for infections caused by members of the family Enterobacteriaceae with advanced resistance to antimicrobial drugs, including producers of ESBL</td>
<td>Bacteria: <strong>Microbiological studies</strong> <em>K. pneumoniae</em> isolates, <em>E. coli</em> <strong>Clinical studies</strong> <em>E. coli, S. typhimurium, S. typhi</em></td>
<td>Intervention <strong>Amoxicillin-clavulanate potassium</strong> <strong>Control group</strong> Fosfomycin–trometamol in two of the <em>E. coli</em> studies</td>
<td>Microbiological success 11 of the 17 studies reported that at least 90% of the isolates were susceptible to fosfomycin Clinical efficacy Measured in four studies. Two studies oral treatment for lower UTI with ESBL-producing <em>E. coli</em> (one prospective and one retrospective) resulted in the treatment group with clinical cure in 75 of the 80 (93.8%) patients included in these studies. Two case reports of infection due to MDR <em>Salmonella</em> spp. Reported treatment was effective with fosfomycin</td>
<td>Low methodological quality (0) This review was included because it is on the topic; however, the conclusions reached are not supported by the study design</td>
</tr>
</tbody>
</table>

Setting International Searches up to January 2009

Participants N=119 Studies: 17 in-vitro microbiological studies, two prospective studies, one retrospective study and two case reports

Inclusion criteria: studies on Enterobacteriaceae isolates with an advanced drug resistance (MDR, carbapenem resistance, or production of ESBLs, AmpC β-lactamases, serine carbapenemases or metallo-β-lactamases) profile and their susceptibility to fosfomycin, and the clinical effectiveness of treatment with fosfomycin for infections with these pathogens
<table>
<thead>
<tr>
<th>Study details</th>
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<th>Results</th>
<th>Quality assessment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Falagas 2012*</td>
<td>Exclusion criteria: abstracts in scientific conferences or studies published in languages other than English, Spanish, French, German, Italian or Greek</td>
<td>To identify and evaluate the available data regarding the susceptibility of recent Gram-negative bacteria to isepamicin, including that of MDR strains of bacteria</td>
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<tr>
<td>Setting</td>
<td>Setting</td>
<td>Intervention</td>
<td>Isepmicin</td>
<td>Microbiological: isepamicin was more effective in four studies than amikacin, six studies reported as effective, one study both groups ineffective. In studies including MDR bacteria, 2/4 reported more effective than amikacin; 1/4 as effective as amikacin; 1/4 both isepamicin and amikacin ineffective</td>
<td>Low methodological quality (0)</td>
</tr>
<tr>
<td>Not reported</td>
<td></td>
<td>Control group</td>
<td>Two clinical studies – amikacin, one clinical study – isepamicin + levofloxacin for prophylaxis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Searches from 2000 to 2010</td>
<td>Participants</td>
<td>N=512</td>
<td>100% clinical and bacteriological response for both the isepamicin and the amikacin arms. Definition of clinical response not stated (e.g. cure, improvement)</td>
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<tr>
<td></td>
<td></td>
<td>Studies=11 microbiological, one RCT, one prospective study, one retrospective study</td>
<td>2. Prophylactic study: acute bacterial prostatitis 1.3%</td>
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<td></td>
<td>Inclusion criteria: either a microbiological (in-vitro) study that evaluated the susceptibility of Gram-negative bacterial isolates (including MDR ones) to isepamicin or a clinical study that evaluated the use of isepamicin, given for the treatment of infections by the aforementioned pathogens or for prophylaxis for this type of infection. In addition, studies deemed relevant should have been published between 2000 and 2010</td>
<td>Inclusion criteria: either a microbiological (in-vitro) study that evaluated the susceptibility of Gram-negative bacterial isolates (including MDR ones) to isepamicin or a clinical study that evaluated the use of isepamicin, given for the treatment of infections by the aforementioned pathogens or for prophylaxis for this type of infection. In addition, studies deemed relevant should have been published between 2000 and 2010</td>
<td>See Table II in the paper for details of studies</td>
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<tr>
<td></td>
<td>Exclusion criteria: studies that examined a sample of fewer than 10</td>
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<tr>
<td></td>
<td>Bacteria: Clinical studies S. epidermidis, E. coli, S. pneumoniae, P. aeruginosa</td>
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<tr>
<td>Study details</td>
<td>Objective and participants</td>
<td>MDR Gram-negative bacteria</td>
<td>Intervention, control and follow-up</td>
<td>Results</td>
<td>Quality assessment</td>
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<tr>
<td>Kaki 2011</td>
<td>isolates or patients, studies referring to synergistic or pharmacodynamic/pharmacokinetic parameters of isepamicin, studies that provided data regarding the susceptibility of isepamicin to micro-organisms other than Gram-negative bacteria or the susceptibility of other aminoglycosides only to Gram-negative bacteria.</td>
<td>Bacteria: P. aeruginosa, A. baumannii, E. coli, Klebsiella spp., ESBL</td>
<td>Overall stewardship intervention: 1. Reductions in antimicrobial utilization (11–38% defined daily dose/1000 patient-days) 2. Lower total antimicrobial costs (US$ 5–10/patient-day) 3. Shorter average duration of antibiotic therapy 4. Less inappropriate use 5. Fewer antibiotic adverse events.</td>
<td>Antibiotic stewardship was not associated with increases in nosocomial infection rates, length of stay or mortality</td>
<td>High methodological quality (+++)</td>
</tr>
</tbody>
</table>

**Setting** International
**Search** January 1996 to December 2010
**Participants** N=not available/not reported for all included studies
Studies: three RCTs, three ITSs, and 18 uncontrolled before–after studies
Inclusion criteria: application of any intervention; to improve antimicrobial utilization; and within an intensive care setting
Exclusion criteria: if no intervention was applied, non-human or non-patient based, non-hospital based, or they did not involve intensive care

**Intervention**
1. Antibiotic restriction/pre-approval
2. Computer-assisted decision support
3. Infectious diseases consultant
4. Re-assessment on pre-specified date
5. Antibiotic de-escalation protocols
6. Antibiotic prophylaxis guideline
7. Antibiotic treatment guideline

**Control group** Not reported, presumably no stewardship

See Table I in the paper for details of studies.
<table>
<thead>
<tr>
<th>Study details</th>
<th>Objective and participants</th>
<th>MDR Gram-negative bacteria</th>
<th>Intervention, control and follow-up</th>
<th>Results</th>
<th>Quality assessment</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Siempos 2007</strong>&lt;sup&gt;a&lt;/sup&gt;</td>
<td>To clarify whether carbapenems are more effective or safer than other broad-spectrum antibiotics for the empirical treatment of patients with HAP</td>
<td><strong>Bacteria:</strong> <em>P. aeruginosa</em> See Table I in the paper for details of studies</td>
<td><strong>Intervention</strong> Carbapenems: 1. Imipenem/ cilastatin (eight studies) 2. Meropenem (four studies) <strong>Control group</strong> Imipenem/ cilastatin compared with: 1. Fluoroquinolones: levofloxacin, ciprofloxacin (three studies) 2. Other beta-lactams: piperacillin/tazobactam, aztreonam, cefepime, ceftazidime (five studies) Meropenem compared with: combination of a cephalosporin (ceftazidime, cefuroxime) with an aminoglycoside (amikacin, gentamicin, tobramycin)</td>
<td>1. All-cause mortality: lower mortality in the carbapenems group (OR 0.72, 95% CI 0.55–0.95) 2. Treatment success (clinical): no difference between groups (OR 1.08, 95% CI 0.91–1.29) 3. Treatment success (microbiological): no difference between groups (OR 1.04, 95% CI 0.72–1.50) 4. Adverse effects: no difference (0.81, 0.46–1.43) <em>P. aeruginosa</em> pneumonia subgroup: lower treatment success (OR 0.42, 95% CI 0.22–0.82) and lower eradication of <em>Pseudomonas</em> spp. strains (OR 0.50, 95% CI 0.24–0.89) in the carbapenems group. Late onset of HAP subgroup: no difference between groups (OR 1.34, 95% CI 0.91–1.97)</td>
<td>High methodological quality (+++)</td>
</tr>
</tbody>
</table>

Not reported

**Setting**

Search

January 1950 to March 2006

Participants

*N* = 2731

Studies: 12 RCTs

Inclusion criteria: randomized controlled clinical trial; studied the role of carbapenems in comparison with other broad-spectrum antibiotics or a combination of antibiotics for the empirical treatment of patients with HAP; assessed the effectiveness, toxicity and mortality of both therapeutic regimens. Included both patients with HAP and patients with community-acquired pneumonia; however, only data regarding patients with HAP were extracted. Trials with both blind and unblind design were included, and only RCTs written in English, French and German

Exclusion criteria: RCTs conducted primarily in neutropenic patients with solid organ tumours or
<table>
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<tr>
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<th>Quality assessment</th>
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<td></td>
<td>haematological malignancies and trials that included fewer than 10 patients with pneumonia who received a carbapenem. Experimental trials and trials focusing on pharmacokinetic and pharmacodynamics parameters. Finally, RCTs comparing the effectiveness and safety of two different carbapenems</td>
<td>P. aeruginosa, Pseudomonas aeruginosa; A. baumannii, Acinetobacter baumannii; K. aerogenes, Klebsiella aerogenes; H. influenza, Haemophilus influenza; P. pyocyain, Pseudomonas pyocyain; K. pneumoniae, Klebsiella pneumoniae; A. aerogenes, Aerobacter aerogenes; E. coli; Escherichia coli; S. typhimurium, Salmonella typhimurium; S. typhi, Salmonella typhi; S. pneumoniae, Streptococcus pneumoniae; S. epidermidis, Staphylococcus epidermidis; MDR, multi-drug resistant; XDR, extensively drug resistant; PDR, pan-drug resistant; RCT, randomized controlled trial; ESBL, extended-spectrum beta-lactamase; HAP, hospital-acquired pneumonia; OR, odds ratio; CI, confidence interval.</td>
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### 4.3.5. Treatment

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<tr>
<th>Study details</th>
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<th>Quality assessment</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Betrosian 2007</strong>&lt;br&gt;RCT</td>
<td>To evaluate the clinical efficacy and safety of high-dose regimen ampicillin/sulbactam for the treatment of VAP from MDR <em>A. baumannii</em>&lt;br&gt;&lt;br&gt;<strong>Participants</strong>&lt;br&gt;N=27&lt;br&gt;Age: not reported&lt;br&gt;Male: 15, female: N=12&lt;br&gt;&lt;br&gt;Inclusion criteria: all patients mechanically ventilated for more than 72 h with positive tracheal aspirates for <em>A. baumannii</em>&lt;br&gt;&lt;br&gt;Exclusion criteria: episodes of VAP in which <em>A. baumannii</em> was isolated in conjunction with another microorganism</td>
<td>Bacteria: <em>A. baumannii</em>&lt;br&gt;Resistant to: ampicillin/sulbactam and susceptible exclusively to colistin (polymyxin E)&lt;br&gt;&lt;br&gt;<strong>Mechanism of resistance:</strong> not reported</td>
<td>Intervention&lt;br&gt;Ampicillin/sulbactam at a rate 2:1 every 8 h, 24 g/12 g daily for seven to 10 days. N=13&lt;br&gt;&lt;br&gt;<strong>Control group</strong>&lt;br&gt;Ampicillin/sulbactam at a rate 2:1 every 8 h, 18 g/9 g daily for seven to 10 days. N=14</td>
<td>Mortality&lt;br&gt;14-day VAP mortality and 30-day all-cause mortality were not significantly different between treatment groups</td>
<td>RCT&lt;br&gt;Low methodological quality (0)</td>
</tr>
<tr>
<td><strong>Betrosian 2008</strong>&lt;br&gt;RCT</td>
<td>To compare the clinical efficacy and safety of high-dose ampicillin/sulbactam vs colistin as monotherapy for the treatment of <em>Acinetobacter</em> spp. VAP</td>
<td>Bacteria: <em>A. baumannii</em>&lt;br&gt;Resistant to: Aminoglycosides, carbapenems,</td>
<td>Intervention&lt;br&gt;Colistin, intravenous 3 MIU every 8 h for eight to 10 days. N=15</td>
<td>Mortality&lt;br&gt;14-day VAP mortality and 28-day all-cause mortality were not significantly different between treatment groups</td>
<td>RCT&lt;br&gt;Low methodological quality (0)</td>
</tr>
<tr>
<td>Study details</td>
<td>Objective and participants</td>
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<td>Intervention, control and follow-up</td>
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</tr>
<tr>
<td>Setting</td>
<td>Participants</td>
<td>cephalosporins, fluoroquinolones</td>
<td>Control group</td>
<td>Clinical success/improvement</td>
<td>Small sample size</td>
</tr>
<tr>
<td>Tertiary (2 ICUs)</td>
<td>N=28</td>
<td></td>
<td>Ampicillin/sulbactam, 9 g (at a rate 2:1) every 8 h for eight to 10 days, administered as follows: three vials (20 mL each) containing 3.0 g of ampicillin/sulbactam diluted in 200 mL of 5% dextrose provided within 1-h duration infusion. N=13</td>
<td>The number of patients with clinical success and clinical failure was not significantly different between treatment groups</td>
<td>++</td>
</tr>
<tr>
<td>Greece</td>
<td>Middle aged 46–64 years, aged 65–79 years</td>
<td>Mechanism of resistance: not reported</td>
<td>Bacterial colonization</td>
<td>The two treatment groups showed no difference in the eradication of <em>A. baumannii</em> isolates (bacteriological success) or bacteriological failure (persistence of <em>A. baumannii</em> isolates (&gt;104 CFU/mL)</td>
<td>+</td>
</tr>
<tr>
<td>Dates not reported</td>
<td>Male: 14, female: 14</td>
<td></td>
<td>Adverse events</td>
<td>There was no difference in the adverse effects experienced by participants</td>
<td>+</td>
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<tr>
<td></td>
<td>Inclusion criteria: ventilated patients for &gt;72 h who developed MDR <em>A. baumannii</em> VAP</td>
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<td></td>
<td>Exclusion criteria: cases of VAP with mixed isolated micro-organisms, combination antibiotic therapy, allergy to beta-lactamase or penicillin, or previous enrolment in similar studies</td>
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<tr>
<td>Setting</td>
<td>Participants</td>
<td>Bacteria: <em>E. coli</em>, <em>Klebsiella</em> spp., <em>Enterobacter</em> spp., <em>P. aeruginosa</em>, <em>Acinetobacter</em> spp., <em>Proteus</em> spp., <em>Serratia</em> spp., <em>C. freundii</em>, <em>M. morgagnii</em></td>
<td>Intervention</td>
<td>Mortality</td>
<td>RCT</td>
</tr>
<tr>
<td>RCT</td>
<td>N=401</td>
<td>Resistant to: ticarcillin, methicillin</td>
<td>Antibiotics for eight days: specific antibiotics, doses and schedules are not reported. Antibiotics were selected by the treating physicians. As per protocol, the initial regimen should have preferably combined at least an aminoglycoside, or a fluoroquinolone and a broad-spectrum beta-lactam antimicrobial agent. N=197</td>
<td>28-day and 60-day all-cause mortality and in-hospital mortality did not significantly differ between the eight- and 15-day regimes</td>
<td>High methodological quality (++)</td>
</tr>
<tr>
<td>Setting</td>
<td>To compare the efficacy of eight days vs 15 days of antibiotic treatment of patients with microbiologically proven VAP</td>
<td>Mechanism of resistance: ESBL</td>
<td>Control group</td>
<td>Clinical success/improvement</td>
<td></td>
</tr>
<tr>
<td>Chastre 2003</td>
<td>Participants</td>
<td></td>
<td></td>
<td>Risk differences (90% CIs) to develop an unfavourable outcome (defined as death, pulmonary infection recurrence, or prescription of a new antibiotic for any reason provided for ≥48 h) were not significantly different between the eight- and 15-day regimes</td>
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<tr>
<td>RCT</td>
<td>N=401</td>
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<tr>
<td>Setting</td>
<td>Middle aged 46–64 years, aged 65–79 years</td>
<td></td>
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<tr>
<td>Tertiary (51 ICUs)</td>
<td>Male: 141, female: 46</td>
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<tr>
<td>France</td>
<td>Inclusion criteria: 1. ≥18 years of age</td>
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<tr>
<td>Dates not reported</td>
<td>2. Clinical suspicion of VAP</td>
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<tr>
<td>May 1999-June 2002</td>
<td>3. Positive quantitative cultures of distal pulmonary secretion samples</td>
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</tbody>
</table>
### Study details

1. **Objective and participants**
   - **MDR Gram-negative bacteria**
   - **Intervention, control and follow-up**
   - **Results**
   - **Quality assessment**

| 4. Instigation within the 24 h following of appropriate empirical antibiotic therapy directed against the micro-organism/s responsible for the infection | Antibiotics for 15 days: specific antibiotics, doses and schedules are not reported. Antibiotics were selected by the treating physicians. As per protocol, the initial regimen should have preferably combined at least an aminoglycoside or a fluoroquinolone and a broad-spectrum beta-lactam antimicrobial agent. N=204 | Non-fermenting Gram-negative bacteria (RR 8.6, 90% CI -5.9 to 23.1) | | |
| Exclusion criteria: | | The rate of and time to (Kaplan-Meier method, log-rank test) pulmonary infection considered to be recurrence, relapses or superinfection was not significantly different between treatment regimes. | | |
| 1. Pregnant | Length of follow-up: three months | Antibiotic use | | |
| 2. Enrolled in another trial | | The number of antibiotic-free days was significantly less for all patients on the eight-day regime, but not for those patients with non-fermenting Gram-negative bacteria. | | |
| 3. Little chance of survival | | No difference was found in the number of patients continuing to receive antibiotics after the end of the trial treatment regimen, or in the number of patients who received an additional course of antibiotics | | |
| 4. Neutropenia | | Antibiotic resistance | For patients who developed recurrent pulmonary infections, those who had received the eight-day treatment of antibiotics had significantly less emergence of MDR pathogens compared with those who had received the 15-day treatment (42.1% vs 62.3% of recurrent infections, respectively; P=0.04) | | |
| 5. Concomitant acquired immunodeficiency syndrome | | | | |
| 6. Immunosuppressants or long-term corticosteroid therapy | | | | |
| 7. Concomitant extrapulmonary infection that required prolonged antimicrobial treatment | | | | |
| 8. Attending physical declined full-life support. | | | | |
| 9. Early-onset pneumonia (within the first five days of mechanical ventilation) | | | | |
| and no antimicrobial therapy during the 15 days preceding infection. | | | | |

<p>| N=204 | | | | |</p>
<table>
<thead>
<tr>
<th>Study details</th>
<th>Objective and participants</th>
<th>MDR Gram-negative bacteria</th>
<th>Intervention, control and follow-up</th>
<th>Results</th>
<th>Quality assessment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cox 1987</td>
<td>To compare the efficacy of norfloxacin vs standard parenteral treatment of non-bacteraemic, hospital-acquired UTI</td>
<td>Bacteria: <em>E. coli</em>, Klebsiella spp., Enterobacter spp., <em>P. aeruginosa</em>, Serratia spp., <em>C. freundii</em>, M. <em>morgagnii</em></td>
<td>Intervention&lt;br&gt;Norfloxacin 400 mg x2/day, minimum treatment seven days. N=52 (46 evaluable patients)&lt;br&gt;&lt;br&gt;Control group&lt;br&gt;Aminoglycosides alone; aminoglycosides and mezlocillin/ticarcillin; aminoglycosides and cephalosporin; aminoglycosides and vancomycin, cephalosporin, cefotaxime alone, administered in accordance with the manufacturers' guidelines. N=52 (48 evaluable patients)</td>
<td>Clinical success/improvement&lt;br&gt;No significant differences were found between norfloxacin and standard parenteral antibiotic treatment in the rate of participants that were clinically cured, showed clinical improvement or had treatment failure</td>
<td>RCT Acceptable methodological quality (+)</td>
</tr>
<tr>
<td>RCT</td>
<td>Setting&lt;br&gt;Secondary (two hospitals) USA</td>
<td>Participants&lt;br&gt;N=104&lt;br&gt;Age: not reported&lt;br&gt;Male: not reported, female: not reported</td>
<td>Resistant to: not reported&lt;br&gt;Mechanism of resistance: not reported</td>
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<td></td>
<td>Setting&lt;br&gt;March 1985–December 1985</td>
<td>Inclusion criteria:&lt;br&gt;1. Hospitalized patients&lt;br&gt;2. &gt;18 years of age&lt;br&gt;3. Documented UTI caused by an organism known or presumed susceptible to norfloxacin</td>
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<td>Exclusion criteria:&lt;br&gt;1. &lt;18 years of age&lt;br&gt;2. Pregnant or not practising an effective means of birth control&lt;br&gt;3. A history of allergic diathesis or an allergy to nalidixic acid, oxolinic acid or norfloxacin&lt;br&gt;4. Functional renal abnormalities or unstable deteriorating renal function&lt;br&gt;5. Comatose or high probability of imminent death&lt;br&gt;6. Serious concurrent infection&lt;br&gt;7. Treated or recently completed treatment with antibiotics&lt;br&gt;8. History or visual disturbances, a psychiatric disorder or central nervous system disease</td>
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<tr>
<td>Study details</td>
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<td>Quality assessment</td>
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<tr>
<td>Giamarellou 1990 RCT</td>
<td>To evaluate the efficacy of monotherapy with pefloxacin in secondary ICU pulmonary infections in comparison with imipenem</td>
<td>Bacteria: <em>E. coli</em>, <em>K. pneumoniae</em>, <em>Enterobacter</em> spp. (various <em>Enterobacteriaceae</em>), <em>P. aeruginosa</em>, <em>A. anitratus</em>, <em>P. mira</em>, <em>S. marcescens</em></td>
<td>Intervention: Pefloxacin intravenously 400 mg, every 8 h for 11.5 (SD 5.8) days. N=35</td>
<td>Mortality: There were three deaths related to sepsis in the imipenem group and one in the pefloxacin group (although the sepsis was not related to the bronchopneumonia, but to an underlying abdominal infection). All-cause mortality was not reported.</td>
<td>RCT Acceptable methodological quality (+)</td>
</tr>
<tr>
<td>Setting Tertiary (one ICU) Greece Dates not reported</td>
<td>Participants: N=71 Adults 19–45 years, middle aged 46–64 years, aged 65–79 years, elderly 80+ years Male: 42, female: 29</td>
<td>Resistant to: aminoglycosides (gentamicine, tobramycin, netilmicin, amikacin), aztreonam, carbapenems (imipenem), cephalosporins (ceftaxime, ceftriaxime, ceftizidime), fluoroquinolones (ciprofloxacin)</td>
<td>Control group: Imipenem intravenously 1 g every 8 h for 12.9 (SD 6.2) days. N=36</td>
<td>Clinical success/improvement: No differences were found in the number of patients cured, the number with superinfection that was cured, the number showing improvement and the number experiencing treatment failure. Bacterial eradication rates were significantly lower in the imipenem group (55.3% vs 82.9%, respectively (<em>P</em>&lt;0.001))</td>
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<td>Inclusion criteria: adult patients presenting serious bacterial infections of the respiratory tract</td>
<td>Mechanism of resistance: not reported</td>
<td>Length of follow-up: duration of treatment</td>
<td>Antibiotic resistance: Resistance development among persisting strains was also significantly different (data not reported, <em>P</em>&lt;0.05)</td>
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<tr>
<td>Huttner 2013</td>
<td>To investigate if intestinal carriage of ESBL-E can be eradicated</td>
<td>Bacteria: <em>Enterobacter</em> spp. (ESBL-E)</td>
<td>Intervention: Colistin sulfate 50 mg (equivalent to 42 mg colistin)</td>
<td>Clinical success/improvement: The rate of eradication of ESBL-E was significantly different between</td>
<td>RCT</td>
</tr>
<tr>
<td>Study details</td>
<td>Objective and participants</td>
<td>MDR Gram-negative bacteria</td>
<td>Intervention, control and follow-up</td>
<td>Results</td>
<td>Quality assessment</td>
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<tr>
<td>RCT Setting Secondary (all inpatient wards of a single hospital) Switzerland June 2009–June 2012</td>
<td><strong>Participants</strong>&lt;br&gt;N=58  Adolescents 13–18 years, adults 19–45 years, middle aged 46–64 years, aged 65–79 years, elderly 80+ years Male: 34, female: 24  Inclusion criteria: aged ≥18 years; ESLB-E-positive rectal swab  Exclusion criteria: patients with active ESLB infection, patients treated with antibiotics active against ESLB-E, pregnancy/breastfeeding, contraindication to the use of study drugs, previous study enrolment and resistance of the colonizing ESLB-E strain to colistin (defined as MIC &gt;2 mg/L)</td>
<td><strong>Resistant to:</strong>&lt;br&gt;cefotaxime, cefotaxime/clavulanic acid, ceftazidime, ceftazidime/clavulanic acid, cefepime, cefepime/clavulanic acid  <strong>Mechanism of resistance:</strong> ESBL</td>
<td>base or 1.26 million units 4x/day) and neomycin sulfate (250 mg equivalent to 178 mg neomycin base 4x/day) for 10 days. In the presence of ESBL-E bacteriuria, the patients were also treated with nitrofurantoin (100 mg 3x/day) for five days. N=27  <strong>Control group</strong> Placebo. N=27  <strong>Length of follow-up:</strong> 28 (SD seven) days</td>
<td>treatment regimes during treatment (day 6: RR 0.40; 95% CI 0.23–0.70) or in the first day after treatment (RR 0.42; 95% CI 0.23–0.76), but did not differ in the end of follow-up  <strong>Treatment adherence</strong> There was no significant difference between groups in the number of patients that adhered to treatment, measured by counting the number of pills on the boxes of study medication  <strong>Adverse events</strong> No statistically significant difference was found between the treatment groups in the number of patients with at least one episode of liquid stool</td>
<td>High methodological quality (+++)</td>
</tr>
<tr>
<td>Moskowitz 2011 RCT Setting Secondary (seven cystic fibrosis centres) USA February 2007–</td>
<td><strong>Participants</strong>&lt;br&gt;N=39  Adolescents 13–18 years, adults 19–45 years Male: 25, female: 14</td>
<td><strong>Bacteria:</strong> <em>P. aeruginosa</em>  <strong>Resistant to:</strong> aminoglycosides, fluoroquinolones  <strong>Mechanism of resistance:</strong> not reported</td>
<td><strong>Intervention</strong> Biofilm testing: biofilm regimens of two antibiotics were selected centrally using a published algorithm, which calculated for each bacterial morphotype the biofilm minimum inhibitory quotient of each drug, defined as achievable serum concentration divided by biofilm MIC. N=20  <strong>Control group</strong> Conventional testing: conventional regimens of two</td>
<td><strong>Antibiotic susceptibility</strong> Participants were assigned to 12 different regimens. The most common regimens included meropenem (52%) and ciprofloxacin (49%). Azithromycin-containing regimens were used for only two participants (5%), both in the biofilm group. No participant received ceftazidime and tobramycin, a combination commonly used in cystic fibrosis clinical practice</td>
<td>Acceptable methodological quality (+) Small sample size</td>
</tr>
<tr>
<td>Study details</td>
<td>Objective and participants</td>
<td>MDR Gram-negative bacteria</td>
<td>Intervention, control and follow-up</td>
<td>Results</td>
<td>Quality assessment</td>
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<tr>
<td>October 2007</td>
<td>Inclusion criteria: diagnosis of cystic fibrosis, history of persistent <em>P. aeruginosa</em> airway infection, clinical stability at the time of screening, ≥14 years with at least one prior course of intravenous antibiotics Exclusion criteria: sputum culture negative for <em>P. aeruginosa</em>, sputum culture positive for <em>B. cepacia</em> complex species, hospitalization or treatment for an acute pulmonary exacerbation, treatment with oral or inhaled antipseudomonal antibiotics, or azithromycin or other macrolides, within 14 days prior to screening</td>
<td>Antibiotics were selected centrally using a published algorithm, which calculated for each bacterial morphotype the conventional minimum inhibitory quotient of each drug defined as achievable serum concentration divided by conventional MIC. N=19</td>
<td>Of the agents tested, meropenem was most active against biofilm-grown bacteria, but antibiotic regimens based on biofilm testing did not differ significantly from regimens based on conventional testing in terms of microbiological and clinical responses</td>
<td>RCT Acceptable methodological quality (+)</td>
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<tr>
<td>Rattanaumpawan 2010</td>
<td>To determine whether nebulized CMS as adjunctive therapy of Gram-negative VAP was safe and beneficial</td>
<td><strong>Bacteria</strong>: <em>E. coli</em> (ESBL +ve) and <em>E. coli</em> (ESBL -ve), <em>K. pneumoniae</em> (ESBL +ve) and <em>K. pneumoniae</em> (ESBL -ve), <em>E. cloacae</em>, <em>P. aeruginosa</em>, <em>A. baumannii</em></td>
<td><strong>Intervention</strong> Systemic antibiotic and nebulized CMS (parenteral) equivalent to 75 mg of colistin base reconstituted in 4 mL of NSS every 12 h via a nebulizer for 10 min. Continued until systemic antibiotic therapy of VAP was ended (decided by physician). N=51 <strong>Control group</strong> Systemic antibiotic(s) plus NSS equivalent to 75 mg of colistin base reconstituted in 4 mL of NSS every 12 h via a nebulizer for 10 min. Continued until systemic antibiotic therapy of VAP was ended. N=49</td>
<td>Mortality Rates of mortality due to VAP and all-cause mortality did not differ between the groups receiving intervention or control <strong>Clinical success/improvement</strong> Favourable microbiological outcome was significantly higher in the intervention group compared with the control group (RR 1.57, 95% CI 1.03–2.37), but no significant difference was observed on clinical outcomes The overall incidence of complications, bronchospasm and renal impairment did not differ between the two treatment groups</td>
<td>RCT Acceptable methodological quality (+)</td>
</tr>
<tr>
<td>Study details</td>
<td>Objective and participants</td>
<td>MDR Gram-negative bacteria</td>
<td>Intervention, control and follow-up</td>
<td>Results</td>
<td>Quality assessment</td>
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<td><strong>Stenderup 1983</strong>&lt;br&gt;RCT&lt;br&gt;<strong>Setting</strong> Community Denmark&lt;br&gt;Dates not reported</td>
<td>To study the use of mecillinam as a prophylactic for travellers’ diarrhoea&lt;br&gt;<strong>Participants</strong> N=74 tourists&lt;br&gt;Adults 19–45 years, middle aged 46–64 years, aged 65–79 years, elderly 80+ years&lt;br&gt;Male: not reported, female: not reported&lt;br&gt;Inclusion criteria: Danish tourists travelling to Egypt and the Far East&lt;br&gt;Exclusion criteria: not reported</td>
<td><strong>Bacteria:</strong> Enterotoxogeni <em>E. coli</em>&lt;br&gt;<strong>Resistant to:</strong> mecillinam, tetracycline, sulfonamide, streptomycin, chloramphenicol, kanamycin, ampicillin, cephalosporin, carbenicillin&lt;br&gt;<strong>Mechanism of resistance:</strong> not reported</td>
<td><strong>Intervention</strong>&lt;br&gt;Mecillinam, 200 g, 1x per day for 25 days. N=38&lt;br&gt;<strong>Control group</strong>&lt;br&gt;Placebo. N=36</td>
<td><strong>Length of follow-up:</strong> duration of treatment</td>
<td>Antibiotic resistance&lt;br&gt;Only 8% of <em>E. coli</em> strains were resistant to three or more antibiotics in the pre-travel samples. Post-travel, after participants had received either mecillinam or placebo, approximately 50% or more of the <em>E. coli</em> was resistant to more than three antibiotics</td>
</tr>
<tr>
<td><strong>Tannock 2011</strong>&lt;br&gt;RCT&lt;br&gt;<strong>Setting</strong> Primary (14 long-term care facilities)&lt;br&gt;New Zealand&lt;br&gt;Dates not reported</td>
<td>To test the efficacy of probiotic strain <em>E. coli</em> Nissle 1917 in reducing the carriage of MDR <em>E. coli</em>&lt;br&gt;<strong>Participants</strong> N=70&lt;br&gt;Age: not reported&lt;br&gt;Male: not reported, female: not reported&lt;br&gt;Inclusion criteria: not reported&lt;br&gt;Exclusion criteria: not reported</td>
<td><strong>Bacteria:</strong> <em>E. coli</em>&lt;br&gt;<strong>Resistant to:</strong> fluoroquinolones (norfloxacin)&lt;br&gt;<strong>Mechanism of resistance:</strong> ESBL</td>
<td><strong>Intervention</strong>&lt;br&gt;Probiotic: strain <em>E. coli</em> Nissle 1917, 5x10⁹-5x10¹⁰ CFU one capsule twice daily for five weeks. N=36&lt;br&gt;<strong>Control group</strong>&lt;br&gt;Placebo starch powder capsule. N=33</td>
<td><strong>Length of follow-up:</strong> five weeks</td>
<td>Clinical success/improvement&lt;br&gt;There was no significant difference between the probiotic and placebo groups in the number of people with faecal and urine samples becoming negative or remaining positive.</td>
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<tr>
<td>Study details</td>
<td>Objective and participants</td>
<td>MDR Gram-negative bacteria</td>
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<td>Quality assessment</td>
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<tr>
<td><strong>Wang 2009</strong></td>
<td>To report the effectiveness of extended-infusion meropenem compared with conventional bolus dosing in the management of HAP due to MDR <em>A. baumannii</em></td>
<td><em>Bacteria: A. baumannii</em></td>
<td><strong>Intervention</strong> Extended intravenous meropenem infusion: 500 mg every 6 h over a 3-h infusion. <em>N</em> = 15</td>
<td><strong>Clinical success/improvement</strong> No significant differences were found between extended-infusion meropenem and conventional bolus dosing in the number of patients with treatment success at days 3, 5 and 7. The rates of relapse also did not significantly differ between the treatment groups</td>
<td><strong>RCT</strong> Acceptable methodological quality (+) Small sample size</td>
</tr>
<tr>
<td><strong>Setting</strong> Tertiary (one ICU) China</td>
<td><strong>Participants</strong></td>
<td><strong>Resistant to:</strong> carbapenems (meropenem)</td>
<td><strong>Control group</strong> Conventional treatment: intravenous meropenem 1 g. every 8 h over a 1-h infusion. <em>N</em> = 15</td>
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<tr>
<td>March 2006–July 2006</td>
<td><em>N</em> = 30</td>
<td><strong>Mechanism of resistance:</strong> not reported</td>
<td><strong>Length of follow-up:</strong> duration of treatment</td>
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<td></td>
<td>Adults 19–45 years, middle aged 46–64 years, aged 65–79 years</td>
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<td>Among strains. None of the isolates were ESBL producers.</td>
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<td>Male: 19, female: 11</td>
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<td></td>
<td>Inclusion criteria: HAP due to MDR <em>A. baumannii</em></td>
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<td></td>
<td>Exclusion criteria: not reported</td>
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<tr>
<td><strong>Xue 2009</strong></td>
<td>To determine the relation of carbapenem restriction with the incidence of MDR <em>A. baumannii</em> in VAP</td>
<td><em>Bacteria: A. baumannii</em></td>
<td><strong>Intervention</strong> Carbapenem restriction policy limiting the use of third-generation carbapenems. Only used when severe sepsis and after consultation with a physician from the Department of Infectious Diseases. <em>N</em> = 12</td>
<td><strong>Mortality</strong> The rates of mortality did not differ significantly between the treatment groups (RR 0.78; 95% CI 0.29–2.12).</td>
<td><strong>RCT</strong> Low methodological quality (0) Small sample size</td>
</tr>
<tr>
<td><strong>Setting</strong> Tertiary (one ICU) China</td>
<td><strong>Participants</strong></td>
<td><strong>Resistant to:</strong> carbapenems</td>
<td><strong>Control group</strong> Conventional treatment: no restrictions of carbapenem (doctors were able to prescribe if necessary). <em>N</em> = 15</td>
<td><strong>Antibiotic resistance</strong> More patients in the conventional group developed a carbapenem-resistant strain of <em>A. baumannii</em>, although the difference was not statistically significant (RR 0.63; 95% CI 0.38–1.04)</td>
<td></td>
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<tr>
<td>June 2007–December 2007</td>
<td><em>N</em> = 26</td>
<td><strong>Mechanism of resistance:</strong> ESBL</td>
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<td></td>
<td>Adults 19–45 years, middle aged 46–64 years, aged 65–79 years</td>
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<td></td>
<td>Male: 15, female: 11</td>
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<tr>
<td></td>
<td>Inclusion criteria: patients receiving mechanical ventilation for more than five days and diagnosed with VAP</td>
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<td></td>
<td>Exclusion criteria: not reported</td>
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<tr>
<td>Study details</td>
<td>Objective and participants</td>
<td>MDR Gram-negative bacteria</td>
<td>Intervention, control and follow-up</td>
<td>Results</td>
<td>Quality assessment</td>
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<td>Exclusion criteria: not reported</td>
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<td>Length of follow-up: duration of treatment</td>
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P. aeruginosa, Pseudomonas aeruginosa; E. coli, Escherichia coli; C. freundii, Citrobacter freundii; M. morgagnii, Morganella morgagnii; A. baumannii, Acinetobacter baumannii; A. anitratus, Acinetobacter anitratus; P. mira, Proteus mira; S.marcescens, Serratia marcescens; B. cepacia, Burkholderia cepacia; MDR, multi-drug resistant; VAP, ventilator-associated pneumonia; ESBL, extended-spectrum beta-lactamase; CMS, colistimethate sodium; RCT, randomized controlled trial; ICU, intensive care unit; UTI, urinary tract infection; HAP, hospital-acquired pneumonia; NSS, nebulized sterile normal saline; CFU, colony-forming unit; SD, standard deviation; RR, risk ratio; CI, confidence interval.
4.4. Systematic Review References

4.4.1. Antimicrobial Stewardship


Lewis GJ, Fang X, Gooch M, Cook PP. Decreased resistance of *Pseudomonas aeruginosa* with restriction of ciprofloxacin in a large teaching hospital’s intensive care and intermediate care units. *Infect Control Hosp Epidemiol* 2012;33:368-373.


4.4.2. Other infection control measures


4.4.3. Selective decontamination


4.4.4. Treatment


4.5. Excluded clinical studies

4.5.1. Case-control study


Fortaleza CMCB, Freire MP, Filho Dde C, de Carvalho Ramos M. Risk factors for recovery of imipenem- or ceftazidime-resistant *Pseudomonas aeruginosa* among patients admitted to a teaching hospital in Brazil. *Infect Control Hosp


4.5.2. Case series/report


4.5.3. Cross-sectional


Iosifidis E, Antachopoulos C, Tsivitanidou M, et al. Differential correlation between rates of antimicrobial drug


4.5.4. In-vitro studies


### 4.5.5. Prospective cohort


### 4.5.6. Surveillance


Behera B, Mathur P. High levels of antimicrobial resistance at a tertiary trauma care centre of India. *Ind J Med Res* 2011; **133**:343–345.


4.5.7. Narrative reviews, commentaries or editorials


Curcio D. Tigecycline for treating ventilator-associated pneumonia: a practical perspective. Diagn Microbiol Infect Dis
2011;69:466–467.


Nseir S. Aerosolized antibiotics are not a good idea – don’t go with the flow: Premum Non Nocere! *Crit Care Med* 2005;33:443–444.


### 4.5.8. Retrospective cohort


4.5.9. Study design not relevant


4.5.10. Controlled before–after studies without a minimum of two intervention and control sites


4.5.11. Interrupted time series studies without at least three data points before and after the intervention


4.5.12. Participants not relevant


Karageorgopoulos DE, Kelesidis T, Kelesidis I, Falagas ME. Tigecycline for the treatment of multidrug-resistant (including


### 4.5.13. Antibiotics used not relevant for the review


### 4.5.14. Not multi-drug-resistant infections


Appendix 5: CPD material

1. Which of the following are appropriate monotherapy meropenem-sparing agents:
   a) Temocillin
   b) Cefixime
   c) Ceftolozane/tazobactam
   d) Fosfomycin
   e) Ceftazidime/avibactam
   Answer a, c, d, e

2. Which of the following are true:
   a) Polymyxins do not require monitoring renal function in the elderly.
   b) Fluoroquinolones can be used to treat urinary infection due to multidrug resistant Gram-negative bacteria
   c) Oral pivmecillinam should be used alone in the treatment of upper urinary infection
   d) Polymyxins should be given in combination with other agents if they are used in treating carbapenem-resistant Enterobacteriaceae.
   e) Co-trimoxazole should be used in treatment of infections due to *Stenotrophomonas maltophilia*
   Answer b, d, e

3. Which of the following are true:
   a) In uncomplicated urinary infection due to a proven ESBL-producing organism, treatment is recommended for 3 days
   b) If infection with MDR GNB is suspected, treat asymptomatic bacteriuria
   c) Give antibiotic prophylaxis for urinary catheter insertion if previous history of symptomatic urinary infections associated with a catheter change or there is trauma during the catheter insertion
   d) Daily antibiotic prophylaxis is preferable to standby antibiotics in recurrent urinary infection
   e) Always send a urine specimen for culture if an antibiotic-resistant organism is suspected AND the patient is asymptomatic
Answer c,

4. Which of the following are true;

   a) Ceftolozane-tazobactam is active against AmpC producing Enterobacteriaceae

   b) Ceftazidime-avibactam is active against AmpC producing Enterobacteriaceae

   c) KPC-producing Klebsiella sp. often produce aminoglycoside methyltransferases conferring pan-aminoglycoside resistance

   d) NDM-producing E. coli are usually mecillinam susceptible

   e) Proteus sp. are usually resistant to fosfomycin

Answer b
Appendix 6: Consultation stakeholders

Antimicrobial Resistance and Hospital Acquired Infection

Advisory Committee (APRHAI)

British Medical Association

British Society of Antimicrobial Chemotherapy

British Infection Association

C. Diff Support

European Society of Clinical Microbiology and Infectious Diseases

Faculty of Intensive Care Medicine

Foundation Trust Network

Hand Hygiene Alliance

Healthcare Infection Society

Infection Prevention Society

Lee Spark Foundation

MRSA Action UK

NHS Confederation

NHS England

NHS Trust Development Authority

Patient’s Association

Public Health England / Wales / Scotland / Northern Ireland

Royal College of Pathologists
Royal College of General Practitioners

Royal College of Nursing

Royal College of Physicians

Royal College of Surgeons

Service User Research Forum Healthcare acquired Infections

UK Clinical Pharmacists Association

Unison
## Appendix 7  Response from Stakeholders in consultation

<table>
<thead>
<tr>
<th>Respondent</th>
<th>Address</th>
<th>Email</th>
<th>Date Rec/d</th>
</tr>
</thead>
<tbody>
<tr>
<td>Conor Doherty</td>
<td>NHS GGC – paed infectious diseases</td>
<td><a href="mailto:Conor.Doherty@ggc.scot.nhs.uk">Conor.Doherty@ggc.scot.nhs.uk</a></td>
<td>23 May 2016</td>
</tr>
<tr>
<td>Ibai Los-Arcos</td>
<td>Infectious Diseases Division, Hospital Universitari Vall d'Hebron Avda. Vall d'Hebron, 119-129 08035 Barcelona. Spain</td>
<td><a href="mailto:bai.losarcos@gmail.com">bai.losarcos@gmail.com</a></td>
<td>01 June 2016</td>
</tr>
<tr>
<td>Prof. Céline PULCINI</td>
<td>Nancy University Hospital, Nancy, France</td>
<td><a href="mailto:celine.pulcini@univ-lorraine.fr">celine.pulcini@univ-lorraine.fr</a></td>
<td>01 June 2016</td>
</tr>
<tr>
<td>Aaron Nagar</td>
<td>Microbiology Department, Antrim Area Hospital, 45 Bush Rd, Antrim, Northern Ireland, BT41 2RL</td>
<td><a href="mailto:Aaron.Nagar@northerntrust.hscni.net">Aaron.Nagar@northerntrust.hscni.net</a></td>
<td>01 June 2016</td>
</tr>
<tr>
<td>Dr Paul Chadwick &amp; Dr Alex Peel</td>
<td>Microbiology Department Salford Royal NHS Foundation Trust Stott Lane, Salford. M6 8HD</td>
<td><a href="mailto:paul.chadwick@srf.t.nhs.uk">paul.chadwick@srf.t.nhs.uk</a>; <a href="mailto:alex.peel@srf.nhs.uk">alex.peel@srf.nhs.uk</a></td>
<td>15 June 2016</td>
</tr>
<tr>
<td>Rebecca Tilley</td>
<td>West Suffolk NHS Foundation Trust, Hardwick Lane, Bury St Edmunds, Suffolk, IP33 2QZ.</td>
<td><a href="mailto:rebecca.tilley@wsh.nhs.uk">rebecca.tilley@wsh.nhs.uk</a></td>
<td>17 June 2016</td>
</tr>
<tr>
<td>Name</td>
<td>Hospital of Infectious Diseases</td>
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<td>Egidia Miftode</td>
<td>Iasi Str O Botez no 2, code 700274, Iasi Romania</td>
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</tbody>
</table>

**British Society for Antimicrobial Chemotherapy**

**Joint Working Party Paper on Multi resistant Gram-negative Infection: Treatment**

**Consultation deadline: Friday 17 June 2016**

- Please use this form for submitting your comments to BSAC. **COMMENTS WILL ONLY BE ACCEPTED ON THIS FORM**
- Please put each comment in a separate row
- Type directly onto the form. Do not paste other tables or figures as they may get lost
- Only comments received on the attached form will be considered.

**How to respond:** Please complete this BSAC response form and submit by email to fdrummond@bsac.org.uk no later than **Friday 17 June 2016**. Comments received after the deadline will not be accepted.

<table>
<thead>
<tr>
<th>Name</th>
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<tbody>
<tr>
<td>Organisation Address &amp; Postcode</td>
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EXAMPLE: Full 16 45 Our comments are as follows ...... Exclude: Reason WP Response

Specific mention made that does not cover neonates and mostly does not deal with paediatric dosage or paediatric-specific issues such prophylaxis of UTI
2) Appropriate empirical treatment and prophylaxis strategies in the face of increasing trimethoprim resistance for paed UTI's is a major issue and not discussed
British Society for Antimicrobial Chemotherapy
Joint Working Party Paper on Multi resistant Gram-negative Infection: Treatment

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References:


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<tr>
<th>Name</th>
<th>Prof. Céline PULCINI</th>
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<tr>
<td>Organisation Address &amp; Postcode</td>
<td>Nancy University Hospital, Nancy, France</td>
</tr>
<tr>
<td>Email</td>
<td><a href="mailto:celine.pulcini@univ-lorraine.fr">celine.pulcini@univ-lorraine.fr</a></td>
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<td>Phone number</td>
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British Society for Antimicrobial Chemotherapy  
Joint Working Party Paper on Multi resistant Gram-negative Infection: Treatment

Consultation deadline:  Friday 17 June 2016

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<table>
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<tr>
<th>Name</th>
<th>Aaron Nagar</th>
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<tbody>
<tr>
<td>Organisation Address &amp; Postcode</td>
<td>Microbiology Department, Antrim Area Hospital, 45 Bush Rd, Antrim, Northern Ireland, BT41 2RL</td>
</tr>
<tr>
<td>Email</td>
<td><a href="mailto:Aaron.Nagar@northerntrust.hscni.net">Aaron.Nagar@northerntrust.hscni.net</a></td>
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<td>Phone number</td>
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British Society for Antimicrobial Chemotherapy  
Joint Working Party Paper on Multi resistant Gram-negative Infection: Treatment  

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How to respond: Please complete this BSAC response form and submit by email to fdrummond@bsac.org.uk no later than Friday 17 June 2016. Comments received after the deadline will not be accepted.

| **Name** | Dr Paul Chadwick, Clinical lead/consultant microbiologist  
Dr Alex Peel, Antimicrobial stewardship lead/consultant microbiologist |
|----------|--------------------------------------------------------------------|
| **Organisation Address & Postcode** | Microbiology Department  
Salford Royal NHS Foundation Trust  
Stott Lane, Salford. M6 8HD |
| **Email** | paul.chadwick@srf.t.nhs.uk;  
alex.peel@srf.nhs.uk |
| **Phone number** | 01612065030 |
| **Conflict(s) of Interest** |  

This guideline is welcomed as a resource to support treatment of MDR Gram negative infections and is supported by an extensive literature review. However, the recommendations in their current form appear as a fairly disjointed and inconsistent collection of statements. For example, the first recommendation starts with the role of temocillin vs Enterobacteria and Burkholderia and the second recommendation is for ampicillin-sulbactam vs Acinetobacter. This is not a logical or helpful sequence for presentation. Some of the recommendations appear as a surprise as they do not relate back to the preceding evidence or discussion. Care should be taken to ensure that this link is made and a justification provided for all recommendations

Perhaps the functionality of the guideline could be improved with a more structured approach to the management of MDR Gram negatives? For example the role of each of the different classes of agents (recommended Y/N + comments) could be systematically presented as a table for each of the common

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<td>Very useful set of comments.</td>
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<td>1. Antibiotics considered have been re-ordered to reflect important issues.</td>
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<td>2. All recommendations checked for relationship to text and evidence</td>
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<td>3. Too many mechanisms to consider all but additional table on mechanisms and activity added.</td>
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<td>resistance mechanisms, if necessary separated into different tables for the different organism groups (e.g. Enterobacteria, non-fermentors).</td>
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<td>783</td>
<td>The conclusion that temocillin may be used as a carbapenem-sparing agent against Enterobacteria is (a reasonable) opinion of the authors but does not follow from the evidence presented. (The same opinion might also have be given for other classes of agent such as polymixins). Consideration should be given to simplifying and rephrasing the recommendation to &quot;temocillin can be used to treat infections due to Enterobacteria, including ESBL and AmpC producers&quot;</td>
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</table>
| Full | 30 | 830 | The recommendation that 'Amoxicillin-clavulanate should not be used to treat infection with known ESBL-producing organism unless sensitivity known' is generally not very helpful for a typical diagnostic laboratory where apparent co-amoxiclav susceptibility will be known either before or at the same time as ESBL production is confirmed.

Alternatively, if the authors are suggesting that a patient with a **history of** ESBL positive UTI/infection should not be given co-amoxiclav until sensitivity for the **current episode** is confirmed, the recommendation should be clearly worded |
| Full | 32 | 883 | The following recommendation is not supported by any evidence linking clinical outcomes to sepsis severity criteria: 'Piperacillin-tazobactam can be considered for use in mild-moderate infections (i.e. not severe sepsis) due to ESBL-producing Enterobacteriaceae if supported by susceptibility results.' The evidence should be provided, the opinion justified, or the recommendation removed. |

Recommended on a case by case basis

Detailed consideration given of this recommendation but given 6+% recurrence rate with ESBL infection previous susceptibility is an important factor in making this choice. Substantial caveats against use of coamoxiclav and piperacillin/tazobactam use in UK added both because of in vitro resistance and prevalence of OXA-1 in UK isolates

Recommendation changed to omit reference to severity of infection
<p>| Full | 32 | 888 | The following recommendation is not supported by any evidence. ‘‘However combination with an aminoglycoside is advisable for severe infections.’ The evidence should be provided, the opinion justified, or the recommendation removed. | Agree. Removed |
| Full | 36 | 986 | It is unclear why there needs to be a separate recommendation for ertapenem: ‘Ertapenem is effective in treatment of infections with multi-resistant Enterobacteriaceae apart from carbapenemase producers’ when this has already been covered by the previous recommendation: ‘Carbapenems should be used to treat serious ESBL-producing Gram-negative infections subject to antibiotic stewardship to minimize the risk of developing resistance’. Is there a reason why the general carbapenem recommendation is not extended to include AmpC resistance? For internal consistency within the document, we suggest merging these two recommendations as follow: ‘carbapenems can be used to treat infections due to ESBL or AmpC producing Enterobacteria’. Ertapenem has different properties and is now recommended for OPAT. AmpC issue now considered |
| Full | 37 | 1010 | The format of the following recommendation is internally inconsistent within the document: ‘Although it retains good efficacy against infections with <em>Pseudomonas aeruginosa</em>, ceftazidime is not recommended for the treatment of other serious infections due to ESBL / AmpC producing Enterobacteriaceae, even if in vitro tests suggest the isolate is susceptible.’ We suggest 1) separating the recommendations for treating Pseudomonas and Enterobacterial infections, 2) rephrasing the recommendation for Enterobacteria as follows: ‘ceftazidime should NOT be used to rephrased |</p>
<table>
<thead>
<tr>
<th>Page</th>
<th>Line</th>
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<tbody>
<tr>
<td>Full</td>
<td>39</td>
<td>1074</td>
<td>Information relating to aztreonam-avibactam, while interesting, does not belong under a heading of ceftazidime-avibactam and is not directly relevant to the guideline – suggest remove</td>
<td>Separate aztreonam section added which houses the experimental combination aztreonam-avibactam</td>
</tr>
<tr>
<td>Full</td>
<td>40</td>
<td>1086</td>
<td>The format of the following recommendation is internally inconsistent within the document: ‘With the exception of infections with metallo-β-lactamase strains, ceftazidime-avibactam, when available, should be used as alternative treatment to carbapenems’.</td>
<td>Rewritten</td>
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<tr>
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<td>1140</td>
<td>The format of the following recommendation is internally inconsistent within the document (and implies that it should be used in preference to carbapenems): ‘Ceftolozane-tazobactam should be used as alternative treatment to carbapenems in treating ESBL-producing Gram negative pathogens (but not carbapenemase producers).’</td>
<td>Rewritten</td>
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<tr>
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<td>1231</td>
<td>There is potential overlap/duplication regarding combination therapy with this recommendation and the recommendation on page 56, line1518. Consider either removing ‘and preferably used in combination with other agents’ and adding a cross reference to the later section</td>
<td>Cross-references inserted where useful</td>
</tr>
<tr>
<td>Full</td>
<td>45</td>
<td>1234</td>
<td>The recommendation with regard to renal function is internally inconsistent within the document as side effects are not systematically considered for other agents. Many important unwanted effects occur for many different antimicrobials and relevant monitoring should be considered as a matter of course by the prescribing clinician (and this might include monitoring colistin levels also, which is not mentioned as a recommendation).</td>
<td>To contain a ready voluminous length Unwanted effects are highlighted where may be specifically over-looked.</td>
</tr>
</tbody>
</table>
| Full | 46 | 1266 | The format of the following recommendation is internally inconsistent within the document: ‘Fluoroquinolones can be used to treat urinary infection due to multidrug resistant Gram-negative bacteria based on susceptibility results.’

We suggest rephrase this recommendation as follows: ‘quinolones can be used to treat complicated urinary tract infections due to Gram negative bacteria’ | Standardised |
| Full | 51 | 1390 | The format of the following recommendation is internally inconsistent within the document: ‘Fosfomycin should be used in treatment of urinary infection due to multiresistant Gram-negative bacteria (oral administration only suitable for lower urinary infection)’

We suggest rephrase as follows: ‘Fosfomycin can be used to treat urinary tract infections due to Gram-negative bacteria (oral administration only suitable for lower urinary infection)’ | Standardised |
| Full | 52 | 1410 | To improve internal consistency within the document, we suggest adding the following additional recommendation (which follows from the preceding evidence): ‘aztreonam should NOT be used to treat infections due to ESBL or AmpC producing Enterobacteria’ | Agreed |
There is a recommendation to use 7 days therapy for ESBL simple UTIs to improve bacteriological clearance. There is no mention of clinical outcomes evidence. Bacteriological clearance does not necessarily correlate well with clinical outcomes (e.g. high prevalence of asymptomatic bacteriuria in certain patient populations). This recommendation could lead to a large increase in ab use if implemented widely and it would need strong clinical evidence before doing so.

Debated at length within WP. Considered that best possible bacteriological clearance should be obtained with proven MDR GNB infection but caveat inserted about clinical relevance of bacteriological cure.

This recommendation: ‘admission for intravenous aminoglycoside therapy’ is potentially confusing as it appears to exclude an inpatient carbapenem option (presumably temocillin or other agents recommended above for Enterobacteria could also be considered).

We suggest rephrase as ‘admission for intravenous therapy with an aminoglycoside or carbapenem (? Or temocillin etc).

Whole section for recommendations recast. Point accepted.

Although the evidence base is weak in many areas, and the authors are to be commended for covering many topic areas, we feel the document does not read like it is focused on an infection specialist dealing with ‘real world’ problems e.g. a patient with KPC bacteraemia with MICs of x,y,z and renal failure and obesity etc – we note that the US has produced flowcharts previously (e.g. Medscape http://www.medscape.com/viewarticle/780065_9 ) see screenshot on following page, and more recent publications - clearly these may be based on minimal evidence but they do provide a start. We wonder whether consideration could be given by the WP to producing similar tools.

* simple flow-charts inserted but subject is too diverse to deal with all possible clinical situations.
Figure 2.
Potential antibiotic combination therapy algorithm for the treatment of carbapenem-resistant Klebsiella pneumoniae infections stratified to site of infection and antibiogram results. *Algorithm would be appropriate for institutions where >50% of isolates exhibit carbapenem MICs in the treatable range with HD therapy (MIC 32 mg/mL). Specific drugs used for empirical therapy should be tailored to the epidemiology of endemic carbapenem-resistant Klebsiella pneumoniae strains. †HD meropenem (8 g daily, administered as prolonged infusion). ‡HD tigecycline (200 mg loading dose, 100 mg once a day), see text regarding the limitations and evidence supporting the use of HD regimens. HD: High-dose.
British Society for Antimicrobial Chemotherapy
Joint Working Party Paper on Multi resistant Gram-negative Infection: Treatment

Consultation deadline: Friday 17 June 2016

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<tr>
<th>Name</th>
<th>Rebecca Tilley</th>
</tr>
</thead>
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<tr>
<td>Organisation Address &amp; Postcode</td>
<td>West Suffolk NHS Foundation Trust, Hardwick Lane, Bury St Edmunds, Suffolk, IP33 2QZ.</td>
</tr>
<tr>
<td>Email</td>
<td><a href="mailto:rebecca.tilley@wsh.nhs.uk">rebecca.tilley@wsh.nhs.uk</a></td>
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<tr>
<td>Phone number</td>
<td>01284 712635</td>
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all departments have junior doctors to assist with this sort of responsibility.

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<td>Would recommend that 1) the term “standby antibiotics” is explained and 2) that advice is given on how a clinician, bearing in mind this is often a GP, would decide which antibiotic would be appropriate as a “standby” option.</td>
<td>Exclude: Needs modification.</td>
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<tr>
<td>There is a superscript β in the flowchart, but it does not appear to refer to anything</td>
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</tr>
<tr>
<td>There is a comment marked ¥, but this symbol does not appear in the flowchart.</td>
<td>Exclude: needs reviewing</td>
</tr>
<tr>
<td>MRGNs are an increasing problem for us but we are not yet seeing many MRGN bacteraemias and CPEs remain very rare locally. The management of sepsis necessarily requires empirical broad-spectrum antibiotic treatment before we have positive microbiology but we are not yet at the stage where our local guidance advises empirical cover for MRGNs unless there are risk factors for this. We are concerned that the recent CQUIN – re: reduction in antibiotic consumption which is particularly targeting piperacillin-tazobactam and carbapenems seems to be at odds with the empirical management of sepsis and if our Trust has any hope of achieving this target (which incidentally uses historic baseline data from a time when MRGNs were far less prevalent) then we would need to be moving empirical therapy back to cephalosorins and quinolones for example. We are reluctant to do this from a C. difficile perspective and from driving resistance mechanisms yet further. We appreciate that this document is not directly related to the CQUIN and that we are venting our frustration but it would be helpful if BSAC could issue a position statement or guidance on this CQUIN and outline the best approach for microbiologists to a) do the right thing in terms of empirical therapy for the septic...</td>
<td>We are also concerned about the potential conflict between antibiotic-use reduction targets and potential mortality in bacteraemia which has similar 30 day mortality to C.difficile. Document extensively revised and your general points incorporated. Thank you</td>
</tr>
</tbody>
</table>
patient, particularly if there is a MRGN risk plus b) reduce the risk of promoting antibiotic resistance plus c) meet contractual obligations. I know we are not the only Trust that is exasperated by the specifics within this DH requirement which seems to totally disregard all the improvements made in recent years with regard to C. difficile and antibiotic stewardship.

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<td>The document discusses using antibiotics such as temocillin, tigecycline, colistin and fosfomycin. EUCAST does not provide guidance on interpretation of temocillin susceptibility either by disk or MIC. Tigecycline needs to be tested via MIC for anything other than E coli. Fosfomycin &amp; colistin need to be tested by MIC. These requirements reduce the turnaround times for results. In addition, the turnaround times for CPE resistance mechanisms/additional sensitivities do not help support optimum patient management. Could PHE Colindale publish its testing methods/MIC interpretations to enable local testing rather than sending isolates to them? Is there a way to expedite EUCAST guidance on temocillin interpretations? Can BSAC offer recommendations to support local business cases for introducing technology that enables faster identification of e.g. CPEs in house as opposed to relying on reference laboratories?</td>
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In practice we now consider that molecular methodology is needed for colistin susceptibility testing and MICs for meropenem with MDR GNB and this has been added. To track the fast changing situation we have now recommended that i) mandatory reporting of carbapenem resistant isolates is introduced ii) isolates are dealt with expeditiously for patient benefit and iii) isolates referred where testing is beyond the scope of local laboratories.
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</tr>
<tr>
<td>Email</td>
<td><a href="mailto:emiftode@yahoo.co.uk">emiftode@yahoo.co.uk</a></td>
</tr>
<tr>
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**British Society for Antimicrobial Chemotherapy**  
**Joint Working Party Paper on Multi resistant Gram-negative Infection: Treatment**

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<tr>
<th>Name</th>
<th>Neil Woodford</th>
</tr>
</thead>
</table>
| Organisation Address & Postcode | Antimicrobial Resistance and Healthcare Associated Infections Reference Unit (AMRHAI)  
Public Health England  
61 Colindale Avenue  
London, NW9 5EQ |
<p>| Email | <a href="mailto:Neil.Woodford@phe.gov.uk">Neil.Woodford@phe.gov.uk</a> |
| Phone number | Tel. +44 (0)20 8327 7255 |
| Conflict(s) of Interest |</p>
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<td>Typos dealt with</td>
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<td>Should there again be a section on the use of sterilising agents or the use of NSAIDs in uncomplicated UTIs</td>
<td>Include</td>
<td>See previous response</td>
</tr>
<tr>
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<td>84</td>
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<td>Typos dealt with</td>
</tr>
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<td>85</td>
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<td>Table sometimes has full stop and at other times does not</td>
<td>Include</td>
<td>Hopefully dealt with</td>
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Accepted manuscript

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