





Draft Genome Sequences of Two Gammaproteobacterial Methanotrophs Isolated from Rice Ecosystems

Katharina Frindte,^a Marina G. Kalyuzhnaya,^b Françoise Bringel,^c Peter F. Dunfield,^d Mike S. M. Jetten,^e Valentina N. Khmelenina,^f Martin G. Klotz,^g J. Colin Murrell,^h Huub J. M. Op den Camp,^e Yasuyoshi Sakai,ⁱ Jeremy D. Semrau,^j Nicole Shapiro,^k Alan A. DiSpirito,^l Lisa Y. Stein,^m Mette M. Svenning,ⁿ Yuri A. Trotsenko,^f Stéphane Vuilleumier,^b Tanja Woyke,^k Claudia Knief^a

Institute of Crop Science and Resource Conservation—Molecular Biology of the Rhizosphere, University of Bonn, Bonn, Germanya; San Diego State University, San Diego, California, USAb; Department of Microbiology, Genomics and the Environment, Université de Strasbourg, UMR 7156 CNRS, Strasbourg, Frances; Department of Biological Sciences, University of Calgary, Calgary, Alberta, Canadad; Department of Microbiology, Faculty of Science, Radboud University, Nijmegen, The Netherlandse; G.K. Skryabin Institute of Biochemistry and Physiology of Microorganisms, Russian Academy of Sciences, Pushchino, Russiaf; Division of Mathematics and Natural Sciences, Queens College of the City University of New York, Flushing, New York, USAa; School of Environmental Sciences University of East Anglia, Norwich, United Kingdomh; Division of Applied Life Sciences, Graduate School of Agriculture, Kyoto University, Kitashirakawa-Oiwake, Kyoto, Japani; Department of Civil and Environmental Engineering, University of Michigan, Ann Arbor, Michigan, USAi; DOE Joint Genome Institute, Walnut Creek, California, USAk; Roy J. Carver Department of Biochemistry, Biophysics and Molecular Biology, Iowa State University, Ames, Iowa, USAl; Department of Biological Sciences, University of Norway, Tromsø, Norwaya

ABSTRACT The genomes of the aerobic methanotrophs "Methyloterricola oryzae" strain 73a^T and Methylomagnum ishizawai strain 175 were sequenced. Both strains were isolated from rice plants. Methyloterricola oryzae strain 73a^T represents the first isolate of rice paddy cluster I, and strain 175 is the second representative of the recently described genus Methylomagnum.

A erobic methanotrophic bacteria play a key role in controlling global climate by reducing the emission of the greenhouse gas methane in ecosystems such as paddy fields (1). Gammaproteobacterial methanotrophs are common inhabitants of rice fields (2, 3). We sequenced the genomes of two gammaproteobacterial isolates from rice plants (4). "Methyloterricola oryzae" strain 73a^T (=LMG 29185=VKM-B-2986) is currently the only cultivated representative of rice paddy cluster I (3, 5), and Methylomagnum ishizawai strain 175 (= LMG 28717=VKM-B-2989) is the second representative of the genus (6).

Genomic DNA was extracted from bacterial cultures using a phenol-chloroform method (7), and draft genome sequences were generated at the DOE Joint Genome Institute. The genome of strain $73a^T$ was sequenced using an Illumina HiSeq 2000, which generated 11,844,428 reads (1.79 Gb). The Pacific Biosciences RS was used for strain 175, and 230,505 filtered subreads (0.73 Gb) were generated. Sequence filtering, genome assembly, and gene annotation were performed as described earlier (8, 9). The final draft of strain $73a^T$ had $302.0\times$ read coverage, contained 74 contigs in 73 scaffolds, was 4.9 Mb in size, and had an average GC content of 61.1%. The draft of strain 175 had $140.8\times$ read coverage, contained 8 contigs in 8 scaffolds, was 5.5 Mb in size, and had an average GC content of 63.0%.

Both strains encode metabolic inventory typical for type I methanotrophs (10). They harbor genes encoding a particulate methane monooxygenase (pmoCAB) and a methane/

Received 28 April 2017 **Accepted** 2 May 2017 **Published** 17 August 2017

Citation Frindte K, Kalyuzhnaya MG, Bringel F, Dunfield PF, Jetten MSM, Khmelenina VN, Klotz MG, Murrell JC, Op den Camp HJM, Sakai Y, Semrau JD, Shapiro N, DiSpirito AA, Stein LY, Svenning MM, Trotsenko YA, Vuilleumier S, Woyke T, Knief C. 2017. Draft genome sequences of two gammaproteobacterial methanotrophs isolated from rice ecosystems. Genome Announc 5:e00526-17. https://doi.org/10.1128/genomeA.00526-17.

Copyright © 2017 Frindte et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0

Address correspondence to Claudia Knief, knief@uni-bonn.de.

This is contribution 13 from the Organization for Methanotroph Genome Analysis (OMeGA).

Frindte et al.

ammonia monooxygenase-related protein (pxmABC) (11). Additionally, the genome of strain 175 encodes a soluble methane monooxygenase (mmoXYBZDCGR). Gene clusters for PQQ-dependent methanol dehydrogenases and PQQ biosynthesis were found in both strains (mxaFJGIRSACKLD, xoxFJ, pqqABCDE, and pqqFG). Formaldehyde oxidation is predicted to proceed via the tetrahydromethanopterin (H_4MPT) pathway (presence of fae, mtdB, mch, fhcABCD, and H_4MPT biosynthesis genes) (12). Additionally, both strains contained genes encoding the tetrahydrofolate-dependent pathway (presence of mtdA, fchA, fhs, and, additionally, a folD gene in strain 73aT). Formate can potentially be oxidized via one (strain 175) or two (strain 73aT) types of formate dehydrogenase.

Both strains may assimilate formaldehyde via the ribulose monophosphate pathway. The cleavage cascade can be realized via fructose-1,6-bisphosphate, and in strain 73a^T additionally via 2-keto-3-deoxy-6-phosphogluconate. Rearrangement of ribulose-5-phosphate can occur by transketolase and transaldolase reactions. A complete serine cycle is unlikely to be present. Both strains have the genes necessary for operational oxidative pentose phosphate and TCA cycle pathways, whereas a complete glycolysis cascade is encoded only in strain 73a^T. RubisCO genes are present (*cbbL* and *cbbS* in strain 175; *cbbM* in strain 73a^T), as well as genes for a complete Calvin-Benson-Bassham cycle.

For nitrogen acquisition, both strains possessed genes encoding ammonium (*amtB*), nitrate (*nasA*), and urea (*urtABCDE*) transporters, as well as urease genes (*ureABCDEFG*). Moreover, *nif* genes were present, suggesting the potential for dinitrogen fixation. The strains may form polyphosphate (*ppk*) and glycogen (*glgAB*, *glgC*, *glgP*, *glgX*, *malQ*, and *pgm*) as storage compounds. Strain 175, in addition, can potentially produce polyhydroxybutyrate (*phbAB* and *phbC*), a characteristic not yet known for type I methanotrophs (13).

Accession number(s). The genome sequences have been deposited in GenBank under the accession numbers JYNS00000000, for *Methyloterricola oryzae* strain 73a^T, and FXAM00000000, for *Methylomagnum ishizawai* strain 175.

ACKNOWLEDGMENTS

We thank all members of the Organization for Methanotroph Genome Analysis (OMeGA) and Genoscope (France) for access to its MicroScope platform for comparative genome analysis (http://www.genoscope.cns.fr/agc/microscope/home). The work conducted by the DOE Joint Genome Institute was supported by the Office of Science of the U.S. Department of Energy under contract no. DE-AC02-05CH11231.

REFERENCES

- Conrad R. 2009. The global methane cycle: recent advances in understanding the microbial processes involved. Environ Microbiol Rep 1:285–292. https://doi.org/10.1111/j.1758-2229.2009.00038.x.
- Conrad R. 2007. Microbial ecology of methanogens and methanotrophs. Adv Agron 96:1–63. https://doi.org/10.1016/S0065-2113(07)96005-8.
- Knief C. 2015. Diversity and habitat preferences of cultivated and uncultivated aerobic methanotrophic bacteria evaluated based on pmoA as molecular marker. Front Microbiol 6:1346. https://doi.org/10.3389/fmicb.2015.01346.
- Frindte K, Maarastawi SA, Lipski A, Hamacher J, Knief C. 2017. Characterization of the first rice paddy cluster I isolate, Methyloterricola oryzae gen. nov., sp. nov. and extended description of Methylomagnum ishizawai. Int J Syst Evol Microbiol, in press.
- Lüke C, Krause S, Cavigiolo S, Greppi D, Lupotto E, Frenzel P. 2010. Biogeography of wetland rice methanotrophs. Environ Microbiol 12: 862–872. https://doi.org/10.1111/j.1462-2920.2009.02131.x.
- Khalifa A, Lee CG, Ogiso T, Ueno C, Dianou D, Demachi T, Katayama A, Asakawa S. 2015. Methylomagnum ishizawai gen. nov., sp. nov., a mesophilic type I methanotroph isolated from rice rhizosphere. Int J Syst Evol Microbiol 65:3527–3534. https://doi.org/10.1099/ijsem.0.000451.
- 7. Sambrook J, Fritsch EF, Maniatis T. 1989. Molecular cloning: a laboratory manual. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
- 8. Hamilton R, Kits KD, Ramonovskaya VA, Rozova ON, Yurimoto H, Iguchi H, Khmelenina VN, Sakai Y, Dunfield PF, Klotz MG, Knief C, Op den Camp

- HJ, Jetten MS, Bringel F, Vuilleumier S, Svenning MM, Shapiro N, Woyke T, Trotsenko YA, Stein LY, Kalyuzhnaya MG. 2015. Draft genomes of gammaproteobacterial methanotrophs isolated from terrestrial ecosystems. Genome Announc 3(3):e00515-15. https://doi.org/10.1128/genomeA.00515-15.
- Kalyuzhnaya MG, Lamb AE, McTaggart TL, Oshkin IY, Shapiro N, Woyke T, Chistoserdova L. 2015. Draft genome sequences of gammaproteobacterial methanotrophs isolated from Lake Washington sediment. Genome Announc 3(2):e00103-15. https://doi.org/10.1128/genomeA.00103-15.
- Trotsenko YA, Murrell JC. 2008. Metabolic aspects of aerobic obligate methanotrophy. Adv Appl Microbiol 63:183–229. https://doi.org/10 .1016/S0065-2164(07)00005-6.
- Tavormina PL, Orphan VJ, Kalyuzhnaya MG, Jetten MS, Klotz MG. 2011.
 A novel family of functional operons encoding methane/ammonia monooxygenase-related proteins in gammaproteobacterial methanotrophs. Environ Microbiol Rep 3:91–100. https://doi.org/10.1111/j .1758-2229.2010.00192.x.
- Vorholt JA. 2002. Cofactor-dependent pathways of formaldehyde oxidation in methylotrophic bacteria. Arch Microbiol 178:239–249. https://doi.org/10.1007/s00203-002-0450-2.
- Pieja AJ, Sundstrom ER, Criddle CS. 2012. Cyclic, alternating methane and nitrogen limitation increases PHB production in a methanotrophic community. Bioresour Technol 107:385–392. https://doi.org/10.1016/j .biortech.2011.12.044.

Volume 5 Issue 33 e00526-17