

Genetic and phenotypic divergence in an island bird: isolation by distance, by colonization or by adaptation?

LEWIS G. SPURGIN,^{*,†‡} JUAN CARLOS ILLERA,[§] TOVE H. JORGENSEN,^{*,¶} DEBORAH A. DAWSON[‡] and DAVID S. RICHARDSON^{*}

^{*}School of Biological Sciences, University of East Anglia, Norwich Research Park, Norwich NR4 7TJ, UK, [†]Behavioural Ecology and Self-organization, Centre for Ecological and Evolutionary Studies, University of Groningen, 9700 CC, Groningen, The Netherlands, [‡]NERC Biomolecular Analysis Facility, Department of Animal and Plant Sciences, University of Sheffield, Sheffield S10 2TN, UK, [§]Research Unit of Biodiversity (UO-CSIC-PA), Oviedo University, Campus of Mieres, Research Building, 5th Floor, C/Gonzalo Gutiérrez Quirós s/n, Mieres, 33600 Asturias, Spain

Abstract

Discerning the relative roles of adaptive and nonadaptive processes in generating differences among populations and species, as well as how these processes interact, is a fundamental aim in biology. Both genetic and phenotypic divergence across populations can be the product of limited dispersal and gradual genetic drift across populations (isolation by distance), of colonization history and founder effects (isolation by colonization) or of adaptation to different environments preventing migration between populations (isolation by adaptation). Here, we attempt to differentiate between these processes using island populations of Berthelot's pipit (*Anthus berthelotii*), a passerine bird endemic to three Atlantic archipelagos. Using microsatellite markers and approximate Bayesian computation, we reveal that the northward colonization of this species ca. 8500 years ago resulted in genetic bottlenecks in the colonized archipelagos. We then show that high levels of genetic structure exist across archipelagos and that these are consistent with a pattern of isolation by colonization, but not with isolation by distance or adaptation. Finally, we show that substantial morphological divergence also exists and that this is strongly concordant with patterns of genetic structure and bottleneck history, but not with environmental differences or geographic distance. Overall, our data suggest that founder effects are responsible for both genetic and phenotypic changes across archipelagos. Our findings provide a rare example of how founder effects can persist over evolutionary timescales and suggest that they may play an important role in the early stages of speciation.

Keywords: Berthelot's pipit, bottleneck, founder effect, microsatellite

Received 16 September 2013; revision received 30 December 2013; accepted 8 January 2014

Introduction

Understanding the processes that drive differentiation among populations is a major aim in evolutionary biology. Most population genetic studies have been carried out using neutral genetic markers, at which variation

across populations has been viewed as the product of mutation, migration and genetic drift (Wright 1943; Hartl & Clark 2007). As migration tends to occur more commonly between neighbouring populations, a relationship between the geographic and genetic distance of pairs of populations – isolation by distance – is expected in many wild systems (Wright 1943, 1946). In contrast, most phenotypic differences across populations are assumed to have arisen in response to differential selection regimes, and drift is considered a null hypothesis against which selection can be tested (Orr 1998).

Correspondence: Lewis G. Spurgin, Fax: +31 (0)50 3635295;

E-mails: L.G.Spurgin@rug.nl, lewisspurgin@gmail.com

[¶]Present address: Department of Bioscience, Aarhus University, 8000 Aarhus C, Denmark

Recently, several authors have questioned whether patterns of neutral genetic differentiation can be understood in terms of migration and drift alone and have suggested that selection can play a role (e.g. Nosil *et al.* 2009; Orsini *et al.* 2013a). In cases where populations exist along environmental gradients, individuals are likely to be adapted to local conditions. This local adaptation may act as a barrier to migration, and subsequent drift will result in neutral genetic structure across populations. This phenomenon, known as ‘isolation by adaptation’, has received both theoretical and empirical support, and a recent review suggests that it may be substantially more common than previously realized (Orsini *et al.* 2013b). Importantly, just as selection may structure neutral DNA, genetic drift may shape populations at the phenotypic level (e.g. Carson 1968; Kolbe *et al.* 2012). However, because drift is usually considered a null hypothesis, its role in shaping differences among populations and species remains poorly understood (Marie Curie SPECIATION Network 2012).

A further related factor that must be considered in studies of population differentiation is colonization history. The founding of a population by a small number of individuals can result in founder effects – genetic and/or phenotypic divergence from the parent population. This can lead to a pattern of ‘isolation by colonization’, whereby differences between populations primarily reflect colonization history, rather than contemporary patterns of migration, gradual genetic drift or selection. Laboratory and field experiments designed to simulate founder events have shown that founder-mediated genotypic and phenotypic divergence can (but do not necessarily) occur and persist in the presence of selection (Moya *et al.* 1995; Travisano *et al.* 1995; Rundle 2003; Kolbe *et al.* 2012). Cases where recent natural or human-mediated introductions have been observed have produced mixed results, with the extent of founder divergence depending on the timing, severity and extent of bottlenecks (Carson 1968; Brinkmann *et al.* 1998; Coyne & Orr 2004; Frantz *et al.* 2009; Kolbe *et al.* 2012). Several recent studies have used modelling approaches to study isolation by colonization, again with mixed results. In humans, patterns of neutral genetic differentiation are largely the product of serial founder effects that arose as small populations colonized out of Africa (Prugnolle *et al.* 2005a). These founder effects have also shaped patterns of morphological variation and resulted in structure at disease resistance genes, both of which are traits under selection (Prugnolle *et al.* 2005b; Manica *et al.* 2007). In silvereyes (*Zosterops lateralis*), a recent island colonizer, sequential founder events produced genetic divergence among populations (Clegg *et al.* 2002a; Estoup & Clegg 2003). At the morphological level, however, selection was

found to be the primary force driving differences among populations (Clegg *et al.* 2002b, 2008).

The presence of isolation by distance can be detected by correlating pairwise levels of genetic structure (e.g. F_{ST}) with pairwise geographic distances between populations, usually using Mantel and partial Mantel tests (Rousset 1997). Similarly, isolation by adaptation can be tested by correlating pairwise F_{ST} values with a matrix of ecological or environmental distance (Nosil *et al.* 2009). However, it will only be possible to differentiate between isolation by distance and isolation by adaptation using this approach when ecological and geographic distances between populations are not strongly correlated. Under isolation by colonization, no relationship is expected between genetic distance and either geographic or ecological distance (Orsini *et al.* 2013b); instead, genetic structure is expected to reflect the colonization and bottleneck history of the populations being studied. Patterns of phenotypic divergence (e.g. Q_{ST}) can be studied in the same way as neutral genetic data. However, there is a lack of studies simultaneously examining isolation by distance, by colonization and by adaptation at the genetic and morphological levels (Orsini *et al.* 2013b). Therefore, while it is clear that these may all occur, their relative importance and how these processes interact are not well understood.

In this study, we investigated patterns of genetic and phenotypic divergence in Berthelot’s pipit (*Anthus berthelotii*), a small passerine endemic to the Canary Islands, Madeira and Selvagens archipelagos in the North

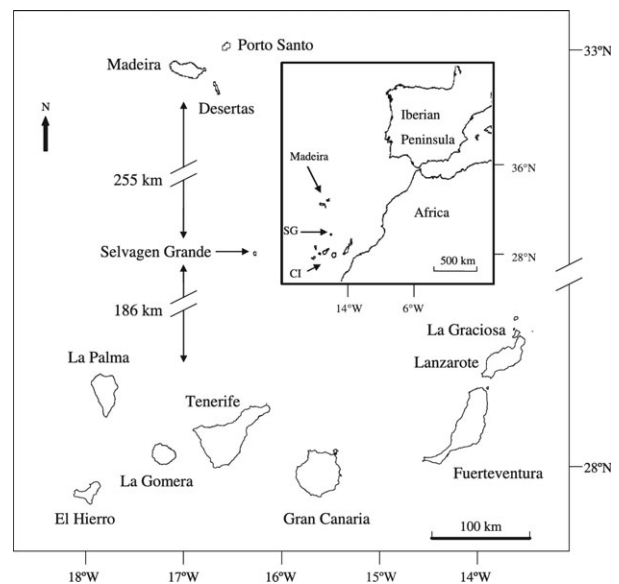


Fig. 1 The location of Berthelot’s pipit populations used in the present study, and (inset) the location of the three archipelagos in relation to the mainland (CI = Canary Islands, SG = Selvagens Grande; reproduced from Illera *et al.* 2007).

Atlantic (Fig. 1). Previous analyses of neutral and functional genetic diversity across Berthelot's pipit populations have revealed very low levels of variation and a recent northward colonization across its range, from the Canary Islands to Madeira and the Selvagens (Illera *et al.* 2007; Spurgin *et al.* 2011). We use a panel of 21 autosomal microsatellite markers and population genetic simulations to test whether the patterns of neutral genetic divergence across the pipit populations are better explained by isolation by distance, by adaptation or by colonization. We then compare patterns of morphological and neutral genetic variation to determine the evolutionary forces driving divergence at the phenotypic level.

Materials and methods

Sampling, morphological and molecular methods

Representative samples (ca. 30 individuals per population) were obtained from all 13 Berthelot's pipit populations in April 2005 (Selvagens), January to March 2006 (Canary Islands) and September 2006 (Madeiran Islands). Birds were caught from multiple localities encompassing the entire range of each population, ensuring that population-level comparisons could be made (see below). Sampled birds from all populations were from low-lying coastal regions, with the exception of birds caught on the mountain of El Teide on Tenerife. These birds exist on a plateau ca. 2000 m above sea level and are separated from the rest of the island by a band of dense forest, which the pipit does not inhabit. For this reason, we considered El Teide a separate thirteenth population. While Berthelot's pipits do exist on alpine plateaus in other islands, such as La Palma and Madeira, these populations are much smaller and were not targeted for trapping. We caught all birds using spring traps baited with *Tenebrio molitor* larvae, fitted each with a uniquely numbered aluminium ring from the relevant national authority, or with a plastic colour ring when these were not available, and took seven morphological measurements (mass, tarsus length, wing length, head length and bill length, width and height; see Illera *et al.* 2007 for details). Blood samples (ca. 40 µl) were taken by brachial venipuncture and stored at room temperature in screw-topped, rubber-sealed, microfuge tubes containing 800 µl of absolute ethanol.

Genomic DNA was extracted using a modified salt extraction method (Bruford *et al.* 1992; Richardson *et al.* 2001) and diluted to a concentration of 10–50 ng/µl. We genotyped all individuals at a panel of 21 microsatellite loci (Primmer *et al.* 1995; Martinez *et al.* 1999; Dawson *et al.* 2010, 2012, 2013) (Table S2, Supporting information). Loci were confirmed unique by comparing their sequences using BLASTN software (Altschul *et al.* 1997).

PCRs were carried out in 2-µl reactions using a method based on Kenta *et al.* (2008). Briefly, 1 µl (10–50 ng) of genomic DNA was added to each PCR well and the liquid evaporated. To this we added 1 µl primer mix (containing all forward and reverse primers in the multiplex reaction at 0.2 µM) and 1 µl 2x QIAGEN Multiplex PCR Master Mix. Samples were overlaid with mineral oil before PCR. An initial hot-start denaturing phase of 95 °C for 15 min was followed by 40 cycles of 94 °C for 30 s, 56 °C for 90 s and 72 °C for 60 s. A final hold of 60 °C for 30 min completed the reaction. PCR products were diluted 1 in 400 and separated on an ABI 3730 DNA analyser. We determined allele sizes using GENEMAPPER version 3.7 (Applied Biosystems).

Genetic analyses

Unless stated otherwise, all statistics and plots were generated in R version 2.15.2 (R Development Core Team 2012). For all variables used in parametric tests, we tested for normality using Shapiro–Wilks tests and used transformations where appropriate.

For each microsatellite locus and population, Hardy–Weinberg equilibrium and linkage disequilibrium were tested using GENEPOP version 4.1 (Raymond & Rousset 1995) and null allele frequencies were estimated using CERVUS version 3.0.3 (Marshall *et al.* 1998). Allele frequencies and expected heterozygosity were calculated using FSTAT version 2.9 (Goudet 1995). Allelic richness was calculated after controlling for differences in sample size, using a rarefaction approach implemented in HP rare (Kalinowski 2004, 2005). We tested for differences in genetic diversity across archipelagos using linear mixed models, implemented in the lme4 and nlme packages in R (Bates *et al.* 2011; Pinheiro *et al.* 2013). Allelic richness and heterozygosity were included as response variables in separate models, with archipelago as a fixed factor and locus identity as a random factor. To assess whether population identity had a significant effect on genetic diversity above and beyond the effect of archipelago identity, we then ran a second mixed model including the same variables, plus population identity as a random factor, and compared the two models using likelihood ratio tests.

To visualize the level of genetic structure across islands and archipelagos, we performed a principal component analysis (PCA) of the 21 microsatellite loci, using the adegenet package in R (Jombart 2008). We also used STRUCTURE (Pritchard *et al.* 2000) to identify the number of distinct genetic clusters (*K*) in our data set. For this, we used a model allowing admixture and correlated gene frequencies and carried out four independent runs for each value of *K* = 1–13. We accepted the value of *K* with the highest average 'log probability of data' as the most

likely number of clusters. For each run, we ran 500 000 steps, with a burn-in of 10 000 steps.

We used two approaches to detect whether populations had undergone genetic bottlenecks. First, we calculated the degree of heterozygosity excess, which occurs due to the loss of rare alleles shortly after bottlenecks (Cornuet & Luikart 1996), in the program BOTTLENECK (Piry *et al.* 1999). We used a two-phase mutation model, with 95% stepwise and 5% nonstepwise mutations. The probability of heterozygosity excess was then calculated using Wilcoxon tests. Second, we calculated Garza and Williamson's index (M) by dividing the number of alleles in a population (k) by the range in allele size (r) (Garza & Williamson 2001). This was modified to $M = k/(r + 1)$ to avoid dividing by zero in monomorphic populations (Excoffier *et al.* 2005).

We used the program DIY-ABC (Cornuet *et al.* 2008) to reconstruct the most likely demographic history of Berthelot's pipit. We constructed the four most plausible model colonization scenarios based on a northward expansion across the three archipelagos (Illera *et al.* 2007). These involved two colonization routes from the Canary Islands, one with a sequential colonization to Selvagens then to Madeira and the other with a simultaneous colonization of Selvagens and Madeira – each with and without bottlenecks associated with the colonization events. The models described an ancestral N_e , as well as the contemporary N_e and the colonization time of each archipelago. For the two scenarios involving bottlenecks, we also included the number of founders for each archipelago and the duration of the bottleneck. Priors were given uniform distributions, informed where possible by knowledge of the past and present distribution of Berthelot's pipit (Table S2, Supporting information). We used the default settings provided by the program as prior information on microsatellite mutation rates. We simulated one million data sets for each scenario and calculated the following summary statistics for observed and simulated data sets: mean number of alleles per locus, mean gene diversity, mean size variance and mean M ratio within each population, as well as pairwise F_{ST} across each pair of populations. The posterior probability of each scenario was estimated using a logistic regression approach implemented in DIY-ABC. We evaluated confidence in our choice of scenario by comparing the match between simulated and observed summary statistics and by calculating type I and II error rates using the standard procedures available in DIY-ABC (Cornuet *et al.* 2008).

Isolation by distance, colonization and adaptation

We tested whether patterns of neutral genetic structure were the product of isolation by distance, by adaptation or by colonization, using a rationale largely based on

that of Nosil *et al.* (2009) and Orsini *et al.* (2013b). Under a scenario of limited dispersal, a linear relationship between pairwise genetic and geographic distance matrices is expected. Under isolation by adaptation, we expect patterns of genetic structure to reflect ecological differences across islands, irrespective of geographic distance. Finally, under isolation by colonization, we expect patterns of genetic structure to mirror the colonization history of the pipit, with highest levels of genetic structure between populations that have been separately colonized by a small number of founders (i.e. the most bottlenecked populations).

We calculated population-level pairwise genetic differentiation as $F_{ST}/(1-F_{ST})$ (Slatkin 1995) in ARLEQUIN version 3.5 (Excoffier & Lischer 2010). Geographic distance was calculated as the closest linear distance between pairs of islands, estimated using Google Earth (<http://earth.google.com>). As a proxy for ecological distance, we used a selection of climate variables. For each island, we extracted data on average monthly precipitation, average temperature in August (the hottest month) and average temperature in January (the coldest month; see Appendix S1 for sources, Supporting information). Our rationale for choosing these variables was based on the extent to which they differed within and among archipelagos, and on their expected relationship with divergence at the morphological traits we had measured. Temperature is the most important predictor of body size in birds globally (Olson *et al.* 2009), and under isolation by adaptation, we therefore predict that birds on islands with similar temperature profiles will be of similar size. Annual rainfall has been shown, via food availability to result in natural selection on beak morphology in birds (Grant & Grant 1996), and has been linked to arthropod abundance in the Canary Islands (Illera & Diaz 2006). We standardized climate variables (v) using the following equation:

$$v_{\text{standardized}} = \frac{v_{\text{island}} - \bar{v}}{\sigma_v}$$

Using the three standardized climate variables for each island, we then calculated Euclidean distances between all pairs of islands, thus obtaining a single distance matrix (hereafter referred to as 'environmental distance').

To obtain a distance matrix that reflects the relative severity of the combined bottleneck effects between two populations, we modified Garza & Williamson's (2001) M ratio as follows:

$$M_{(\text{population pair})} = -\log(M_{(\text{pop1})} \times M_{(\text{pop2})})$$

Thus, $M_{(\text{population pair})}$ is highest when both populations are bottlenecked (and M is low) and is therefore hereafter termed 'bottleneck severity'.

We tested separately whether genetic distance was related to geographic distance, bottleneck severity and environmental distance using Mantel tests, implemented in the Ecodist package in R (Goslee & Urban 2007). Importantly, environmental and geographic distance in this system are not correlated (Mantel test, $R = 0.05$, $P = 0.3$), making it possible to assess the relative importance of these variables. To control for the potentially confounding effect of hierarchical structure (i.e. of archipelago-level effects) on relationships between distance matrices, we ran partial Mantel tests, including a coding variable that identified which archipelagos were included in each pairwise comparison e.g. Canary Islands-Canary Islands, Canary Islands-Madeira, etc).

Morphometric analyses

To visualize the level of morphological structure across archipelagos, we conducted a PCA of six of the seven morphological measurements (excluding mass, which fluctuates too much over short timescales in small passerines) and plotted the first two principal components (PC1 and PC2). We tested how morphological variation was partitioned within and across archipelagos; for each PC as the dependent variable, we first ran a linear model, including age, sex and archipelago identity as fixed factors, and we then ran a mixed model, including the same variables, plus population identity as a random factor. To assess whether island identity had a significant effect on morphology above and beyond the effect of archipelago identity, we compared the linear mixed models to the linear models using likelihood ratio tests.

We tested whether morphological differences between populations were the product of isolation by distance, by colonization or by adaptation using the same rationale as for the genetic data. We calculated Euclidean distances in morphology between all pairs of populations, using the mean values per population of the six morphometric traits (i.e. we obtained a single matrix of Euclidean distances in six-dimensional space). Morphometric traits were standardized in the same manner as environmental variables (equation 2). We tested whether morphometric distance was related to geographic distance, bottleneck severity and environmental distance using Mantel tests and partial Mantel tests (accounting for archipelago-level differences). We also directly tested whether morphometric and genetic distance matrices were related, again using Mantel and partial Mantel tests.

Results

In total, 371 individuals were genotyped at 21 unique microsatellite loci (sample sizes for each population

provided in Table 1). No loci were in Hardy-Weinberg disequilibrium in more than two of the 13 populations typed, no loci had estimated null allele frequencies greater than 0.2, and no loci were in linkage disequilibrium after Bonferroni correction for multiple tests. Allele frequencies are given in Table S3 (Supporting information). Allelic richness and heterozygosity both differed significantly across archipelagos (Fig. 2), with highest levels of variation found in the Canary Islands and lowest levels in Selvagens ($F = 27.9$; $P < 0.001$). Including population identity as a random factor did not significantly improve the model fit for either allelic richness or heterozygosity models (likelihood ratio tests, both P -values > 0.99), suggesting that differences in genetic diversity were predominantly across, compared to within, archipelagos (Fig. 2). The two tests employed to detect genetic bottlenecks yielded concordant results: there was no evidence for a bottleneck in any of the Canary Island populations, whereas there was a bottleneck signal in the Selvagens, and in all three Madeiran Islands (Table 1).

A PCA of the 21 microsatellite loci revealed high levels of structure across, but not within, archipelagos (Fig. 3A). Likewise, Bayesian clustering revealed $K = 3$ as the most likely number of genetic clusters, and these clusters corresponded to the three archipelagos (Figure S1, Supporting information). There was an overall relationship between pairwise genetic and geographic distance matrices (Mantel test, $R = 0.39$, $P = 0.007$), suggesting a pattern of isolation by distance; however,

Table 1 Tests for genetic bottlenecks in Berthelot's pipit populations: Wilcoxon tests for heterozygote excess (Piry *et al.* 1999) and M (Garza & Williamson 2001). Values highlighted in bold are those indicative of a bottleneck ($P \leq 0.05$ for the Wilcoxon tests and $M < 0.68$ for the Garza-Williamson statistic). The sample size for all genetic analyses (N) is also provided

Island	P (Hz excess)	M	N
Canary Islands			
La Graciosa	0.86	0.78	23
Lanzarote	0.54	0.78	30
Fuerteventura	0.44	0.82	30
Gran Canaria	0.71	0.82	29
Tenerife (El Teide)	0.58	0.75	29
Tenerife (coast)	0.58	0.82	30
La Gomera	0.33	0.78	30
La Palma	0.90	0.81	28
El Hierro	0.31	0.75	30
Selvagens			
Selvagem Grande	0.01	0.47	30
Madeira			
Madeira	0.02	0.62	26
Deserta Grande	0.01	0.60	30
Porto Santo	0.03	0.57	28

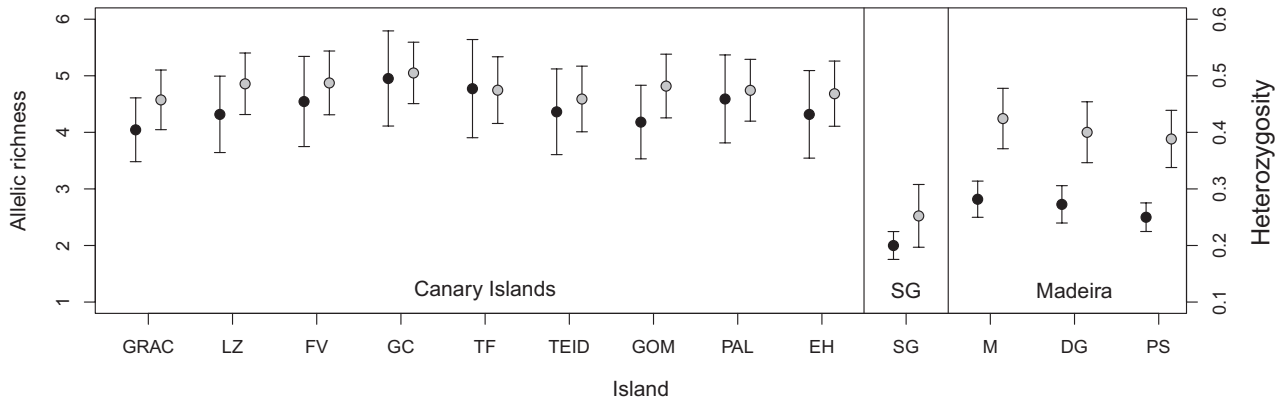


Fig. 2 Allelic richness (black dots) and heterozygosity (grey dots) across 13 Berthelot's pipit populations. Populations are coded as follows: GRAC = La Graciosa, LZ = Lanzarote, FV = Fuerteventura, GC = Gran Canaria, TF = Tenerife, TEID = El Teide, GOM = La Gomera, PAL = La Palma, EH = El Hierro, SG = Selvagem Grande, DG = Deserta Grande, M = Madeira, PS = Porto Santo.

this relationship disappeared when we controlled for archipelago-level effects (partial Mantel test, $R = 0.05$, $P = 0.69$; Fig. 4A). Genetic distance was strongly related to bottleneck severity ($R = 0.86$, $P < 0.001$), and this relationship remained highly significant after controlling for archipelago effects ($R = 0.89$, $P < 0.001$; Fig. 4B). Genetic distance was not related to environmental distance, regardless of whether archipelago effects were accounted for ($R = -0.14$, $P = 0.89$ and $R = -0.14$, $P = 0.90$ respectively; Fig. 4C).

The most highly supported demographic scenario in DIY-ABC analyses involved simultaneous colonization of Madeira and Selvagens both directly from the Canary Islands, with bottlenecks associated with the founder events (scenario 1, posterior probability = 0.96; Table 2). Posterior estimates from scenario 1 suggested moderate contemporary N_e in all three archipelagos and that Madeira and Selvagens were colonized from the Canary Islands between ca. 1000 and 26 000 years ago (Table 3). While estimates of the bottleneck N_e were very broad (4–49) given the prior distribution (1–50), the simulations did suggest that Madeira and Selvagens remained at a low N_e for a long time following colonization, with the length of the bottleneck estimated at between 200 and 12 000 years (Table 3). DIY-ABC was unable to estimate the colonization time from the mainland to the Canary Islands with any degree of accuracy (25 000–2 000 000 years ago), most likely because of a lack of samples from the ancestor of Berthelot's pipit. Using scenario 1, all but one of our simulated summary statistics were within the range or our observed summary statistics at the $P > 0.05$ level (the M ratio in the Canary Islands was the only exception to this). Rates of type I and II error were low, 0.044 and 0.040, respectively.

Variation across archipelagos for each of the six morphological traits is shown in Figure S2 (Supporting

information). As with the genetic data, morphometric variation was greater across than within archipelagos. The first two components from a PCA of the morphological traits explained 83% and 12% of the total variation, respectively. PC1 was moderately negatively correlated with all six original variables and thus represents body size. PC2 was highly negatively correlated with bill width and height and thus represents bill shape. We detected highly significant differences across archipelagos in both PC1 ($F = 47.6$, $P < 0.001$) and PC2 ($F = 46.8$, $P < 0.001$). Madeiran birds were associated with increased body size (decreased PC1) with no change in bill shape, whereas on Selvagens birds exhibited decreased overall size and increased bill height and width (decreased PC2) (Fig. 3B). Archipelago-level differences in both PC1 and PC2 exceeded population-level differences (Fig. 3C); however, including population identity as a random factor did result in a significantly improved model for PC1 (likelihood ratio test, $P < 0.001$), but not for PC2 ($P = 0.07$). This effect on PC1 was caused by an increased body size in the mountainous population on El Teide, in Tenerife (Fig. 3C).

Population-level pairwise morphological distance was not related to geographic distance between populations ($R = 0.17$, $P = 0.19$; Fig. 4D), nor to environmental distance ($R = -0.14$, $P = 0.91$; Fig. 4F). However, like genetic distance, morphological distance was strongly related to bottleneck severity ($R = 0.52$, $P < 0.001$; Fig. 4E), and this relationship remained highly significant after controlling for archipelago effects ($R = 0.52$, $P < 0.001$). Finally, pairwise genetic and morphological distance matrices were strongly correlated ($R = 0.59$, $P < 0.001$), and again this relationship remained strong and significant when controlling for archipelago effects ($R = 0.51$, $P < 0.001$; Figure S3, Supporting information), and for geographic distance ($R = 0.58$, $P < 0.001$).

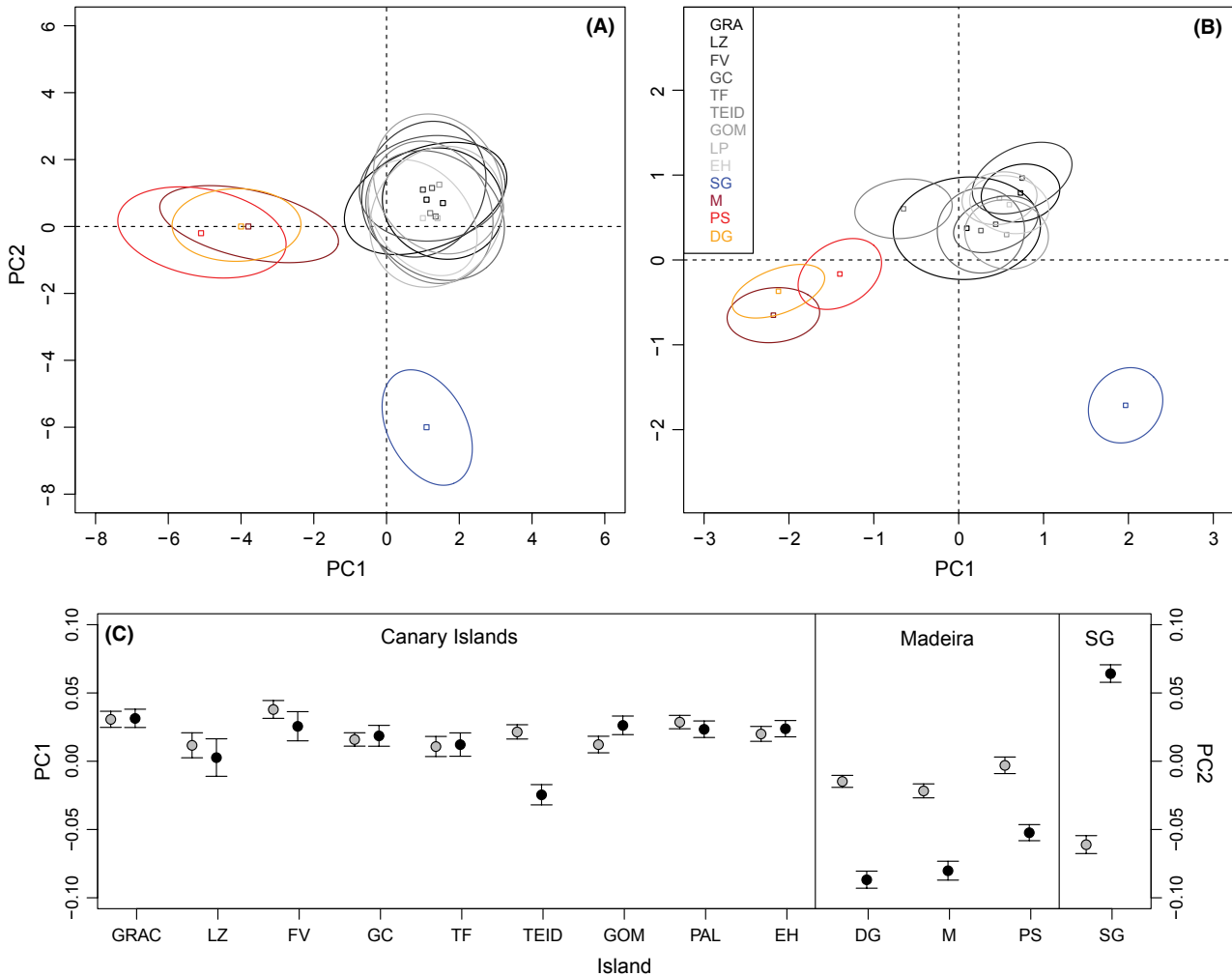


Fig. 3 Principal component analysis (PCA) of (A) microsatellite and (B) morphological data from 371 Berthelot's pipits. For the morphological PCA, PC1 is negatively related to overall body size and PC2 is negatively related to bill height and width. We have avoided inverting the axes to enable comparison with the microsatellite PCA. Points are colour-coded by archipelago (greys = Canary Islands, reds = Madeira, blue = Selvagens). Ellipses represent 95% confidence intervals for individual populations. (C): Mean \pm SE PC1 (black dots) and PC2 (grey dots) from the morphological PCA across individual populations.

Discussion

Here, we show that these latter two archipelagos were colonized independently from the Canary Islands between 1000 and 26 000 years ago. Both basic population genetic analyses and Bayesian simulations indicate that the patterns of structure observed across the island populations are the result of these dispersal events, with genetic bottlenecks having occurred in the two most recently colonized archipelagos. Similarly, explicit tests for isolation by distance, colonization and adaptation revealed that only colonization history was a significant predictor of genetic structure across populations. We also found that patterns of morphological variation closely mirror patterns of structure at the neutral

microsatellite loci and are consistent with the bottleneck history of the pipit, but not with a scenario of selection or gradual genetic drift. Together, our data suggest that founder effects have been the predominant force shaping genetic and morphological variation in this species.

Revisiting our previous study (Illera *et al.* 2007) with additional loci and new analytical methods has changed our interpretation of the Berthelot's pipit population history. The most striking result was the detection of genetic bottlenecks in the recently colonized populations (Table 1), which were not observed when five microsatellite loci were used (Illera *et al.* 2007). Simulations have highlighted that the two most commonly used methods for detecting genetic bottlenecks (Cornuet & Luikart 1996; Garza & Williamson 2001) are sensitive to

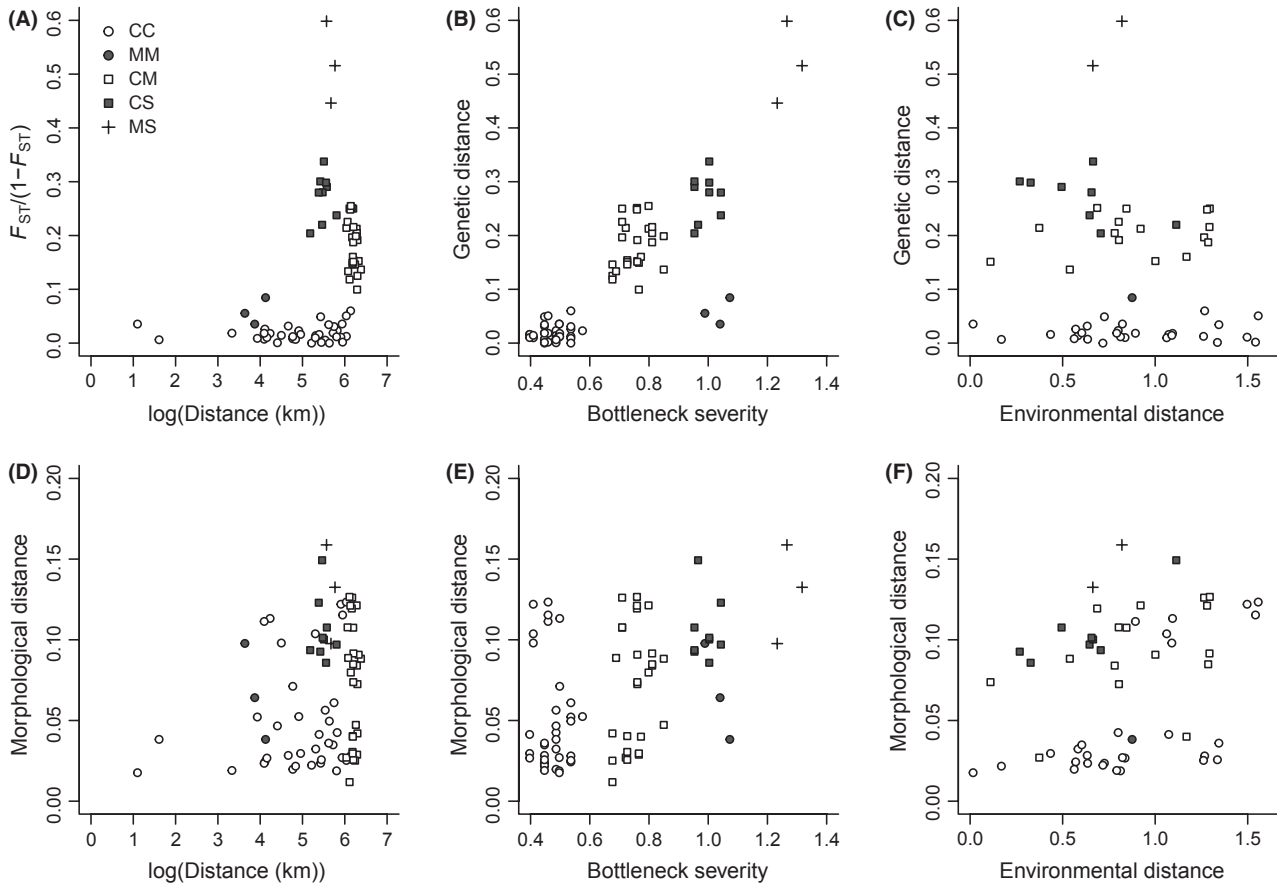


Fig. 4 Isolation by distance, by colonization and by adaptation across Berthelot's pipit populations. Top row: pairwise genetic distance in relation to (A) geographic distance, (B) bottleneck severity and (C) environmental distance (see text for details). Bottom row: pairwise morphological distance in relation to the same three variables as in the top row (D, E and F, respectively). Different point characters represent pairwise comparisons between the different archipelagos (CC = Canary Islands–Canary Islands; CM = Canary Islands–Madeira; CS = Canary Islands–Selvagens; etc.).

Table 2 Model choice and performance from approximate Bayesian computation analyses on 371 Berthelot's pipit individuals. The best-supported scenario is highlighted in bold

Scenario	Posterior probability	Outlier summary statistics		
		$P < 0.05$	$P < 0.01$	$P < 0.001$
1	0.96	1	0	0
2	0.03	2	1	1
3	0.01	2	0	1
4	0.00	2	1	0

type II error, depending on the number of loci used and the variability of these loci (Williamson-Natesan 2005) – our findings here provide a striking example of how this can occur, and how it can influence our interpretations. In the light of this, that we did not detect a bottleneck in the Canary Islands does not mean that one did not occur there either, but it does

suggest that if a bottleneck did occur, it was older and/or less severe than in the Selvagens or Madeiran populations, which is in accordance with our colonization scenario.

The other result that contrasts with our previous study is that the apparent pattern of isolation by distance was no longer observed when archipelago-level effects were controlled for (Fig. 4A). This suggests that patterns of neutral genetic structure across Berthelot's pipit populations are in fact not the simple product of limited migration and gradual genetic drift. We also suggest that our patterns of neutral genetic structure are not the product of environmental differences limiting migration across archipelagos. Our evidence for this is the following: (i) we found no relationship between divergence at our selected environmental variables and genetic distance (Fig. 4C); (ii) our own observations indicate that ecological differences (e.g. food availability) are greater within compared to across archipelagos; and (iii) we do not expect a relationship between

Table 3 Mean and quantiles of posterior distribution samples for parameters estimated in DIY-ABC analyses of Berthelot's pipit colonization history. Parameters were estimated using the best-supported scenario (scenario 1, see Table 2) of an initial colonization of the Canary Islands, followed by simultaneous colonization from the Canary Islands directly to both Madeira and Selvagens, with bottlenecks associated with each colonization event

Parameter	Mean	$Q_{0.05}$	$Q_{0.95}$
N_e			
Canary Islands	4080	1260	8200
Madeira	2,930	795	7400
Selvagens	428	304	494
Colonization time (years ago)			
Canary Islands	315 200	24200	1 908 000
Madeira and Selvagens	8520	1504	26 640
N_e during bottleneck			
Madeira and Selvagens	28	4	49
Bottleneck duration (years)			
Madeira and Selvagens	4080	220	12 240

genetic structure and bottleneck history (Fig. 4B) if adaptation is the primary driver of differences among populations. It can be very difficult to distinguish between true isolation by distance, isolation by colonization and isolation by adaptation, and we echo recent advice that more care needs to be taken in differentiating between these phenomena (Meirmans 2012; Orsini *et al.* 2013b).

In contrast to what one would expect from selection (Clegg *et al.* 2002b, 2008), the divergence observed at different morphological traits strongly reflected patterns of neutral genetic divergence (Fig. 3). As with our genetic data, most of the differences in morphology were across archipelagos, despite there being greater variation in ecological parameters within populations, or among populations within archipelagos, than there is among archipelagos. Morphological divergence was not related to geographic or environmental differences between populations, but was strongly related to the bottleneck history of the pipit (Fig. 4). While it will never be possible to rule out that some environmental or ecological variables that we have not considered, such as diet, are driving morphological (and genetic) differences across archipelagos, the most parsimonious explanation is that isolation by colonization has been the predominant force driving both genetic and phenotypic divergence in Berthelot's pipits. Importantly, the relationship between morphometric divergence and bottleneck history was strongly significant even after controlling for archipelago-level effects, suggesting that differences in morphology within, as well as among, archipelagos were largely driven by neutral processes.

An exception to this is the increased body size in the mountain population on El Teide, on the island of Tenerife (Fig. 3C). This population is situated >2000 m above sea level, and temperatures frequently drop below freezing in winter, contrasting starkly with the other populations, which are predominantly low-lying, subtropical and coastal. In passerine birds, body size and altitude are positively related, as suggested by Bergmann's rule (Blackburn & Ruggiero 2001). Thus, it seems probably that the colder weather on El Teide has selected for larger body size in Berthelot's pipits. We note, however, that overall differences in body size due to founder effects observed across the archipelagos far outweigh the differences between El Teide and the other Canarian populations (Fig. 3C).

Morphological changes in island birds constitute some of the classic examples of natural selection in the wild and can occur over remarkably short timescales (Grant 1986). It is somewhat surprising, therefore, that the apparent founder effects observed across Berthelot's pipit populations have not yet been obscured by natural selection. This may be because i) with the exception of El Teide (and the other, smaller isolated mountain populations on Madeira and La Palma, which we did not sample), this species occupies uniform habitats with benign and stable climate. Additionally, very few other insectivorous bird species can be found in the same habitat as Berthelot's pipits (Illera 2007), suggesting that interspecific competition is low. However, although natural selection does not appear to have been the primary driver of morphological change in Berthelot's pipits, it is almost certain that natural selection and sexual selection have acted on other genotypic and phenotypic traits than the ones we measured here. Indeed, we have previously shown that these pipits face different levels of pathogen-mediated selection across populations (Spurgin *et al.* 2012) and that selection is involved in differentiation at major histocompatibility complex (MHC) genes (Spurgin *et al.* 2011). This species therefore provides an excellent opportunity to study how selection and drift interact at the genotypic and phenotypic levels.

In addition to a lack of (or weak) selection on morphological change, it is thanks to a fortuitous evolutionary time frame and a lack of gene flow across archipelagos (Illera *et al.* 2007) that we have been able to observe founder effects so clearly in this study. In very recently separated populations and experimental studies, in which it is possible to directly observe founder events, it is unclear how long any observed founder effects will persist (Kolbe *et al.* 2012). On the other hand, in populations or species that have been separated for very long periods of time, a combination of selection, mutation and drift will have produced

differences so great that it becomes difficult to disentangle the signature of founder effects from other evolutionary forces (e.g. Hoeck *et al.* 2010; Aleixandre *et al.* 2013). The fleeting nature of founder effects may be one reason that their prevalence and importance in evolution are poorly understood (Templeton 2008). By studying moderately diverged populations, we have been able to show that founder effects can indeed persist over evolutionary timescales, and our results suggest that under certain circumstances, founder effects may play an important role in the initial stages of differentiation and speciation. Further targeted research using systems where founder speciation is predicted to play a role will help us to better understand its evolutionary importance.

Acknowledgements

We are grateful to the friends who provided assistance and accommodation during fieldwork. Local governments in the Canary Islands and Madeira also provided accommodation; the Spanish Environment Ministry, the Canary government and the Natural Park of Madeira gave permission to perform the sampling work, and the Portuguese Navy helped with transport. Genotyping was carried out at the NERC Biomolecular Analysis Facility in Sheffield (NBAF-S). We thank T. Burke, G. Horsburgh and A. Krupa for discussions and assistance with genotyping. This work was funded by PhD and NBAF grants awarded to DSR from the UK Natural Environment Research Council, and a Spanish postdoctoral fellowship (Ramón y Cajal) to JCI.

References

- Aleixandre P, Montoya JH, Milá B (2013) Speciation on oceanic islands: rapid adaptive divergence vs. cryptic speciation in a Guadalupe island songbird (Aves: Junco). *PLoS ONE*, **8**, e63242.
- Altschul SF, Madden TL, Schäffer AA *et al.* (1997) Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Research*, **25**, 3389–3402.
- Bates D, Maechler M, Bolker B (2011) lme4: Linear Mixed-Effects Models Using S4 Classes, R package version 0.999375-42. Available from: <http://cran.r-project.org/web/packages/lme4/index.html>.
- Blackburn TM, Ruggiero A (2001) Latitude, elevation and body mass variation in Andean passerine birds. *Global Ecology and Biogeography*, **10**, 245–259.
- Brinkmann B, Klintschar M, Neuhuber F, Hühne J, Rolf B (1998) Mutation rate in human microsatellites: influence of the structure and length of the tandem repeat. *The American Journal of Human Genetics*, **62**, 1408–1415.
- Bruford MW, Hanotte O, Brookfield JFY, Burke T (1992) Single-locus and multilocus DNA fingerprinting. In: *Molecular Genetic Analysis of Populations: A Practical Approach* (ed. Hoelzel A), pp. 225–269. IRL Press, Oxford.
- Carson HL (1968) The population flush and its genetic consequences. In: *Population Biology and Evolution* (ed. Lewontin RC), pp. 123–137. Syracuse University Press, Syracuse, New York.
- Clegg SM, Degnan SM, Kikkawa J *et al.* (2002a) Genetic consequences of sequential founder events by an island-colonizing bird. *Proceedings of the National Academy of Sciences*, **99**, 8127–8132.
- Clegg SM, Degnan SM, Moritz C *et al.* (2002b) Microevolution in island forms: the roles of drift and directional selection in morphological divergence of a passerine bird. *Evolution*, **56**, 2090–2099.
- Clegg SM, Frentiu FD, Kikkawa J, Tavecchia G, Owens IP (2008) 4000 years of phenotypic change in an island bird: heterogeneity of selection over three microevolutionary time-scales. *Evolution*, **62**, 2393–2410.
- Cornuet JM, Luikart G (1996) Description and power analysis of two tests for detecting recent population bottlenecks from allele frequency data. *Genetics*, **144**, 2001.
- Cornuet JM, Santos F, Beaumont MA *et al.* (2008) Inferring population history with DIY ABC: a user-friendly approach to approximate Bayesian computation. *Bioinformatics*, **24**, 2713–2719.
- Coyne JA, Orr HA (2004) *Speciation*. Sinauer Associates, Sunderland, Massachusetts.
- Dawson DA, Horsburgh GJ, Küpper C *et al.* (2010) New methods to identify conserved microsatellite loci and develop primer sets of high cross-species utility; as demonstrated for birds. *Molecular Ecology Resources*, **10**, 475–494.
- Dawson DA, Horsburgh GJ, Krupa AP *et al.* (2012) Microsatellite resources for Passeridae species: a predicted microsatellite map of the house sparrow *Passer domesticus*. *Molecular Ecology Resources*, **12**, 501–523.
- Dawson DA, Ball AD, Spurgin LG *et al.* (2013) High-utility conserved avian microsatellite markers enable parentage and population studies across a wide range of species. *BMC Genomics*, **14**, 176.
- R Development Core Team (2012) *R: A Language and Environment for Statistical Computing*. R version 2.15.2. R Foundation for Statistical Computing, Vienna, Austria. Available at: <http://www.R-project.org/>.
- Estoup A, Clegg S (2003) Bayesian inferences on the recent island colonization history by the bird *Zosterops lateralis lateralis*. *Molecular Ecology*, **12**, 657–674.
- Excoffier L, Lischer HE (2010) Arlequin suite ver 3.5: a new series of programs to perform population genetics analyses under Linux and Windows. *Molecular Ecology Resources*, **10**, 564–567.
- Excoffier L, Estoup A, Cornuet JM (2005) Bayesian analysis of an admixture model with mutations and arbitrarily linked markers. *Genetics*, **169**, 1727–1738.
- Frantz A, Cellina S, Krier A, Schley L, Burke T (2009) Using spatial Bayesian methods to determine the genetic structure of a continuously distributed population: clusters or isolation by distance? *Journal of Applied Ecology*, **46**, 493–505.
- Garza JC, Williamson EG (2001) Detection of reduction in population size using data from microsatellite loci. *Molecular Ecology*, **10**, 305–318.
- Goslee SC, Urban DL (2007) The ecodist package for dissimilarity-based analysis of ecological data. *Journal of Statistical Software*, **22**, 1–19.
- Goudet J (1995) FSTAT (version 1.2): a computer program to calculate F-statistics. *Journal of Heredity*, **86**, 485–486.

- Grant PR (1986) *Ecology and Evolution of Darwin's Finches*. Princeton University Press, Princeton, New Jersey.
- Grant BR, Grant PR (1996) High survival of Darwin's finch hybrids: effects of beak morphology and diets. *Ecology*, **77**, 500–509.
- Hartl DL, Clark AG (2007) *Principles of Population Genetics*. Sinauer Associates, Sunderland.
- Hoeck PE, Bollmer JL, Parker PG, Keller LF (2010) Differentiation with drift: a spatio-temporal genetic analysis of Galápagos mockingbird populations (*Mimus* spp.). *Philosophical Transactions of the Royal Society B: Biological Sciences*, **365**, 1127–1138.
- Illera JC (2007) Bisbita Caminero *Anthus berthelotii*. In: *Atlas de Las Aves Nidificantes en el Archipiélago Canario (1997–2003)* (ed. Lorenzo JA), pp. 344–347. Dirección General de Conservación de la Naturaleza-Sociedad Española de Ornitología, Madrid.
- Illera JC, Diaz M (2006) Reproduction in an endemic bird of a semiarid island: a food-mediated process. *Journal of Avian Biology*, **37**, 447–456.
- Illera JC, Emerson BC, Richardson DS (2007) Population history of Berthelot's pipit: colonization, gene flow and morphological divergence in Macaronesia. *Molecular Ecology*, **16**, 4599–4612.
- Jombart T (2008) adegenet: a R package for the multivariate analysis of genetic markers. *Bioinformatics*, **24**, 1403–1405.
- Kalinowski ST (2004) Counting alleles with rarefaction: private alleles and hierarchical sampling designs. *Conservation Genetics*, **5**, 539–543.
- Kalinowski ST (2005) hp rare 1.0: a computer program for performing rarefaction on measures of allelic richness. *Molecular Ecology Notes*, **5**, 187–189.
- Kenta T, Gratten J, Haigh NS *et al.* (2008) Multiplex SNP-SCALE: a cost-effective medium-throughput single nucleotide polymorphism genotyping method. *Molecular Ecology Notes*, **8**, 1230–1238.
- Kolbe JJ, Leal M, Schoener TW, Spiller DA, Losos JB (2012) Founder effects persist despite adaptive differentiation: a field experiment with lizards. *Science*, **335**, 1086–1089.
- Manica A, Amos W, Balloux F, Hanihara T (2007) The effect of ancient population bottlenecks on human phenotypic variation. *Nature*, **448**, 346–348.
- Marie Curie SPECIATION Network (2012) What do we need to know about speciation? *Trends in Ecology and Evolution*, **27**, 27–39.
- Marshall TC, Slate J, Kruuk LEB, Pemberton JM (1998) Statistical confidence for likelihood-based paternity inference in natural populations. *Molecular Ecology*, **7**, 639–655.
- Martinez JG, Soler JJ, Soler M, Møller AP, Burke T (1999) Comparative population structure and gene flow of a brood parasite, the great spotted cuckoo (*Clamator glandarius*), and its primary host, the magpie (*Pica pica*). *Evolution*, **53**, 269–278.
- Meirmans PG (2012) The trouble with isolation by distance. *Molecular Ecology*, **21**, 2839–2846.
- Moya A, Galiana A, Ayala FJ (1995) Founder-effect speciation theory: failure of experimental corroboration. *Proceedings of the National Academy of Sciences*, **92**, 3983–3986.
- Nosil P, Funk DJ, Ortiz-Barrientos D (2009) Divergent selection and heterogeneous genomic divergence. *Molecular Ecology*, **18**, 375–402.
- Olson VA, Davies RG, Orme CL *et al.* (2009) Global biogeography and ecology of body size in birds. *Ecology Letters*, **12**, 249–259.
- Orr HA (1998) Testing natural selection vs. genetic drift in phenotypic evolution using quantitative trait locus data. *Genetics*, **149**, 2099–2104.
- Orsini L, Mergeay J, Vanoverbeke J, De Meester L (2013a) The role of selection in driving landscape genomic structure of the waterflea *Daphnia magna*. *Molecular Ecology*, **22**, 583–601.
- Orsini L, Vanoverbeke J, Swillen I, Mergeay J, Meester L (2013b) Drivers of population genetic differentiation in the wild: isolation by dispersal limitation, isolation by adaptation and isolation by colonization. *Molecular ecology*, **22**, 5983–5999.
- Pinheiro J, Bates D, DebRoy S, Sarkar D, Team RDC (2013) nlme: Linear and Nonlinear Mixed Effects Models. R package version 3.1-110.
- Piry S, Luikart G, Cornuet JM (1999) Computer note. BOTTLENECK: a computer program for detecting recent reductions in the effective size using allele frequency data. *Journal of Heredity*, **90**, 502–503.
- Primmer CR, Møller AP, Ellegren H (1995) Resolving genetic relationships with microsatellite markers: a parentage testing system for the swallow *Hirundo rustica*. *Molecular Ecology*, **4**, 493–498.
- Pritchard JK, Stephens M, Donnelly P (2000) Inference of population structure using multilocus genotype data. *Genetics*, **155**, 945–959.
- Prugnolle F, Manica A, Charpentier M, Guégan JF, Guernier V, Balloux F (2005a) Pathogen-driven selection and worldwide HLA class I diversity. *Current Biology*, **15**, 1022–1027.
- Prugnolle F, Manica A, Balloux F (2005b) Geography predicts neutral genetic diversity of human populations. *Current Biology*, **15**, R159–R160.
- Raymond M, Rousset F (1995) GENEPOP (Version-1.2) - Population genetics software for exact tests and ecumenicism. *Journal of Heredity*, **86**, 248–249.
- Richardson DS, Jury FL, Blaakmeer K, Komdeur J, Burke T (2001) Parentage assignment and extra-group paternity in a cooperative breeder: the Seychelles warbler (*Acrocephalus hellensis*). *Molecular Ecology*, **10**, 2263–2273.
- Rousset F (1997) Genetic differentiation and estimation of gene flow from F-statistics under isolation by distance. *Genetics*, **145**, 1219–1228.
- Rundle HD (2003) Divergent environments and population bottlenecks fail to generate premating isolation in *Drosophila pseudoobscura*. *Evolution*, **57**, 2557–2565.
- Slatkin M (1995) A measure of population subdivision based on microsatellite allele frequencies. *Genetics*, **139**, 457–462.
- Spurgin LG, Van Oosterhout C, Illera JC *et al.* (2011) Gene conversion rapidly generates major histocompatibility complex diversity in recently founded bird populations. *Molecular Ecology*, **20**, 5213–5225.
- Spurgin LG, Illera JC, Padilla DP, Richardson DS (2012) Biogeographical patterns and co-occurrence of pathogenic infection across island populations of Berthelot's pipit (*Anthus berthelotii*). *Oecologia*, **168**, 691–701.
- Templeton AR (2008) The reality and importance of founder speciation in evolution. *BioEssays*, **30**, 470–479.

- Travisano M, Mongold JA, Bennett AF, Lenski RE (1995) Experimental tests of the roles of adaptation, chance, and history in evolution. *Science*, **267**, 87–90.
- Williamson-Natesan EG (2005) Comparison of methods for detecting bottlenecks from microsatellite loci. *Conservation Genetics*, **6**, 551–562.
- Wright S (1943) Isolation by distance. *Genetics*, **28**, 139–156.
- Wright S (1946) Isolation by distance under diverse systems of mating. *Genetics*, **31**, 39–59.

D.S.R. obtained funding for and set up the study. D.S.R., J.C.I. and T.H.J. supervised the study. J.C.I., L.G.S. and D.S.R. performed fieldwork. L.G.S., J.C.I. and D.A.D. performed molecular work. L.G.S. performed analyses and drafted the manuscript; all authors contributed to the writing of the manuscript.

Data accessibility

Microsatellite, morphological and climate data sets have been posted on Dryad (doi:10.5061/dryad.n1506).

Supporting information

Additional supporting information may be found in the online version of this article.

Appendix 1 Sources for climate variables used to calculate 'environmental distance' across Berthelot's pipit populations.

Table S1 Demographic scenarios used for approximate Bayesian computation (ABC) analyses (see Fig. 1 for visual representation).

Table S2 Twenty-one microsatellite loci characterised in the Berthelot's pipit.

Table S3 Allele frequencies at 21 microsatellite loci across island populations of Berthelot's pipit.

Fig S1 Output from a Bayesian clustering algorithm on 371 Berthelot's pipit individuals, implemented in STRUCTURE.

Fig S2 Variation (mean \pm SE) in morphological traits across island population of Berthelot's pipits.

Fig S3 Pairwise morphological distance in relation to genetic distance in Berthelot's pipit populations.